

Epitope-focused vaccine design

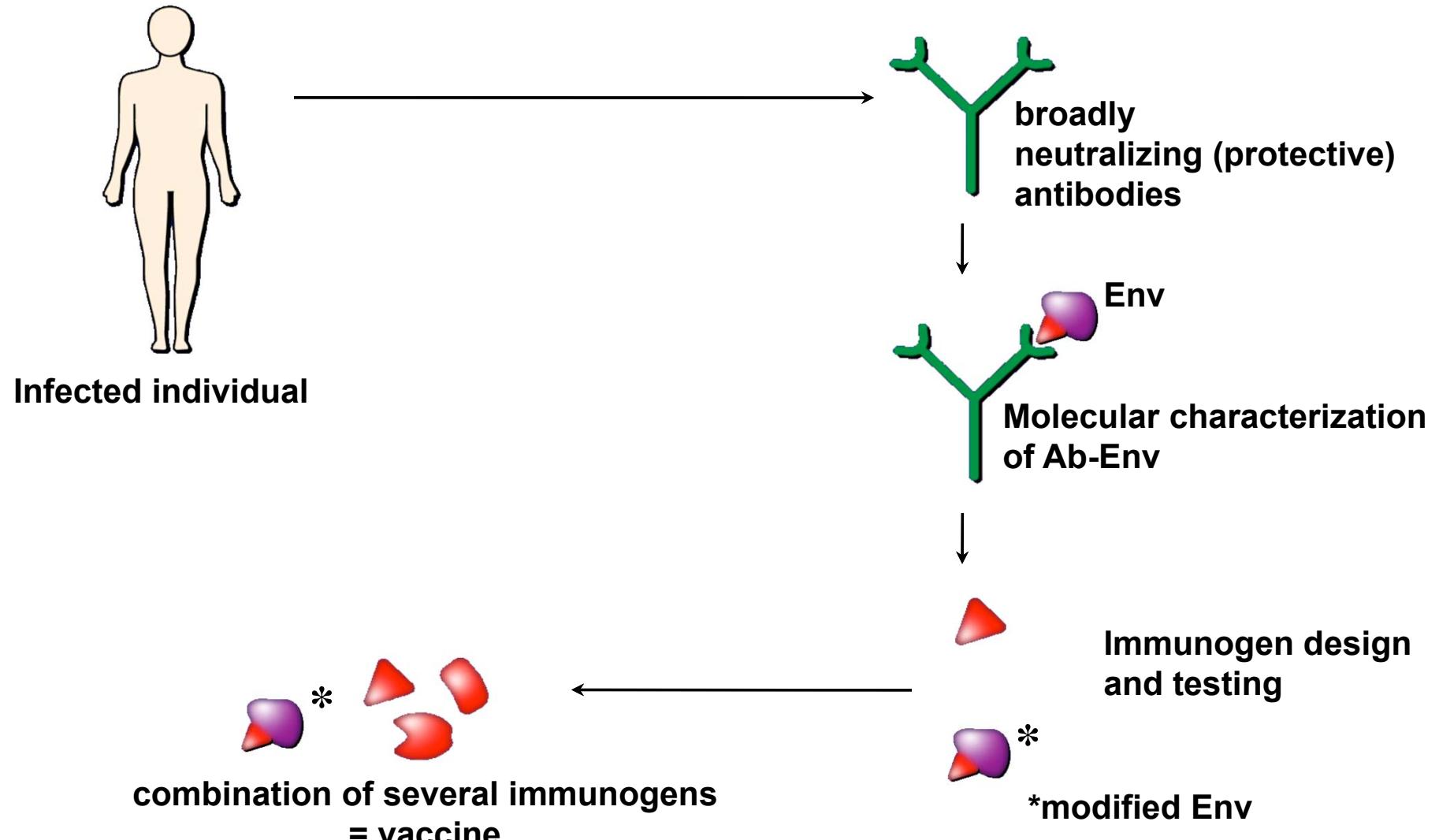
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2016 Global Vaccine and Immunization Research Forum
(GVIRF)

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Vaccine reverse engineering

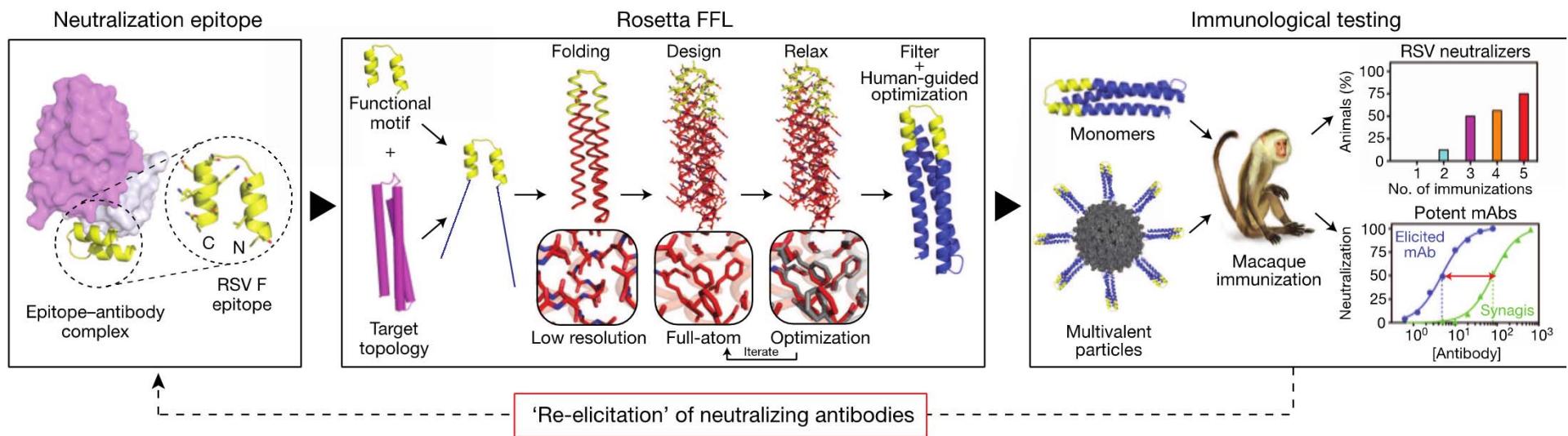


(adapted from Burton, Nat. Rev. Immunol., 2:706, 2002)

Three stories

- Proof of principle for epitope-focused vaccine design:
Epitope-scaffolds for motavizumab elicit potent
neutralization of RSV in NHPs
- Toward an HIV vaccine based on the CD4-binding site:
germline targeting to initiate induction of VRC01-class
broadly neutralizing antibodies (bnAbs)
- Proof of principle for elicitation of HIV bnAbs starting from
human germline B cells: vaccine induction of PGT121-class
glycan-dependent bnAbs by germline targeting and
reductionist boosting

Proof of concept for epitope-focused vaccine design: Epitope-scaffolds induce potent neutralization of RSV in NHPs



- A. Potent polyclonal serum neutralizing responses.**
- B. Rhesus mAbs isolated from an immunized macaque are more potent than Synagis**
- C. Rhesus mAbs recapitulate Mota structural specificity.**

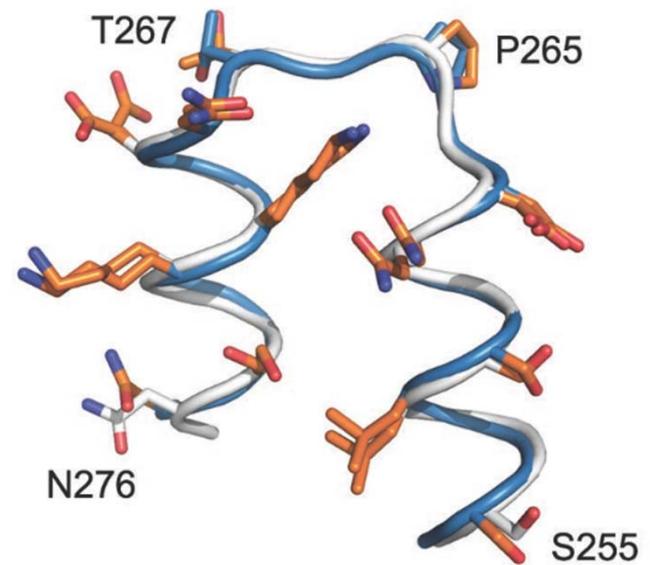
“Re-elicitation” of a neutralizing specificity: Vaccine-elicited mAb targets the same epitope structure as the humanized mAb that guided vaccine design



Guide structure
(Motavizumab+epitope
e)

