

Optimization of a solid-liquid extraction method for blackberry fruits bioactive compounds using a Box-Behnken design.

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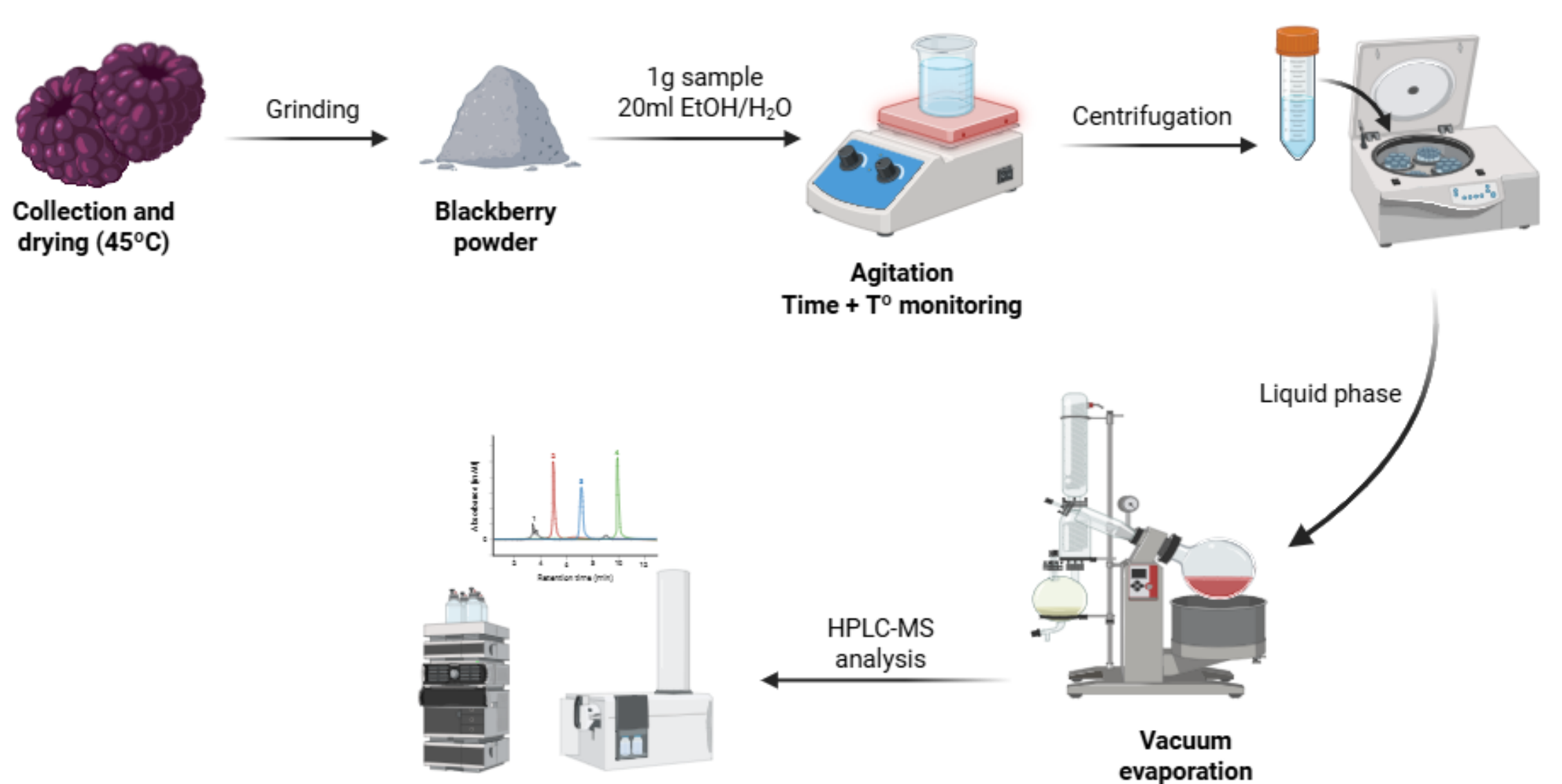
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INTRODUCTION

Blackberries (*Rubus fruticosus*) are rich in phenolic compounds with notable antioxidant, anti-inflammatory, and antimicrobial properties, making them valuable as functional food ingredients^{1,2}. This study optimized the solid-liquid extraction method using GRAS solvents (ethanol and water), with total phenolic content as dependent variable. Response Surface Methodology (RSM) was employed to enhance the efficiency and sustainability of the extraction process for potential food and nutraceutical applications.

