



**A STUDY OF ANTIBACTERIAL ACTIVITY OF ANDROGRAPHIS PANICULATA LEAF  
EXTRACTS AGAINST SOME CLINICAL PATHOGENIC BACTERIA'S**

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**ABSTRACT**

The use of plant extracts for antimicrobial activity and other diseases have been observed to be promising remedies since ancient time in Chinese medicine, Ayurveda, Arabic, and Unani medicine. The plants have traditionally furnish a source of hope for novel drug compounds, as plant herbal mixtures have made large endowment to human health and well being. The use of plant extracts with known antimicrobial properties can be of appreciable significance for therapeutic treatment. Presently, the research has been initiated to study the antibacterial activity of chloroform, and methanol extracts of *Andrographis paniculata* to emphasize the potential of herbal components in the field of medical science to kill various dreadful pathogens. The agar well diffusion method was followed to evaluate the antibacterial activity of leaf and stem extracts of *A. paniculata* against *Bacillus subtilis*, *S. aureus*, *Enterococcus*, *P. aeruginosa*, *K. pneumonia* and *E. coli*. The result revealed that all the doses of both extracts of *A. paniculata* potentially inhibited the growth of all the pathogens tested. Hence, the present investigation evaluates the potential anti bacterial activity of methanol extract of leaf and stem extract of *A. paniculata*.

**KEYWORDS:** *Andrographis paniculata*, antimicrobial activity, pathogens, methanol, leaf and stem bark extract.

**1. INTRODUCTION**

Plants produce a wide range of bioactive molecules, making them rich source of different types of medicines. Most of the drugs used today are acquired from natural sources or semi synthetic derivatives of natural products used in the traditional systems of medicine (Sukanya, et. al., 2009). Medicinal plants are finding their way into pharmaceuticals, cosmetics, and nutraceuticals. In pharmaceutical field medicinal plants are largely used for the broad range of substances present in plants which have been used to treat infectious as well as chronic diseases (Okigbo, et. al., 2009). The drugs already in use to treat infectious disease are of concern because drug safety remains a huge global issue. Almost all of the synthetic drugs cause side effects and also most of the microbes developed resistant against the synthetic drugs. To alleviate this problem, antimicrobial compounds from potential plants should be explored. These drugs from plants are fewer side effects, less toxic, scanty and also cost effective. They are effective in the treatment of infectious diseases while simultaneously mitigating many of the side effects that are often associated with synthetic antimicrobials (Kadhim, et. al., 2016). Treatment with medicinal plants having antibacterial activity is potentially beneficial alternative and promising source of pharmaceutical agents (Sridevi,

et.al., 2010). Plants are rich in a wide variety of secondary metabolites of phytochemical constituents such as tannins, alkaloids and flavonoids, which act against different diseases (Govind and Madhuri, 2010). In addition, plant derived phytomedicines provide a cheaper source for treatment and significant accuracy than chemotherapeutic agents (Punitha, et. al., 2008). *Andrographis paniculata*, commonly known as 'King of Bitter', is a small, annual, branched and erect plant belongs to the family Acanthaceae. It grows abundantly in south eastern Asia including India, Sri Lanka, Java, Pakistan, Indonesia and Malaysia. It prefers to grow well in a diversity of habitats such as moist, shady areas, hill slopes, plains, farms, seashores, waste lands and dry or wet lands (Prajapati, 2003). It is rich in a wide variety of phytochemical constituents such as diterpenes, flavonoids and lactones (Chang, et. al., 1987). *A. paniculata* is extensively used in Ayurveda, Unani and Siddha medicine as home remedy for various diseases in Indian traditional system as well as in tribal medicine applications. The therapeutic value of Kalmeg is due to its mechanism of action by enzyme induction.