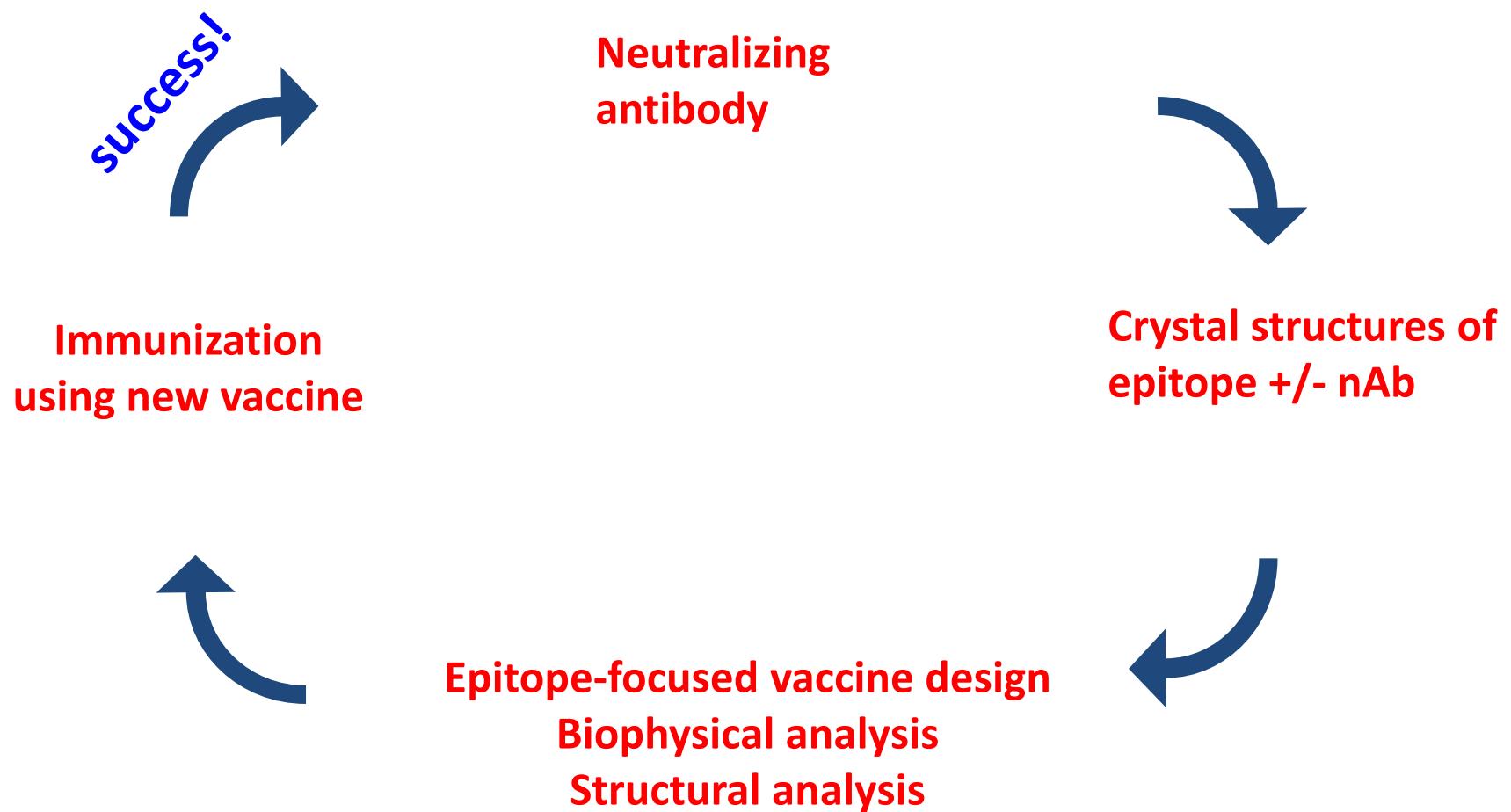


Structure of
vaccine-elicited mAB
(17HD9+epitope)



- ✓ epitope structures superimpose (RMSD = 0.5 Å)
- ✓ 85% of buried side-chains are shared

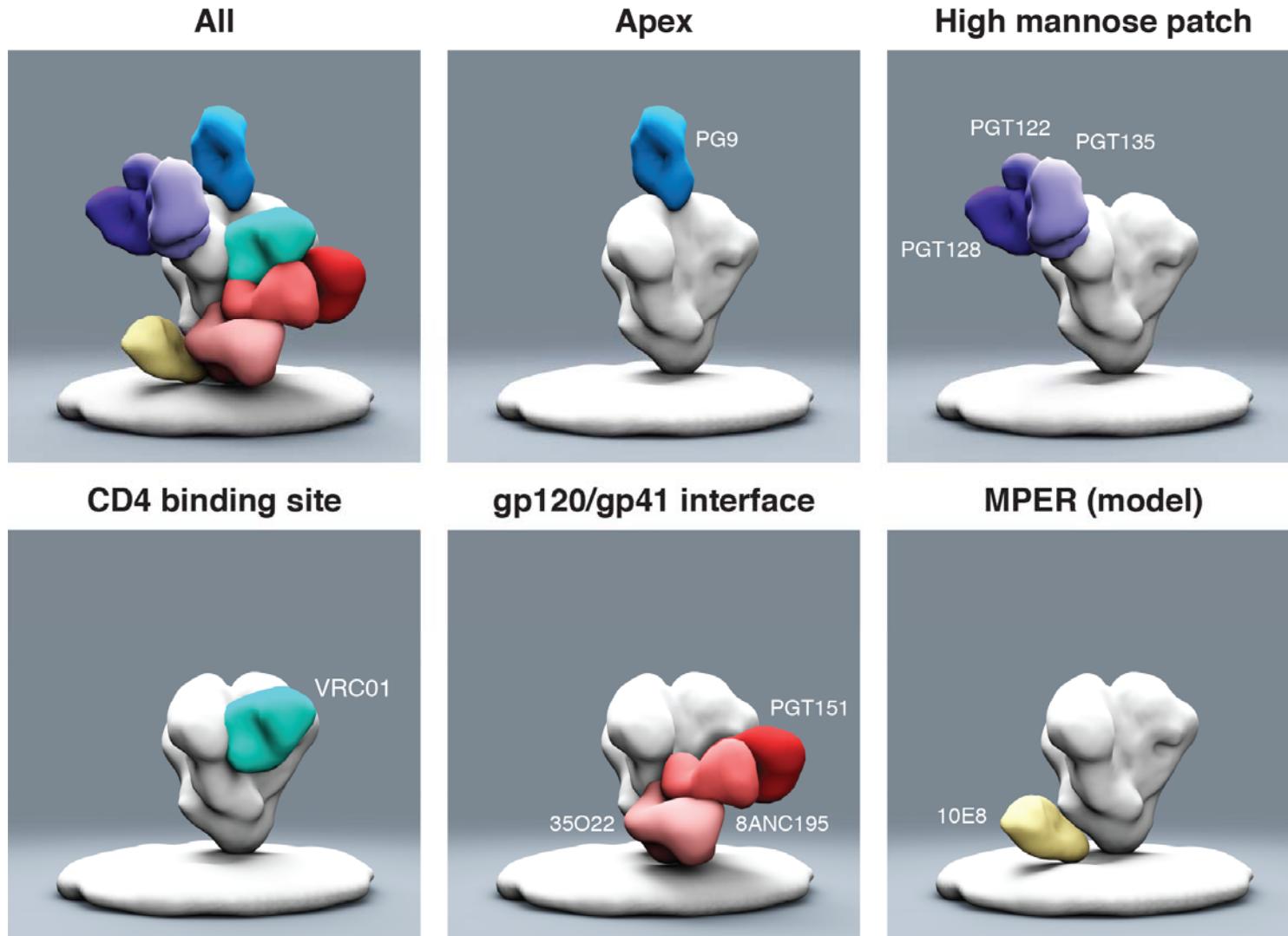
“Closing the loop” of Reverse Vaccine Engineering



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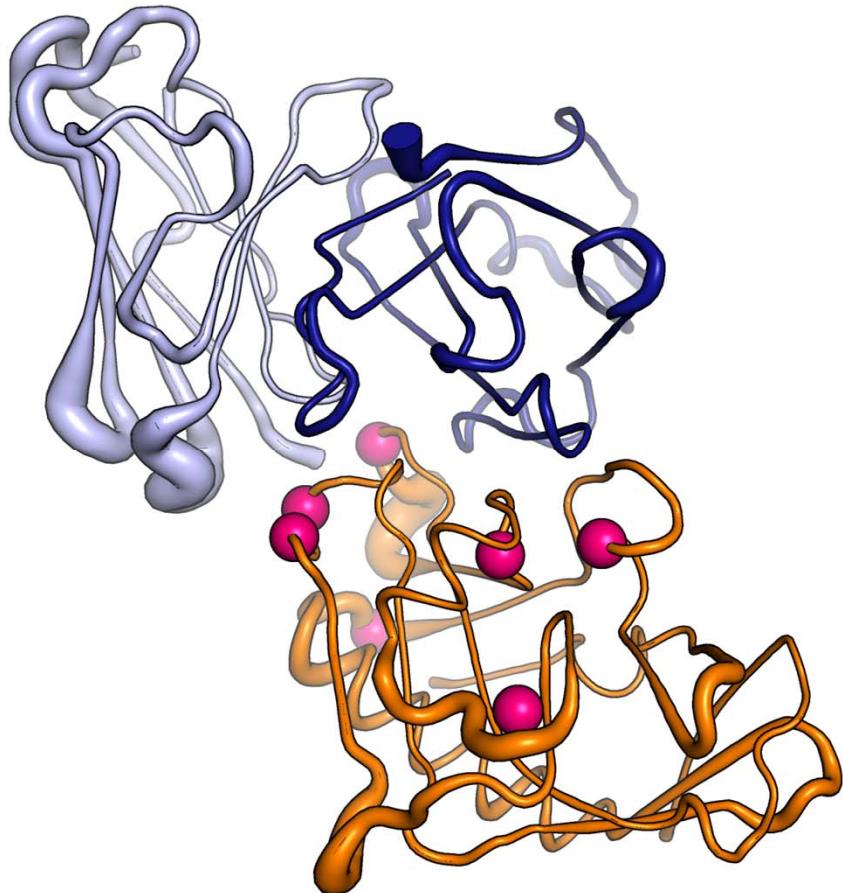
Prototype HIV bnAbs: binding regions



