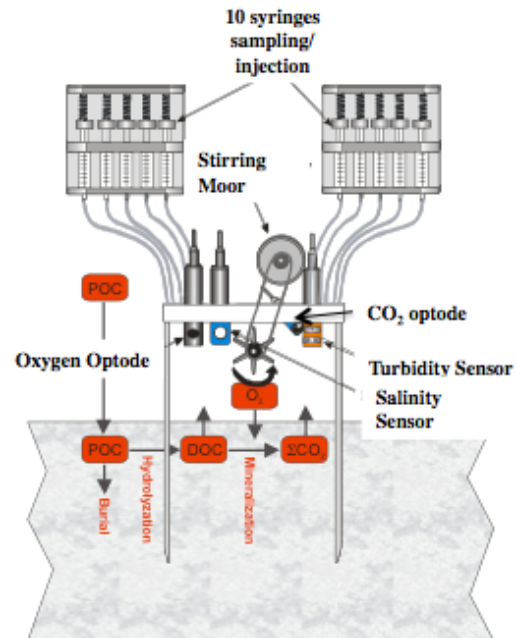
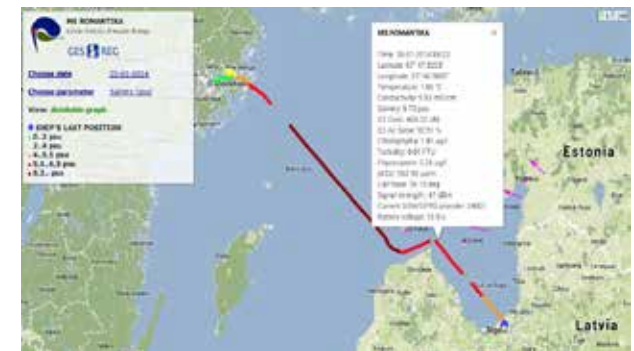


Oxygen measurements: why and when?

Anders Tengberg (anderste@chem.gu.se) Associate Professor at the University of Gothenburg and at the Chalmers University of Technology, Sweden, Scientific Advisor/Product Manager at Aanderaa since 1997.



Outline

9:00-10:30

- Why should you measure Oxygen (O_2)?
- How is O_2 related to other parameters?
- Are we measuring at the right frequencies?
- Methods to analyze and measure O_2

11:00-13:00

- What affects O_2 sensors/methods to evaluate performance?
- How good is Winkler titration?
- Field data, value of combined measurements
- Summary
- Practical Demonstration and Hands-On

Why should you measure Oxygen?

1. Most important parameter and “easy” to measure
2. Crucial for higher organisms (fish, bottom living)
3. Crucial for biological, chemical & geological processes
4. Oceanographic tracer
5. Climate related, geologically has O₂ been constant?

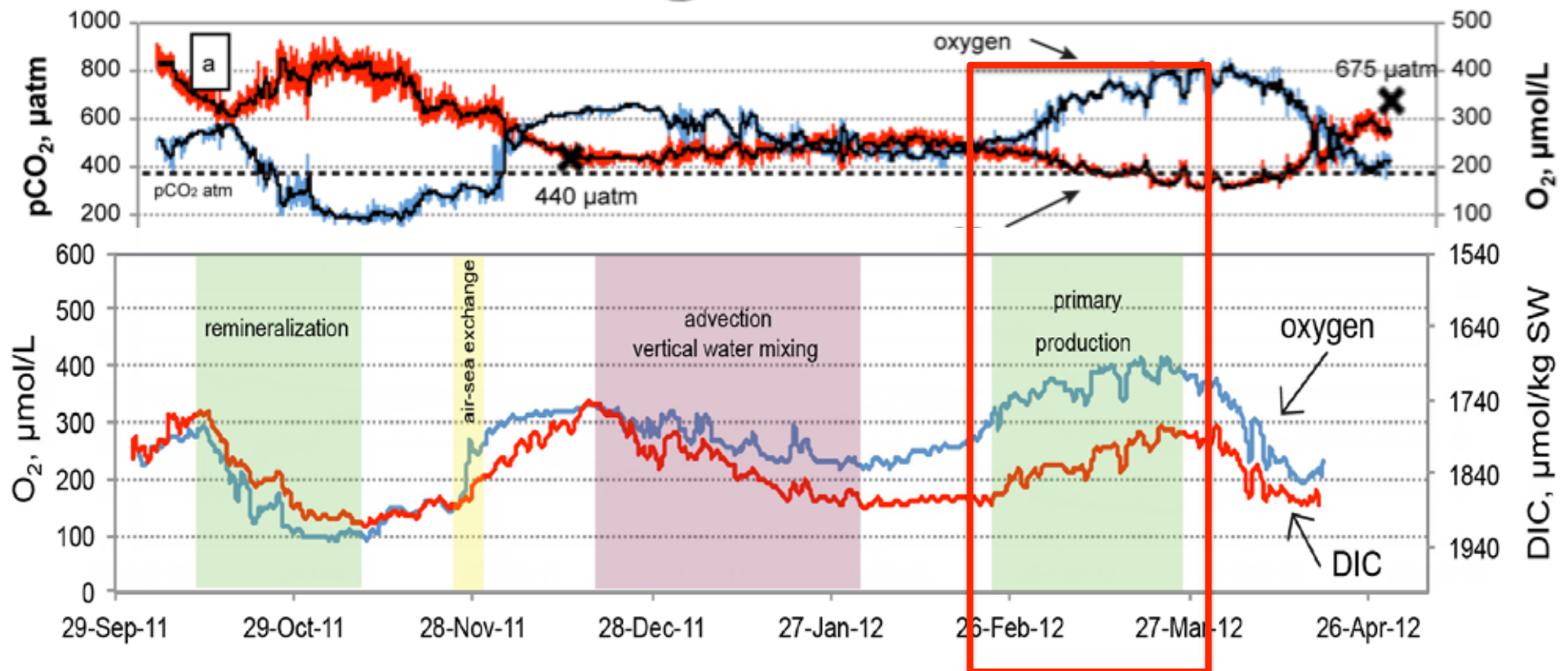
How is Oxygen related to other parameters?

1. Temperature: higher Temp less O₂
2. Salinity: higher Sal lower O₂
3. Currents: higher currents more turbulence/more mixing
4. pH/pCO₂: depends on process in O₂ rich conditions
in general higher pH → lower pCO₂ → higher O₂
5. ChlA → higher O₂; Turbidity → lower O₂; CDOM → lower O₂
Light → higher O₂

pCO₂ optode and O₂ optode at the depth of 12.6



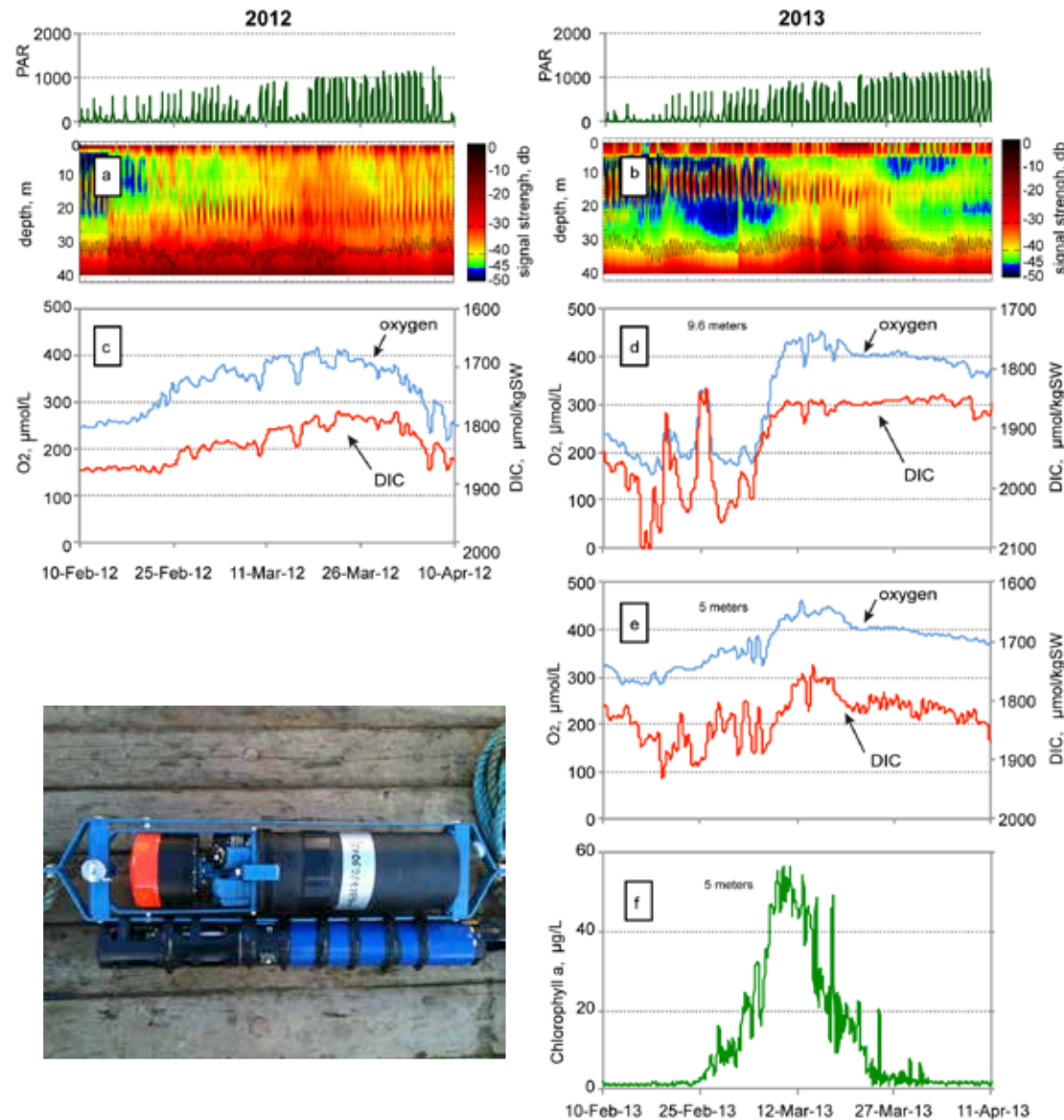
Salinity derived Alkalinity



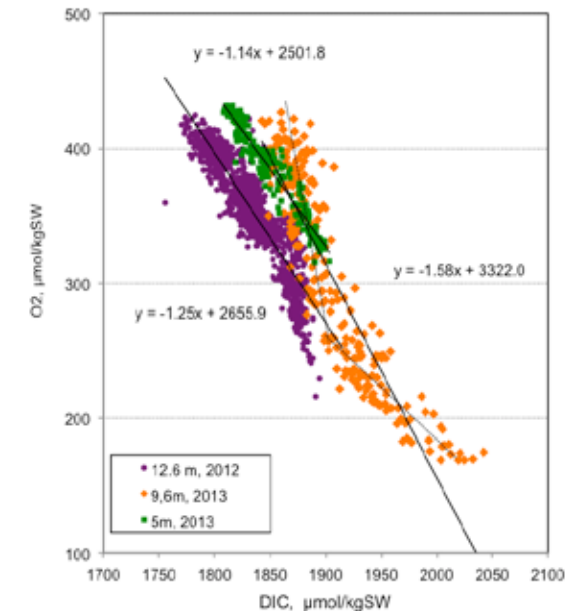
Net primary production (NPP) rates during spring bloom

- In 2012: 1.79 g C m⁻²
- In 2013: 2.10 g C m⁻²

Combining sensors gives increased understanding

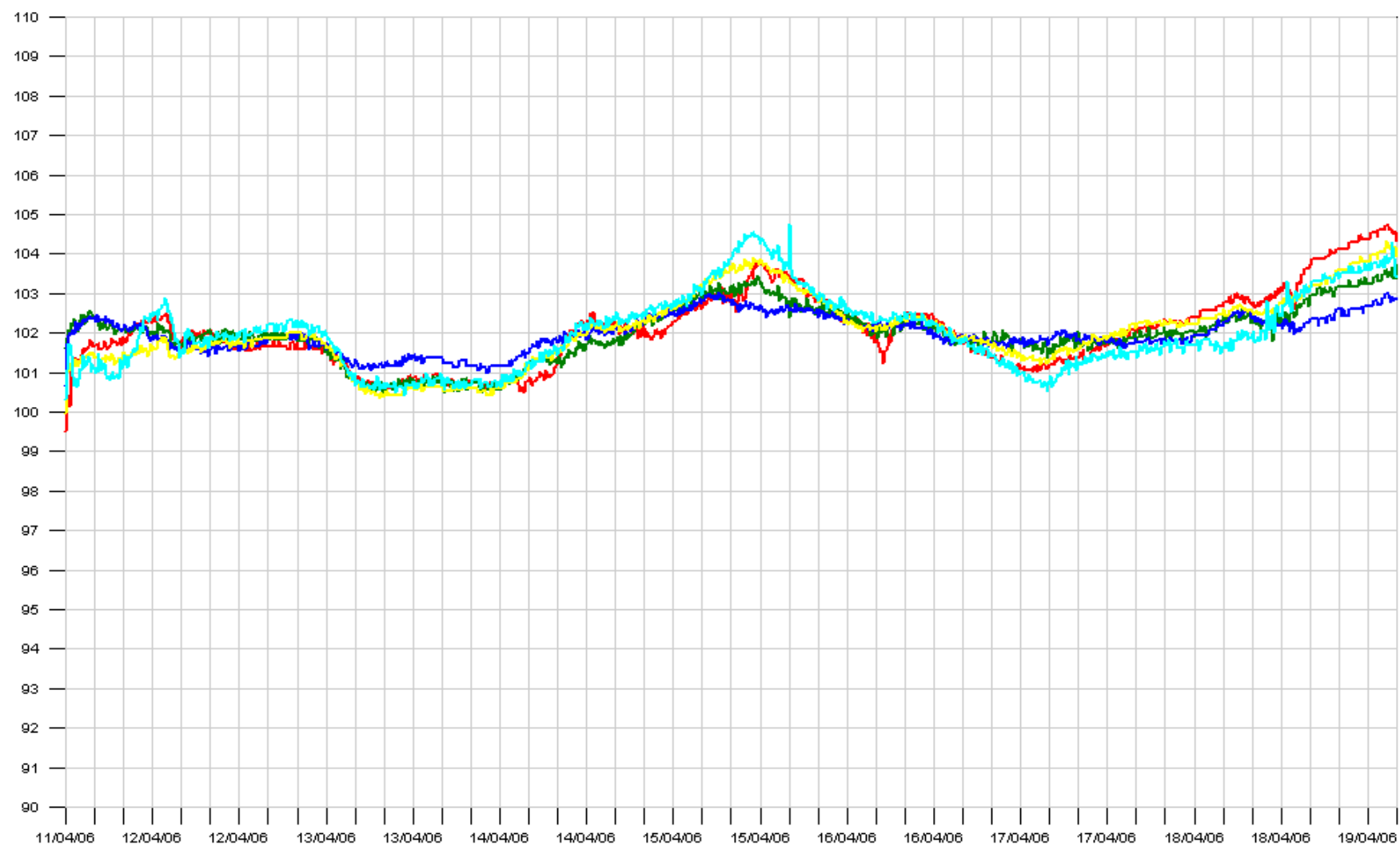


Redfield ratios -C:O₂



Atamanchuk, Kononets, Thomas, Hovdenes, Tengberg and Hall (2015)
Continuous long-term observations of the carbonate system dynamics in the water column of a temperate fjord.
Journal of Marine Systems 148, 272–284

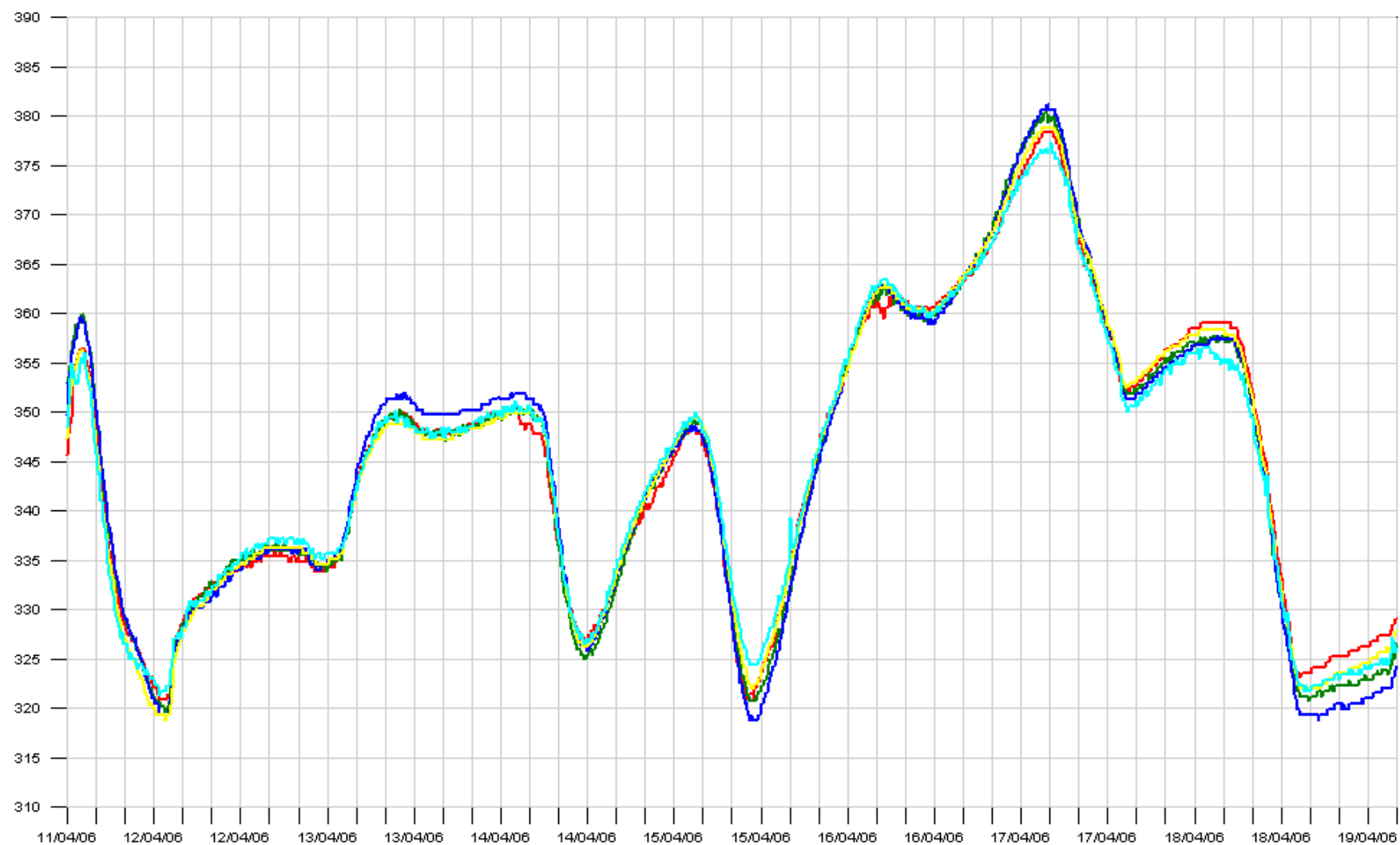
Optode postcheck in lab, airbubbled water (% saturation)



25.02 (above surf.) [% Sat] 26.02 (0.6 m) [% Sat] 27.02 (1.4 m) [% Sat] 28.02 (1.9 m) [% Sat] 29.02 (4.8 m) [% Sat]