MATERIALS AND METHODS



Equation 1. Second order polynomial equation for RSM.

Y = β₀ + ∑ β_i X_i + ∑ β_{ii} X_{ii}² + ∑ ∑ β_{ij} X_i X_j

RESULTS

The Box-Behnken design revealed that ethanol concentration (β₁), extraction time (β₂), and temperature (β₃) significantly affected total phenolic content (TPC). The response surface model showed a high predictive accuracy (R² = 0.9981) with no significant lack of fit (p > 0.05). The strongest effects on TPC were observed for ethanol concentration, followed by time and temperature, with significant linear and quadratic terms. Optimal extraction conditions were determined as 50% ethanol, 14.5 h, and 50 °C, yielding 31.1 ± 4.9 mg gallic acid equivalents (GAE)/g dry weight (predicted value of 31.66 ± 0.75 mg GAE/g d.w.). A total of 34 phenolic compounds were characterized by UPLC-QTOF-MS, including phenolic acids, flavonoids, ellagitannins, lignans, and anthocyanins, with cyanidin-3-O-glucoside (1635.15 μg/g d.w.), cyanidin-3-O-rutinoside (505.34 μg/g d.w.), and sanguin H6 isomer (35.09 μg/g d.w.) being the most abundant.

CONCLUSIONS

Response Surface Methodology successfully optimized the extraction of antioxidant phenolic compounds from blackberry fruits. The model demonstrated excellent predictive accuracy and robustness, defining optimal conditions (50% ethanol, 14.5 h, 50 °C) that maximize extraction efficiency while maintaining sustainability. Additionally, UPLC-QTOF-MS characterization revealed a rich phenolic profile, with cyanidin-3-O-glucoside, cyanidin-3-O-rutinoside, and sanguin H6 isomer identified as the major compounds. These findings support the development of high-value blackberry extracts for functional food and nutraceutical applications.

REFERENCES

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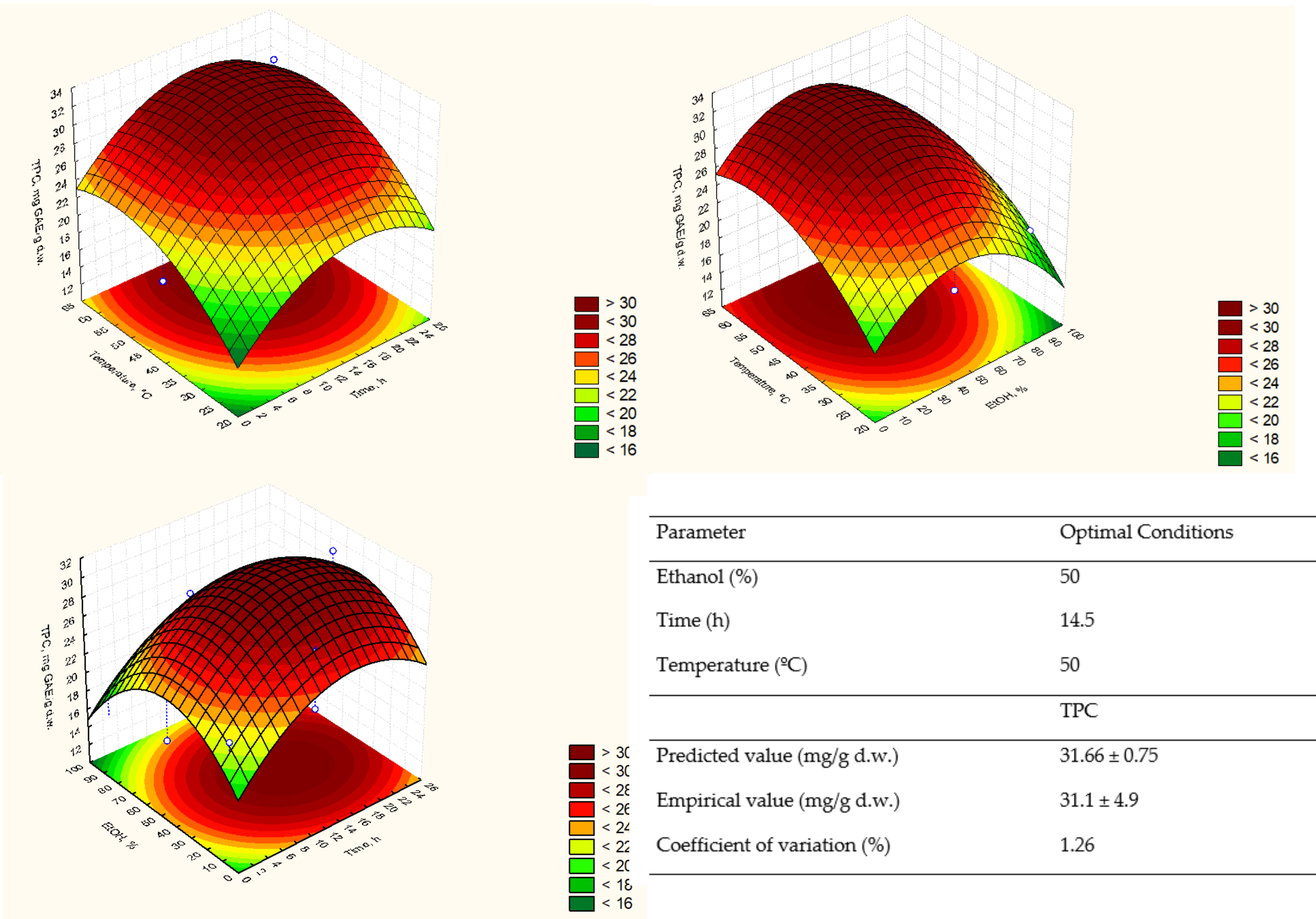


