



## HERBS AND PHYTOCHEMICALS WITH ANTIPARKINSONIAN ACTIVITY

**Rakesh K. Goyal\* and Janardhan Singh**

\*Faculty of Pharmaceutical Sciences, Pt. B.D. Sharma University of Health Sciences, Rohtak 124 001, India.  
Department of Pharmacology, Pt. B.D. Sharma Post Graduate Institute of Medical Sciences, Rohtak 124 001, India.

**\* Corresponding Author: Rakesh K. Goyal**

Faculty of Pharmaceutical Sciences, Pt. B.D. Sharma University of Health Sciences, Rohtak 124 001, India.

Article Received on 10/05/2016

Article Revised on 30/05/2016

Article Accepted on 20/06/2016

### ABSTRACT

Parkinson's disease, a most common neurodegenerative disorder, is characterized by bradykinesia, muscular rigidity, tremors and impairment of postural balance. Pathophysiology of the disease is not clear so far. Conventional drugs (levodopa / carbidopa, catechol o-methyl transferase (COMT) inhibitors, monoamine oxidase-B (MAO-B) inhibitors, etc) fail to cure the disease and can cause motor fluctuations (on/off phenomenon) along with other serious side effects. Herbal plants and their phytochemicals might potentially offer a novel neuroprotective approach; and are reported to produce beneficial effects in this disease. The present review describes some plants and their extracts & phytochemicals exhibiting beneficial pharmacological properties relevant to Parkinson's disease treatment.

**KEYWORDS:** Parkinsonism, Medicinal plants, Levo dopa, Macuna pruriens, Genistein.

### INTRODUCTION

Parkinson's disease (PD) is the second most common neurodegenerative disorder after Alzheimer's disease, affecting 1% of the population over the age of 65 years and 4-5% of the population over the age of 85 years.<sup>[1,2]</sup> The pathophysiological hallmark of PD is the loss of the pigmented dopaminergic neurons in the substantia nigra pars compacta region of midbrain that culminates in the major clinical symptoms of PD.<sup>[3]</sup> Progressive loss of dopamine containing neurons is a feature of normal aging, however, most people do not lose the 70-80% of dopamine neurons required to cause symptomatic PD. Without treatment PD progresses over 5-10 years to a rigid, akinetic state. Death frequently results from complications of immobility. PD is characterized by bradykinesia, muscular rigidity, tremors, and impairment of postural balance leading to disturbances of gait.<sup>[3]</sup> Other non motor features include sleep disorders, depression, cognitive impairment, mood fluctuations,<sup>[4]</sup> psychosis and dementia.<sup>[5]</sup> Treatments of PD include levodopa, dopamine receptor agonists, catechol o-methyl transferase (COMT) inhibitors, monoamine oxidase-B (MAO-B) inhibitors, anticholinergics and amantadine.<sup>[6]</sup> Levodopa dramatically improves the motor symptoms of PD and remains the gold standard antiparkinsonian treatment.<sup>[7]</sup> However, its chronic use is frequently associated with dyskinesia or motor fluctuations (on/off phenomenon). Current treatments are mainly symptomatic and can temporarily slow down disease progression, but can not halt this. To date there is a lack of effective preventive strategies for PD. Therefore, safe

and effective treatment strategies are urgently required for management of PD. Plant extracts have a wide range of medicinal properties and has been used to treat many types of diseases. *Lycium Chinensis* Miller, plant has been used as an anti aging therapy and a treatment of neurodegenerative diseases<sup>[8]</sup> and recent research has confirmed neuroprotective effect of the fruits of plant in a rat model.<sup>[9]</sup> Herbal plants and their phytochemicals might potentially offer a novel neuroprotective approach in a neurodegenerative diseases and might be developed for therapeutic use. This review describes some plants and their extracts & phytochemicals exhibiting beneficial pharmacological properties relevant to PD treatment.