Averaging: 1

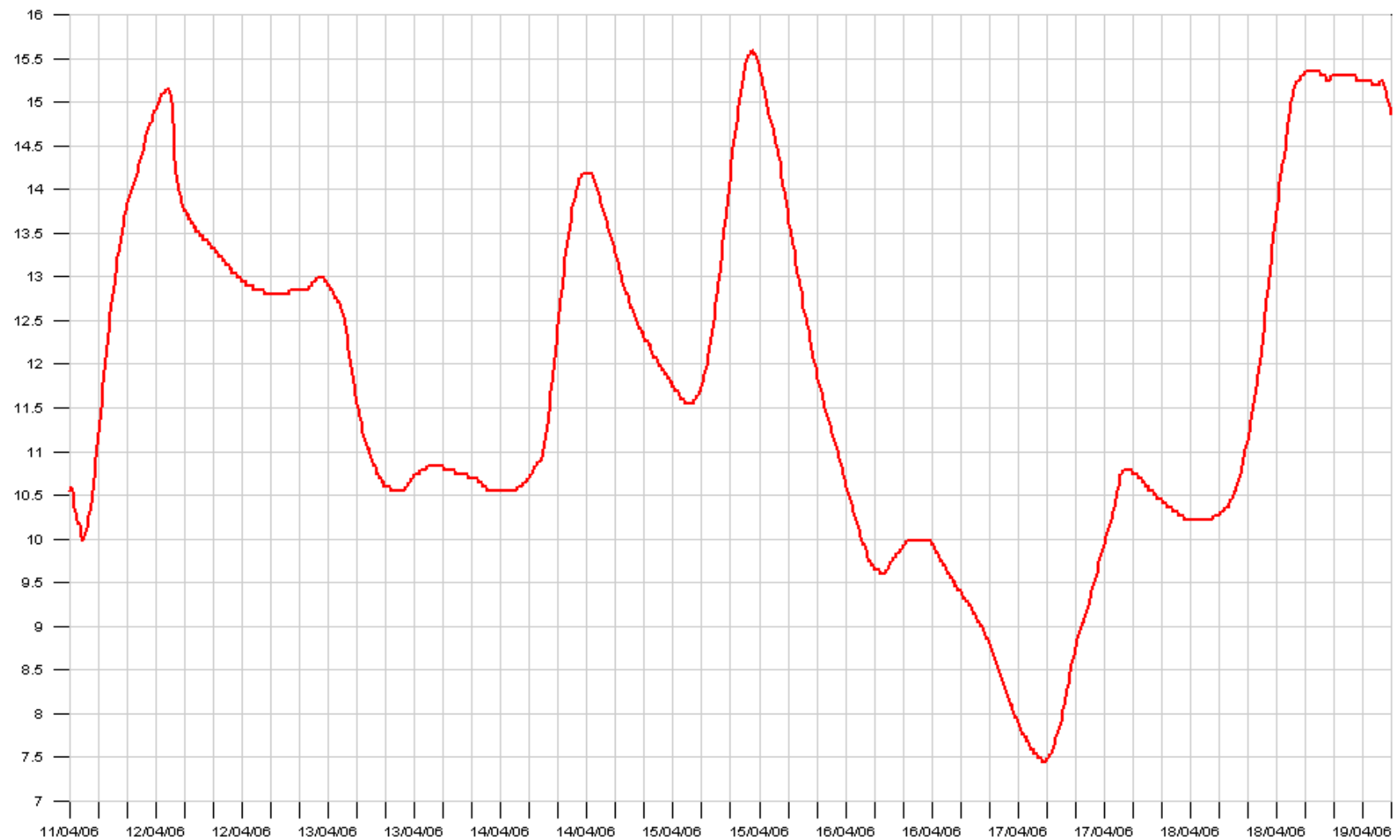
Optode postcheck in lab, airbubbled water



15. SalCompO2 (above surf.) [μM] 16. SalCompO2 (0.6 m) [μM] 17. SalCompO2 (1.4 m) [μM] 18. SalCompO2 (1.9 m) [μM] 19. SalCompO2 (4.8 m) [μM]

Averaging: 1

Optode post check in lab, airbubble water, Temperature



— 3. Temp. (0.8m) [deg.C]

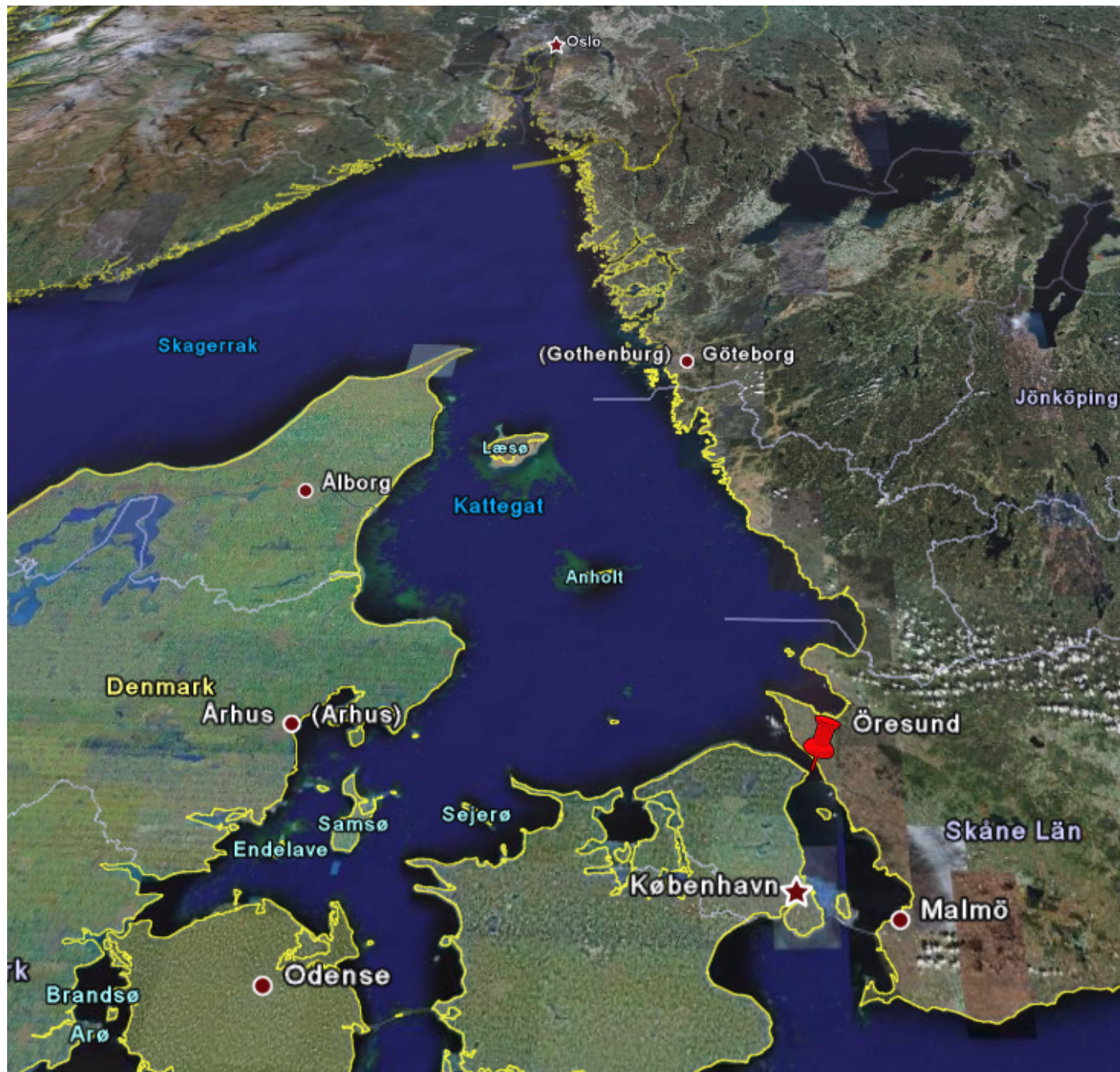
Averaging: 1

What affects O₂ concentration?

1. Salinity and Temperature (Garcia and Gordon, 1992, L&O)
2. Primary production & bubbles, surface waters ~105%
3. Decomposition of organic matter
4. Chemical oxidation, mainly at the bottom
5. Air pressure, at surface
6. Hydrothermal vents, lower O₂

All sensors (electrochemical & optical) measure partial pressure (PO₂)

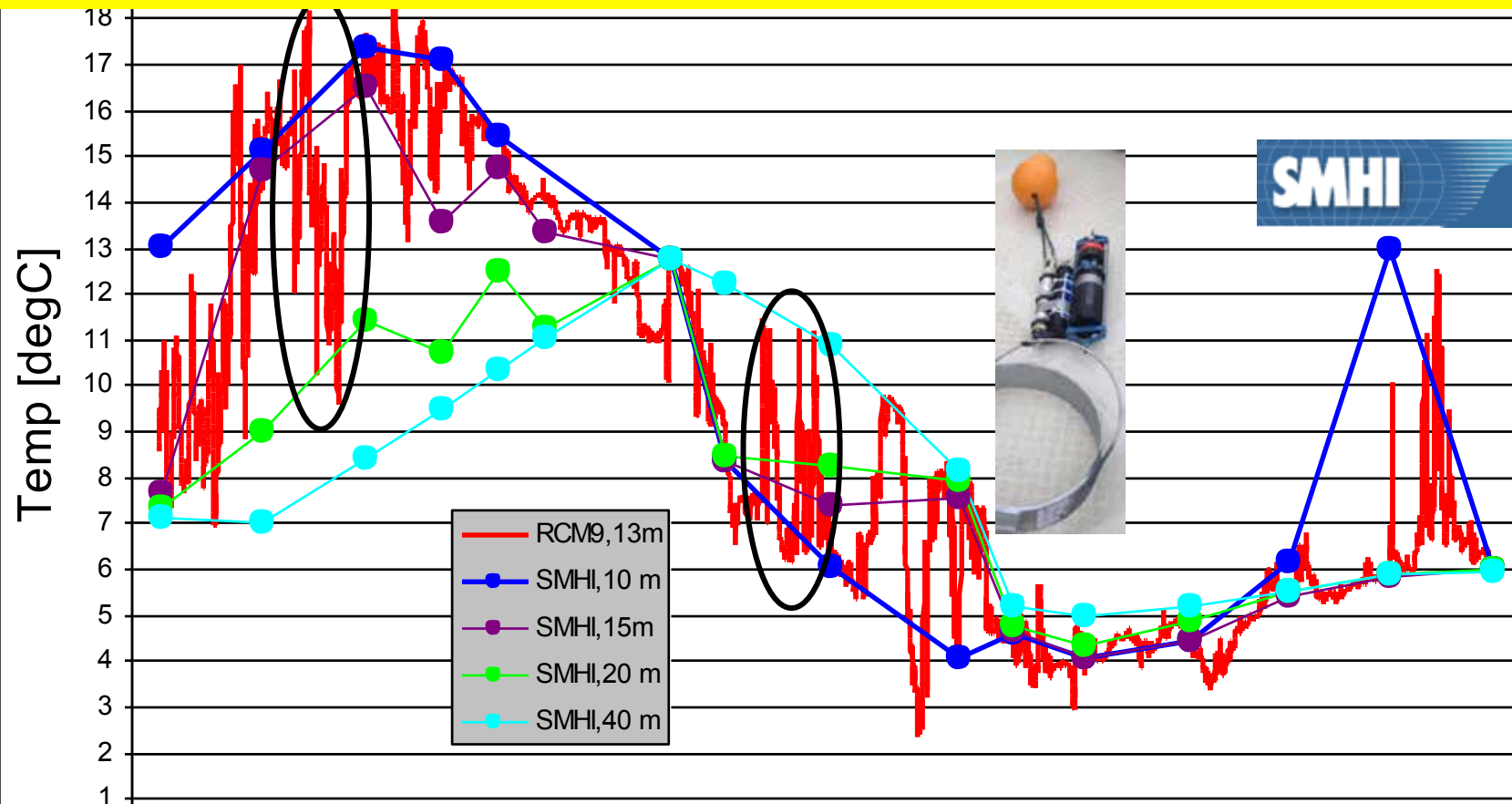
Outside the photic zone there is now known process that produce O₂



Natural variations, are we measuring at the right frequencies?

Temperature, June 12, 2007 to June 10 2008

Nyquist criteria → To understand changes Measurements should be done at twice the frequency that they occur

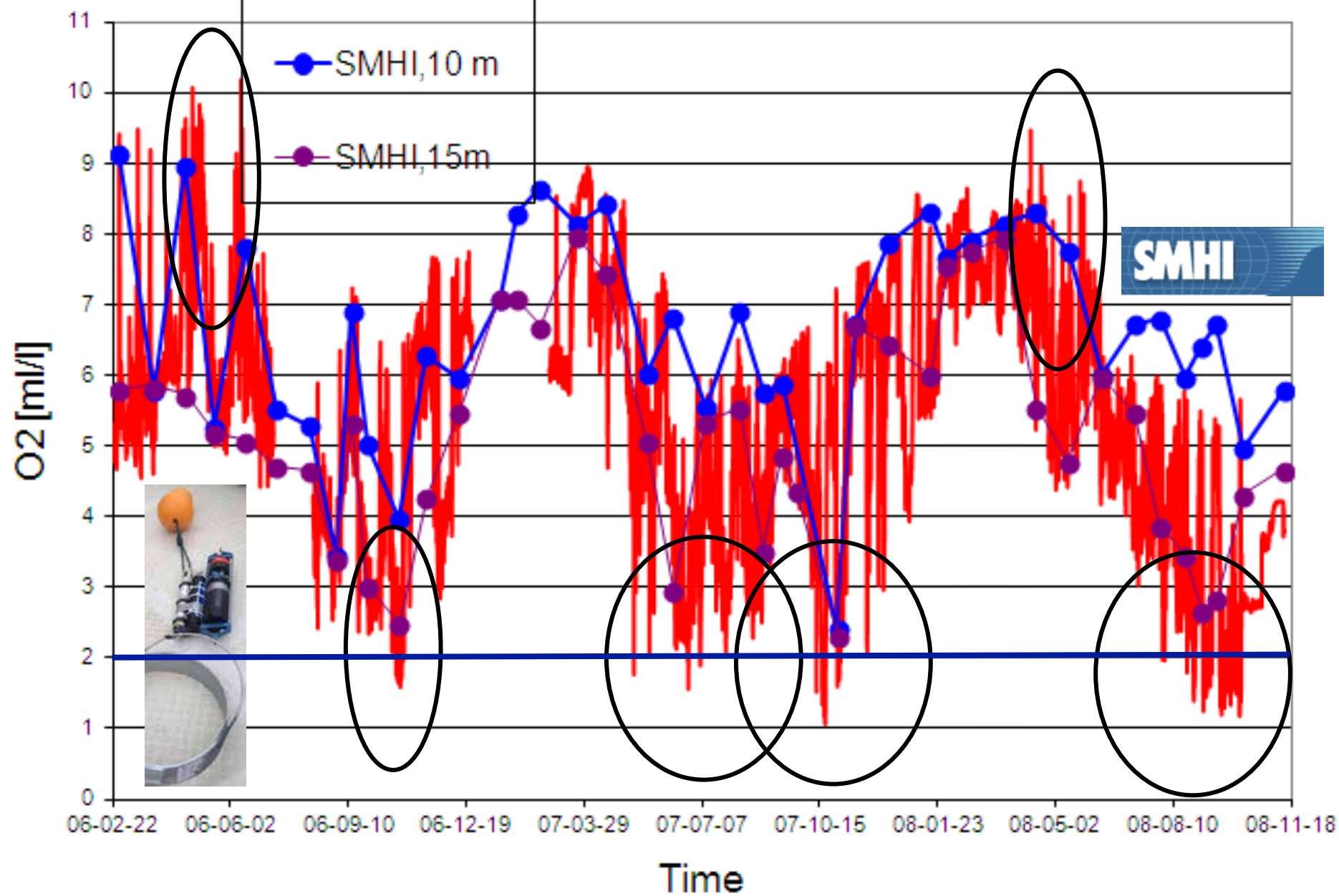


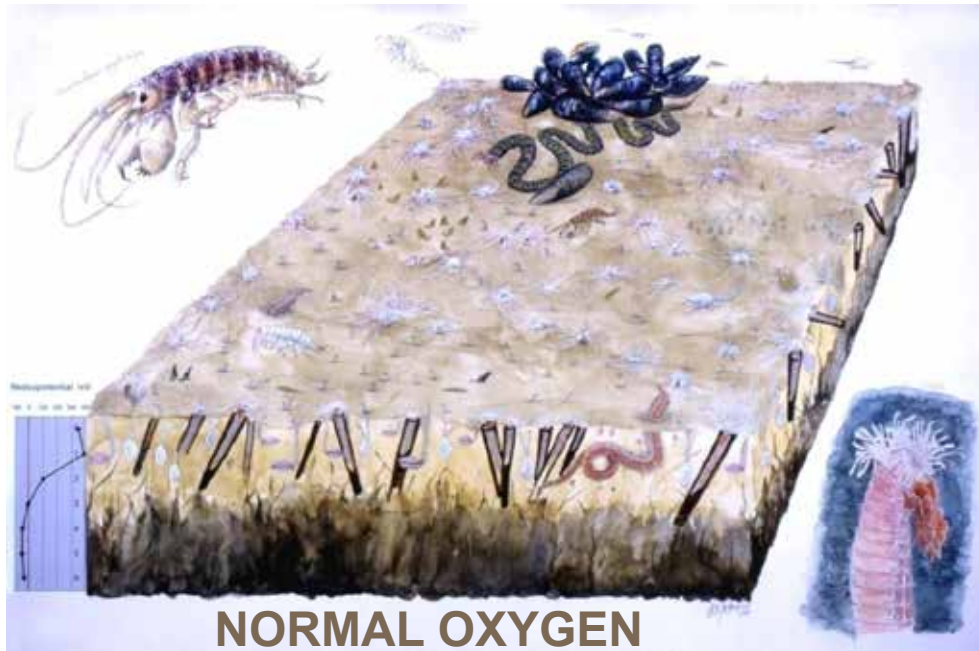
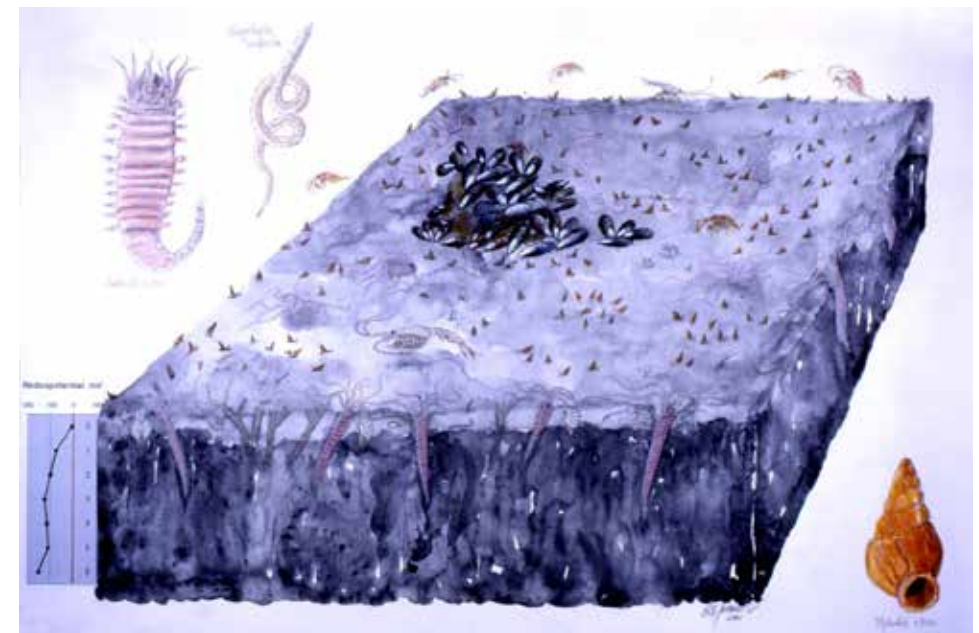
With weekly sampling maximal difference from one week to another was 9°C

The average difference was 1.5°C.