It is a powerful cold property herb, used in fevers and to dispel toxins from the body. It is used to treat gastrointestinal tract and upper respiratory infections, fever,

herbs, sore throat, hepatitis and a variety of other chronic and infectious diseases (Chopra, 1956). The herbs and its isolates like, isoandrographolide, neoandrographolide, andrographolide, isoandrographolide are reported to possess anti-inflammatory activity (Liu, 2007) hepatoprotective (Shukla, 1992), anti - diabetic (Umamaheswari, et. al., 2007), anti - malarial (Misra, et. al., 1992), anti - microbial (Singha, 2003), piles and gonorrhoea (Prajapati, 2003). Herbal drugs in disease management are attain success, because they are cost effective, eco-friendly and have minimal side effects (Ahilan, et. al., 2010). Hence, this work made an attempt to study the antimicrobial activity of *Andrographis paniculata* against various pathogenic microorganisms.

## 2. MATERIALS AND METHODS

### 2.1 Collection of *A. paniculata*

The experimental plant species, *A. paniculata* was purchased from the local herbal market. The plant was authenticated and the voucher specimen was deposited in the herbarium of Central Laboratory Facility, Chhattisgarh council of Science and Technology, Raipur.

### 2.2 Preparation of plant powder

Fresh *A. paniculata* plants were washed thoroughly in tap water followed by distilled water and were then shade dried until all the water content was lost completely. Dried plants were crushed and powdered using blender. Fine powder was obtained after sieving and stored in airtight container until further use.

### 2.3 Preparation of experimental plant extracts

The plant powder was extracted with methanol solvent with an increasing polarity. The successive extraction was done by a cold maceration process for seven days with regular agitation. After seven days of cold maceration process it was filtered through sterile muslin cloth and the solvent was evaporated using soxhlet apparatus. The residues obtained after evaporation were stored at -20°C until used for experimentation.

### 2.4 Test microorganisms

To evaluate the antimicrobial activity of *A. paniculata* extracts, six species/ strains of microorganisms were

selected, namely *Bacillus subtilis*, *S. aureus*, *Enterococcus*, *P. aeruginosa*, *K. pneumonia* and *E. coli*. All these bacterial strains were collected from clinical lab and sub cultured in nutrient agar medium and used for antimicrobial susceptibility test.

### 2.5 Antibacterial assay

The potential antibacterial activity of *A. paniculata* extract was studied through agar well diffusion method (Murry, et. al., 1995). The sterile petri dishes were filled with 25ml of agar and allowed the agar to get solidified. Prior to streaking the plates with bacterial culture, 5mm diameter wells were punched in the medium using a sterile borer. After the agar gets solidified the bacterial cultures were inoculated by spreading in the petri plates using sterile cotton swabs. Then 0.1ml of plant extract in peptone water was directly applied to the well made on the surface of agar containing bacterial lawn. Positive control was maintained with different antibiotics and wells containing solvent alone was maintained as negative control. The inoculated plates were incubated overnight at 37°C for allowing the bacterial growth and the diameter of zone of inhibition was measured in mm.

## 3. RESULTS

In the present investigation, methanol leaf and stem bark extract of *A. paniculata* were studied against *Bacillus subtilis*, *S. aureus*, *Enterococcus*, *P. aeruginosa*, *K. pneumonia* and *E. coli* using agar well diffusion method. The results showed that the extract of *A. paniculata* has a concentration dependent antibacterial activity with more sensitivity for Gram negative bacteria than Gram positive bacteria used in the study. The extracts of *A. paniculata* showed considerable antibacterial activity at all the four concentrations 100, 75, 50 and 25mg/ml (Table 1 and 2). Table 3 shows the sensitivity of the tested bacteria to the standard antibiotics. Examination of this study clearly revealed that methanol extracts of *A. paniculata* act as a significant growth inhibitor against broad spectrum of pathogens and act as a potent antimicrobial activator.