Figure 1. Response surface plots showing combined effects of process variables for TPC (mg GAE/g d.w.): temperature (°C)—time (h) (a), temperature (°C)—% EtOH (b) and % EtOH—time (h) (c). Optimal conditions of extraction and predicted and empirical values of the model (n=3)

Table 4. Identification and quantification of phenolic compounds present in blackberry extract.

Peak No.	Retention Time (Min)	m/z Exp.	m/z Calc.	Error (ppm)	Molecular Formula	Score	Proposed Compound	Quantification (μg/g d.w.)
Phenolic acids and derivatives								
3	0.433	333.057	333.061	-4.8	C ₁₆ H ₁₄ O ₈	93.9	Jaboticabin	27.68 ± 0.26
7	4.979	223.06	223.0606	-3.6	C ₁₁ H ₁₂ O ₅	97.4	Sinapic acid	17.83 ± 0.12
9	5.476	385.111	385.1135	-7.3	C ₁₇ H ₂₂ O ₁₀	92.7	Sinapic acid hexoside	13.35 ± 0.03
16	6.639	183.025	183.0293	-2.4	C ₈ H ₈ O ₅	99.2	Methylgallic acid	19.18 ± 0.07
20	7.707	433.041	433.0407	-0.2	C ₁₈ H ₁₄ O ₁₂	84.5	Ellagic acid-pentoside	27.98 ± 0.02
21	7.888	433.041	433.0407	2.7	C ₁₈ H ₁₄ O ₁₂	92.9	Ellagic acid-pentoside isomer	25.62 ± 0.16
24	8.726	300.999	300.9984	3.3	C ₁₄ H ₆ O ₈	100	Ellagic acid	19.46 ± 0.13
29	9.276	447.056	447.0564	-0.9	C ₂₀ H ₁₆ O ₁₂	94	Ellagic acid 2-rhamnoside	20.89 ± 0.10
32	9.917	315.012	315.0141	-7.0	C ₁₅ H ₈ O ₈	86	3-O-Methylellagic acid	10.41 ± 0.05
Flavonoids and derivatives								
6	4.367	463.085	463.0877	-5.8	C ₁₁ H ₂₀ O ₁₂	95.6	Quercetin-O-hexoside	3.37 ± 0.09
10	5.487	577.14	577.1405	-1.6	C ₃₀ H ₂₆ O ₁₂	85.4	B-type procyanidin dimer	7.57 ± 0.12
11	5.716	315.123	315.1232	-1.9	C ₁₈ H ₂₆ O ₅	85.8	4-hydroxy-5,7,4'-trimethoxyflavan	5.89 ± 0.01
14	6.335	289.071	289.0712	0.7	C ₁₅ H ₁₄ O ₆	87.9	Epicatechin	7.90 ± 0.03
15	6.517	577.136	577.1346	3.1	C ₃₀ H ₂₆ O ₁₂	86.5	B-type procyanidin dimer isomer	7.94 ± 0.05