Estimated that global warming increase temp with about 0.06°C/year

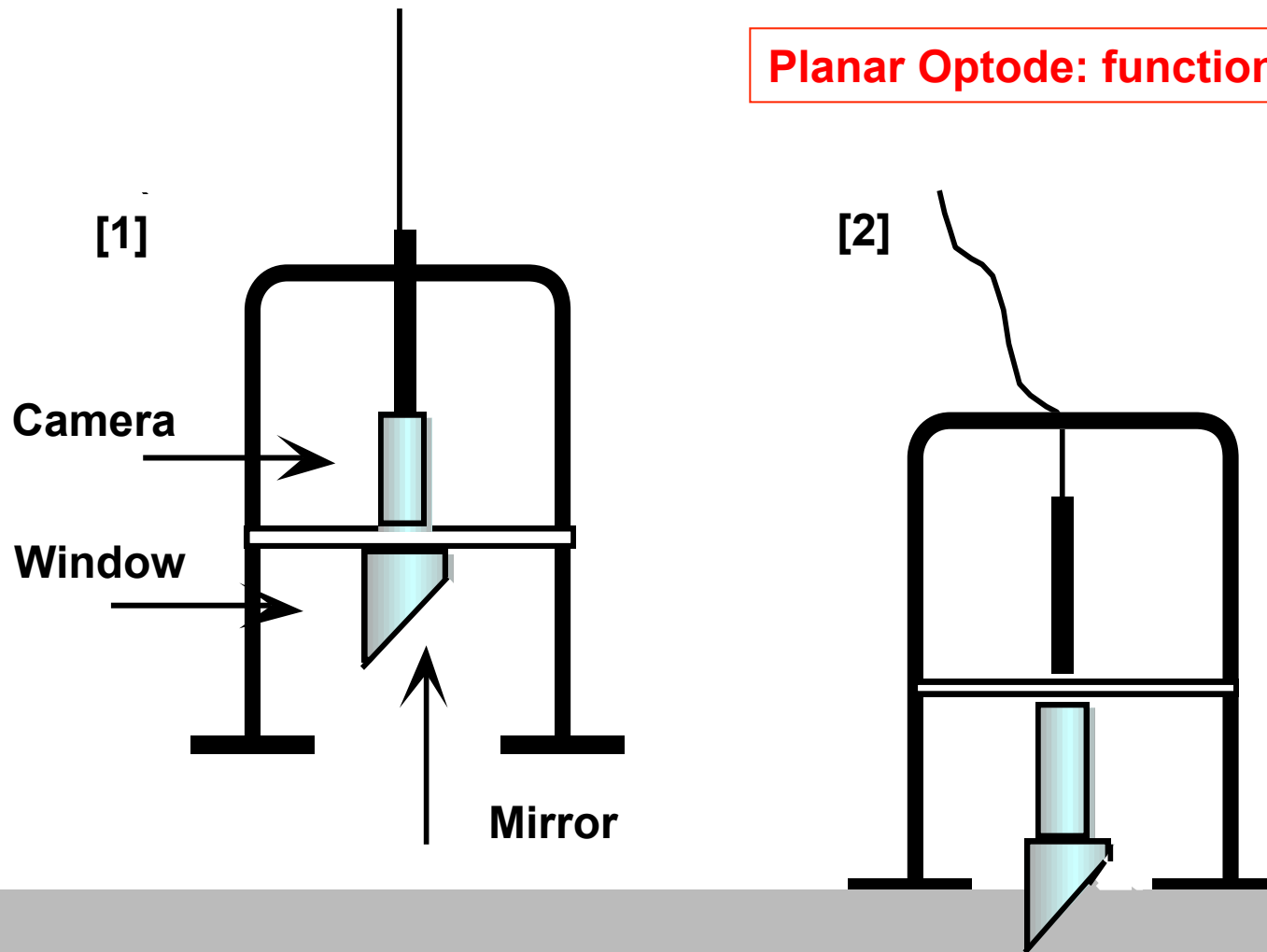
Oxygen





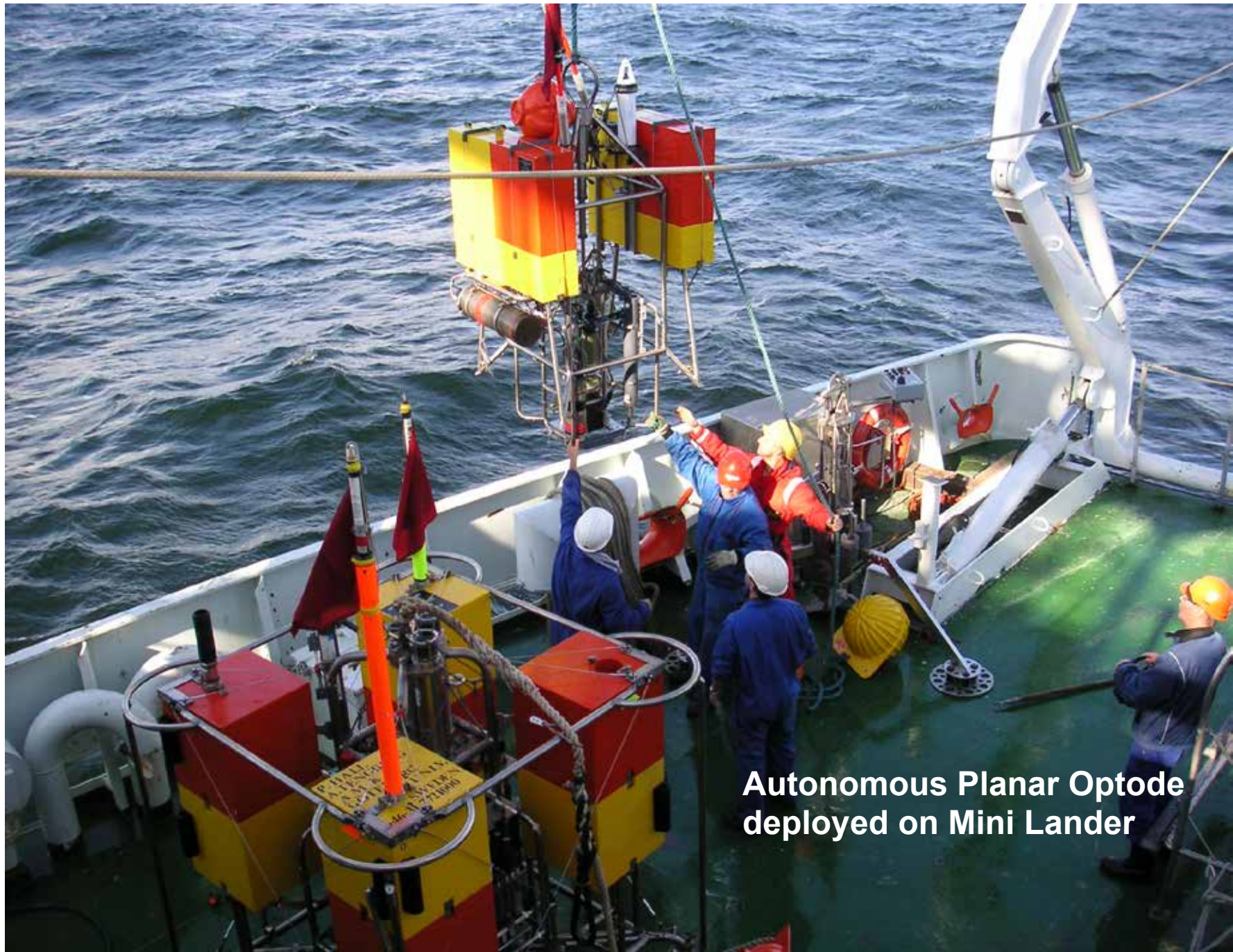
AFTER ANOXIA

Planar Optode: functioning

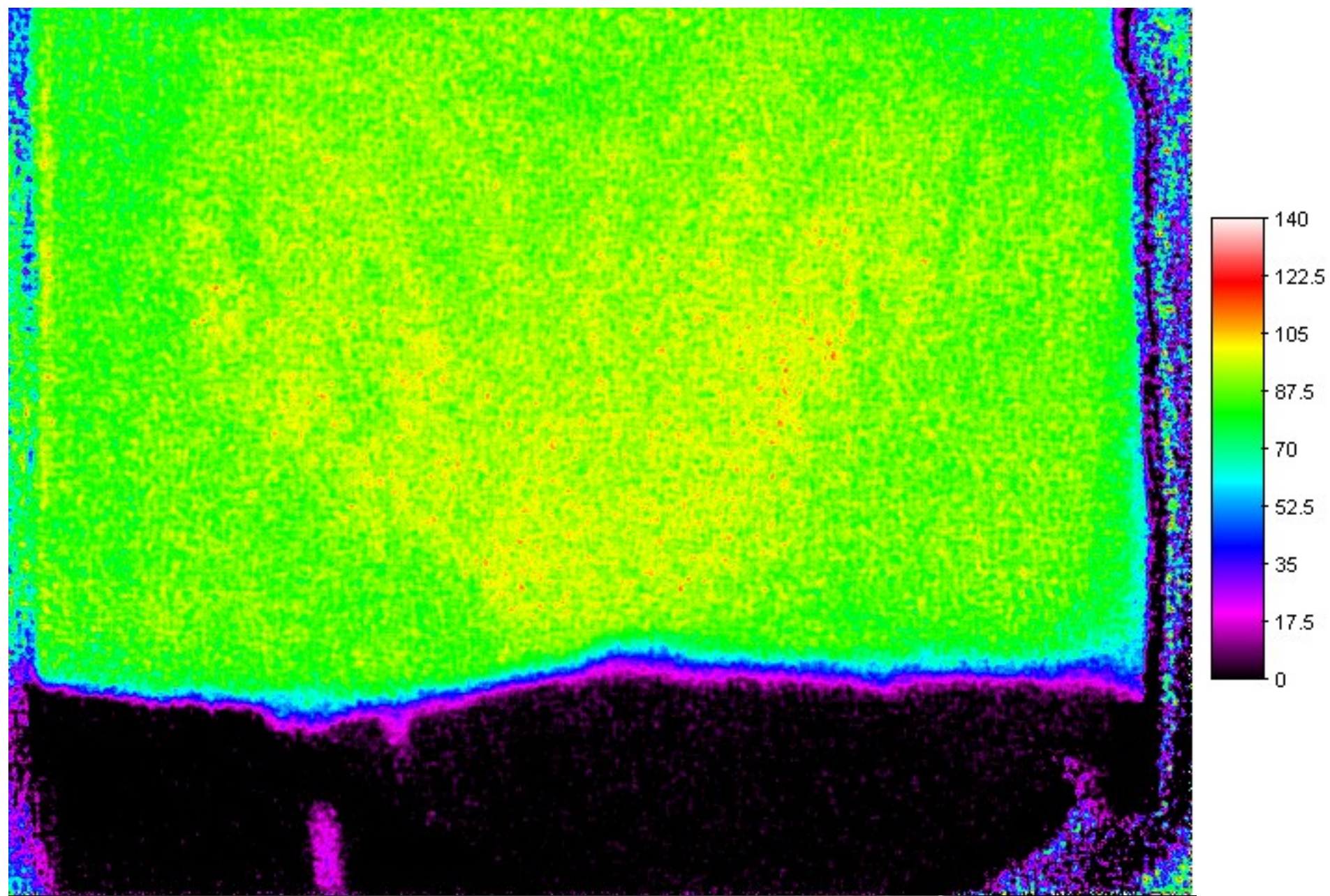


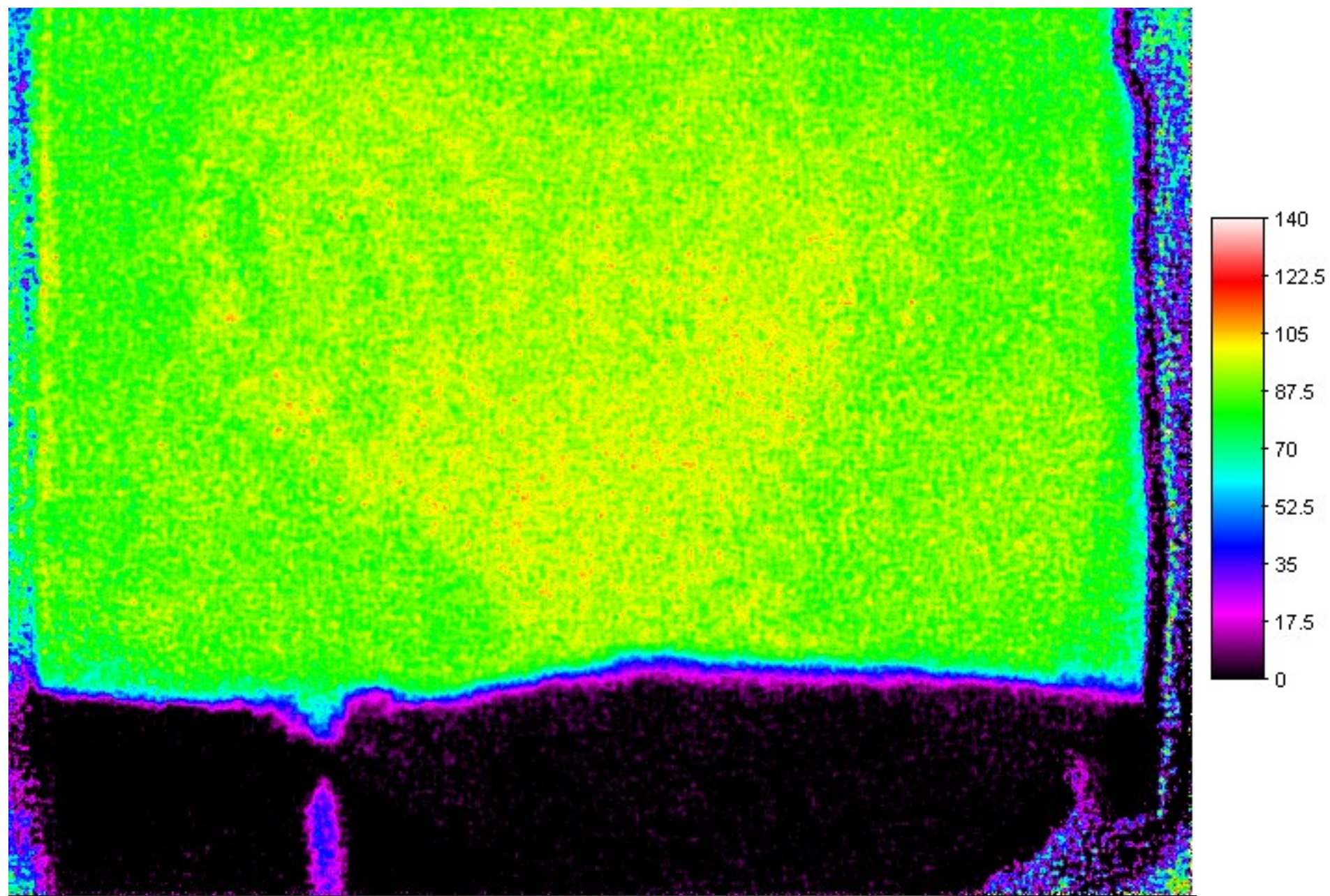
Sediment

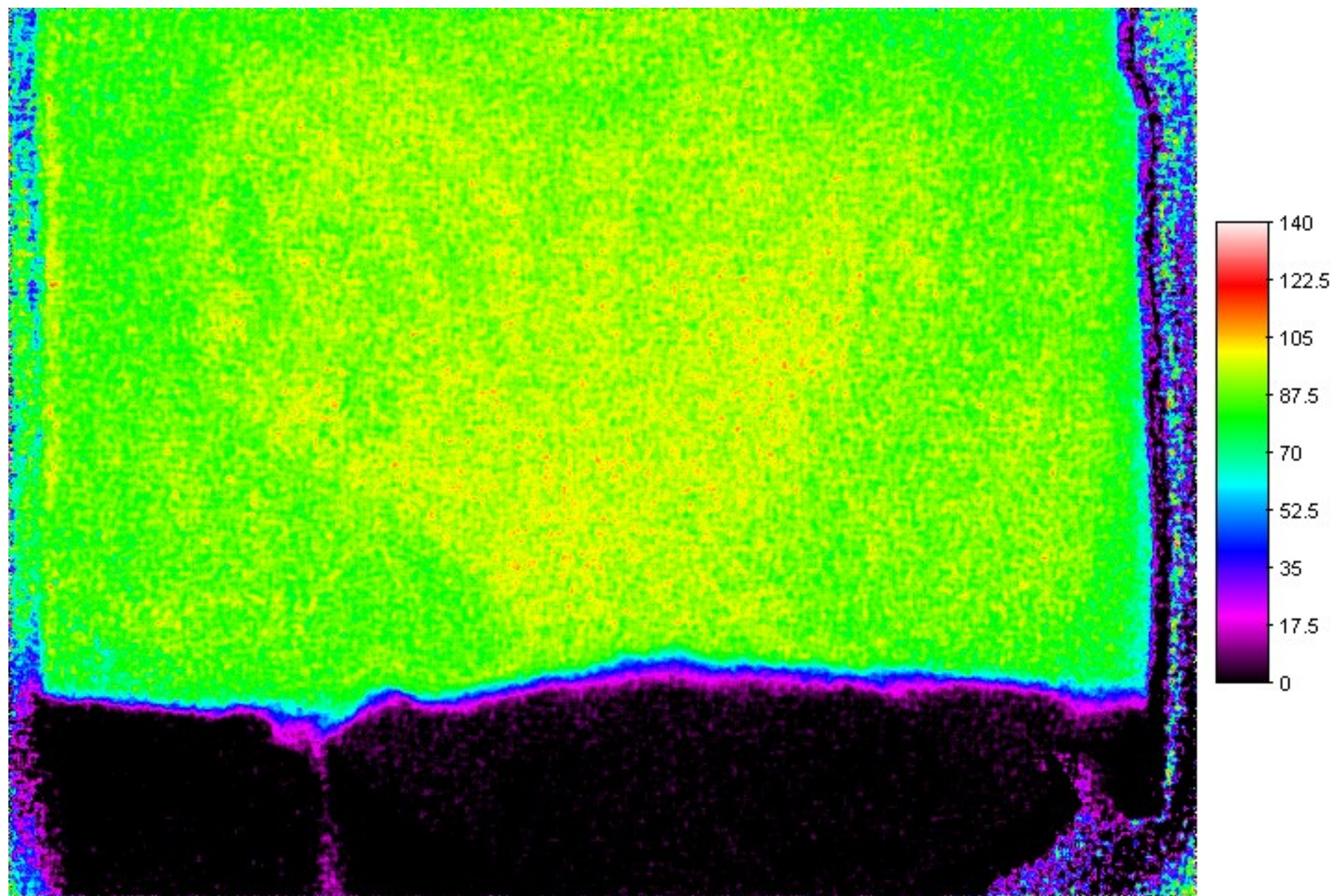
GLUD R.N, A. TENGBERG, M. KÜHL, P. HALL, I. KLIMANT and G. HOLST (2001) An in situ instrument for planar O₂ optode measurements at benthic interfaces. *Limnology and Oceanography*, 46(8): 2073-2080