**Table 2: The study of anti-bacterial activities of *Andrographis paniculata* leaf extracts using disk diffusion Method (Mean± SE).**

SI	Name of Bacteria	Zone of Inhibition (In MM)			
		100%	75%	50%	25%
1	Gram Positive (+)				
	<i>Bacillus subtilis</i>	3.66 ± 0.78	8.33 ± 0.94	8.66 ± 1.05	6.33 ± 0.64
	<i>S. aureus</i>	4.33 ± 0.84	8.33 ± 0.65	7.33 ± 0.66	8.33 ± 0.74
	<i>Enterococcus</i>	5.66 ± 0.97	8.33 ± 0.74	8.33 ± 0.94	10.33 ± 0.74
2	Gram Negative (-)				
	<i>P. aeruginosa</i>	4.66 ± 0.87	9.66 ± 1.05	11.33 ± 0.94	7.33 ± 0.66
	<i>K. pneumoniae</i>	6.33 ± 1.02	8.66 ± 0.74	9.33 ± 1.05	10.66 ± 0.74
	<i>E. Coli</i>	5.33 ± 0.94	8.66 ± 0.66	10.33 ± 0.74	9.33 ± 1.10

**Table 3: The study of anti-bacterial activities of standard antibiotics using disk diffusion method.**

SI	Name of Bacteria	Zone of Inhibition (In MM)			
		NX10	OF5	E15	GEN10
1.	Gram Positive (+)				
	<i>Bacillus subtilis</i>	25	29	22	20
	<i>S. aureus</i>	30	25	10	23
	<i>Enterococcus</i>	34	28	13	25
2.	Gram Negative (-)				
	<i>P. Aeruginosa</i>	35	32	15	30
	<i>K. Pneumoniae</i>	30	25	10	18
	<i>E. Coli</i>	31	30	15	32

#### 4. DISCUSSION

Medicinal plants are the prime sources of new medicines and may constitute an alternative to the usual drugs. Medicinal and aromatic plants are used on a wide scale in medicine against drug resistant bacteria (Tepe, et al., 2004). In this study all the *A. paniculata* extracts exhibited varying degree of inhibitory activity against the growth of all the microorganisms tested except *Pseudomonas aeruginosa*. This result was supported by many of the researchers who already reported that *A. paniculata* as potent antimicrobial activator. Mishra et al. (2013) reported that 75% methanol extract of *A. paniculata* leaves was found to be active against *S.aureus*, *E. faecalis* and *M. tuberculosis*. Zaidan et al. (2005) have reported that the water extracts of *A. paniculata* possess a potential antibacterial activity towards both gram positive and gram negative bacteria. According to the results of Humnabadkar and Kareppa (2012), the aqueous extracts of *A. paniculata* showed maximum antibacterial activity against *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Hosamani et al. (2011), have reported that the acetone and alcohol extracts of *A. paniculata* with higher inhibitory activity against *Bacillus subtilis* and *Staphylococcus aureus*. Research conducted on other plants also showed positive result on antimicrobial activity. Turker et al. (2009), examined the aqueous and alcoholic extracts of *Nuphar lutea*, *Nymphaea alba*, *Stachys annua*, *Genista lydia*, *Vinca minor*, *Fragaria* herbs of Turkey with antibacterial activity against *A. hydrophila*, *Enterococcus faecalis*, *Lactococcus garvieae*, *Streptococcus agalactiae* and *Yersinia ruckeri* bacteria isolated from fish. Mahesh and Satish (2008).

In our present investigation, methanol leaf and stem bark extract of *A. paniculata* were studied against *Bacillus subtilis*, *S. aureus*, *Enterococcus*, *P. aeruginosa*, *K. pneumonia* and *E. coli* using agar well diffusion method. The results showed that the extract of *A. paniculata* has a concentration dependent antibacterial activity with more sensitivity for Gram negative bacteria than Gram positive bacteria used in the study. The extracts of *A. paniculata* showed considerable antibacterial activity at all the four concentrations 100, 75, 50 and 25mg/ml (Table 1 and 2). Table 3 shows the sensitivity of the tested bacteria to the standard antibiotics. Examination of this study clearly revealed that methanol extracts of *A. paniculata* act as a significant growth inhibitor against broad spectrum of pathogens and act as a potent antimicrobial activator.

