



THE LEVEL OF IL-10, IL-17 AND S.IGA IN PATIENTS WITH *ENTAMOEBA HISTOLYTICA*

*Hwaida SH. AL-Mahdawy, *Dr. Farhan Abood Risan, **Dr. Khalil I. Abd Mohammed

*College of Health and Medical Technology / Middle Technical University, Iraq.

**College of Dentistry / Baghdad University/Baghdad, Iraq.

*Author for Correspondence: Dr. Hwaida SH. AL-Mahdawy

College of Health and Medical Technology / Middle Technical University, Iraq.

Article Received on 29/10/2015

Article Revised on 20/11/2015

Article Accepted on 11/12/2015

ABSTRACT

The study was carried out during the period from the beginning of (November / 2013 - November / 2014) for detection of *Entamoeba histolytica* in patients with age range from (3-60) year who attended to AL-Yarmouk teaching hospital and AL-Tifil central hospital. The diagnosis done by microscopic examination. A total of 200 suspected patient there was 120 infected with the parasite diagnosed by the direct examination method, a blood sample was taken from each one, as well as (60) healthy controls were involved. The study included the measurement of Interleukin -10, Interleukin-17 and Secretory- IgA (S.IgA) activities by Enzyme linked immuno sorbent assay (ELISA). The results indicated: The prevalence of *Entamoeba histolytica* by using microscopic examination was 145 (72.5%) in comparison to 120 (82.75%) by using Triage (Micro parasite panel test). The level of Interleukin -10, Interleukin-17 and Secretory IgA increased significantly ($P < 0.05$) in sera patients in comparison to healthy control, but there is no-significant ($P > 0.05$) differences between the gender in both groups.

KEYWORDS: *Entamoeba histolytica* , IL-10 , IL-17 , S.IgA.

INTRODUCTION

Entamoeba histolytica is a protozoan parasite that causes amoebic dysentery and liver abscess. The disease is still one of the major health problems and predominantly affects individuals of lower socioeconomic status who live in developing countries.^[1, 2, 3] Infections can be intestinal, extra intestinal, or both. Most cases are intestinal and asymptomatic. Symptoms, when occur, are multiple and varied, ranging from mild abdominal discomfort and diarrhea (often with blood and mucus) alternating with periods of remission or constipation, to severe illness with fever, chills, and significant bloody or mucoid diarrhea ("amoebic dysentery"). Amoebic colitis may be confused with inflammatory bowel disease such as ulcerative colitis.^[4]

IL-10 is encoded by the IL-10 gene^[5]. IL-10 signals through a receptor complex consisting of two IL-10 receptor-1 and two IL-10 receptor 2 proteins.^[6] Consequently, the functional receptor consists of four IL-10 receptor molecules. IL-10 binding induces STAT3 signalling via the phosphorylation of the cytoplasmic tails of IL-10 receptor 1 + IL-10 receptor 2 by JAK1 and Tyk2 respectively.^[6] In humans, IL-10 is encoded by the IL10 gene, which is located on chromosome 1 and comprises 5 exons^[5] and is primarily produced by monocytes and to a lesser extent lymphocytes, namely

type 2 T helper cells (Th2), mastocytes, CD4⁺ CD25⁺ Foxp3⁺ regulatory T cells, and in a certain subset of activated T cells and B cells. IL-10 can be produced by monocytes upon PD-1 triggering in these cells^[7]. IL-10 is released by cytotoxic T-cells to inhibit the action of NK cells during the immune response to viral infection.^[5]

Interleukin 17 is a cytokine that acts as a potent mediator in delayed-type reactions by increasing chemokine production in various tissues to recruit monocytes and neutrophils to the site of inflammation, similar to Interferon gamma. IL-17 is produced by T-helper cells and is induced by IL-23 which results in destructive tissue damage in delayed-type reactions.^[8] Interleukin -17 as a family functions as a pro inflammatory cytokine that responds to the invasion of the immune system by extracellular pathogens and induces destruction of the pathogen's cellular matrix. Interleukin 17 acts synergistically with tumor necrosis factor and interleukin-1.^[9] IgA is produced in mucosal linings than all other types of antibody combined.^[10] Between three and five grams are secreted into the intestinal lumen each day.^[11] This accumulates up to 15% of the total immunoglobulin produced in the entire body.^[12]

IgA has two subclasses (IgA1 and IgA2) and can exist in a dimeric form called secretory IgA (sIgA). In its

secretory form, IgA is the main immunoglobulin found in mucous secretions, including tears, saliva, sweat, colostrums and secretions from the genitourinary tract, gastro intestinal tract, prostate and respiratory epithelium. It is also found in small amounts in blood. The secretory component of sIgA protects the immunoglobulin from being degraded by proteolytic enzymes, thus sIgA can survive in the harsh gastrointestinal tract environment and provide protection against microbes that multiply in body secretions.^[13] S.IgA can also inhibit inflammatory effects of other immunoglobulins.^[14]