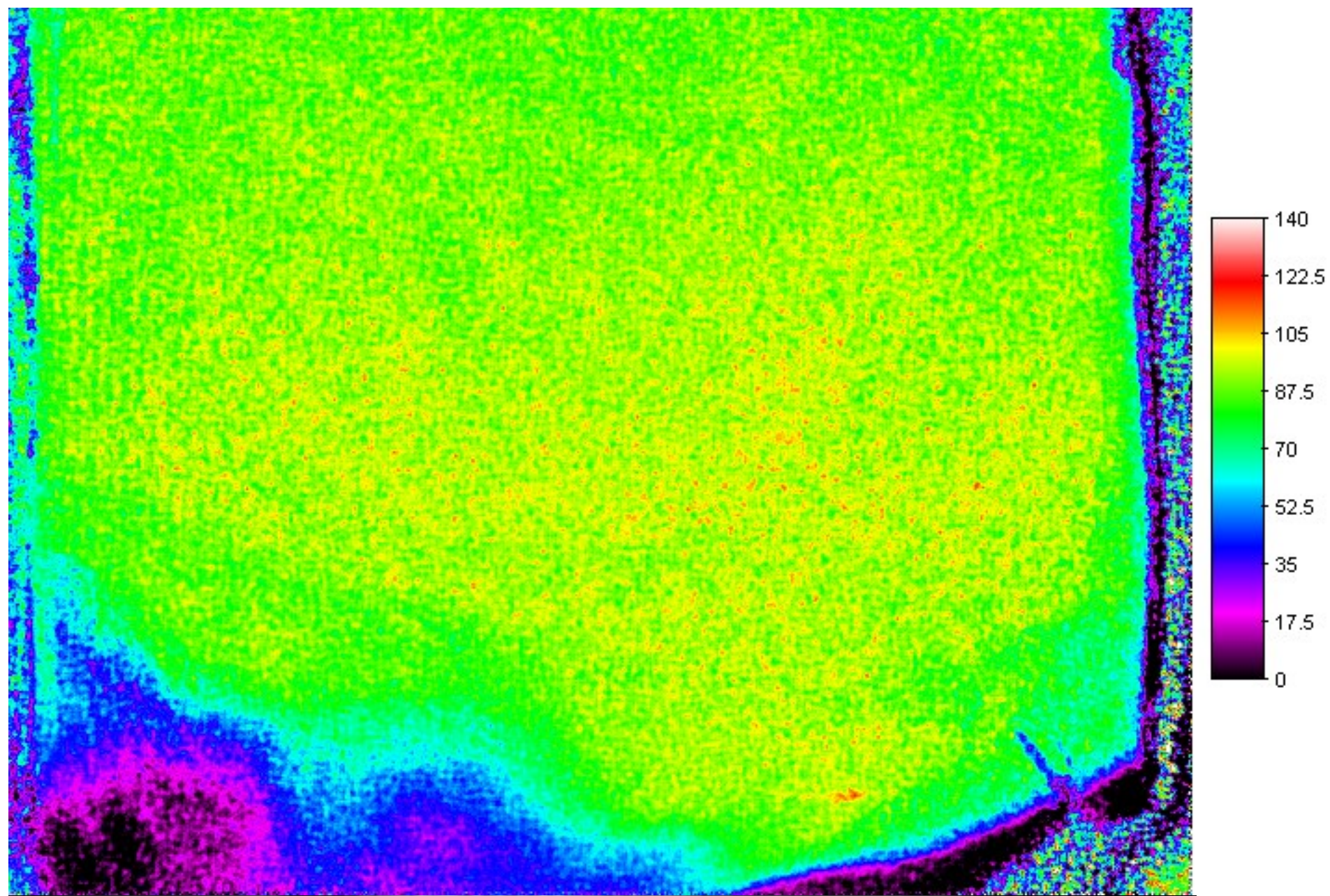


**Autonomous Planar Optode
deployed on Mini Lander**









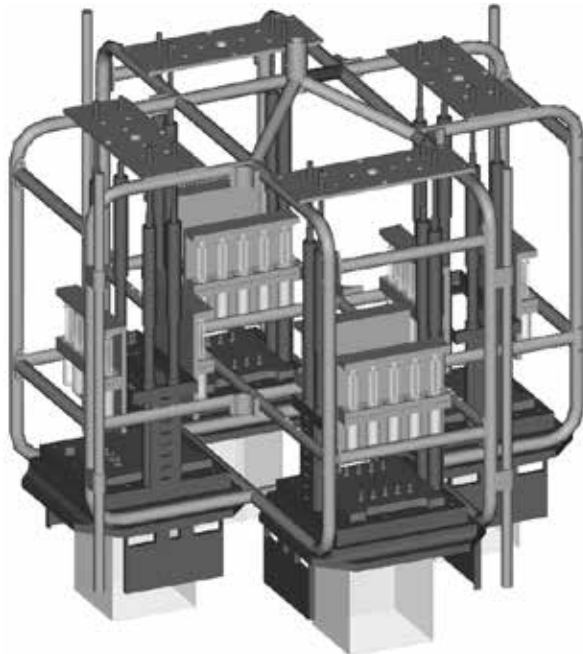
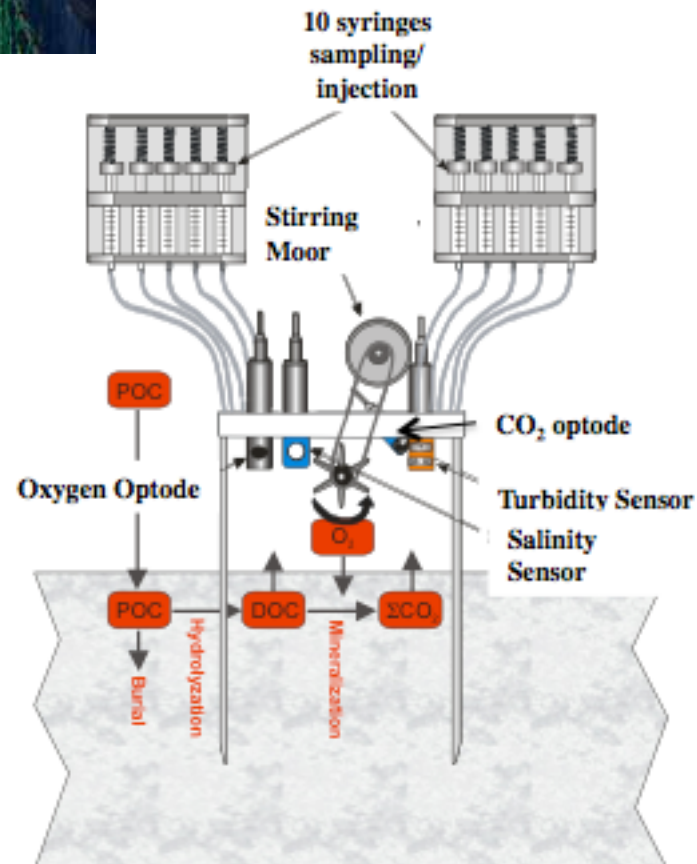


Tools: Autonomous Benthic landers

Our landers:

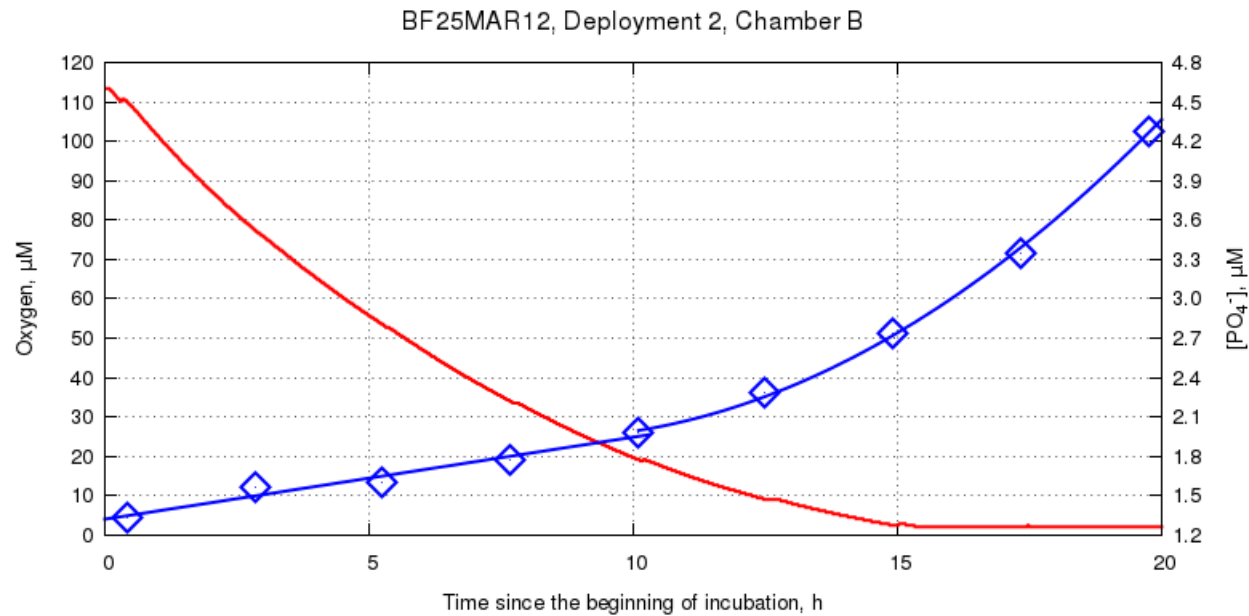
- 250-300 deployments
- About 800 incubations
- Deployed at 5-5600 m
- 40 water samples
- 15-25 sensors
- 95 % success rate

Inner frame
(4 incubations chamber modules)



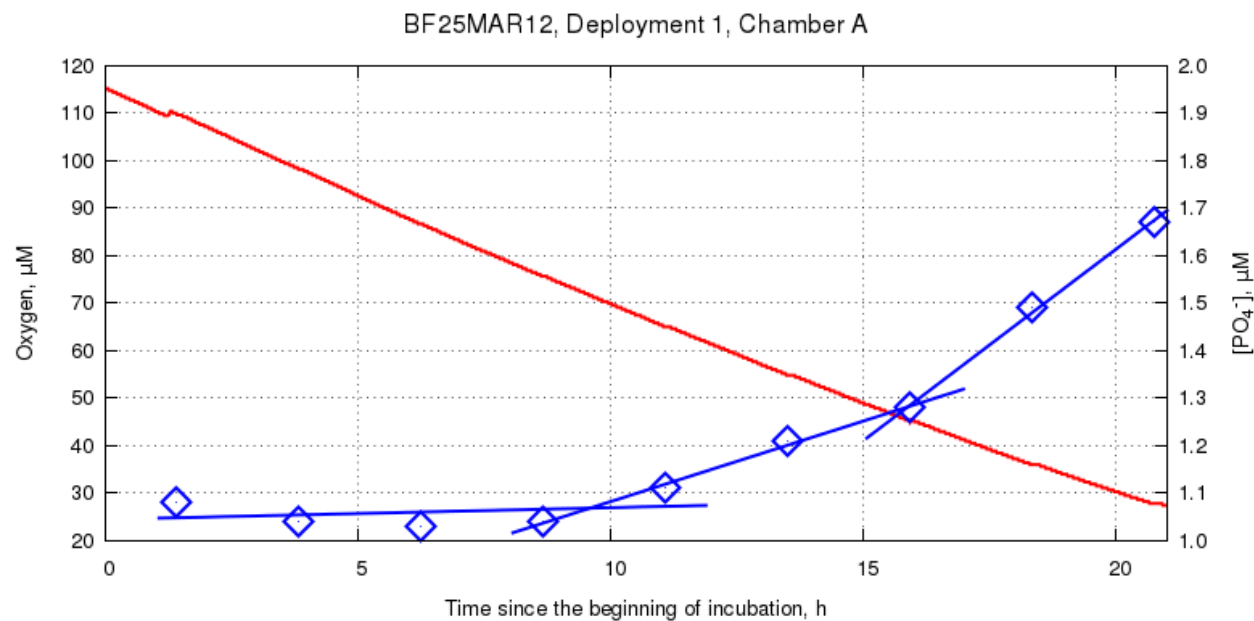
Measured Fluxes

- **Oxygen**
- Total Carbonate
- Silicate
- **Phosphate**
- Nitrate+Nitrite
- Ammonium
- DOC
- Alkalinity
- pH
- Fe and Mn
- Turbidity
- pCO₂



Red: Oxygen conc.
Blue: PO_4 conc.
in benthic chambers

- For phosphate fluxes: Oxygen concentration more important than resuspension



Ways to measure Oxygen in Water

1. Chemical: Winkler titration (Winkler, 1888)
Accuracy: about 2%; Precision: about 0.5 %

**Reference
method**

2. Electro-chemical: By sensors (Clark, 1959)
**Accuracy: absolute measurements difficult,
needs frequent calibration.**
Precision: about 0.5 %

3A Optical: By sensors (Klimant et al., 1995)
**Intensity based: Accuracy: absolute
measurements difficult, needs frequent calibration.**
Precision: about 0.5 %

3B. Optical: By sensors (Holst et al., 1995)
Lifetime based: Accuracy: about 1 %;
Precision: about 0.5 %

Summary

9:00-10:30

- Why should you measure Oxygen (O_2)?

Crucial parameter

- How is O_2 related to other parameters?

Related to most other parameters

- Are we measuring at the right frequencies?

Nyquist criteria: sample at twice the frequency as changes occur

- Methods to analyze and measure O_2

Winkler, Electrochemical sensors, Optical foil based sensors

11:00-13:00

- What affects O_2 sensors/methods to evaluate performance?

- How good is Winkler titration?

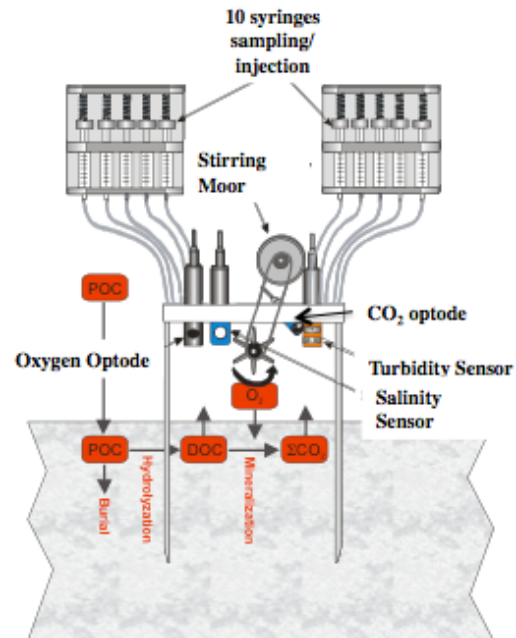
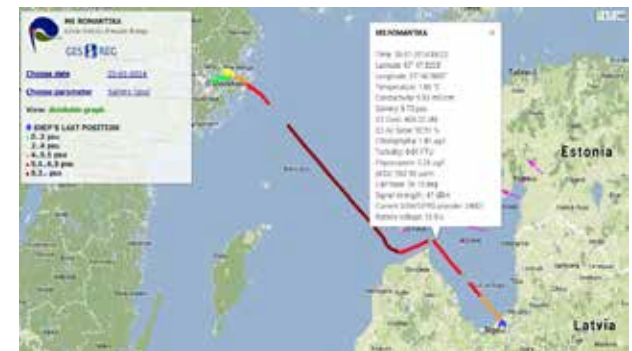
- Field data, value of combined measurements

- Summary

- Practical Demonstration and Hands-On

Oxygen measurements: how?

Anders Tengberg (anderste@chem.gu.se) Associate Professor at the University of Gothenburg and at the Chalmers University of Technology, Sweden, Scientific Advisor/Product Manager at Aanderaa since 1997.



Ways to measure Oxygen in Water

1. Chemical: Winkler titration (Winkler, 1888)
Accuracy: about 2%; Precision: about 0.5 %

Reference
method

2. Electro-chemical: By sensors (Clark,)
Accuracy: absolute

Precision: about 0.5 %



Careful with manufacturer specifications! Do they give field or lab specifications? Is response time for air or water? Is accuracy for the hole range of O₂ Conc and Temp? Can they show data to sustain claimed accuracy/precision and response time? What about publications from independent sources?

Precision: about 0.5 %



BERNTSSON, TENGBERG, HALL & JOSEFSSON (1997) Multivariate experimental methodology applied to the calibration of a Clark type oxygen sensor. *Analytica and Chimica Acta*, 355: 43-53

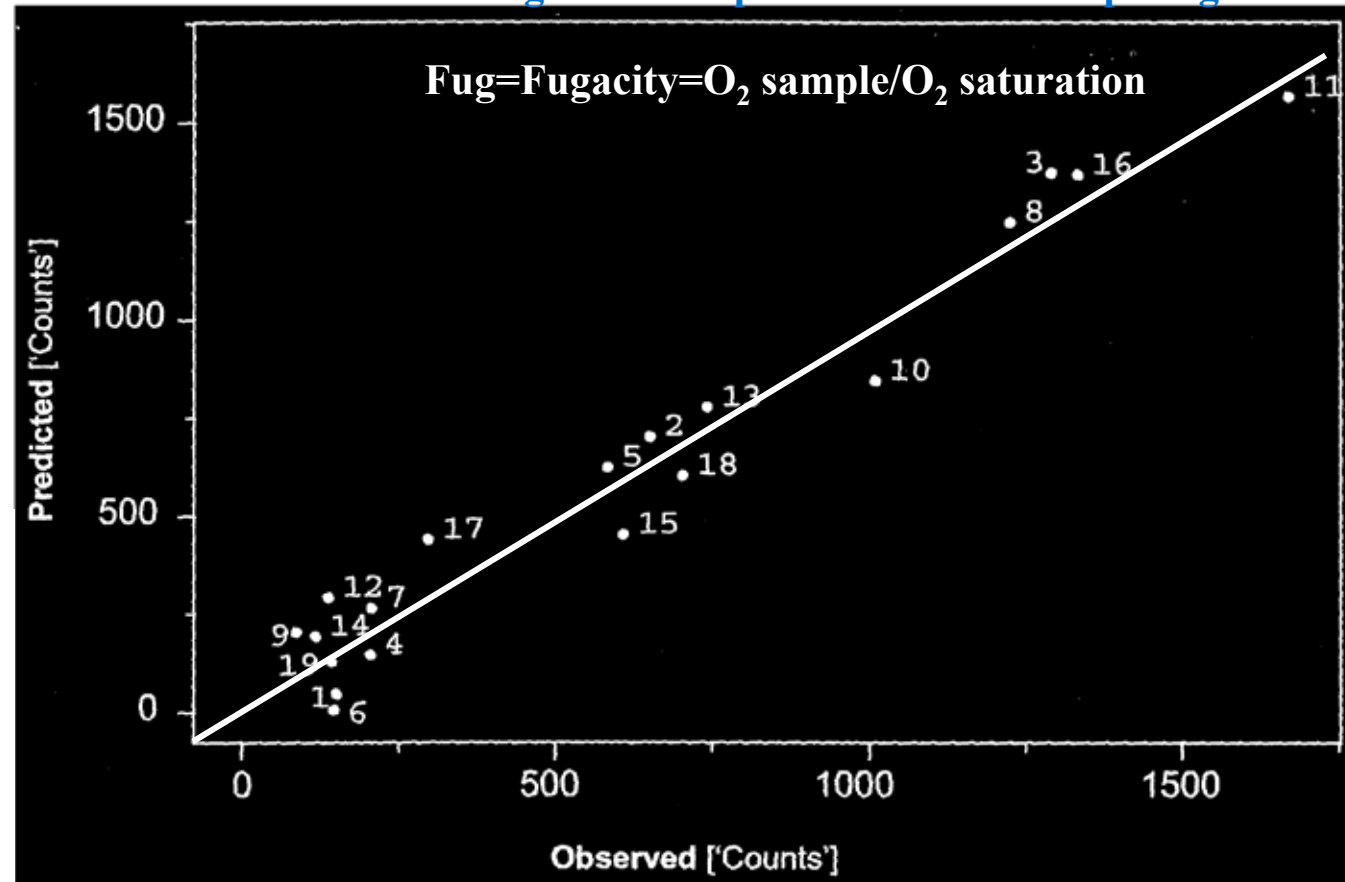
Cross-term



$$\text{Counts} = -142 + \text{Fug} * 716 + \text{Temp} * 7.8 + \text{Stir} * 1.9 + \text{Temp} * \text{Fug} * 27.8$$

Which Parameters will influence the response of an Electro-chemical sensor?