Generally gram positive bacteria were more sensitive to plant extracts because of the presence of a mesh-like peptidoglycan layer which is more accessible to permeation by the extracts (Tajkarimi, et. al., 2010). The resistance of the gram negative bacteria could be attributed to its cell wall structure. Gram negative bacteria have a powerful permeability barrier, composed of a thin lipopolysaccharide exterior membrane, which could restrict the penetration of the extruding plant extract. It has been discussed earlier that gram negative bacteria are usually more resistant to the plant originated antimicrobials and even show no effect, compared to gram positive bacteria (Stefanello, 2008).

Results obtained from this study, indicated that, the plant extracts showed the strongest antimicrobial activity than the control. Further studies are needed for these potent plant extracts to evaluate the other parameters of antimicrobial activity (e.g., toxicity, in vivo efficacy, antiviral and antiparasitic and antimycobacterial activity).

#### 5. CONCLUSION

*A. paniculata* has been used in Ayurveda, Unani and Siddha systems of medicine from ancient times. It has wide spectrum of pharmacological activities either in the form of powder, extracts or in its isolated compounds with minimum side effects; several products fortified with extract or isolated compounds have been launched in national and international markets for various diseases.

In our present investigation, methanol leaf and stem bark extract of *A. paniculata* were studied against *Bacillus subtilis*, *S. aureus*, *Enterococcus*, *P. aeruginosa*, *K. pneumonia* and *E. coli* using agar well diffusion method. The results showed that the extract of *A. paniculata* has a concentration dependent antibacterial activity with more sensitivity for Gram negative bacteria than Gram positive bacteria used in the study. The extracts of *A. paniculata* showed considerable antibacterial activity at all the four concentrations 100, 75, 50 and 25mg/ml (Table 1 and 2). Table 3 shows the sensitivity of the tested bacteria to the standard antibiotics. Examination of this study clearly revealed that methanol extracts of *A. paniculata* act as a significant growth inhibitor against broad spectrum of pathogens and act as a potent antimicrobial activator. The enriched *A. paniculata* plant extracts are further

studied for their antibacterial properties against selective human pathogens.

In this context, the present study was carried out to find out the antibacterial potential of various solvent based extract of *A. paniculata* against selective human pathogens. The antibacterial activity of *A. paniculata* may be due to the presence of active principle called andrographolide. In future, the improvements of active principle andrographolide content in *A. paniculata* using plant growth promoting rhizobacteria (PGPR) are studied. The enriched *A. paniculata* plant extracts are further studied for their antibacterial properties against selective human pathogens.