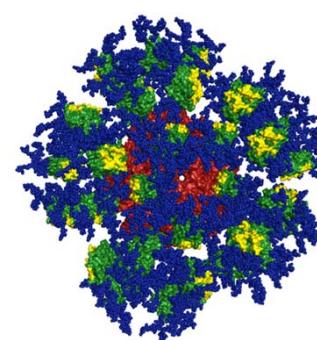
A. Ward & C. Corbaci

Development of VRC01-class Germline-Targeting Immunogen

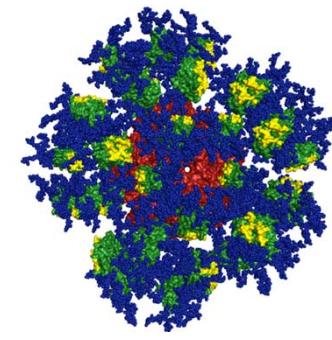
eOD-GT6 bound to GL-VRC01



Self Assembling Nanoparticles

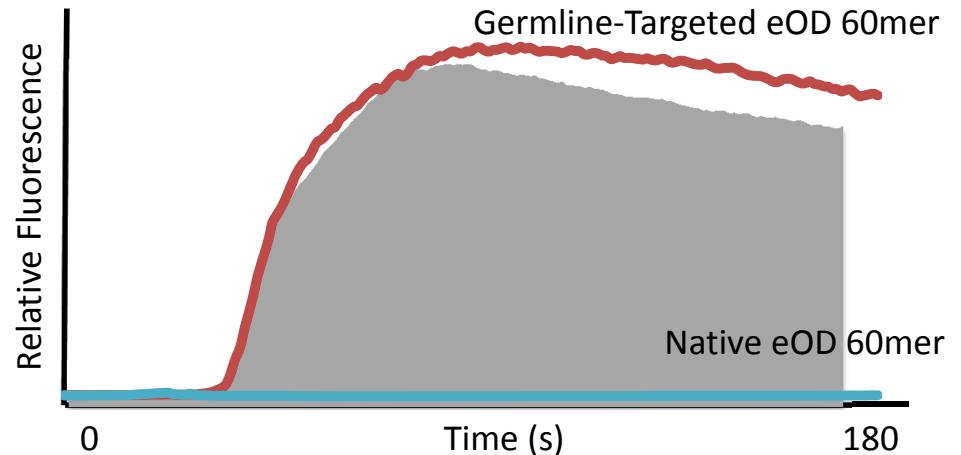


Germline-Targeted
eOD 60mer



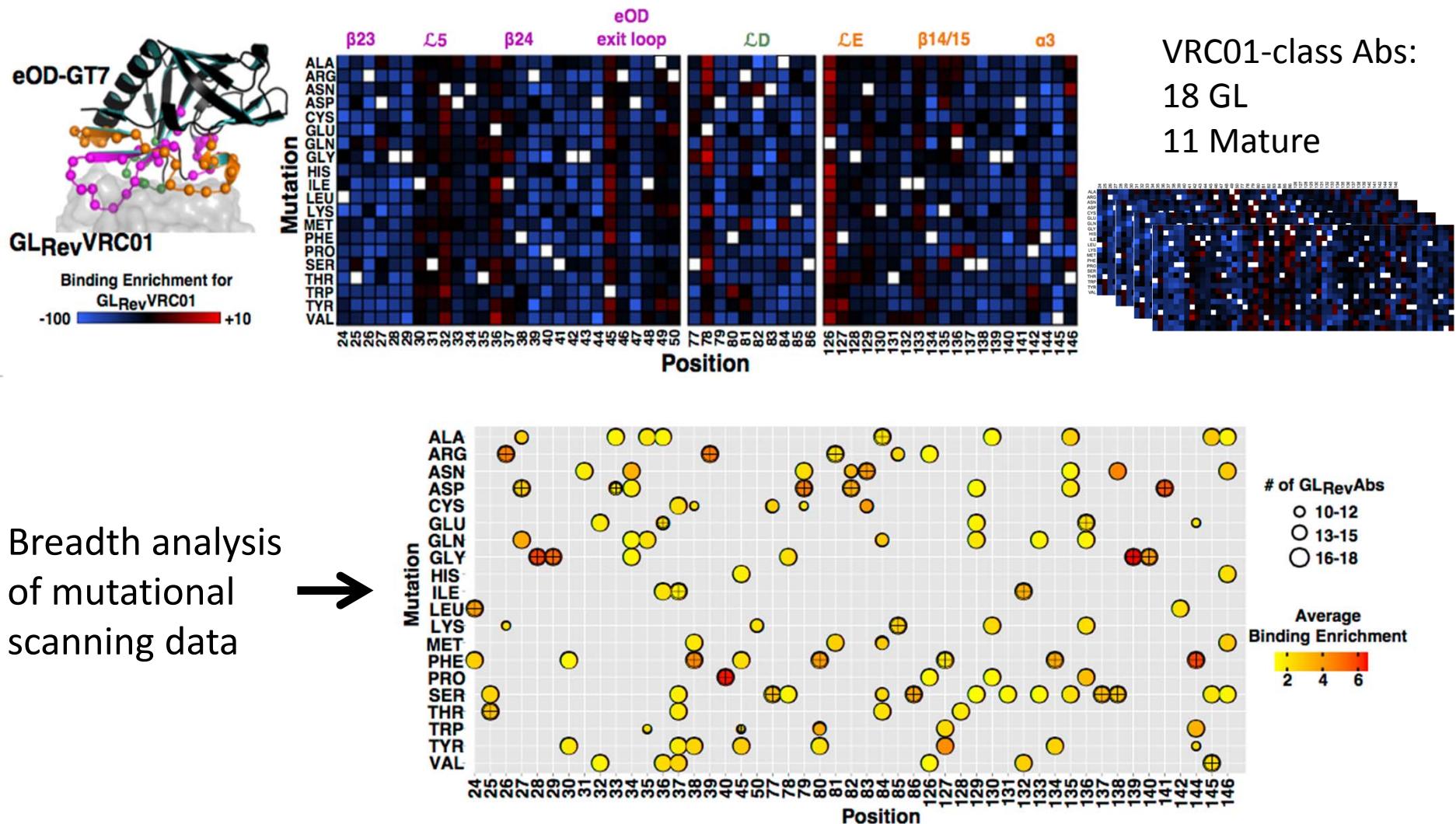
eOD 60mer
(Native CD4bs)

In Vitro Germline VRC01 B Cell Activation

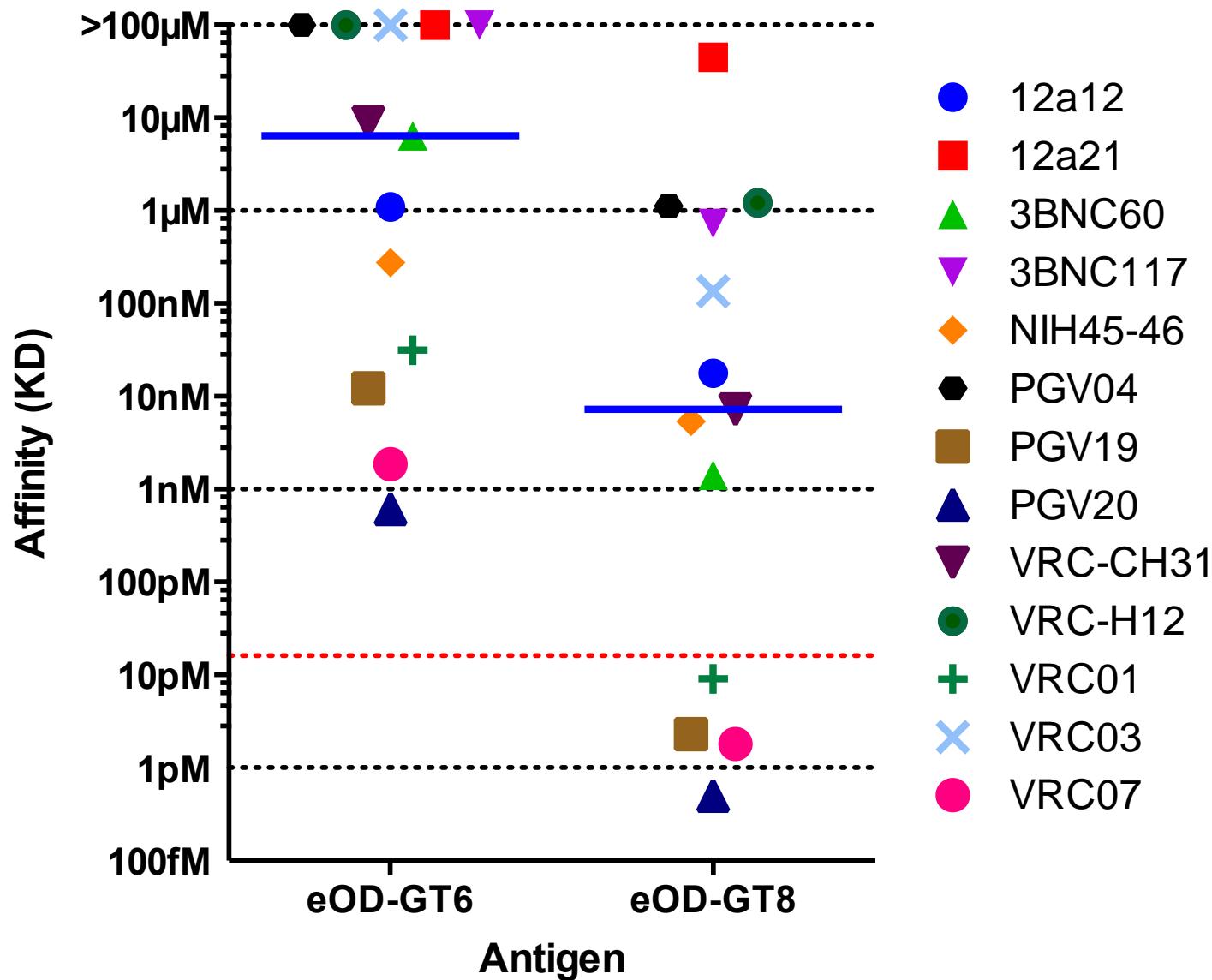


Jardine, Julien, Menis et. al., Science, 2013

Improvement of germline-targeting immunogen by deep mutational scanning and multtarget optimization



eOD-GT8 has improved binding to VRC01-class germline antibodies compared to eOD-GT6