- **O₂: Yes**
- **pH: No Effect**
- **Sal: Yes, cross-term**
- **Temp: Yes, cross-term**
- **Pres: Yes, not fully reversible**
- **Stirr: Yes**
- **Aging=drift: Yes, electrolyte & membrane decompose**

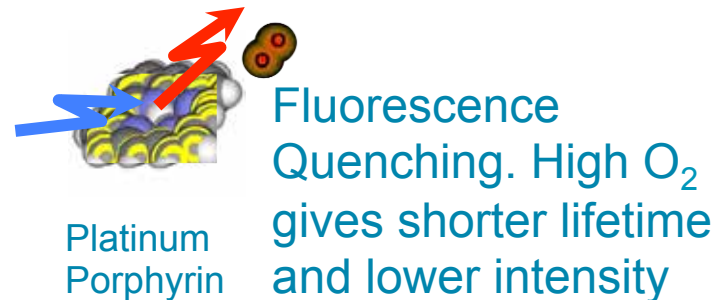


Every sensor individual

Optode Measuring Principle Based on Fluorescence Quenching Detection of lifetime

Note: Oxygen Optodes are measuring partial pressure of oxygen.

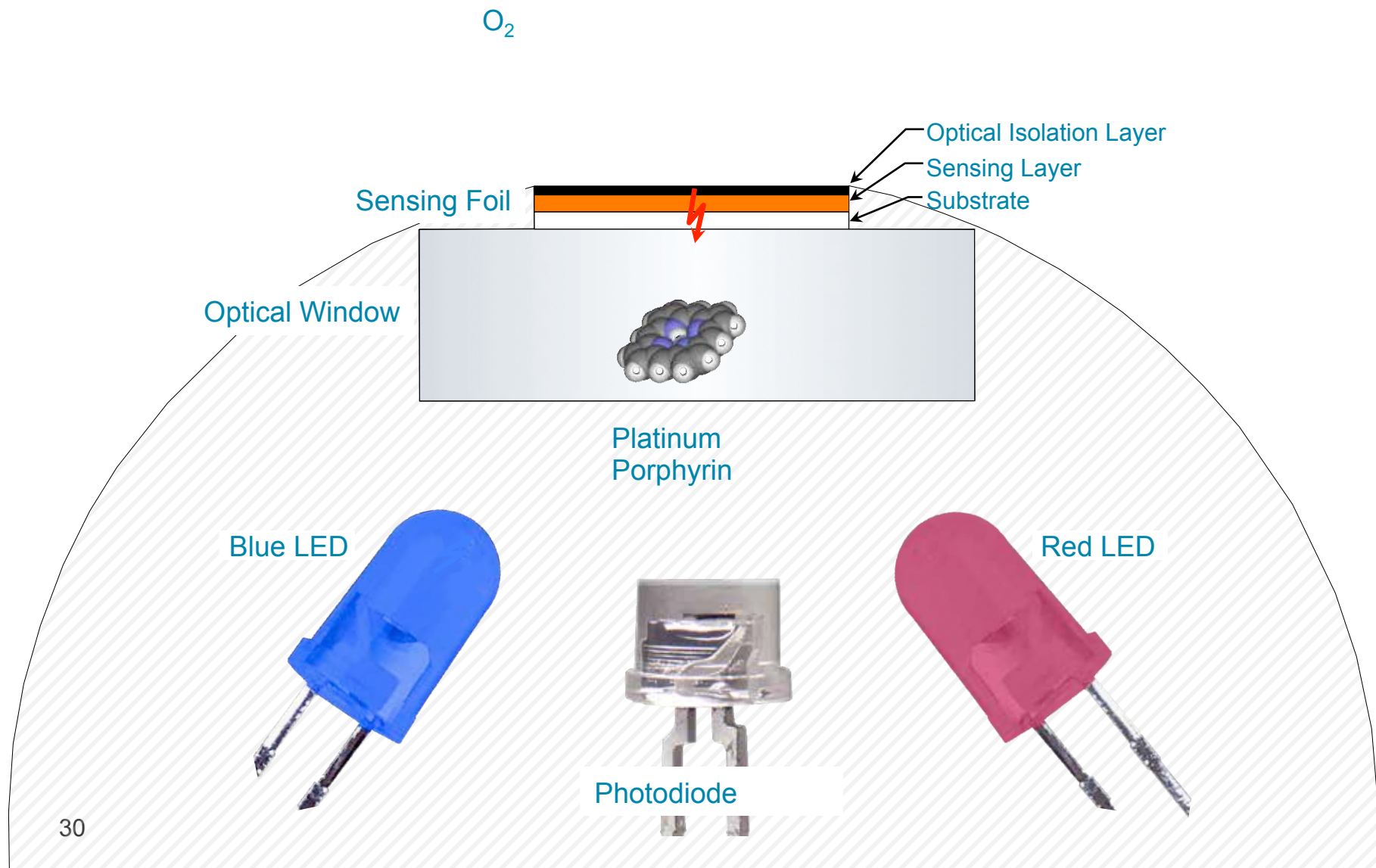
- 100 % in sea water at 10°C and 35 ppt corresponds to $282 \mu\text{M} = 9.0 \text{ mg/l} = 6.3 \text{ ml/l}$
 - 100 % in fresh water at 10°C corresponds to $353 \mu\text{M} = 11.3 \text{ mg/l} = 7.9 \text{ ml/l}$
- **If salinity variable it should be measured and compensated for**



Literature:

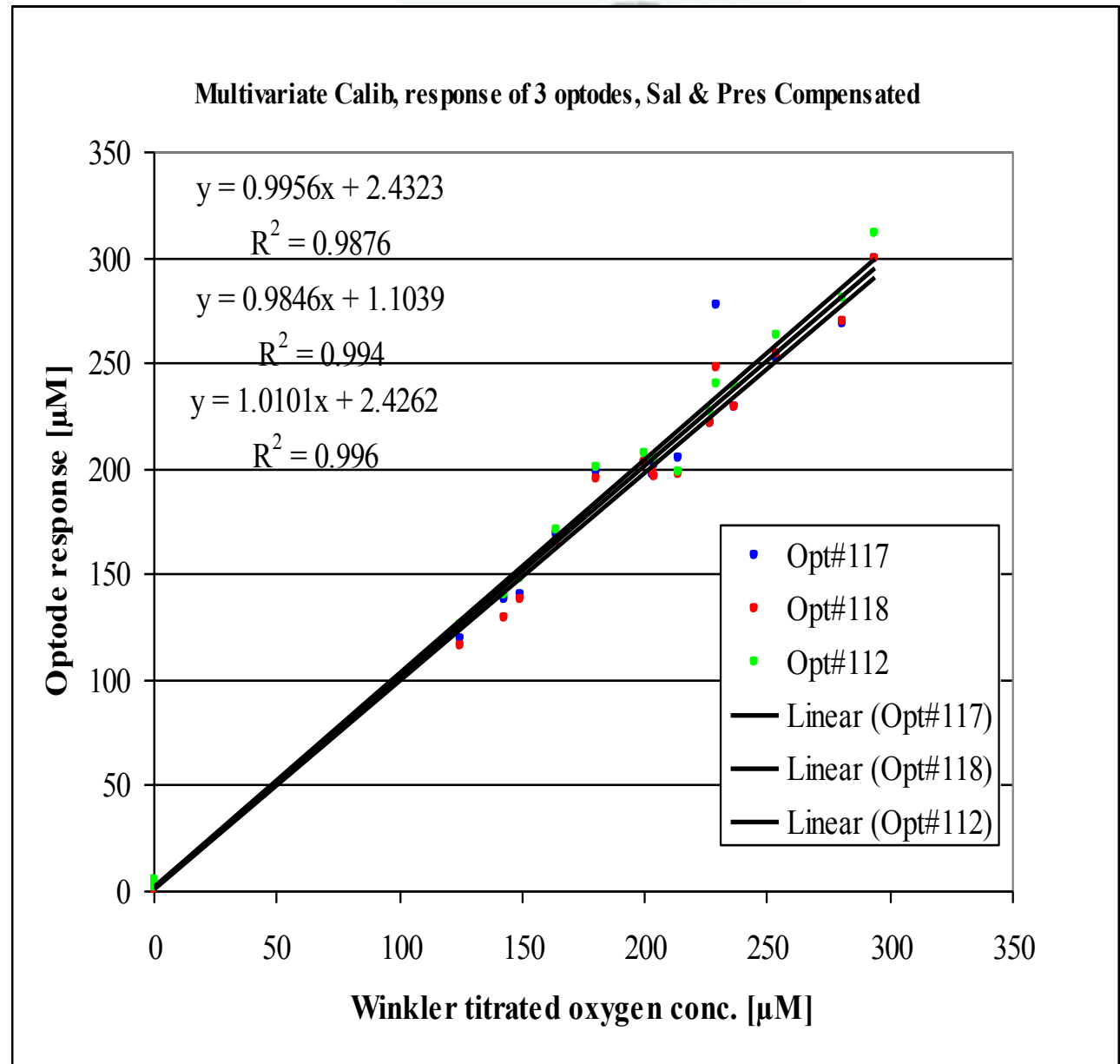
- Dynamic Fluorescence Quenching. Kautsky, 1939
- Klimant I., V. Meyer, and M. Kohls. 1995. Fibre-optic oxygen microsensors, a new tool in aquatic biology. Limnology and Oceanography 40: 1159-1165.
- Tengberg A., J. Hovdenes, J. H. Andersson, O. Brocandel, R. Diaz, D. Hebert, T. Arnerich, C. Huber, A. Körtzinger, A. Khripounoff, F. Rey, C. Rönning, S. Sommer and A. Stangelmayer (2006). Evaluation of a lifetime based optode to measure oxygen in aquatic systems. Limnology and Oceanography, Methods, 4, 7-17.

Optics

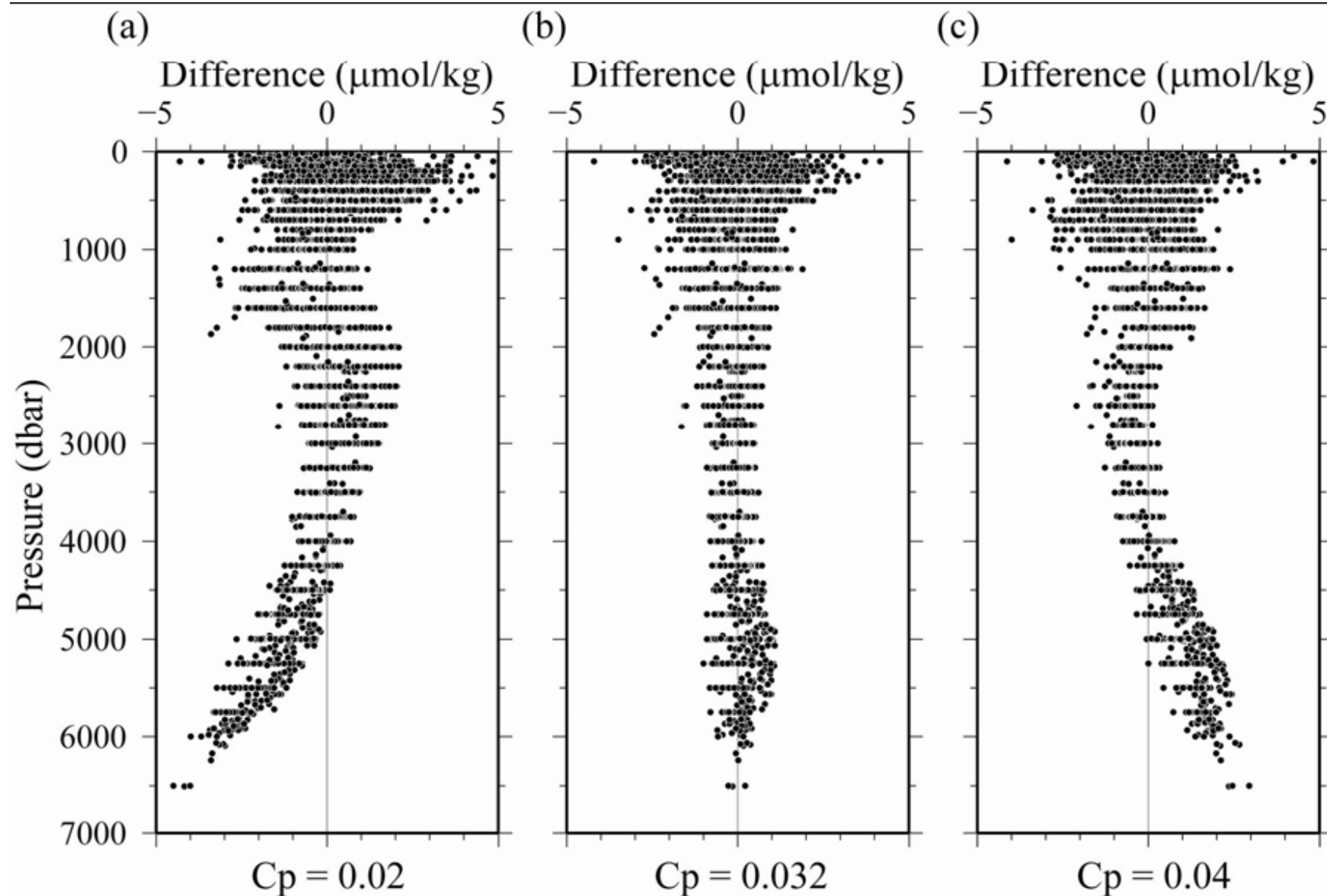


Which Parameters will influence the response of an Oxygen Optode?

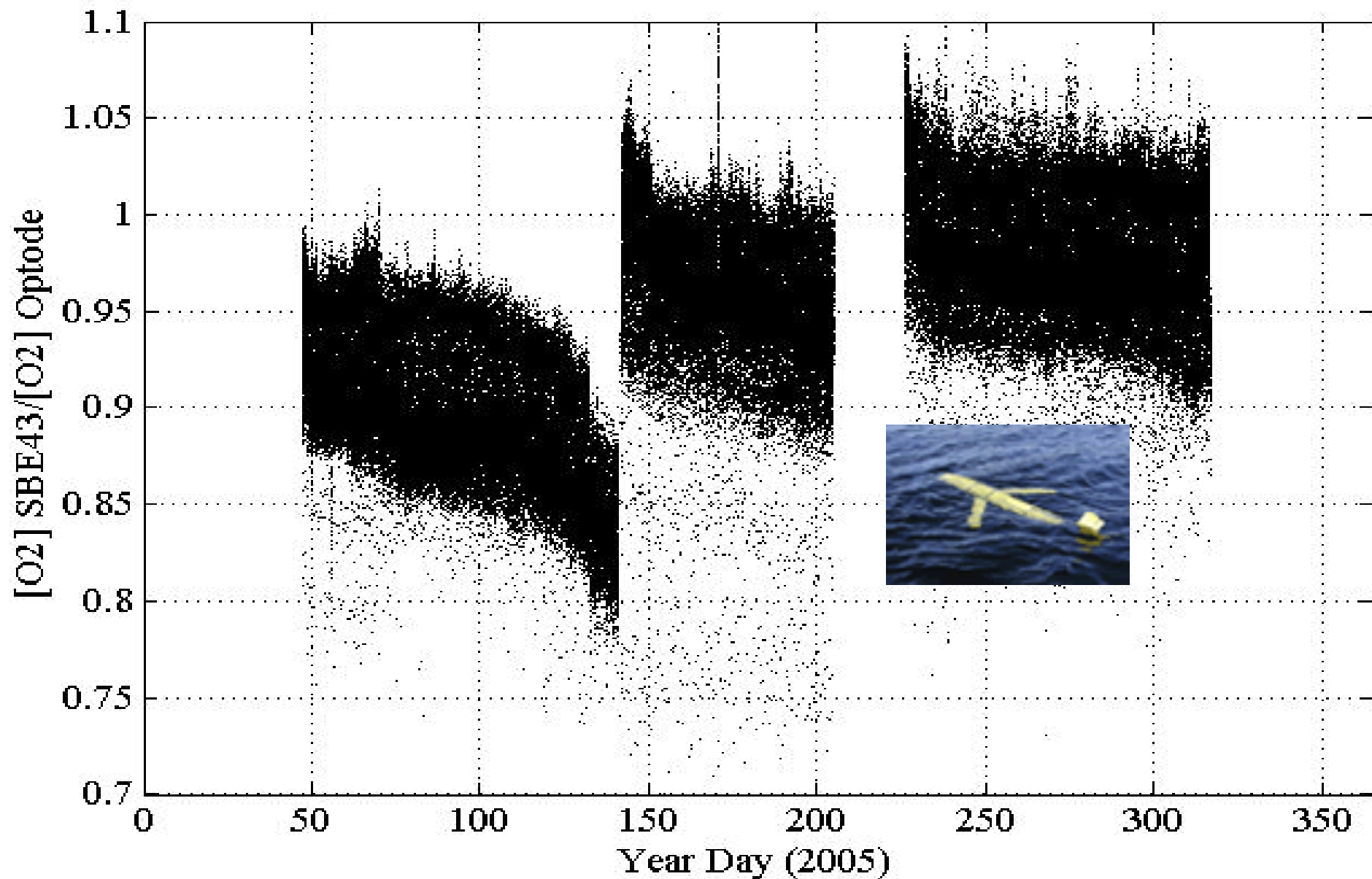
- **O₂: Yes**
- **pH: No Effect**
- **Sal: Yes, simple formula**
- **Temp: No Effect**
- **Pres: Yes, about 4% per 1000 m, fully reversible**
- **Stirr: No Effect**
- **Aging=drift: No Effect**



Tengberg, Hovdenes, Andersson, Brocandel, Diaz, Hebert, Arnerich, Huber, Körtzinger, Khripounoff, Rey, Rönning, Sommer & Stangelmayer (2006). Evaluation of a life time based optode to measure oxygen in aquatic systems. *Limnology and Oceanography, Methods*, 4, 7-17.



Uchida H., T. Kawano , I. Kaneko and M. Fukasawa (2008) In-situ calibration of optode-based oxygen sensors. *Journal of Atmospheric and Oceanic Technology*, 25: 2271-2281.