## 6. REFERENCES

- Ahilan B, Nithyapriyadarshini A, Ravaneshwaran K. Influence of certain herbal additives on the growth, survival and disease resistance of glod fish, *Carassius auratus* (Linnaeus). *Journal of Veterinary Animal Sciences*, 2010; 6(1): 5-11.
- Chang HM, But PPH. *Pharmacology and application of Chinese material medica*, Chinese Medicinal Material Research Centre, The Chinese University of Hong Kong, Singapore. World Scientific Publishing Co. Pte. Ltd, 1987; 2: 918-928.
- Chopra RN, Nayar SL, Chopra IC. *Glossary of Indian medicinal plants*. NISCOM, CSIR, New Delhi, 1956. Liu J, Wang ZT, Ji LL. In vivo and in vitro antiinflammatory activities of neoandrographolide, *The American Journal of Chinese Medicine*, 2007; 35: 317- 328.
- Govind P, Madhuri S. Significance of fruits and vegetables in malnutrition cancer. *Plant Archives*, 2010; 10(2): 517-522.
- Hosamani PA, Lakshman HC, Kumar SK, Rashmi C, Hosamani. Antimicrobial activity of leaf extract of *Andrographis paniculata* wall. *Science Research Reporter*, 2011; 1(2): 92-95.
- Humnabadkar SS, Kareppa BM. In vitro study of antibacterial activity of *Andrographis paniculata* against clinically important pathogens. *International Journal of Advanced Biological Research*, 2012; 2(4): 584-586.
- Kadhim WA, Kadhim MJ, Hameed IH. Antibacterial activity of several plant extracts against *Proteus* species. *International Journal of Pharmaceutical and Clinical Research*, 2016; 8(12): 1673-1684.
- Mahesh B, Sathis S. Antimicrobial activity of some important medicinal plant against plant and human pathogens. *World Journal of Agricultural Science*, 2008; 4: 839-843.
- Mishra PK, Rahul Kunwar S, Anamika G, Adya C, Rahul P, Shree Prakash T et al. Antimicrobial activity of *Andrographis paniculata* (Burm.f.) wall ex Nees leaves against clinical pathogens. *Journal of Pharmacy Research*, 2013; 7: 459-462.
- Misra P, Pal NL, Guru PY, Katiyar JC, Srivastava V, Tandon JS. Antimalarial activity of *Andrographis paniculata* (Kalmegh) against *Plasmodium berghei* NK 65 in *Mastomys natalensis*. *International Journal of Pharmacognosy and Phytochemistry*, 1992; 30: 263-274.
- Murry PR, Baron EJ, Pfaller MA, Tenover FC, Tenover FC. *Manual of clinical Microbiology*, 6th Edition, ASM Press, Washington, DC, 1995; 15-18.
- Okigbo RN, Anuagasi CL, Amadi JE. Advances in selected medicinal and aromatic plants indigenous to Africa. *Journal of Medicinal Plants Research*, 2009; 3(2): 86-95.
- Prajapati ND, Purohit SS, Sharma AK, Kumar T. *A hand book of medicinal plants, A complete source Book*. Agrobios, Jodhpur, India, 2003; 45-46.
- Punitha SMJ, Babu MM, Sivaram V, Shankar VS, Dhas SA, Mahesh TC et al. Immunostimulating influence of herbal biomedicines on non-specific immunity in grouper *Epinephelus tauvina* juvenile against *Vibrio harveyi* infection. *Aquaculture*, 2008; 16: 511-523.
- Shukla B, Visen PKS, Patnaik GK, Dhawan BN. Choloretic effect of andrographolide in rats and guinea pigs. *Planta Medica*, 1992; 58: 146-148.
- Singha PK, Roy S, Dey S. Antimicrobial activity of *Andrographis paniculata*. *Fitoterapia*, 2003; 74: 692-694.
- Sridevi M, Kondala Rao B, Sathiraju D. Sensitivity of bacteria isolated from Champarathi estuary to some medicinal plants of Vizianagaram district, East Coast of India. *Drug Invention Today*, 2010; 2(7): 366-368.
- Stefanello M<sup>É</sup>A, Cervi AC, Ito IY, Salvador MJ, Wisniewski A, Simionatto EL. Chemical composition and antimicrobial activity of essential oils of *Eugenia chlorophylla* (Myrtaceae). *Journal of Essential Oil Research*, 2008; 20(1): 75-78.
- Sukanya SL, Sudisha J, Hariprasad P, Niranjana SR, Prakash HS, Fathima SK. Antimicrobial activity of leaf extracts of Indian medicinal plants against clinical and phytopathogenic bacteria. *African Journal of Biotechnology*, 2009; 8(23): 6677-6682.
- Tajkarimi MM, Ibrahim SA, Cliver DO. Antimicrobial herb and spice compounds in food. *Food Control*, 2010; 21(9): 1199-1218.
- Tepe B, Daferera D, Sokmen M, Polissiou M, Sokmen A. In vitro antimicrobial and antioxidant activities of the essential oils and various extracts of *Thymus eigi* M. Zohary et P. H. Davis. *Journal of Agricultural and food chemistry*, 2004; 52: 1132-1137.
- Turker H, Yildirim B, Karakas FP. Antibacterial activities of extracts from some Turkish endemic plants on common fish pathogens. *Turk Journal of Biology*, 2009; 33: 73-78.
- Umamaheswari S, Mainzen, Prince PS. Antihyperglycaemic effect of 'Ilogen-Excel', an ayurvedic herbal formulation in streptozotocin-induced diabetes mellitus. *Acta Poloniae Pharmaceutica*, 2007; 64: 53-61.

24. Zaiden MR, Noor Rain A, Badrul AR, Adlin A, Norazah A, Zakiah I. In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. *Tropical Biomedicine*, 2005; 22: 165-170.