MATERIALS AND METHODS

Studied groups

The study carried out during the period from (November 2013- November 2014), the age of patients extended from (3 – 60) years, two studied groups were involved:

- Suspected patients: Blood and stool samples were obtained from a total of 200 patients clinically suspected with amoebic dysentery that had been examined and defined as suspected cases by specialized physician; the samples were collected from (Al-Yarmouk teaching hospital & Al-Tifl central hospital) in Baghdad.

- Healthy Control: Blood and stool samples from a total 60 healthy control group were involved from Al-Yarmouk teaching hospital staff and from different places in Baghdad; they were examined and defined as healthy, with no history of amoebic dysentery.

Samples collection

Stool sample from each patient was collected in a clean, dry tight cover container and examined with a half an hour. The samples were examined for the presence of *E. histolytica*.

Stool sample examination

Macroscopic examination

It was performed by observing the consistency of stool, presence of blood, mucous and other substances.

Microscopic examination

For each stool sample, wet mount preparation slide was examined by clean, dry slides by obtaining one drop of normal saline and small amount of stool from different places of stool by using clean wooden stick, especially when blood or mucous were noticed, then mixed gently with normal saline and covered with cover slip, the slide

was examined under the low (10x) and high power (40x) of microscope.^[15]

Blood samples

Five mL of Venous blood was obtained from each patient and collected in sterilized screw cap plastic tube, blood samples were left for 30 min. at room temperature, then centrifuge at 3000 rpm for five minute, then the serum for each sample was collected in Eppendorf tubes and stored in deep freeze at -20°C until the time for using. The current study included some Immunological aspects: One hundred twenty clinical patients of *E.histolytica* and (60) healthy control involved in the study. The level of IL-10, IL-17 and S.IgA examined by Enzyme Linked Immunosorbant Assay (ELISA) according to.^[16]

Statistical analysis

The statistical Analysis (T - test) was used to compare between means in studied groups according to.^[17]

RESULTS AND DISCUSSION

Interleukin -10 (IL-10)

The level of IL-10 show significant difference ($P < 0.05$) in patients and control, while the results showed non-significant difference ($P > 0.05$) between the gender in both groups. The level was (491.93 ± 12.45) mg/mL, (460.20 ± 14.47) mg/mL in males and females of patients group respectively and (110.05 ± 13.33) mg/mL, (187.33 ± 18.01) in males and females of control group respectively Table (1).

The result in line with ^[18] who found that the malnutrition decreases T-cell function and cytokines production, also the ability of lymphocytes, to respond appropriately to cytokines. Another study done by ^[19], which detected that IL-10 serum levels were significantly increased only in amoebic liver abscess patients, but not in other groups, the invasion of the liver tissue by *E.histolytica* elicits the anti-inflammatory immune response. IL-10 is capable of inhibiting synthesis of pro-inflammatory cytokines such as IFN- γ , IL-2, IL-3, TNF α and GM-CSF made by cells such as macrophages and regulatory T-cells. It also displays a potent ability to suppress the antigen-presentation capacity of antigen presenting cells; however, it is also stimulatory towards certain T cells (Th2) and mast cells and stimulates B cell maturation and antibody production.^[20]

Table 1: The level of IL-10 (pg /mL) in serum of *E.histolytica* patients and healthy control

Group	Mean $\pm$ S.D Gender	
	Male	Female
Patients	491.93 $\pm$ 12.45	460.20 $\pm$ 14.47
Control	110.05 $\pm$ 13.33	187.33 $\pm$ 18.01
T-test value	38.51 *	50.06 *

* ($P < 0.05$)

Interleukin -17(IL-17)

The level of IL-17 in patients serum infected with *E.histolytica* showed significant increasing ($P<0.05$) in comparison with healthy control, while the results between the gender showed no significant difference ($P>0.05$) in both groups. The level was (326.66 ± 5.58) mg/mL, (266.93 ± 9.47) mg/mL in males and females of patients group respectively and (225.04 ± 10.88) mg/mL, (192.93 ± 12.29) mg/mL in males and females of healthy control group respectively Table (2).