Nicholson D., S. Emerson and C. C. Eriksen (2008) Net community production in the deep euphotic zone of the subtropical North Pacific gyre from glider surveys. *Limnol. Oceanogr.*, 53(5, part 2), 2226–2236.

Long term data from 69 Optodes on Argo floats

Yuichiro Takeshita et al., Poster presented at AGU fall meeting, San Francisco, Dec 2010.
Now published in: Takeshita et al. (2013) A climatology-based quality control procedure for profiling float oxygen data. J. Geophysical Res, Vol. 118, 1–11.

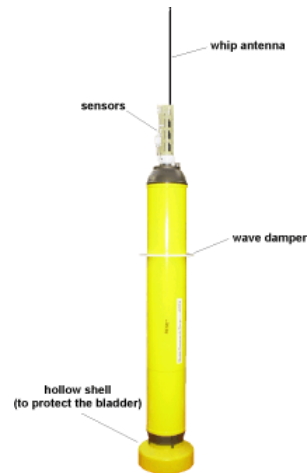
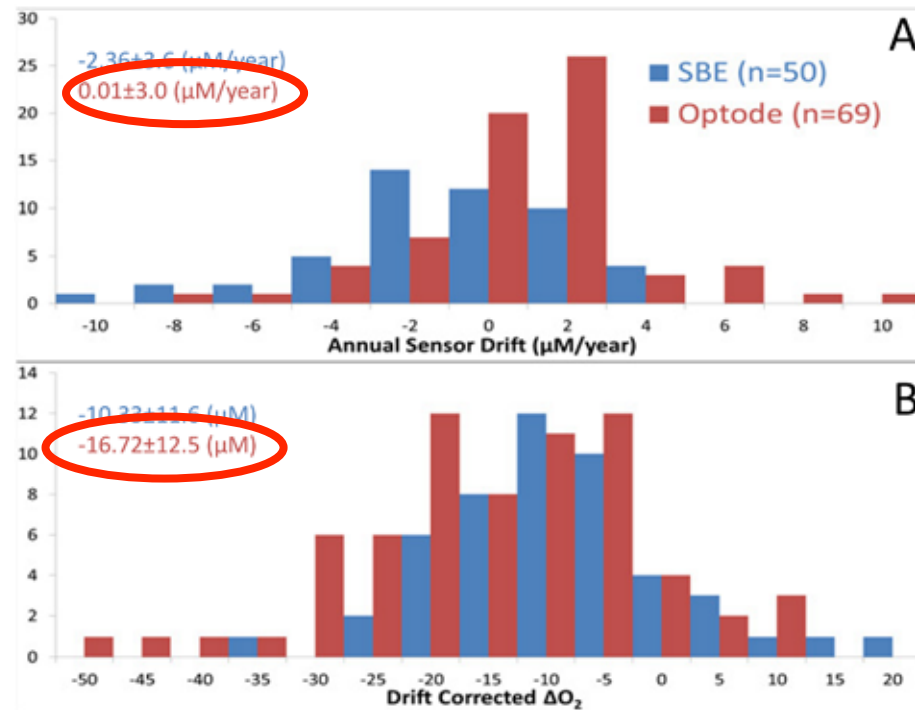
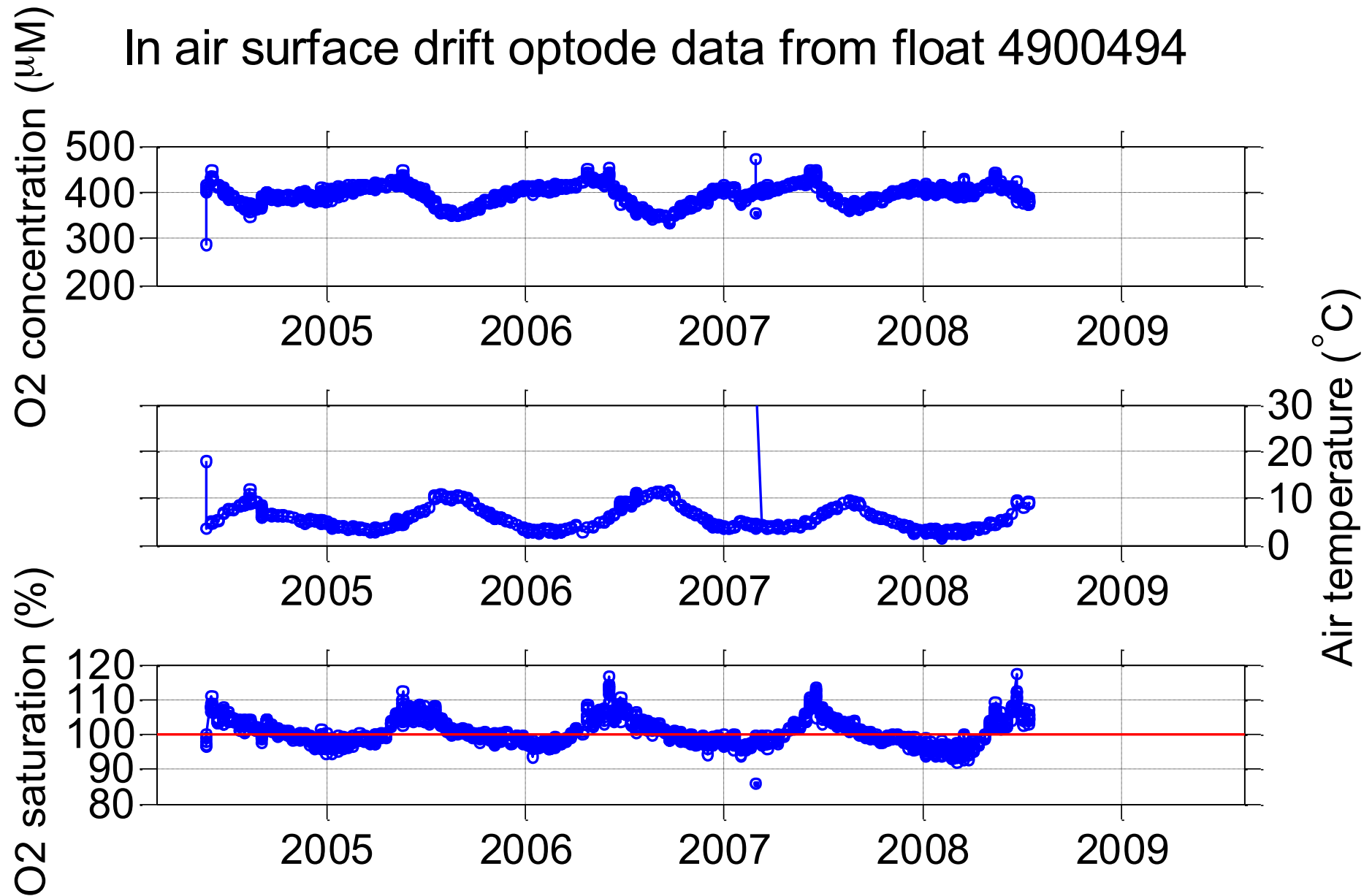


Figure 2. Histogram of sensor drift (A) and the Drift Corrected Average Deep ΔO_2 (B) for the two different types of oxygen sensors. The mean $\pm$ std. dev. is labeled on the plot with its corresponding color.

- Conclusion: Optodes stable but gave lower values
- Reason: Foils bleach in ambient light (especially fluorescent) and if sampling is done at high frequency

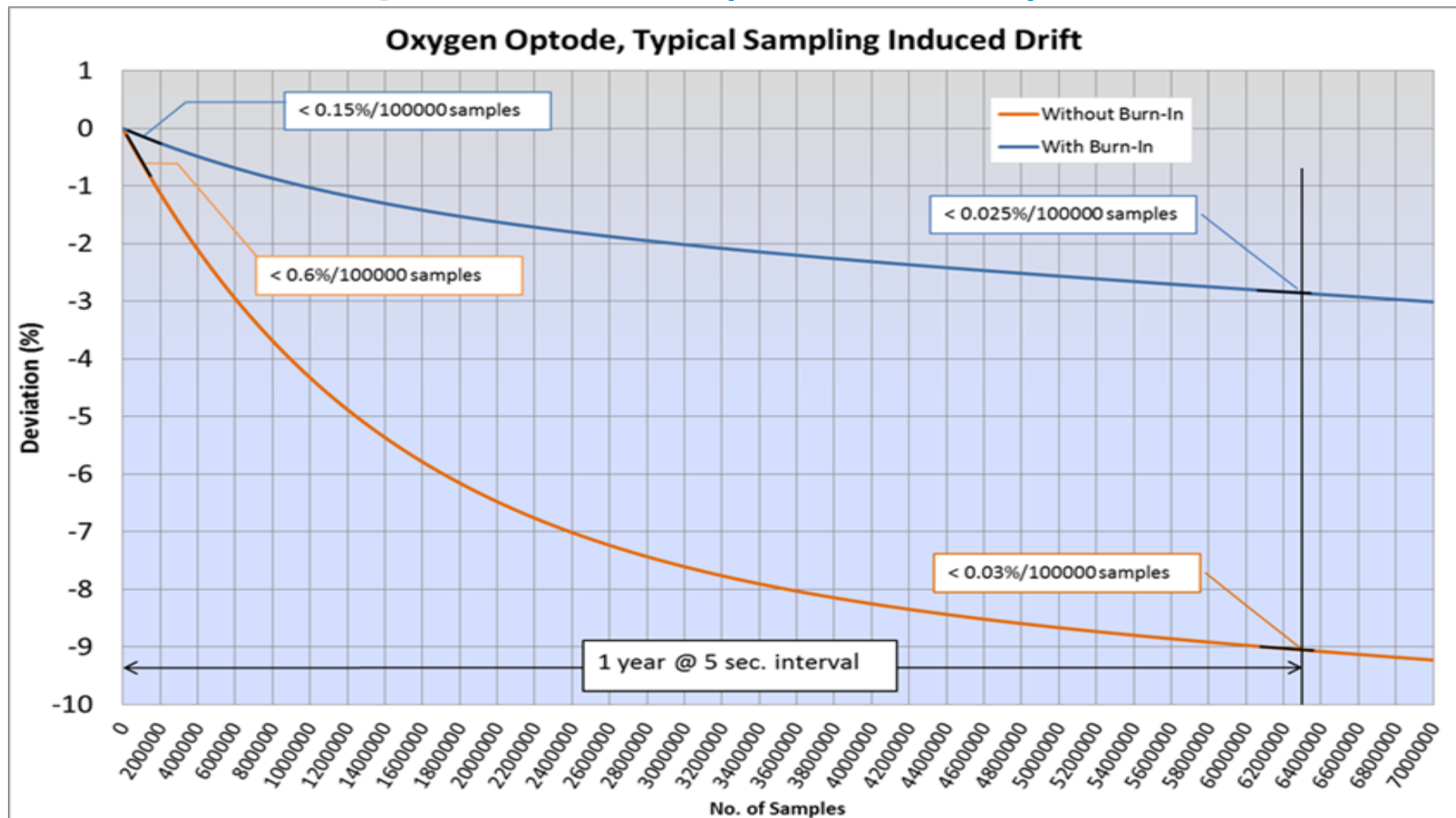
When storing sensors use black protection cap!

In air surface drift optode data from float 4900494



**Denis Gilbert et al., Argo Science Workshop 3,
Hangzhou, China, March 27, 2009**

Optode drift in relation to number of measurements and pre-treatment (with burn-in) or not



All Multipoint calibrated Foils are Pre-matured ► Better Accuracy

► Better Stability Drift < 0.15 μ M/100,000 samples

If not mechanically damaged foils get better over time

Multipoint Calibration System

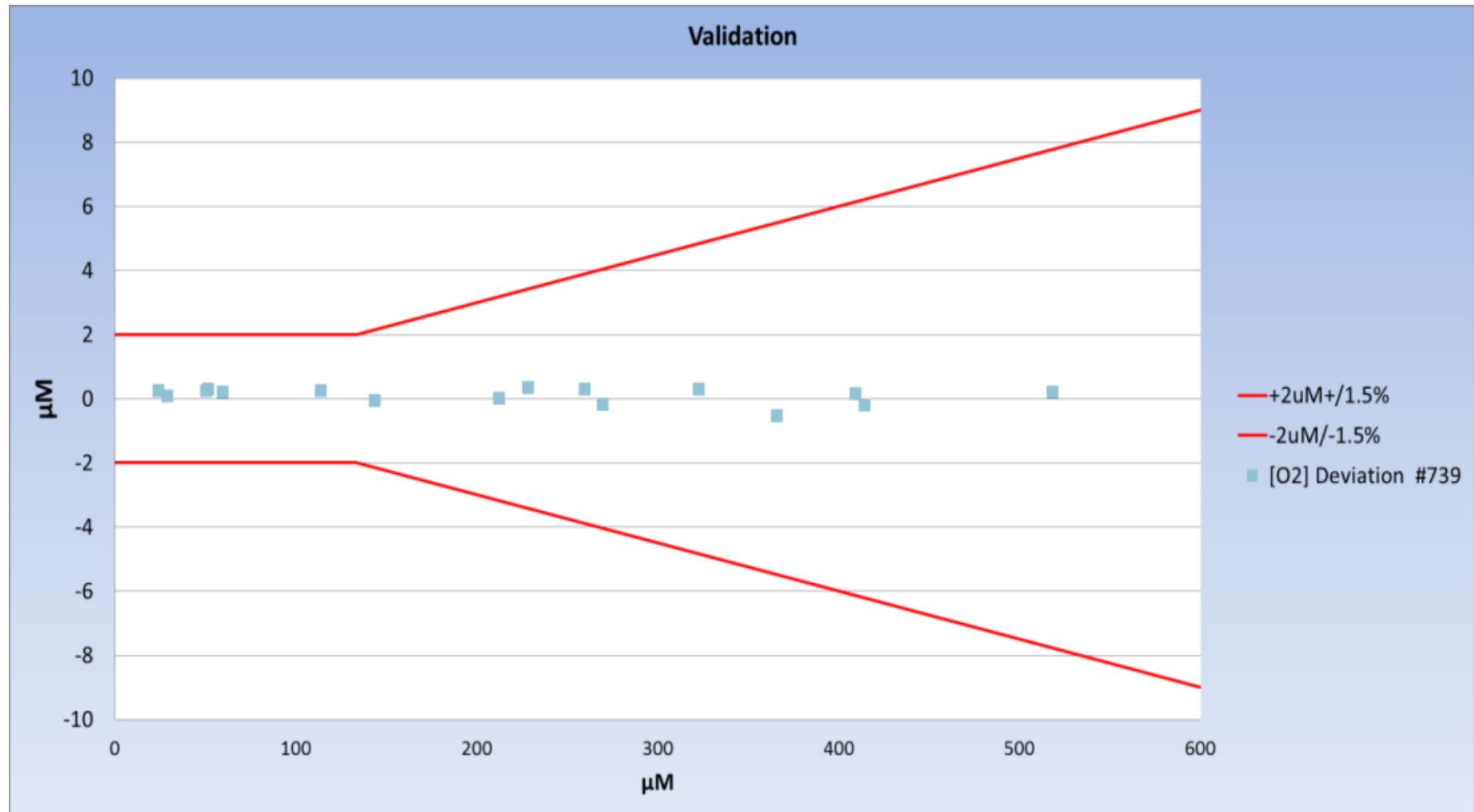
- Gas injection by use of mass flow controllers
- Automatic System 40 point calibration & 20 point subsequent verification
- Operational since August 2012
- 3 parallel reference optodes in system
- Automatic Winkler system from SI Analytics for frequent verification
- International inter-comparison of calibration facilities
- Absolute accuracy better than 1.5%/±2.5 µM



**Multipoint
calibration only for
MkII: 4330 & 4831**



Validation



Accuracy over entire range of O_2 and Temp: $\pm 2.5 \mu\text{M}$ or $\pm 1.5\%$. Field resolution: $\pm 0.2 \mu\text{M}$

Multipoint Calibration + red LED referencing gives the highest accuracy on the market

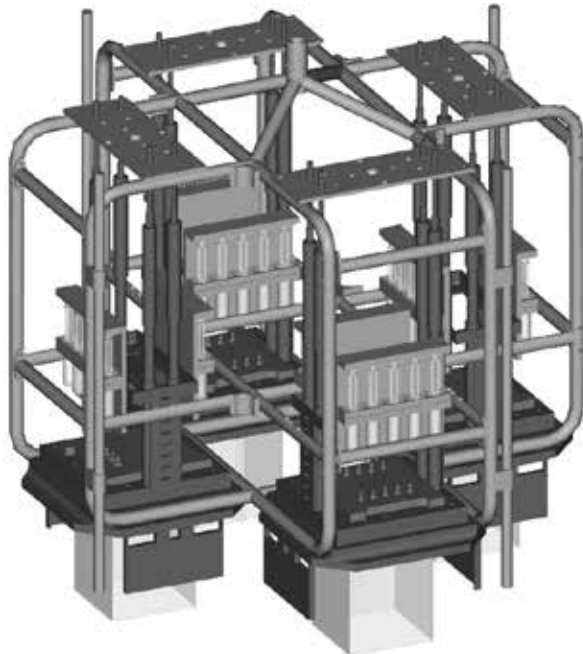
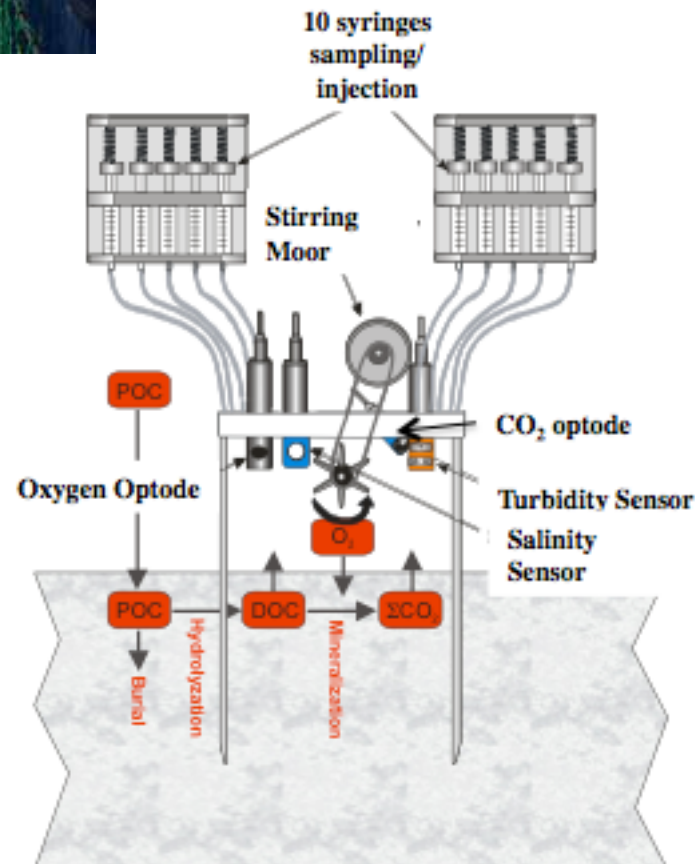


Tools: Autonomous Benthic landers

Our landers:

- 250-300 deployments
- About 800 incubations
- Deployed at 5-5600 m
- 40 water samples
- 15-25 sensors
- 95 % success rate