25	8.801	609.148	609.1456	3.1	C ₂₇ H ₃₀ O ₁₆	99.8	Rutin	8.46 ± 0.15
26	9.046	463.089	463.0877	1.9	C ₂₁ H ₃₀ O ₁₂	99.6	Quercetin 3-galactoside	5.63 ± 0.09
27	9.051	609.146	609.1456	0.7	C ₂₇ H ₃₀ O ₁₆	99.6	Rutin isomer	5.50 ± 0.18
28	9.213	463.085	463.0877	-5.0	C ₂₁ H ₃₀ O ₁₂	97.5	Quercetin-O-glucoside	3.96 ± 0.02
30	9.577	477.066	477.0669	-2.7	C ₃₁ H ₁₈ O ₁₃	98.2	Quercetin 3-glucuronide	4.01 ± 0.01
34	10.199	505.1	505.0982	3.6	C ₁₉ H ₂₂ O ₁₃	99.5	Quercetin-O-acetylhexoside	3.24 ± 0.11
Ellagitannins								
19	7.519	935.079	935.0791	-0.1	C ₆₁ H ₂₈ O ₂₆	100	Casuarictin	21.02 ± 0.23
22	8.29	934.075	934.0712	2	C ₆₁ H ₂₈ O ₂₆ (X2)	83.6	Sanguin H6	25.99 ± 0.18
23	8.381	934.076	934.0712	1.5	C ₆₁ H ₂₈ O ₂₆ (X2)	89.5	Sanguin H6 isomer	35.09 ± 0.27
Lignans								
33	9.984	571.218	571.2179	-0.7	C ₃₀ H ₃₆ O ₁₁	89.6	Kadsurarin	28.17 ± 0.14
35	10.509	571.213	571.2179	-7.9	C ₃₀ H ₃₆ O ₁₁	85.7	Kadsurarin isomer	12.37 ± 0.06
36	10.74	341.137	341.1389	-5.6	C ₂₀ H ₂₂ O ₅	99.9	Kadsurenin B	8.85 ± 0.10
Anthocyanins (MS+)								
12	5.859	449.107	449.1084	-2.2	C ₂₁ H ₂₁ O ₁₁	99.9	Cyanidin-3-O-glucoside	1635.15 ± 13.24
17	6.691	595.165	595.1663	-2.9	C ₂₇ H ₃₁ O ₁₅	92.3	Cyanidin-3-O-rutinoside	505.34 ± 5.37
18	7.268	419.098	419.0919	0.7	C ₂₀ H ₁₉ O ₁₀	98.6	Cyanidin-3-O-arabinoside	145.05 ± 1.63
38	11.499	465.104	465.1033	1.1	C ₂₁ H ₂₁ O ₁₂	96.3	Delphinidin-3-O-galactoside	32.46 ± 0.97
39	11.501	611.16	611.1612	-2.3	C ₂₇ H ₃₁ O ₁₆	92.9	Cyanidin-3,5-O-diglucoside	132.11 ± 2.08
40	11.672	465.104	465.1033	1.9	C ₂₁ H ₂₁ O ₁₂	99.4	Delphinidin-3-O-glucoside	63.56 ± 1.05
41	11.83	611.159	611.1612	2.1	C ₂₇ H ₃₁ O ₁₆	91.6	Cyanidin-3-O-sophoroside	94.96 ± 1.58
43	13.572	465.106	465.1033	5.4	C ₂₁ H ₂₁ O ₁₂	82.4	Delphinidin-3-O-galactoside	11.38 ± 0.46