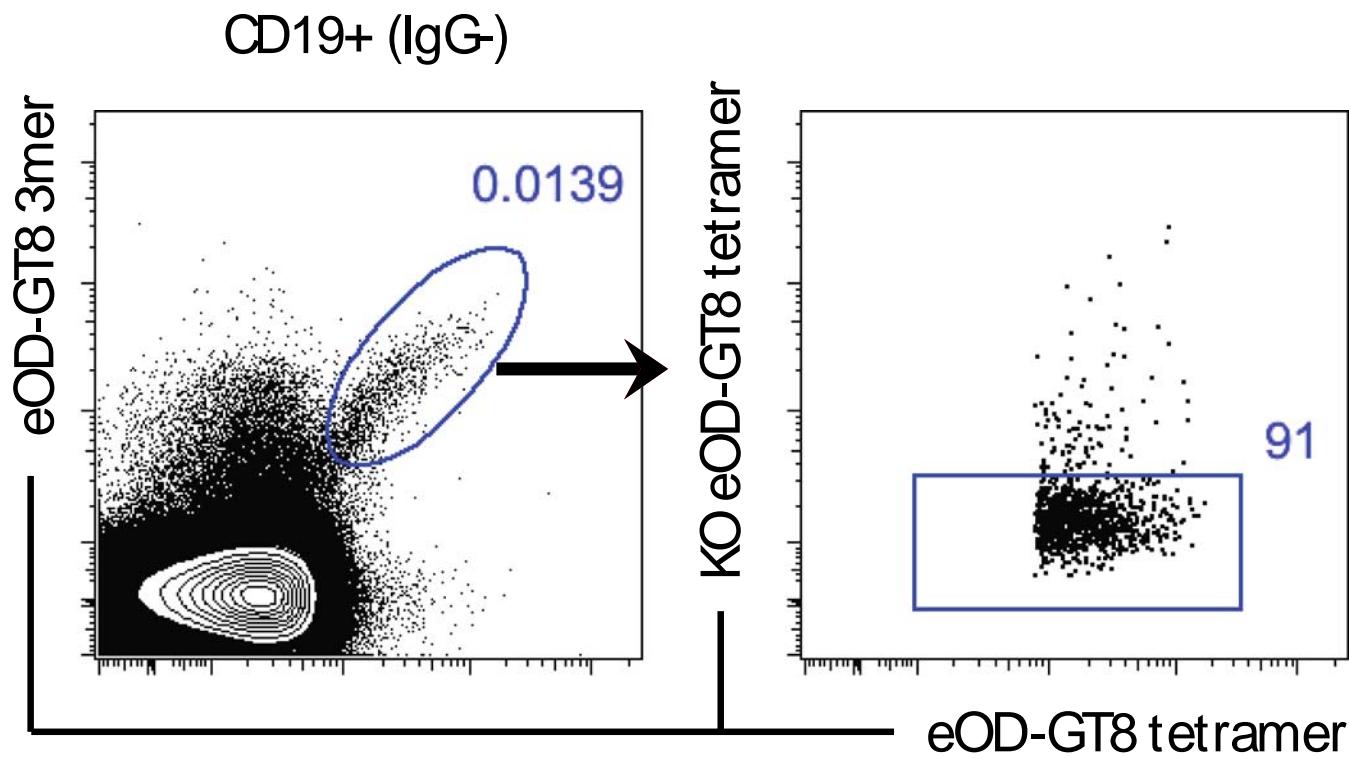
True vs germline-reverted VRC01-class precursors

Design and validation of germline-targeting immunogens has been based on “germline-reverted” precursors that use CDRH3 and CDRL3 loops from mature bnAbs.

This raises several important questions:

- what do “true” VRC01-class human precursors look like?
- How frequently do they occur in HIV-naïve humans?
- Do “true” VRC01-class precursors bind to eOD-GT8?
 - With sufficient affinity to allow B cell activation?

Sorting GT8 specific naïve human B cells



eOD-GT8 isolates VRC01-class precursors from 1 in 2.4 million human naïve B cells

Donor	B cells Screened (millions)	VRC01-class naive B cells	VH1-2 (*02/*03/*04) + 5aa CDRL3
1	1.6	1	
2	2.1	1	
3	0.9	0	
4	5.4	2	
5	0.6	0	
6	0.5	0	
7	1.8	0	
8	14.4	4	
9	7.8	6	
10	4.5	2	
11	7.0	1	
12	5.9	1	
13	1.1	2	
14	6.7	5	
15	1.3	1	
Total	61.6	26	

Frequency: 1 in 2.4 million

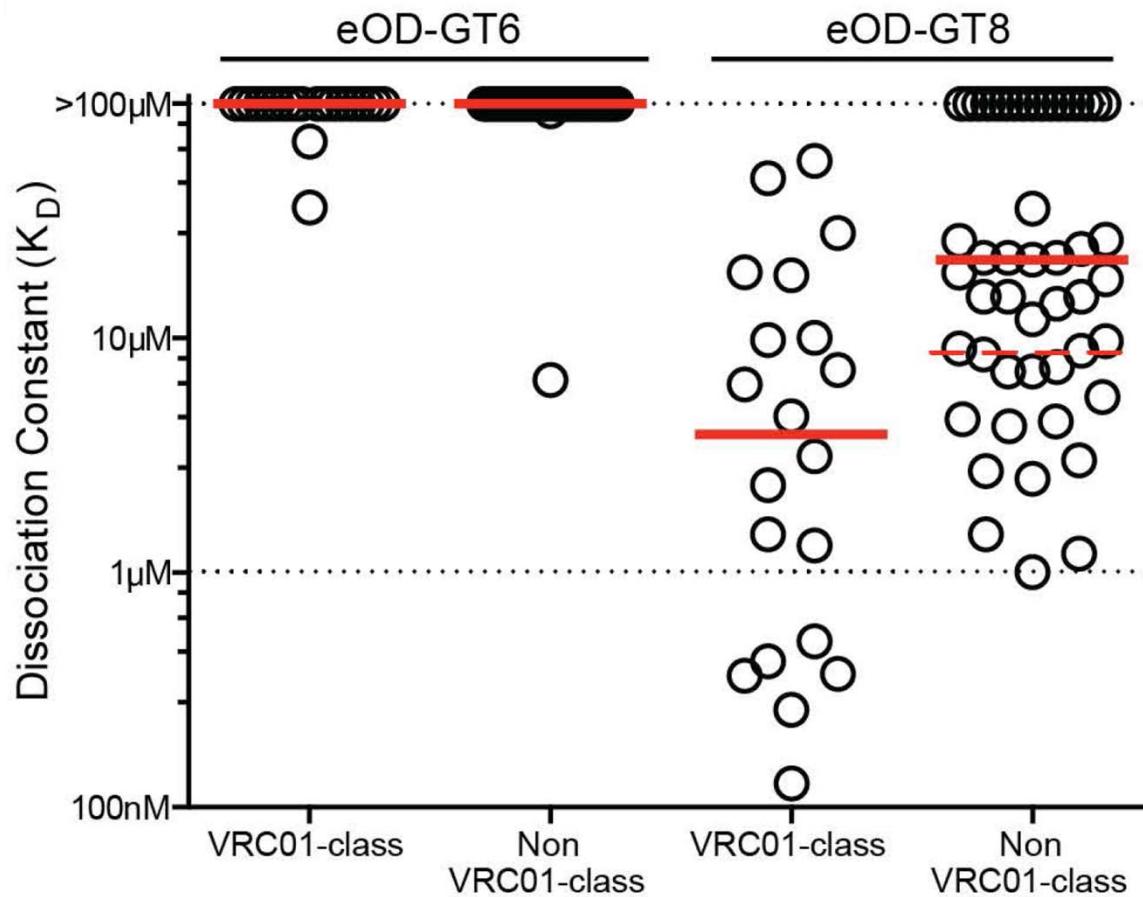
VRC01-class precursors that bind eOD-GT8 are common in humans

- 10^{10} - 10^{11} B cells in adult human
- ~50 million B cells per human lymph node
- 65-75% are naïve B cells
- 96% of humans are hetero/homozygous for VRC01-class VH1-2 alleles (*02, *03, *04)

A frequency of 1 VR01-class precursor in 2.4 million naïve B cells means that:

- nearly all humans have 2700 to 31,000 VRC01-class precursors that bind eOD-GT8
- each human lymph node has ~15 VRC01-class precursors that bind eOD-GT8

eOD-GT8 binds with 0.1-30 μ M affinity to VRC01-class precursors (and has higher affinity for VRC01-class compared to non-VRC01-class)



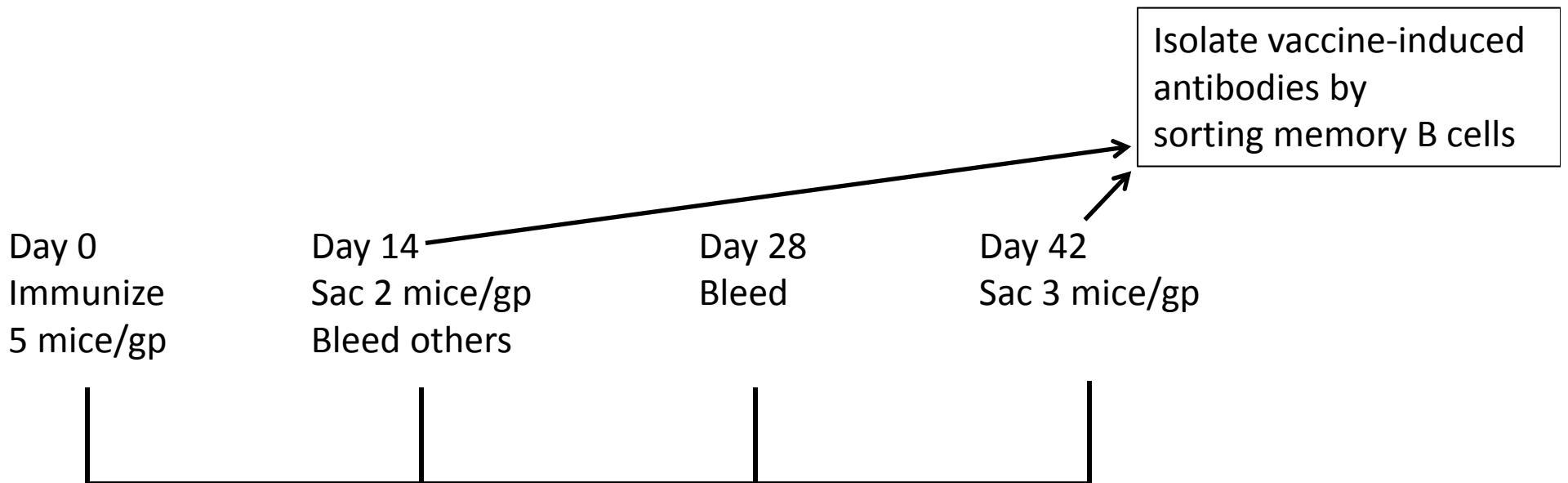
This is promising for *in vivo* activation and VRC01-class memory B cell generation during human vaccination:

- ◆ Batista, Neuberger JEM 1998
- ◆ Dal Porto, Shlomchick JEM 2002
- ◆ Shih, Nussenzweig Nat Imm 2002
- ◆ Paus, Brink JEM 2006