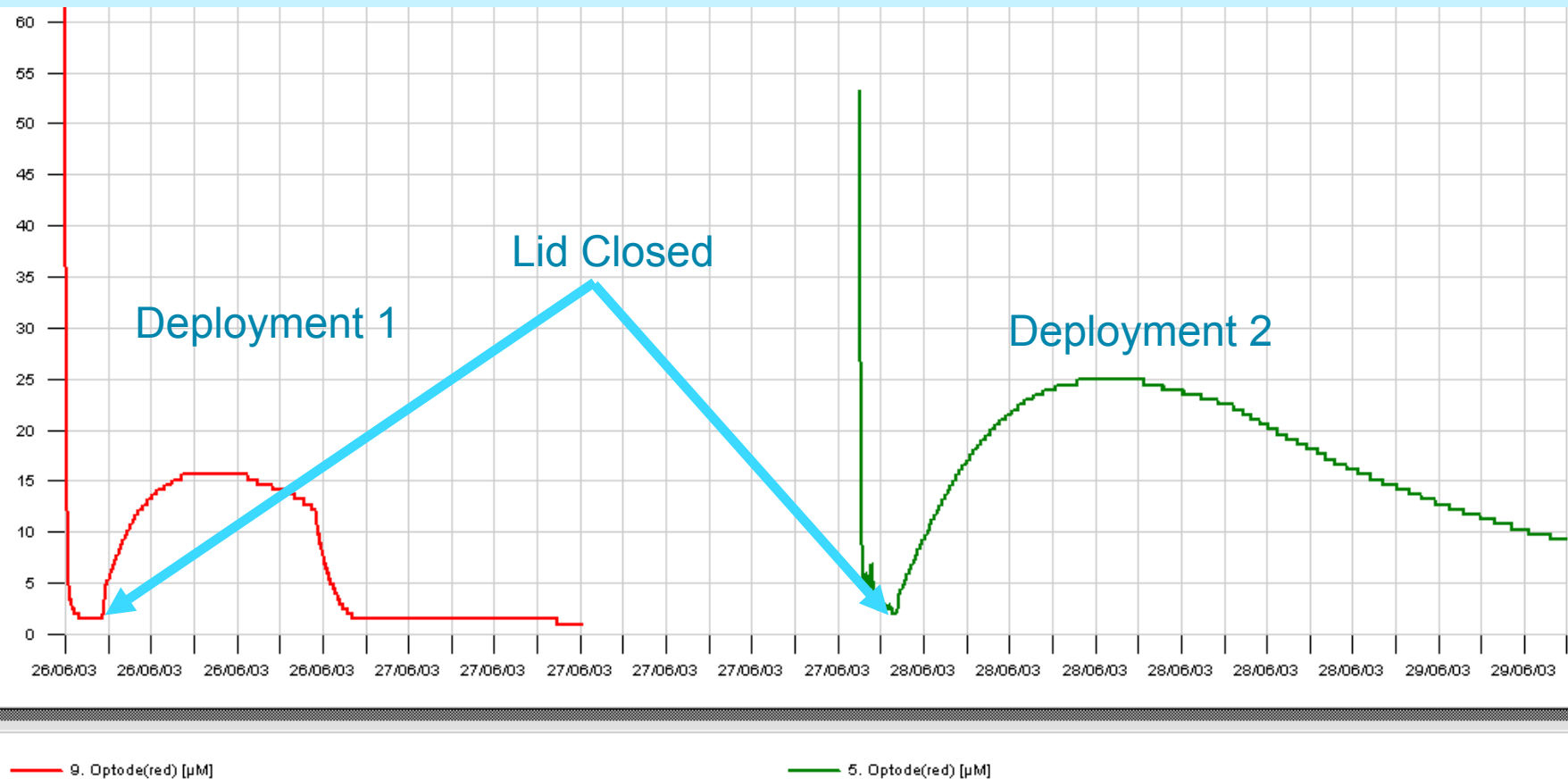
Inner frame
(4 incubations chamber modules)



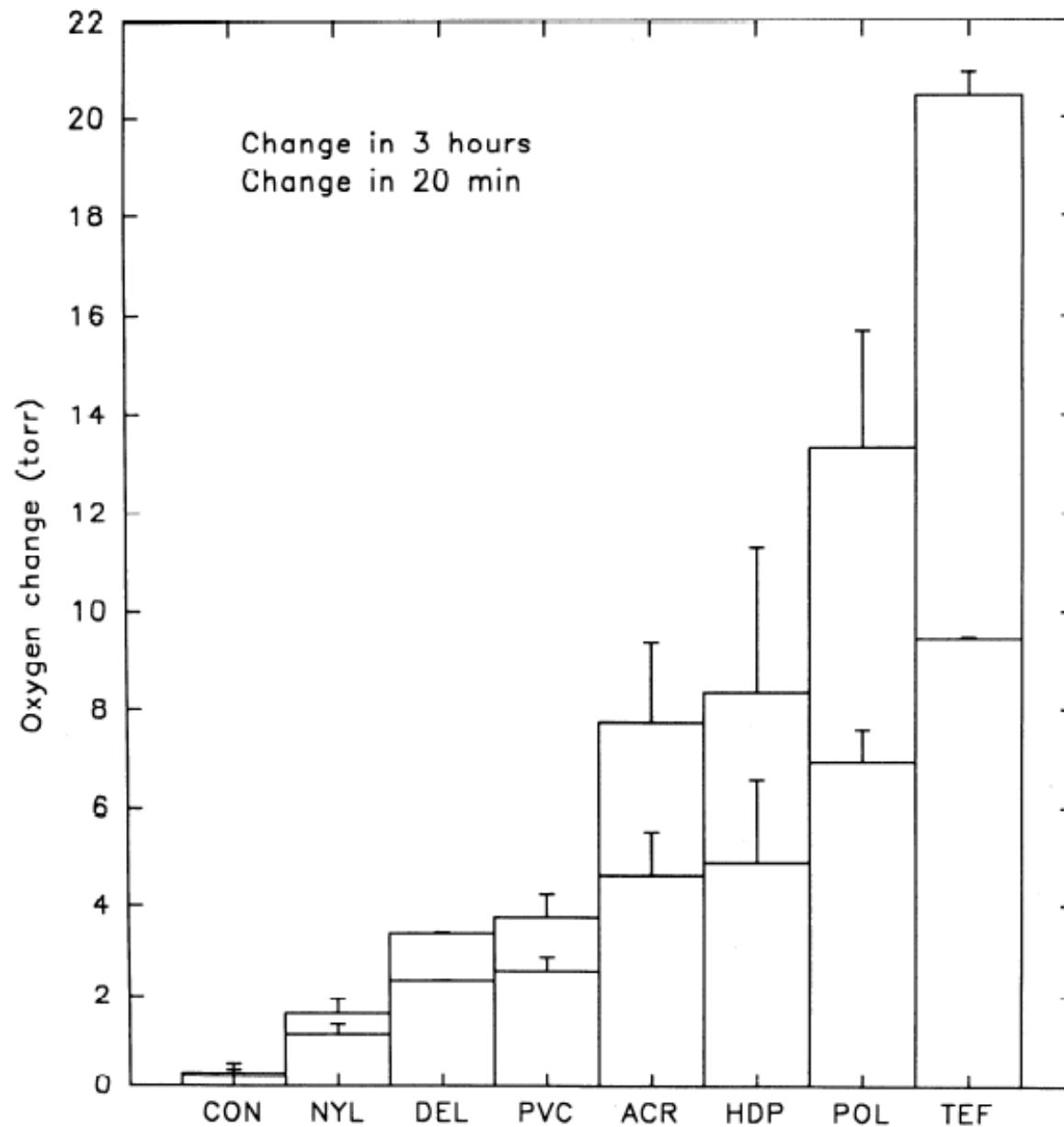
Measured Fluxes

- **Oxygen**
- Total Carbonate
- Silicate
- **Phosphate**
- Nitrate+Nitrite
- Ammonium
- DOC
- Alkalinity
- pH
- Fe and Mn
- Turbidity
- pCO₂

Plastic/Polycarbonate incubators dissolve high amounts of oxygen. Have to ventilated for hours prior to incubation in Hypoxic environments → Artifacts



Averaging: 1

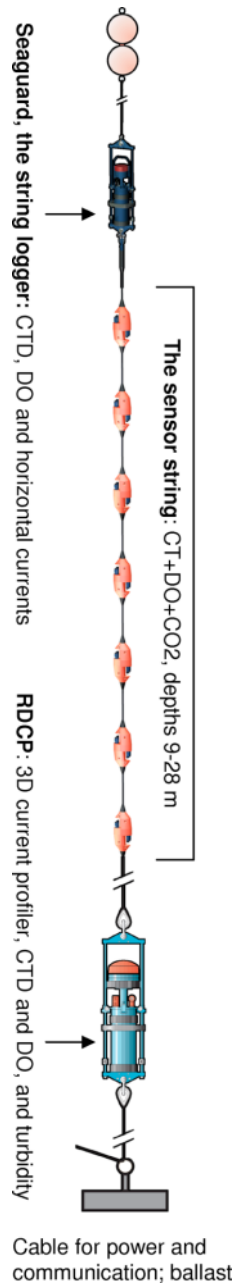
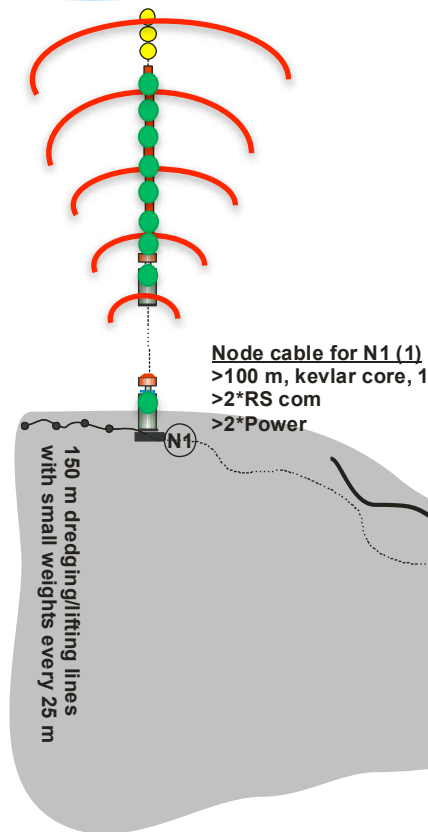
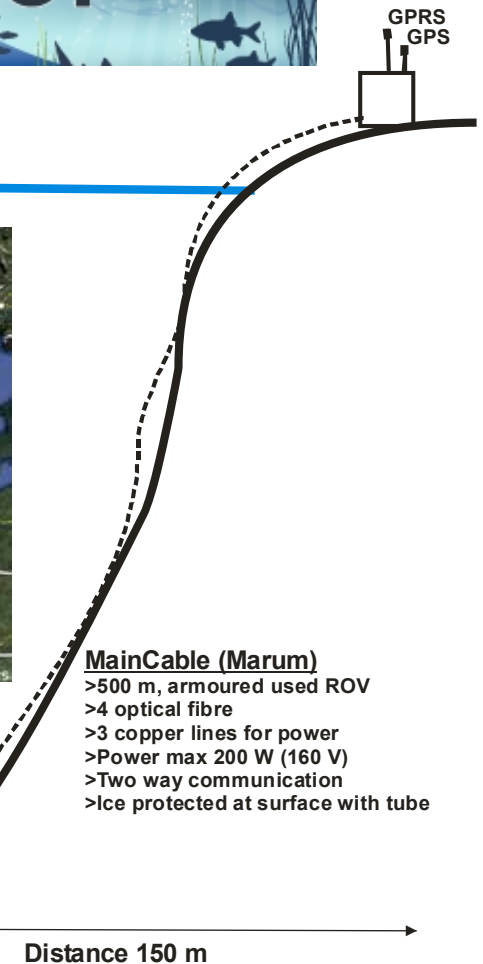


Stevens, E. Don (1992) Use of plastic materials in oxygen-measuring systems. *Journal of Applied Physiology*, 72(2): 801-804.

Plastic Materials have “memory” effects on O₂. Plastic materials should be avoided for profiling and if high accuracy/precision is needed.

Koljoe fjord observatory

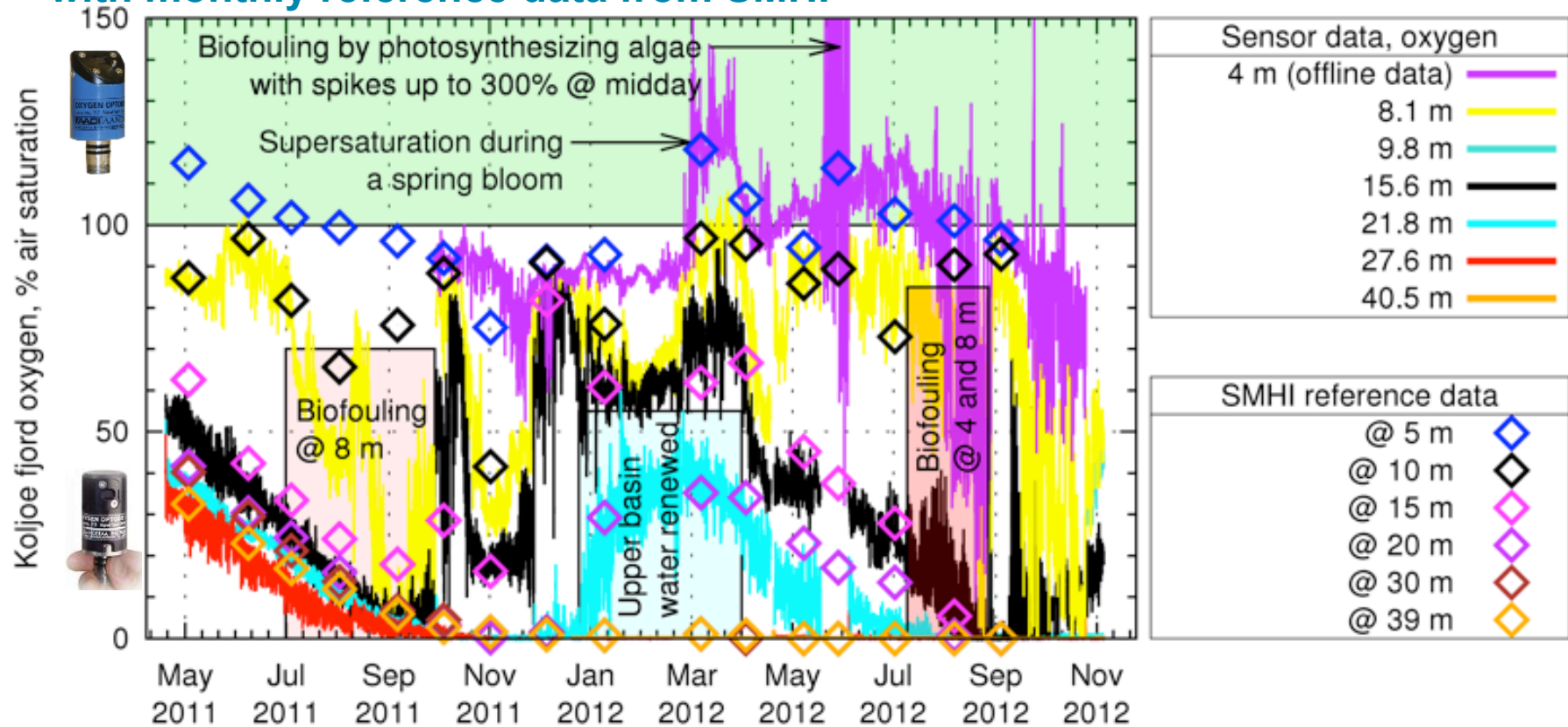
- Installed April 2011
- Real time: <http://koljofjord.cmb.gu.se>
- 40-50 sensors
- Quality control by monthly sampling (SMHI)



Cable for power and communication; ballast



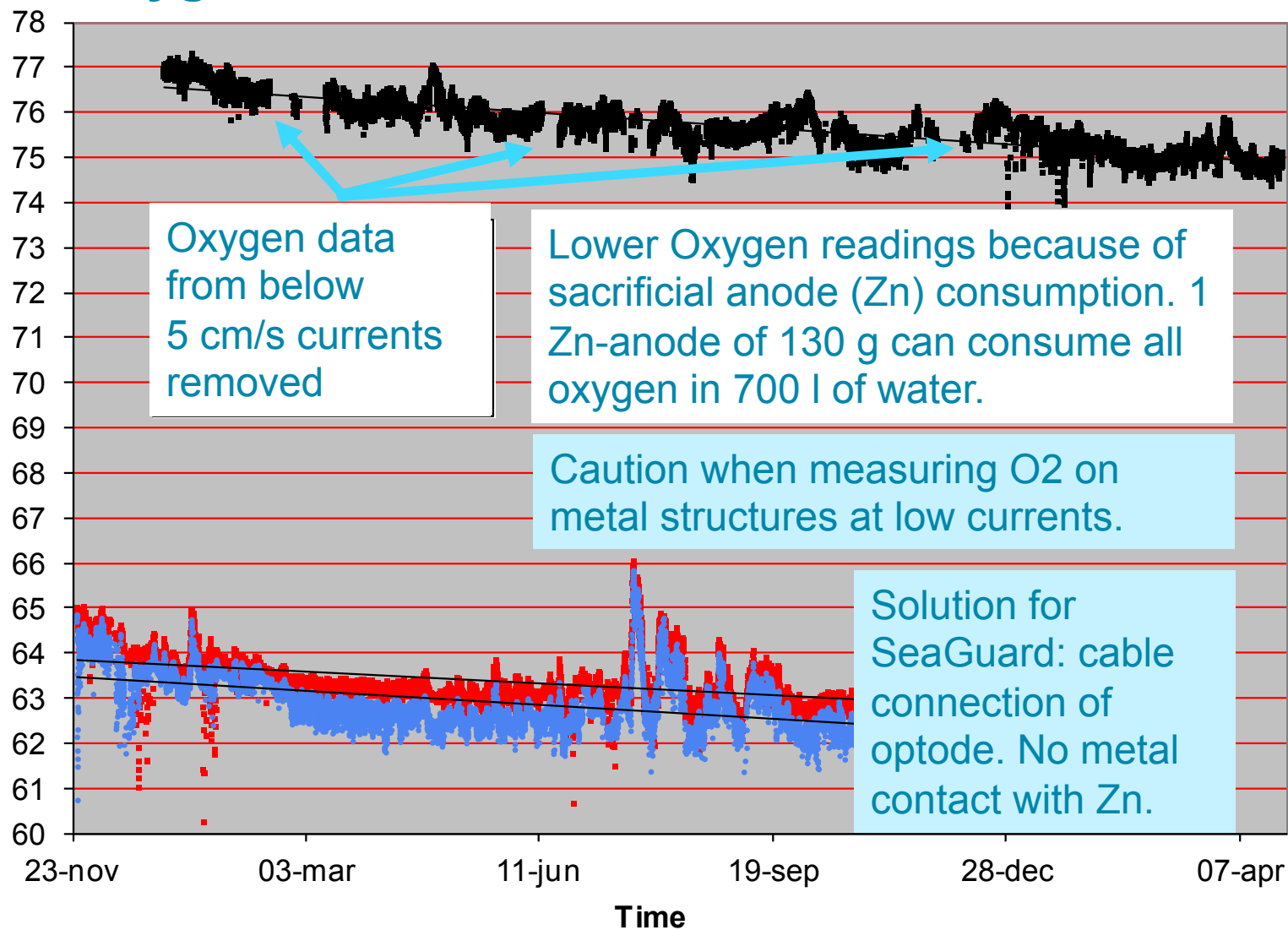
Examples of data: 18 months oxygen recordings, with monthly reference data from SMHI