### METHODS

**Data Collection:** The data for present review was collected from internet, online journals and from various Ayurvedic texts like database on medicinal plants used in Ayurveda, central council for research in Ayurveda and Sidha, Department of ISH & H, Ministry of Health & Family Welfare, Govt. of India, New Delhi, Volumes 1-8, Reviews on medicinal plants, Volumes 1-9 (ICMR, New Delhi). Most of the papers reviewed herein pertinent to herbal medicine research were published in internationally recognized peer reviewed journals. Some of the medicinal plants / extracts and their isolated active phytochemicals reported to be useful for the prevention and treatment of neurodegenerative disorders particularly parkinson's disease are:

**1. *Bacopa monnieri* L (Scrophulariaceae)** Eng- Thyme leaved gratiola, Hindi- Brahmi. Small creeping, marshy with branched 10-25 cm long. Distributed throughout India, ascending to an altitude of 1320 m in marshy places, Bangladesh, Pakistan and Sri Lanka. Alcoholic extract of the plant increased learning performance in rats and activity attributed to saponin mixture containing bacosides A, B & others. Clinical reports showed antianxiety and adaptogenic effects. It is a brain tonic<sup>[10]</sup> and possess antiapoptotic,<sup>[11]</sup> antioxidant<sup>[12]</sup> and memory enhancing properties. *B. monnieri* has been shown to reduce level of oxidative stress in fruit flies and thereby inhibiting dopamine depletion with decreased mortality rate.<sup>[13]</sup> In a *Caenorhabditis elegans* transgenic model of PD, positive results (decreased alpha synuclein protein, preventive dopaminergic neurodegeneration and restoring lipid contents) has been reported in *B. monnieri* treated 6-hydroxydopamine (6-OHDA) models of Parkinson's disease.<sup>[14]</sup> Therefore, *B. monnieri* has its proven potential as an antiparkinsonian agent.

**2. *Camellia sinensis* Linn (Theaceae)** Eng- Tea plant, Hindi- Chai. A variable evergreen shrub or a small tree found in Assam and hilly regions to the East and South of it. It is mainly cultivated in the hilly districts of North Bengal & South India. The natural product (-)-Epigallocatechin-3-gallate (EGCG) is the major polyphenolic bioactive component of *C. sinensis* and has been recognized as potent neuroprotective agent against oxidative stress, neuroinflammation, protein aggregation, autophagy and neuronal cell death in vitro as well as in vivo.<sup>[15]</sup> Anti-inflammatory and antioxidant activity of *C. sinensis* has been reported by many authors.<sup>[16]</sup> EGCG administered in mice (25 mg/kg, po) prevented loss of dopaminergic neurons in the substantia nigra and preserved striatal levels of dopamine.<sup>[17]</sup> EGCG prevented the accumulation of iron and alpha-synuclein in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated mice. These effects have been attributed to the antioxidant activity and iron-chelating properties of EGCG, respectively.<sup>[18]</sup> Pinto et al 2015<sup>[19]</sup> have demonstrated neuroprotective effect of *C. sinensis* on the striatal 6-OHDA model of PD in rats by decreased rotational behavior, increased locomotor activity, antidepressive effects, and improvement of cognitive dysfunction, as compared to the untreated 6-OHDA-lesioned group. Further, they showed that *C. sinensis* reversed, at least partly, the behavioral changes observed in the untreated 6-OHDA lesioned animals, at very low doses and reversed the decreased locomotor activity observed in the untreated lesioned group; and similar results had been reported with the forced swimming test and water maze test, indicative of antidepressant and spatial learning effects, respectively. EGCG was found to have a potential therapeutic effect against lipopolysaccharide induced neurotoxicity in male Sprague Dawley rats via reducing TNF $\alpha$  and NO inflammatory mediators and preserving dopamine (DA) level in midbrain.<sup>[20]</sup>