Test of germline-targeting immunogens in VRC01 gH mice

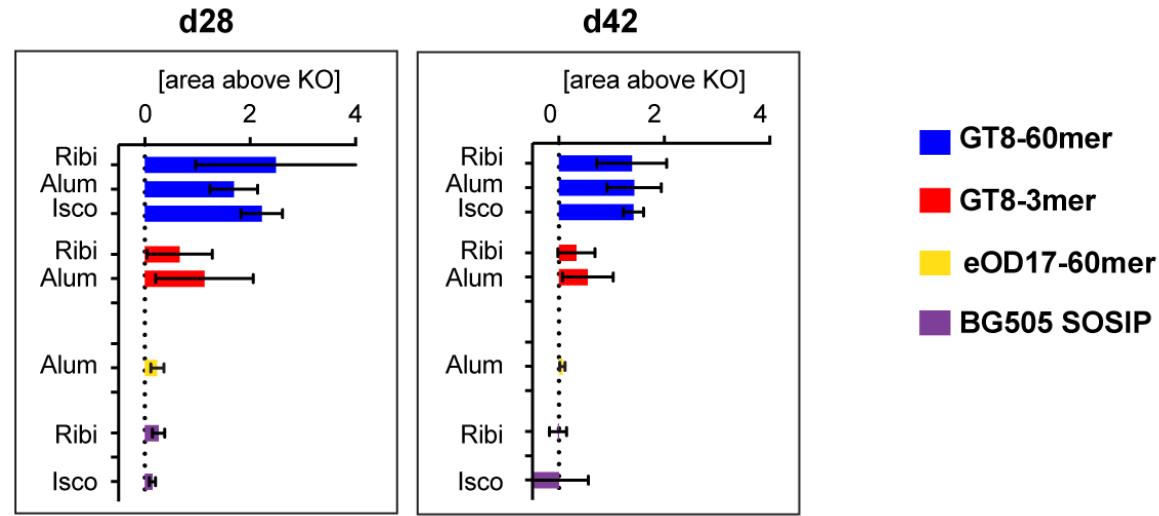
(David Nemazee)

Can eOD-GT8 activate appropriate B cells and select productive mutations, to produce memory B cells that could be boosted by a more native-like immunogen?

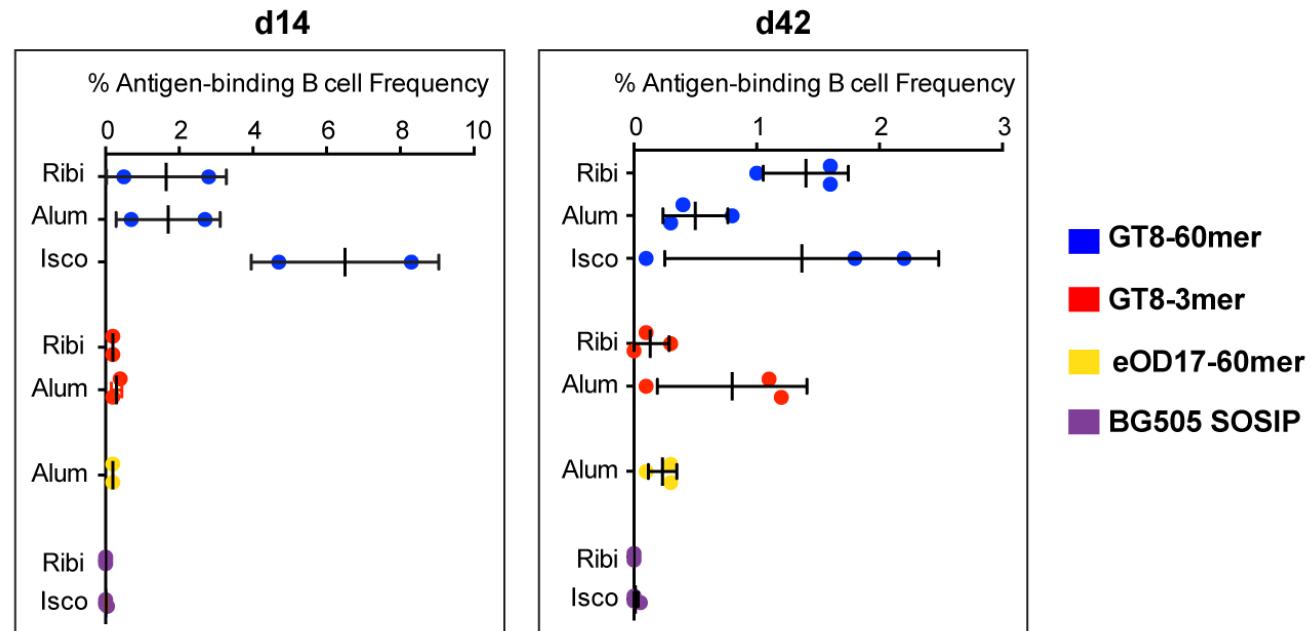


Serum and B cell responses in VRC01 gH mice showed robust responses to the eOD-GT8 60mer