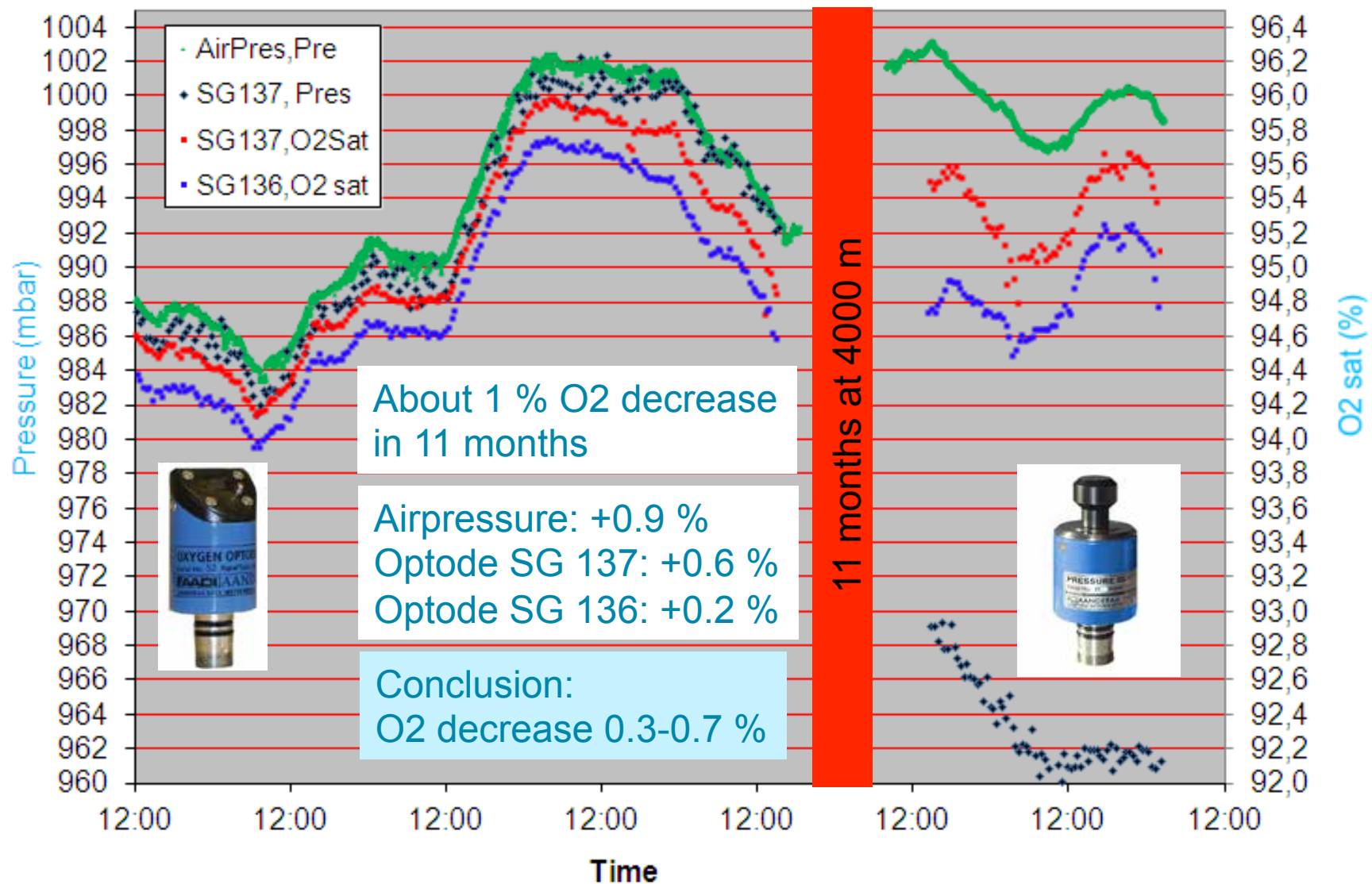
O₂ Saturation (%)

Oxygen at 4000 m

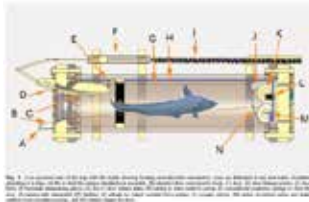


Lo Bue N. et al. (2011) Anomalies of oxygen measurements performed with Aanderaa optodes. J of Operational Oceanography. Volume 4 No. 118, 1–11.

Pre & Post Deployment data. Air Pressure Ship + SG Pressure + Oxygen Saturation before and after deployment on two Seaguards



Incubators

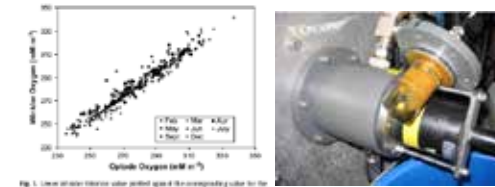


Drazen et al (2005), Almroth et al (2012),
Wikner et al (2013)

Oxygen Optodes

Examples of Scientific Papers

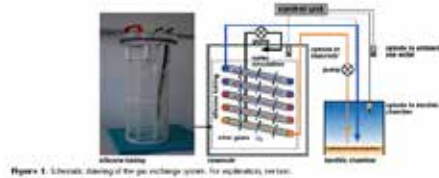
Ferry boxes



Hydes et al (2009)

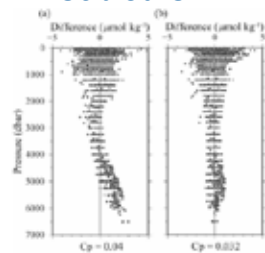
Tengberg et al (2006)

Gas Exchange Chamber



Sommer et al (2008)

Cabled CTD



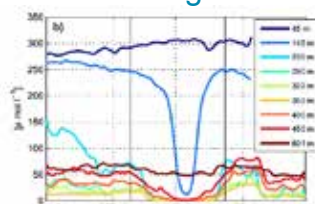
Uchida et al (2008)

Gliders



Nicholson et al (2008)

Moorings



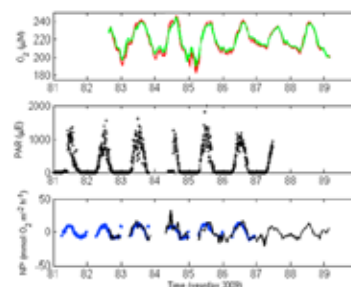
Stramma et al (2014),
Viktorsson et al (2012)

Boys



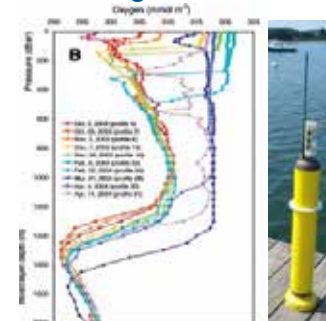
Jannash et al (2008),
Bushinsky & Emerson (2013)

Gradients



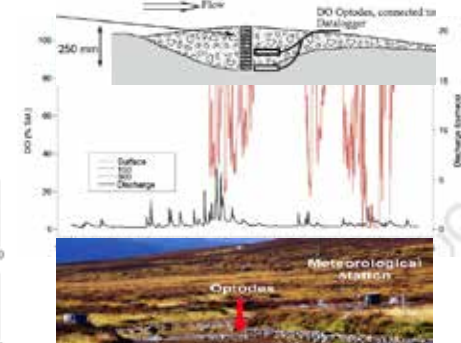
McGillis et al (2011),
Champenois and
Borges (2012)

Argo floats



Körtzinger et al (2004, Nature)
Johnson et al (2010, Nature)
Fiedler et al (2013)
Takeshita et al (2013)

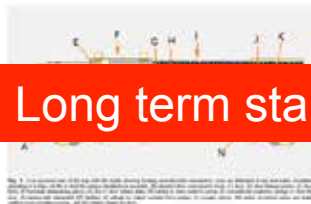
Rivers/Hydrology/Hyporheic



Birkel et al (2013),
Malcolm et al (2006, 2008,
2010), Soulsby et al (2008)

Incubators

Long term stable



Oxygen Optodes

Examples of Scientific Papers

Ferry boxes

No O₂ consumption & Robust



Drazen et al (2005), Almroth et al (2012),
Wikner et al (2013)

Not freezing sensitive

berg et al (2006)

Hydes et al (2009)

Gas Exchange Chamber

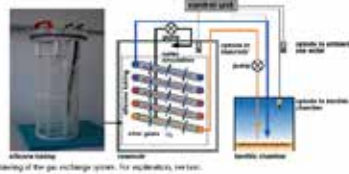


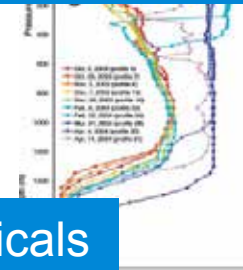
Figure 1. Schematic drawing of the gas exchange system, for incubation, see text.

Sommer et al (2008)

Good for hot water monitoring

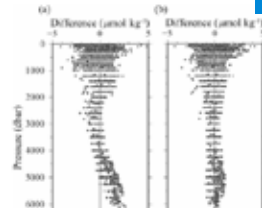


Arctic floats



Zinger et al (2004, Nature)
Johnson et al (2010, Nature)
Fiedler et al (2013)
Takeshita et al (2013)

Cabled CTD



Not sensitive to H₂S and most other chemicals

High accuracy & low noise

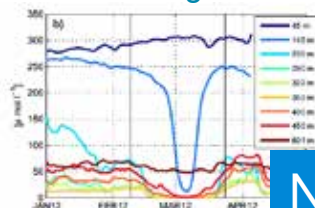
Uchida et al (2008)

Gliders



Nicholson et al (2008)

Moorings



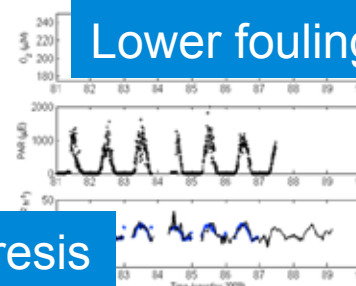
Stramma et al (2014),
Viktorsson et al (2012)

Boys



Jannasch et al (2006),
Bushinsky & Emerson (2013)

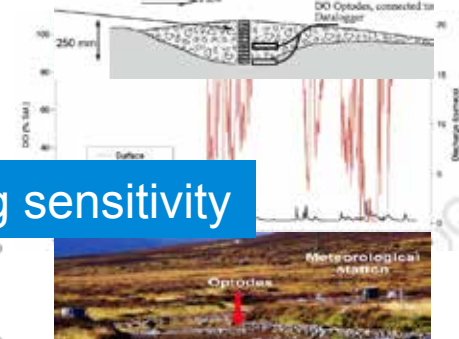
Gradients



McGillis et al (2011),
Champenois and
Borges (2012)

Lower fouling sensitivity

Rivers/Hydrology/Hyporheic



Birkel et al (2013),
Malcolm et al (2006, 2008,
2010), Soulsby et al (2008)

No pressure hysteresis



INEE



Centre d'Études
Biologiques de
Chizé



Marine Predators Team



Coriolis
OPERATIONAL OCEANOGRAPHY



Contribution of the Elephant Seal to monitor the oxygen content of the Southern Ocean

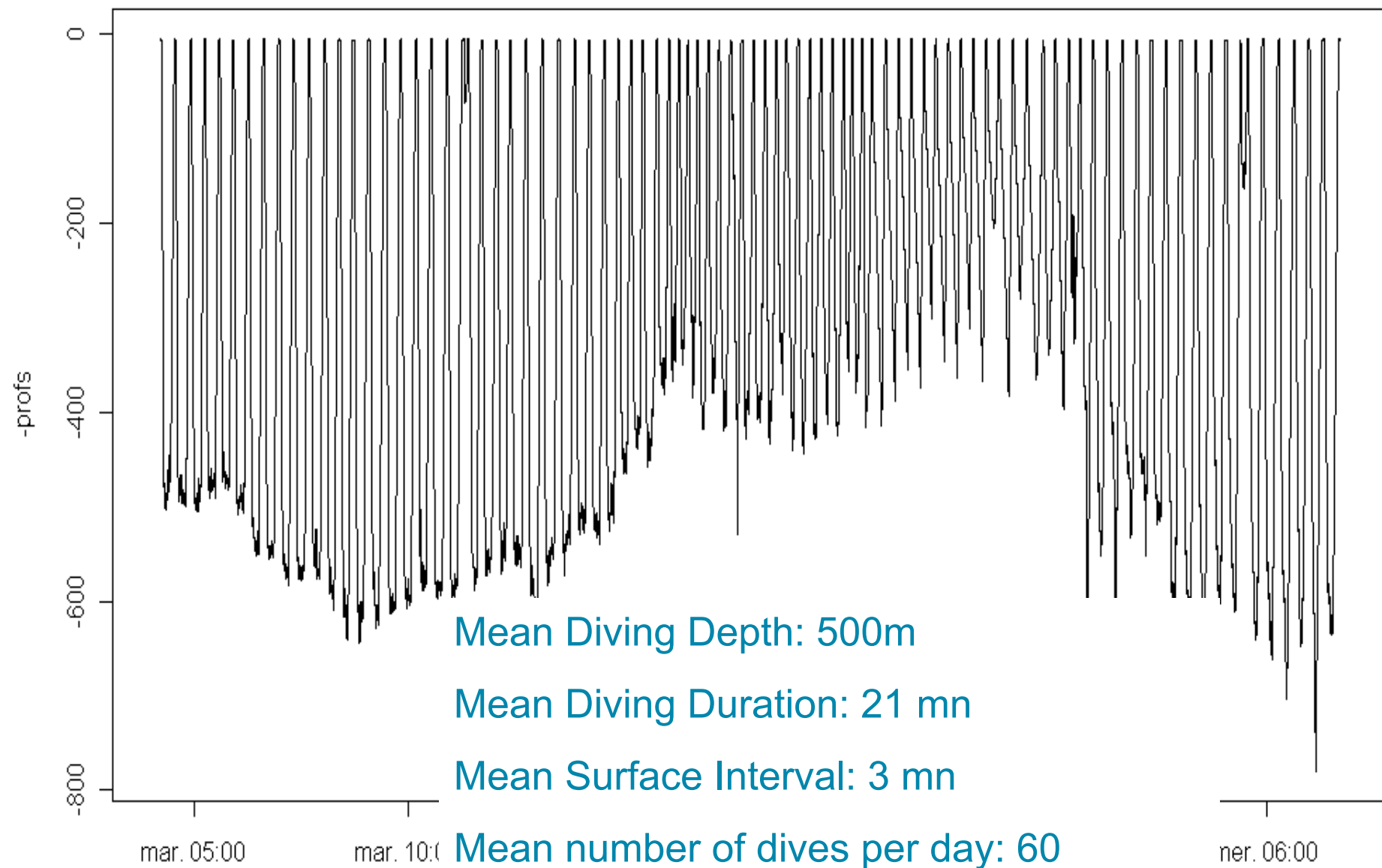
Christophe Guinet, CEBC-CNRS



ANR
AGENCE
NATIONALE
DE LA
RECHERCHE



FONDATION
TOTAL

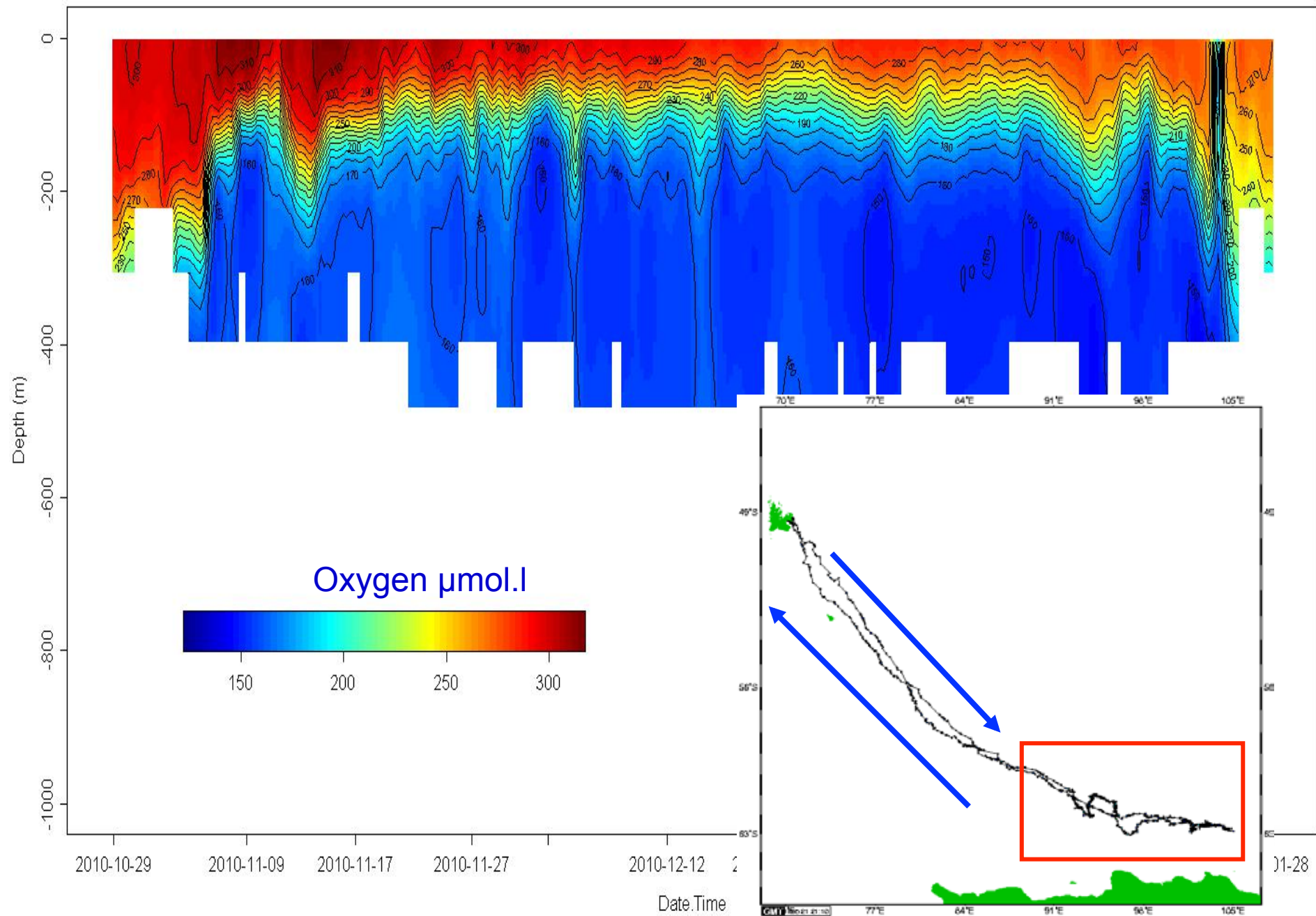


0-2 km between dives

25 Oct -17 Nov 2010

17 Nov 2010 -10 Jan 2011

10 - 28 Jan 2011



Summary

9:00-10:30

- Why should you measure Oxygen (O_2)?

Crucial parameter

- How is O_2 related to other parameters?

Related to most other parameters

- Are we measuring at the right frequencies?

Nyquist criteria: sample at twice the frequency as changes occur

- Methods to analyze and measure O_2

Winkler, Electrochemical sensors, Optical foil based sensors

11:00-13:00

- What affects O_2 sensors/methods to evaluate performance?

Multivariate methods good tool for evaluation

- How good is Winkler titration?

Depends on person, contamination at low O_2 from plastic bottles

- Field data, value of combined measurements

Combined measurements gives better certainty, O_2 should always be combined with pCO_2 , pH, ChIA

- Practical Demonstration and Hands-On