**3. *Centella asiatica* Linn (Apiaceae)** Eng- Indian Penny Wort, Hindi- Mandookaparni. A perennial herb commonly found in moist areas and in crop fields and other waste places throughout India upto an altitude of 600m. Results of a double blind trial of Mandookaparni on mentally retarded children showed a very significant increase in general ability and behavioural pattern when drug was given for a short period of 12 weeks.<sup>[21]</sup> Traditionally *C. asiatica* has been used as a brain tonic in Ayurveda & Chinese medicine. *C. asiatica* has been shown to possess antioxidative effect and reduce mitochondrial dysfunction induced by fungal neurotoxin (3-nitro propionic acid) in mice by affective malondialdehyde (MDA) and radical oxygen species.<sup>[22]</sup> In experimentally induced PD in rats, *C. asiatica* extract protected the mitochondrial damage. Chloroform methanol extract of *C. asiatica* showed free radical scavenging effect in monosodium glutamate (MSG) stressed rats.<sup>[23]</sup> Neuroprotective effect of *C. asiatica* extract has also been reported in old aged rat brain via decreasing protein carbonyl content and lipid peroxidation. Aqueous extract of *C. asiatica* has been shown to reverse the neurotoxic effect in MPTP-induced Parkinsonism in aged Sprague- Dawley rats.<sup>[24]</sup>

**4. *Erigeron breviscapus*.** Lopez & Calvo 2011<sup>[25]</sup> demonstrated neuroprotective effects of the *Erigeron breviscapus* and *C. sinensis* against hydrogen peroxide induced toxicity in PC-12-cells.

**5. *Ginkgo biloba* L (Ginkgoaceae)** Eng-Maiden hair tree, Hindi- Balkuwari. Tree with pyramidal form, reaching a height of 30 m; leaves petiolar, lamina fan-shaped, bilobed; dioecious; mature seeds orange-coloured and are about the size of an apricot. *Ginkgo biloba* has been shown to possess neuroprotective, antioxidative and iron chelating properties in 6-hydroxydopamine (6-OHDA) induced parkinsonian in rats.<sup>[26]</sup> EGb761 is a standardized extract of the leaves of the *G. biloba* tree and is characterized by its main fractions, the flavonols (mainly isorhamnetin, kaempferol and quercetin: 22–27%) and the terpene lactone (5–7%), These two fractions are thought to be, at least partly, responsible for potential neuroprotective properties of EGb761.<sup>[27]</sup> Other active constituents comprises ginkgolides ABC & M, sesquiterpenic trilactone and bilobalide. Bilobalide reduce damage caused by global brain ischemia & glutamate-induced excitotoxic neuronal death.<sup>[28]</sup> Bilobalide has also been reported significantly to restore the behavioral changes induced by 6-OHDA and to inhibit loss of tyrosine hydroxylase-positive neurons, decreased the activation of NF-kappaB, and protected dopaminergic neurons from apoptosis in 6-OHDA induced rat model of PD significantly.<sup>[29]</sup> EGb761 also inhibited oxidative stress induced by MPTP model of parkinsonism in mice<sup>[30]</sup> and significantly decreased drug induced rotation and produced significant restoration of striatal DA and its metabolites in rats,<sup>[31]</sup> the beneficial effects of EGb761 in parkinsonism may be due to its potent MAO inhibitory activity which prevents

degradation of DA and increase its availability. When given concurrently, EGB761 reduced neurotoxic effects of levodopa.<sup>[32]</sup> Presently standardized extracts are widely prescribed in Europe & US for symptomatic treatment of Alzheimer's disease, cerebral insufficiency & improvement of memory. A clinical trial concluded that the *G. biloba* extract EGB 761, specifically in a dose of 240 mg daily, was both safe and effective in the treatment of patients with dementia associated with neuropsychiatric features.<sup>[33]</sup>