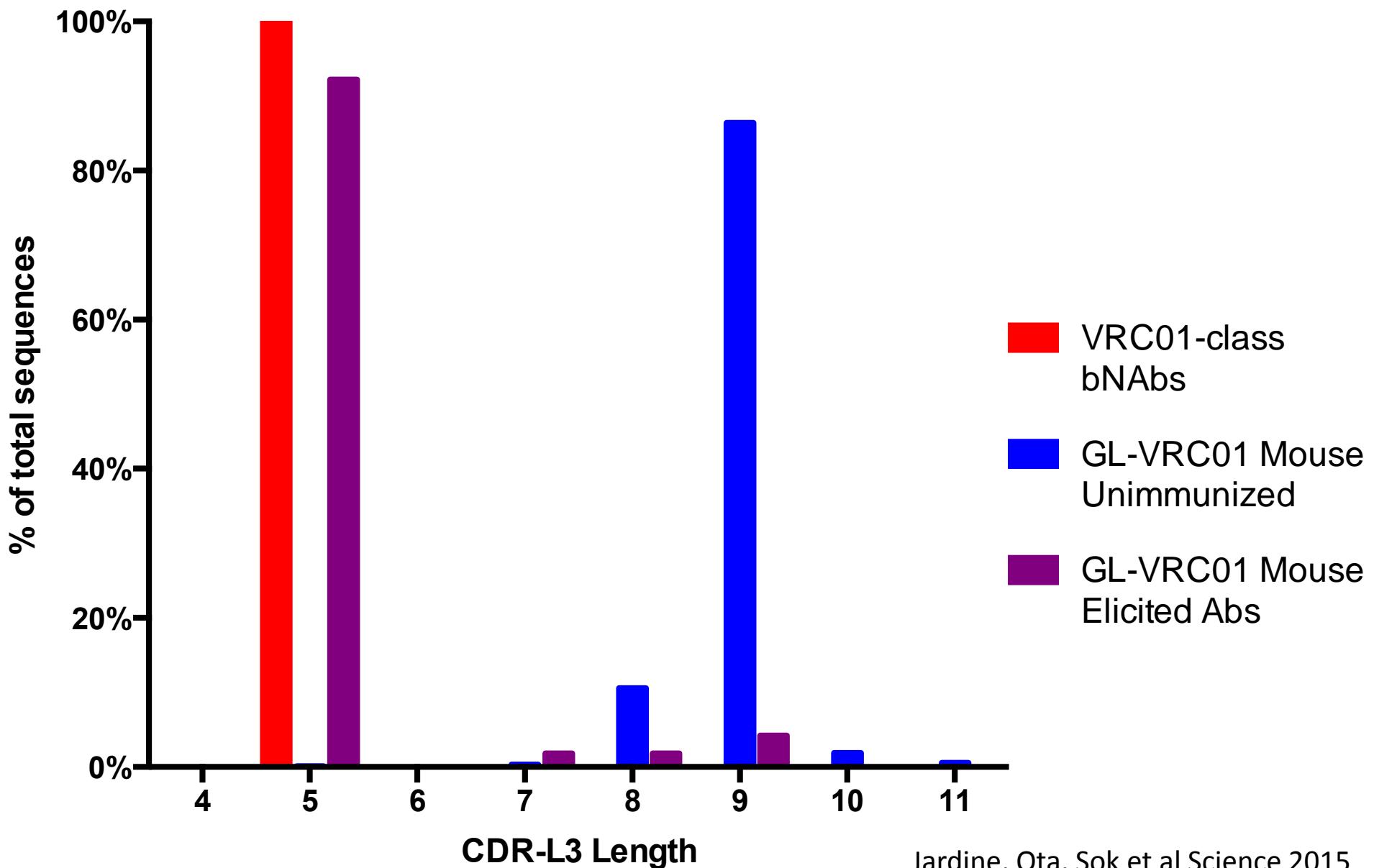
ELISA responses



Epitope-specific B cell frequencies



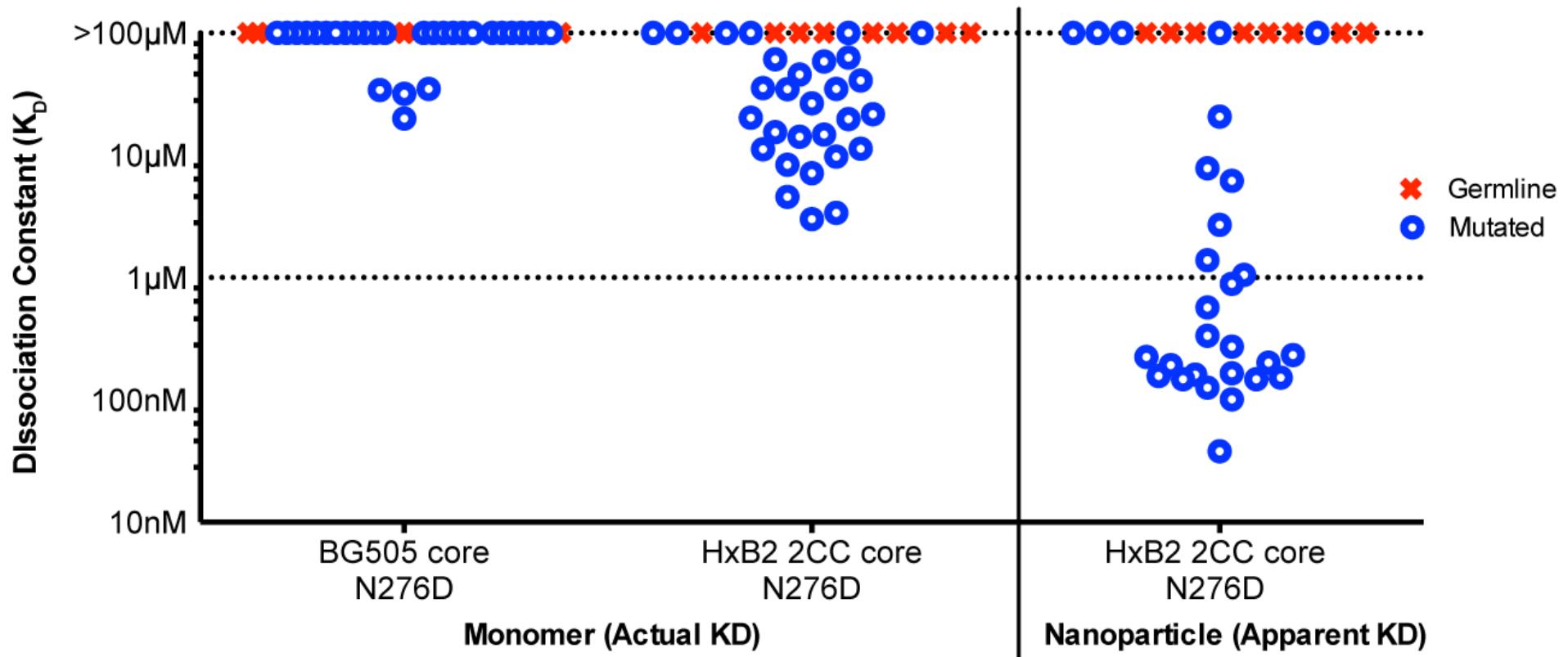
eOD-GT8 60mer reliably elicits Abs with 5AA CDR-L3



Jardine, Ota, Sok et al Science 2015

eOD-GT8 60mer induced Abs bind to near-native CD4bs in candidate boost immunogens

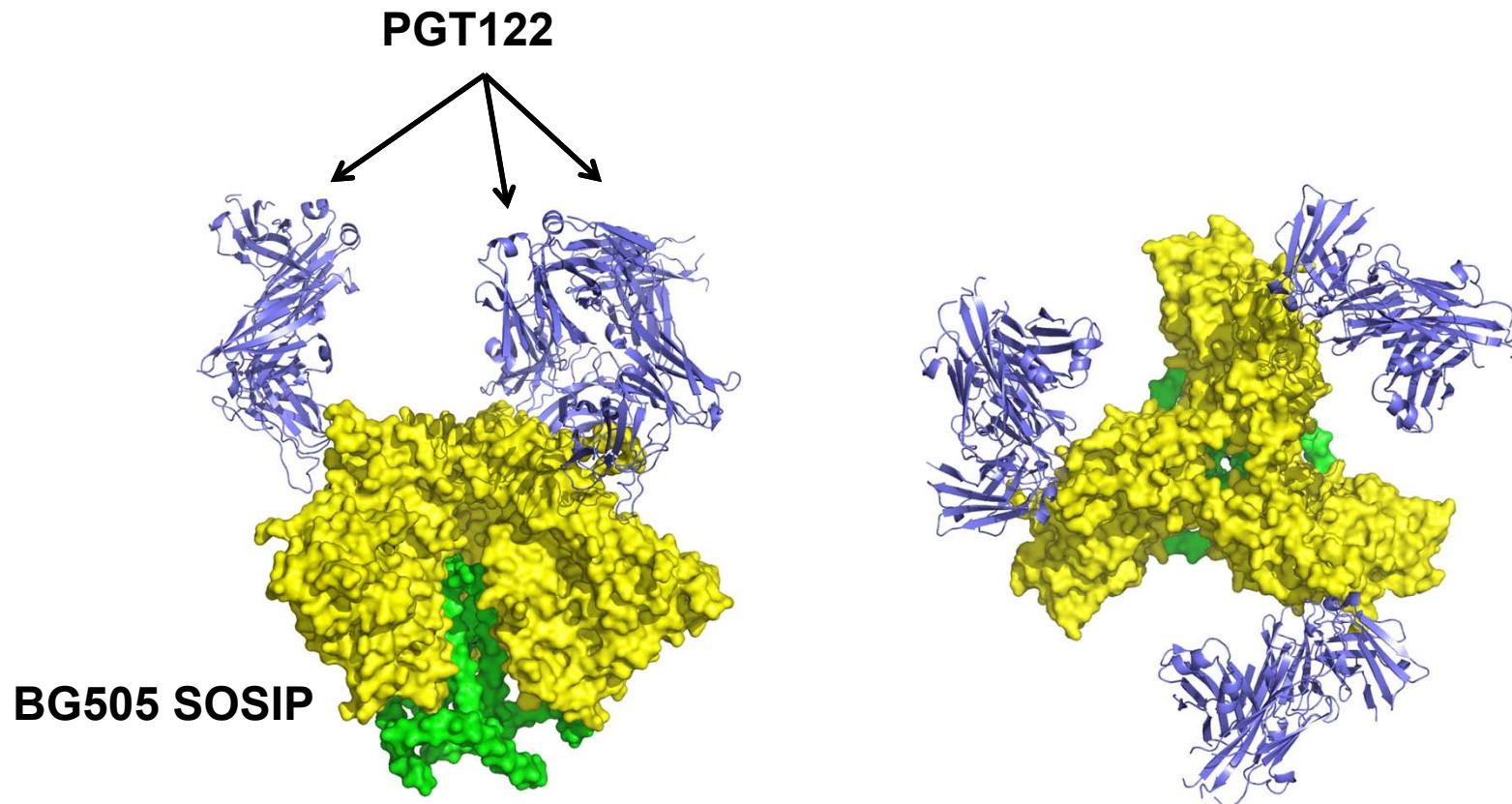
- GT8-induced monoclonal Abs were isolated by sorting of memory B cells from day 42
- Antibody affinities for boost candidates were measured by SPR



Three stories

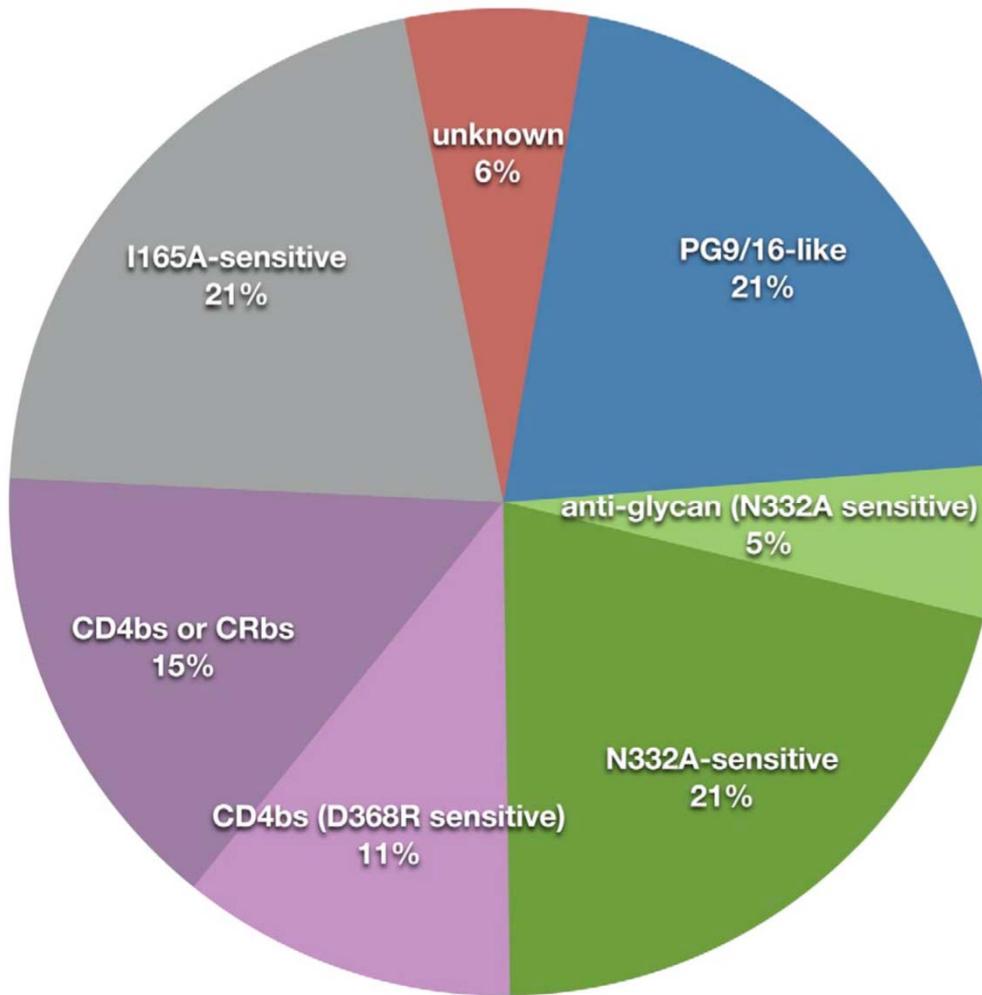
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PGT121-class glycan-dependent bnAbs by germline
targeting and reductionist boosting