These results were agreed with another study ^[21] who found that persistent *E.histolytica* infection is associated with elevated levels of Th-17 cytokines. The present study is disagreeing with other study shows that IL-17 exhibited little effect during the infection with *E.histolytica*.^[22] The mechanism of IL-17 mediated protection involves neutrophil recruiting to sites, regulation of dendritic cells function and Th1 responses through cytokines and chemokines induced by IL-17.^[23,24]

Table 2: The level of IL-17 (pg /mL) in sera of *E.histolytica* patients and healthy control group

Group	Mean $\pm$ S.D Gender	
	Male	Female
Patients	326.66 $\pm$ 5.58	266.93 $\pm$ 9.47
Control	225.04 $\pm$ 10.88	192.93 $\pm$ 12.29
T-test value	21.88 *	33.08 *

*($P<0.05$)

Secretory- IgA

The level of secretory IgA showed significant increasing ($P<0.05$) in patients with *E.histolytica* and healthy control, while the results indicated no significant difference ($P>0.05$) in the gender of both groups. The level of secretory- IgA was (5.46 ± 0.03) mg/mL, (5.42 ± 0.03) mg/mL in males and females respectively of patients group and (3.34 ± 0.04) mg/mL, (3.23 ± 0.03) mg/mL in males and females respectively of healthy control group Table (3) & Figure (3).

These results were in agree with previous study denoted that the highest level of specific secretory anti-amoebic IgA were observed in the patients infected with intestinal amoebiasis and liver amoebic abscess ^[25], another study was done by ^[26, 27] which found that in the infection with *E.histolytica* antigen specific secretory

immunoglobulin A (IgA) antibodies have been found to mediate protection against intestinal infection by *E.histolytica*. Secretory -IgA serves as lines of defense against microorganisms through a mechanism called immune exclusion; S.IgA adherence selectively to Microfold cell (M. cells) in intestinal Peyer's patches, thus mediating the trans epithelial transport of the Ab molecule from the intestinal lumen to underlying gut-associated organized lymphoid tissue.

In Peyer's patches, S.IgA bind and is internalized by dendritic cells in the sub- epithelial dome region. When used as carrier for antigens in oral immunization. S.IgA induces mucosal and systemic responses associated with production of anti-inflammatory cytokines and limits activation of dendritic cells.^[28]

Table 3: The level of S.IgA (µg /mL) in sera of *E.histolytica* patients and healthy control group

Group	Mean $\pm$ S.D Gender	
	Male	Female
Patients	5.46 $\pm$ 0.03	5.42 $\pm$ 0.03
Control	3.34 $\pm$ 0.04	3.23 $\pm$ 0.03
T-test value	0.109 *	0.115 *

*($P<0.05$)

REFERENCES

- Simonetta, G.; Giovanni, S.; Francico, R. and Mariella, A. Amebic infections due to the Entamoeba histolytica – Entamoeba dispar complex. A study of the incidence in a remote rural area of Ecuador. Am. J. Trop. Med. Hyg., 2002; 67: 123-127.
- ElBakri, A.; Samie, A.; Ezzedine, S. and Abu Odeh, R. Differential detection of Entamoeba histolytica, Entamoeba dispar and Entamoeba moshkovskii in fecal samples by nested PCR in the United Arab Emirates (UAE).J. Sprig. Intern. Pub., 2013; 58(2): 185-190.
- Zahida, T.; Mushtaq, H.; Lashari, A. and Fariha, A. Institute of Pure and Applied Biology, Bahauddin Zakariya University, Multan. J. Cell Anim. Biol., 2013; 7(6): 73-76.
- Guide to Surveillance, Reporting and control. Amoebiasis. Massachusetts Department of public Health, Bureau of Communicable disease Control., 2006; 29-35.
- Eskdale, J.; Kube, D.; Tesch, H. and Gallagher, G. Mapping of the human IL -10 gene and further characterization of the 5'flankingsequence. Immunogenetics, 1997; 46(2): 1208.

6. Mosser, D.M. and Zhang, X. Interleukin-10: new perspectives on an old cytokine. *Immunol. Rev.*, 2008; 226(1): 205–18.
7. Said, E. A.; Dupuy, F. P.; Trautmann, L.; Zhang, Y.; Shi, Y.; El-Far, M.; Hill, B. J.; Noto, A.; Ancuta, P.; Peretz, Y.; Fonseca, S. G. Programmed death-1-induced interleukin-10 production by monocytes impairs CD4⁺ T cell activation during HIV infection. *Nat. Med.*, 2010; 16(4): 452–9.
8. Kuby, T. Janis; Kindt, T. J.; Goldsby, R. A. and Osborne, B. A. (2007). *Kuby immunology*. San Francisco: W.H. Freeman. p. 396. ISBN 1-4292-0211-4.
9. Chiricozzi, A.; Guttman-Yassky, E.; Suárez-Fariñas, M.; Nograles, K.E.; Tian, S.; Cardinale, I.; Chimenti, S. and Krueger, J.G. J. Invest. Dermatol., Integrative responses to IL-17 and TNF- α in human keratinocytes account for key inflammatory pathogenic circuits in psoriasis. *J. Inves. Dermatol*, 2011; 131(3): 677-87.
10. Fagarasan, S. and Honjo, T. Intestinal IgA Synthesis: Regulation of Front-line Body Defenses. *Nature Reviews Immunology*, 2003; 3(1): 63–72.
11. Brandtzaeg, P.; and Pabst, R. Let's go mucosal: communication on slippery ground. *Trends Immunology*, 2004; 25(11): 570–577.
12. Macpherson, A. J. and Slack, E. The functional interactions of commensal bacteria with intestinal secretory IgA. *Current Opinion in Gastroenterology*, 2007; 23(6): 673–678.
13. Junqueira, L. C. and Jose, C. (2003). *Basic Histology*. McGraw-Hill. ISBN. 0-8385-0590-2.
14. Holmgren, J.; Czerkinsky, C. Mucosal immunity and vaccines. *Nature Medicine*, 2005; 11(4s): S45–S53.
15. Frances, F. and Marshall B. D. (2009). *A Manual of Laboratory and Diagnostic Tests*. Lippincott William & Wilkins. 8th ed. 286-290.
16. Cusabio Biotech. (2007-2015 www.cusabio.com).
17. SAS., (2012). *Statistical Analysis System, User's Guide*. Statistical. Ver. 9.1 th ed. SAS. Inst. Inc. Cary. N.C.USA.
18. Soboslay, P. T.; Hamm, D. M.; Pfafflin, F.; Fendt, J.; Banla, M.; Banla, M. and Schulz-Key, H. Cytokines and chemokines responses in patient's co-infected with *Entamoeba histolytica* / *dispar*, *Necator americanus* and *Mansonella perstans* and changes after anti-parasite treatment. *Microbes. Infect.*, 2009; 8(1): 238-247.
19. Hannah, B.; Claudia, M.; Thomas, J.; Norbert, B.; Levan, A.; Jörg. B. and Hannelore, L. Immune markers characteristic for asymptotically infected and diseased *Entamoeba histolytica* individuals and their relation to sex. *BMC. Infect. Dis.*, 2014; 14: 621.
20. Sikka, G.; Miller, K. L.; Steppan, J.; Pandey, D.; Jung, S. M.; Fraser, C. D.; Ellis, C.; Ross, D. Interleukin - 10 knockout frail mice develop cardiac and vascular dysfunction with increased age. *Experimental Gerontology*, 2013; 48(2): 128–35.
21. Guo, X.; Stroupt, S. E. and Houpt, E. R. (2008). Persistence of *Entamoeba histolytica* infection in CBA mice owes to intestinal IL-4 production and inhibition of protective IFN γ . *Mucosal Immunol*, 2008; 1(2): 139-146.
22. Xiaoti, G.; Lisa, B.; David, M.; William, A.; Petri, J. and Eric, R. (2011). CD⁺ and CD⁺ T cell- and IL-17- mediated protection against *Entamoeba histolytica* induced by a recombinant vaccine. *NIH-P manuscript. USA*. 17; 29(4): 772-777.
23. Bai, H.; Cheng, J. and Gao, X. (2009). IL-17 / Th 17 promote type 1 T cell immunity against pulmonary intracellular bacterial infection through modulating dendritic cell function. *J. Immunol*. 2009; 183(9): 5886-5895.
24. Zhang, X.; Gao, L.; and LEI, L. (2009). A My D88-dependent early IL-17 production protects mice against airway infection with the obligate intracellular pathogen *Chlamydia muridarum*. *J. Immunol*. 15; 183 (2): 1291-300.
25. Sehgal, R.; Devi R.; Singh, K.; Ganguly, N. K. and Mahajan, R. C. Secretory - IgA as a marker of invasive amoebiasis. *RIF*, 2010; 1(5): 235-238.
26. Haque, R.; Duggal, P. and Ali, I. M. (2002). Innate and acquired resistance to amebiasis in Bangladeshi children. *J. Infect. Dis.*, 2002; 186: 547-52.
27. Ravidin, J.; Abd-Alla, S.; Wells, S.; Reddy, S. and Jackson, T. Intestinal anti lectin immunoglobulin - Antibody response and immunity to *Entamoeba dispar* infection following cure of amebic liver abscess. *Infect. Immun.*, 2003; 71: 6899-6905.
28. Blaise C., Round trip Ticket for Secretory IgA: Role in Mucosal Homeostasis. *J. Immunol*, 2007; 178 (1): 27-32.