PGT121-class interaction with native-like trimer defined



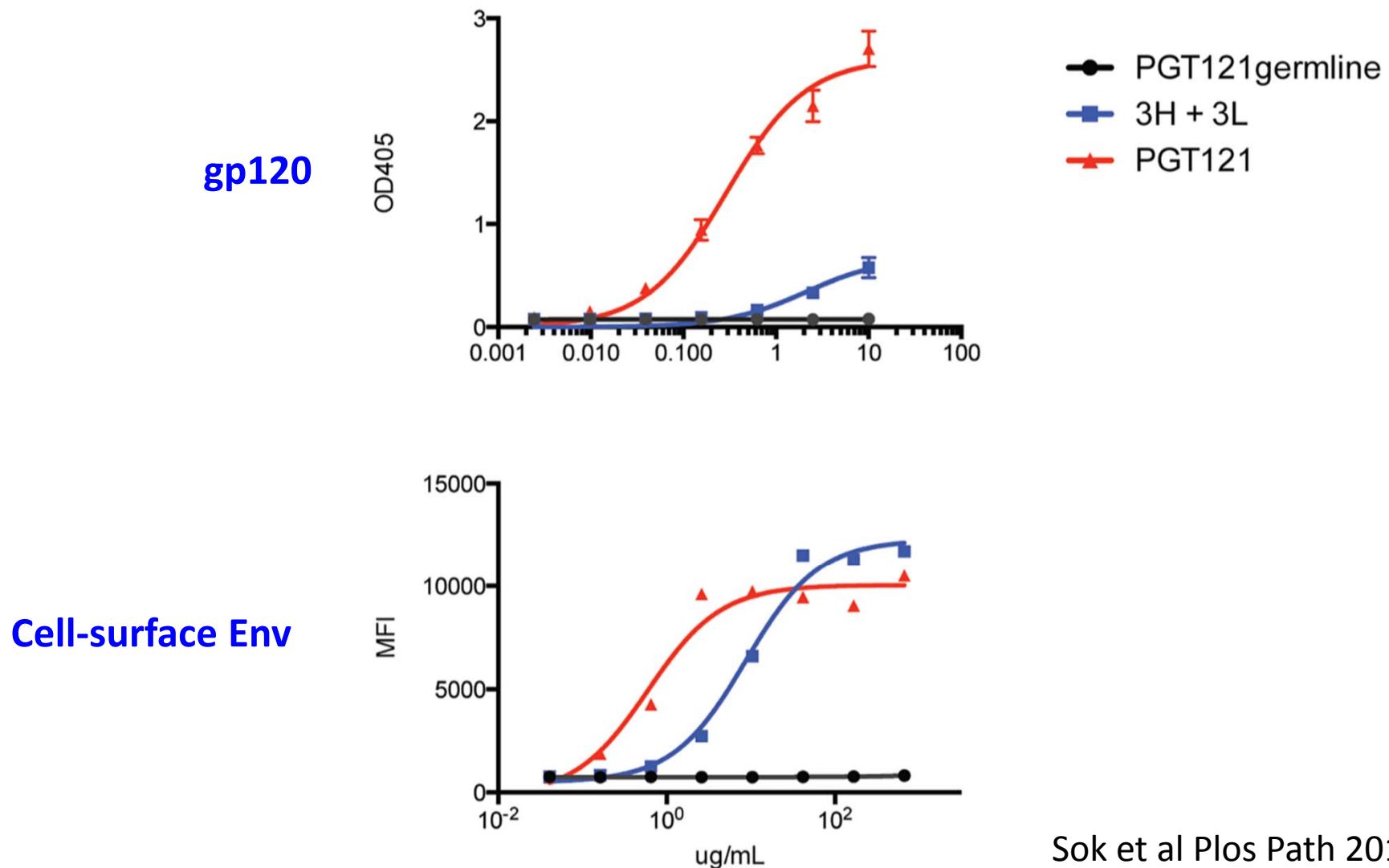
Julien et al Science 2013

PGT121-class bnAbs among the most common from infection



Walker et al Plos Path 2010

Barrier to elicitation: PGT121 germline-reverted Abs lack detectable affinity for gp120 or cell-surface Env



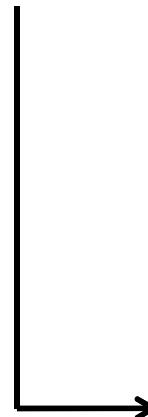
Sok et al Plos Path 2013

Mammalian display/ directed evolution overview

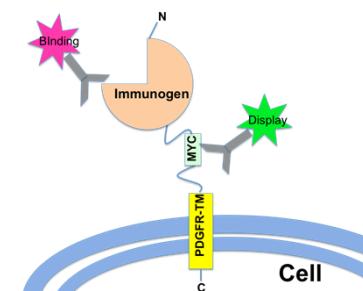
1. Produce lentivirus containing antigen library



2. Transduce 293T cells
Low moi (<0.1)



3. Induce expression (Doxycycline)



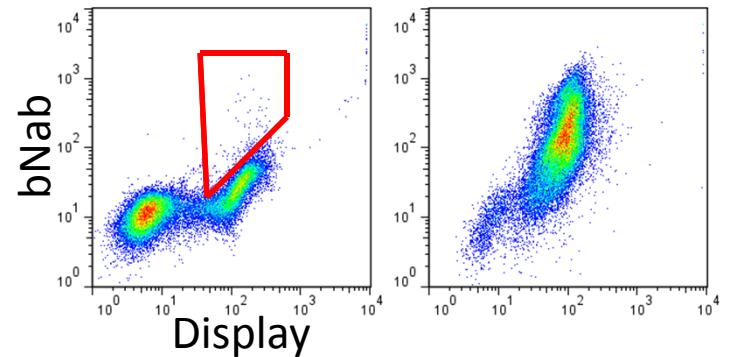
6. Biophysical Characterization



5. Purify genomic DNA, PCR and sequence

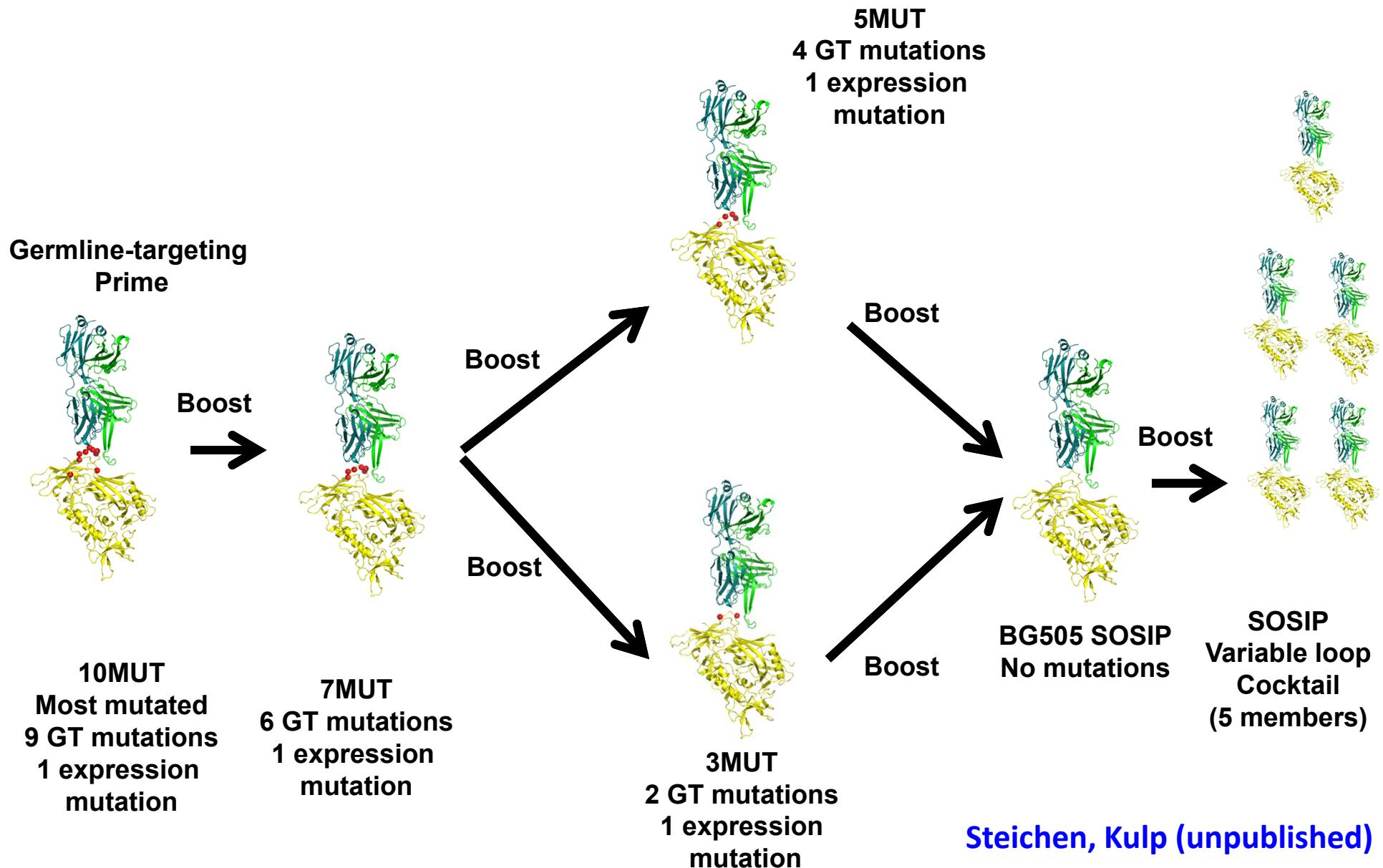


4. Sort cells



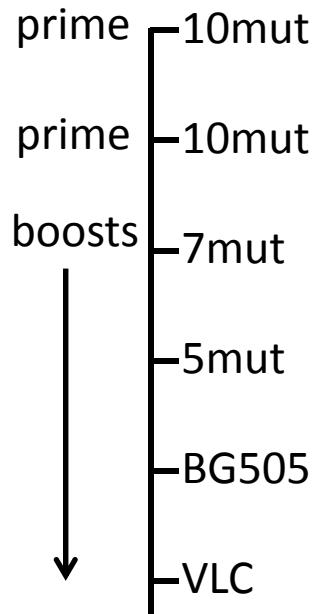
Jon Steichen (unpublished)

Reductionist germline-targeting/boosting strategies to induce PGT121-like bnAbs



Elicitation of tier 2 cross-neutralizing antibodies by reductionist vaccine design in PGT121 gHgL mouse (with Nussenzweig, Burton)

Immunizations
(All SOSIP except 1st)



Neut measured
from purified
serum IgG

Escolano, Steichen, et al. (unpublished)

Specificity of tier 2 cross-neutralization from gHgL mouse tracks closely With PGT121 “ancestor” 3H3L

Virus	clade	MJ6	3H3L	9H3L	PGT121	IC50 ug/mL
6535.3	B	1019	0.043	0.03	0.002	<0.01
TRO.11	B	<25	1.445	0.071	0.005	0.01 - 0.1
CAAN5342.A2	B	1800	0.914	0.236	0.007	0.1 - 1.00
RHPA4259.7	B	<25	>50	0.073	0.015	>1.00
JR-CSF	B	~10	>50	0.277	0.02	
AC10.0.29	B	<25	>50	2.643	0.024	
PVO.4	B	<25	>50	>50	0.098	
SC422661.8	B	<25	>50	>50	0.038	
QH0692.42	B	<25	>50	>50	0.302	
WITO4160.33	B	<25	>50	>50	0.334	
REJO4541.67	B	<25	>50	>50	4.774	
SC05_8C11_2344	B(T/F)	<25	0.386	0.475	0.019	
WEAU_d15_410_50:B(T/F)		<25	1.329	0.3	0.026	
6240_08_TA5_4622	B(T/F)	<25	4.137	0.145	0.033	
1012_11_TC21_3257B(T/F)		<25	>50	0.506	0.003	
1006_11_C3_1601	B(T/F)	<25	>50	14.62	0.002	
1056_10_TA11_182eB(T/F)		<25	>50	2.237	0.004	
1054_07_TC4_1499	B(T/F)	<25	>50	34.74	0.064	
6244_13_B5_4576	B(T/F)	<25	>50	>50	0.061	
62357_14_D3_4589	B(T/F)	<25	>50	>50	2.597	
IAVI C22	C	4429	0.012	0.008	0.002	
DU156.12	C	2877	0.026	0.053	0.004	
HIV-001428-2.42	C	<25	>50	19.74	0.014	
ZM53M.PB12	C	<25	>50	>50	0.001	
Du172.17	C	<25	>50	>50	0.033	
Du422.1	C	<25	>50	>50	0.039	
HIV-16055-2.3	C	<25	>50	>50	0.153	
ZM214M.PL15	C	<25	>50	>50	0.46	
ZM135M.PL10a	C	<25	>50	>50	0.716	
CAP45.2.00.G3	C	<25	>50	>50	1.634	
ZM109F.PB4	C	<25	>50	>50	8.639	
Ce1176_A3	C(T/F)	>5000	0.026	0.078	0.013	
7030102001E5(Rev-)C(T/F)		<25	5.56	1.619	0.009	
1394C9G1(Rev-)	C(T/F)	<25	17.108	1.081	0.264	
Ce704809221_1B3	C(T/F)	<25	>50	11.58	0.025	
246F C1G	C(T/F)	<25	>50	>50	0.041	
ZM247v1(Rev-)	C(T/F)	<25	>50	>50	0.028	
CNE20	BC	2073	0.006	0.005	0.003	
CNE21	BC	<25	2.226	0.256	0.007	
CNE52	BC	<25	>50	3.188	2.045	
CNE19	BC	<25	>50	>50	0.008	
CNE17	BC	<25	>50	>50	7.6	
92RW020	A	1138	0.01	0.012	0.002	
Q23.17	A	>5000	0.007	0.021	0.001	
191084 B7-19	A	<25	38.078	3.994	0.011	
0260.v5.c36	A	<25	>50	15.05	0.53	
0330.v4.c3	A	<25	>50	>50	0.05	
T250-4	CRF02_AG	<25	0.202	0.25	0.001	
235-47	CRF02_AG	<25	>50	>50	0.137	
T251-18	CRF02_AG	<25	>50	>50	29.016	
P1981_C5_3	G	>5000	0.011	0.015	0.001	
X2131_C1_B5	G	<25	8.487	0.224	0.004	
X2088_c9	G	<25	31.256	7.019	0.003	
X1193_c1	G	<25	29.989	11.29	0.016	
X1254_c3	G	<25	1.271	1.832	0.014	
A07412M1.vrc12	D	<25	1.532	0.173	0.009	
6811.v7.c18	CD	3417	0.003	0.001	0.001	
6480.v4.c25	CD	1120	0.033	0.015	0.001	
6952.v1.c20	CD	<25	>50	>50	0.056	
3817.v2.c59	CD	<25	>50	>50	18.888	
3301.v1.c24	C	<25	0.793	0.432	0.008	
3103.v3.c10	ACD	<25	0.062	0.03	0.009	
0815.v3.c3	ACD	<25	0.622	1.549	0.025	

Escolano, Steichen, et al. (unpublished)

Neutralization breadth of best MJ6 mAbs is similar to 3H3L

	MJ6-1	MJ6-2	MJ6-3	3H3L	PGT121	IC50 ug/mL
6535	0.009	0.014	0.026	0.043	0.002	<0.01
92RW020	0.004	0.012	0.022	0.01	0.002	0.01 - 0.1
IAVI C22	0.003	0.008	0.011	0.012	0.002	0.1 - 1.00
Q23	0.005	0.016	0.024	0.007	0.001	>1.00
DU156	0.011	0.024	0.059	0.026	0.004	
P1981	0.004	0.007	0.016	0.011	0.001	
X2088	0.037	0.609	0.316	31.3	0.003	
191084B7	0.712	0.862	>50	38.1	0.011	
JR-CSF	0.334	0.739	>50	>50	0.02	
BG505 T332N	0.55	0.598	>50	0.064	0.026	
T250	>50	>50	>50	0.202	0.001	
HIV-001428	>50	>50	>50	>50	0.014	
PV0.4	>50	>50	>50	>50	0.098	
ZM53	>50	>50	>50	>50	0.001	
CNE19	>50	>50	>50	>50	0.008	
R1166	>50	>50	>50	>50	>50	
MLV	>50	>50	>50	>50	>50	

Conclusions/Outlook

- ◆ A key challenge for HIV vaccine design is immuno-focusing to bnAb epitopes
- ◆ Similar challenges for other antigenically highly variable pathogens such as influenza and hepatitis C viruses, and related challenges for dengue virus
- ◆ RSV scaffold immunogens:
Re-capitulation of Mota neutralization specificity provides proof of principle that epitope-focused vaccine design can achieve immuno-focusing with high precision
- ◆ Vaccines to induce bnAbs against HIV:
Hypothesize that (a) germline-targeting is needed to consistently activate bnAb precursors in vaccine recipients and (b) structure-guided boosting strategies are needed to guide SHM to produce bnAbs.

VRC01 example: Germline-targeting eOD-GT8 60mer has promise to initiate induction of VRC01-class bnAbs. To be tested in humans (IAVI/BMGF). Reductionist boosting strategies to induce VRC01-class bnAbs being tested in various transgenic mice.

PGT121 example: Demonstrated proof of principle for vaccine-induction of HIV bnAbs starting from human germline B cells, by PGT121 germline-targeting and reductionist boosting (using engineered SOSIP trimers) in PGT121 gHgL mice. **A major milestone for HIV vaccine development.** Precursor frequency in humans remains a question.

Acknowledgments– RSV scaffolds

Schief lab

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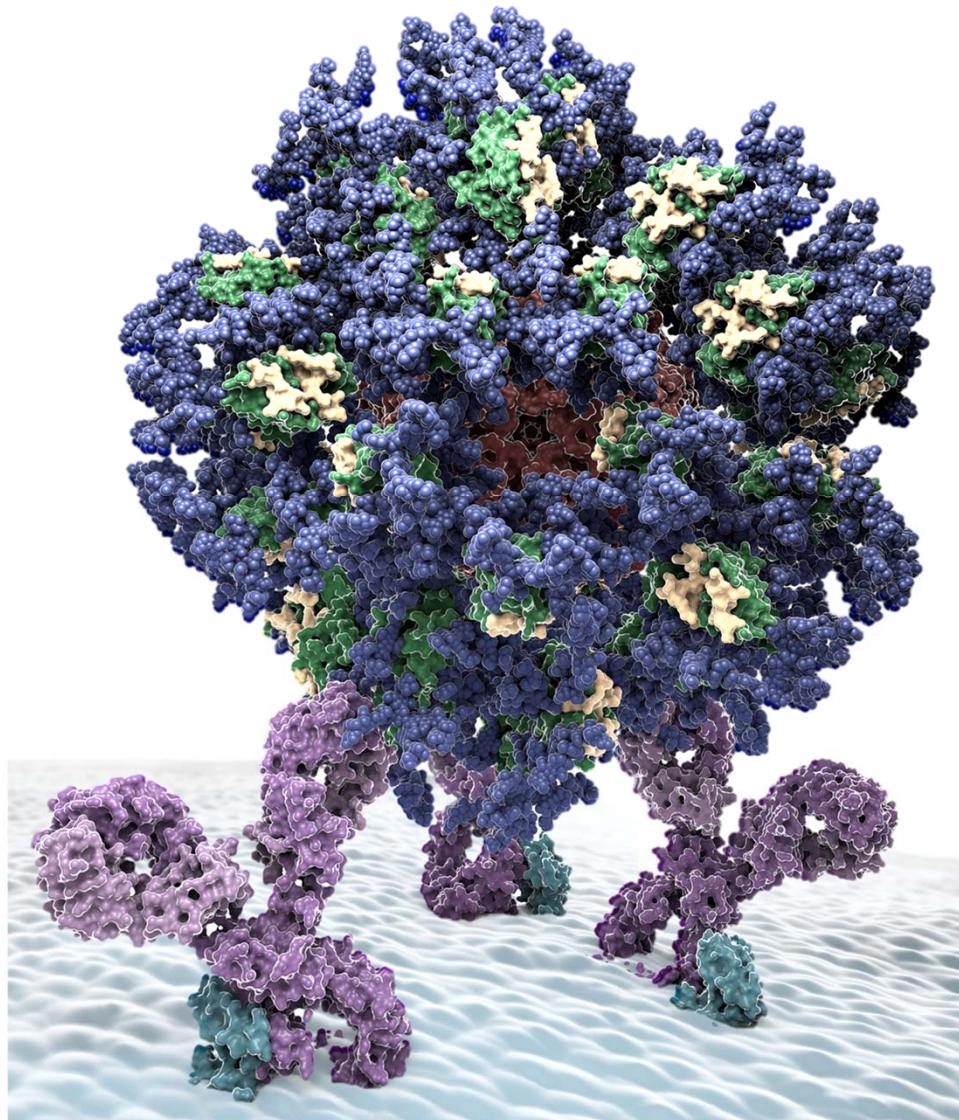
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Ryan Tingle

Jordan Willis

Burton lab

Laura McCoy

Bryan Briney

Devin Sok

Nemazee lab

Taka Ota

Deepika Bhullar

Nussenzweig lab

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