**6. *Ganoderma lucidum*** is a fungus that grows from the tops of stumps or submerged logs and is found in most parts of the world. Extract of *Ganoderma lucidum* inhibited Staurosporine induced apoptosis by 30-50% in a dose dependent manner. The oil from *ganoderma lucidum* spores showed neuroprotective effect on pathological changes in the substantia nigra and behaviors of MPTP induced model of PD in mice. Treatment with ganoderma spores oil increased survival of dopamine neurons in the substantia nigra and levels of dopamine in the striatum, attenuated involuntary motor symptoms.<sup>[34]</sup> *Ganoderma lucidum* extract has been shown to protect dopaminergic neurons through inhibiting the production of inflammatory mediators by activated microglia.<sup>[35]</sup> Neuroprotective effect of extract may be due to free radical scavenging properties of this plant.<sup>[36]</sup>

**7. *Glycyrrhiza glabra* Linn (Fabaceae) Eng-** Liquorice, Hindi- Mulhatti. It is a hardy herb attaining height upto 2m. It is distributed in the subtropical and warm temperate regions of the world. In India it is reported to be cultivated in Baramulla, Srinagar, Jammu, Dehradun, Delhi & South India. Pharmacological studies demonstrated antioxidant, anti-inflammatory & neuroprotective properties of this plant.<sup>[37]</sup>

**8. *Lycium chinense* Mill (solanaceae):** Common name Wolfberry, It is a deciduous perennial shrub to 5' tall. *L. chinensis* Miller, a traditional herbal medicine used in China, Korea and Japan has been shown to have hypotensive, hypoglycemic and antipyretic effects in animal studies. Aqueous methanol extract of *L. chinense*, exhibited inhibitory effect on MAO-B in rat brain homogenates and therefore has been proposed as a good candidate for use in delaying the progressive degeneration caused by neurological diseases.<sup>[38]</sup> This plant has also been used as an antiaging therapy and a treatment of neurodegenerative diseases.<sup>[8]</sup> Recently neuroprotective effect from the fruit of this plant in a rat model of trimethyltin induced learning and memory impairment has been confirmed.<sup>[9]</sup> *Lycium chinense* Miller extracts have been shown to produce beneficial effects in Parkinson's disease by attenuating rotenone induced toxicity in PC12 cells.<sup>[39]</sup>

**9. *Macuna pruriens* Linn (Fabaceae) Eng-** Common cowitch, Hindi- Kaunch Herbaceous twining annuals, pods 5-10 cm long curved, distributed all over India upto

100m in Himalyas & in Andaman, Nicobar islands. It has been used for treating parkinsonism in Ayurvedic system of medicine.<sup>[40]</sup> Studies showed that seeds of *M. pruriens* contain levodopa as one of the active constituents. Pharmacological studies demonstrated that L-dopa free fraction of seed exhibited potent antiparkinsonian effect in mice (200mg/Kg, ip). In a clinical trial in 62 patients with parkinson's disease (46 males & 16 females, mean age 59±9 years) treated for 12 weeks with HP200 powder (7.6g orally). Statistically significant reductions in Hoehn & Yahr stage and Unified Parkinson's Disease Rating Scale (UPDRS) scores were seen from base line to the end of the 12 weak treatment.<sup>[41]</sup> *M. pruriens* has been shown to antagonize motor symptoms of parkinson's disease in 6-OHDA induced rat model of PD.<sup>[42]</sup> *M. pruriens* reported to reduce dyskinesia in MPTP monkey model<sup>[43]</sup> and hemi parkinsonian rat model.<sup>[44]</sup> It has shown to be as effective as pure levodopa /carbidopa in the treatment of Parkinson's disease.<sup>[45]</sup> In 1978, Vaidya et al.<sup>[46]</sup> reported the beneficial effects of *M. pruriens* in patients with PD in an open clinical trial, however no further information is available.

**10. *Panax ginseng* C.A. Mayer (Araliaceae) *P. ginseng*** root is the most popular traditional medicine in China, Korea and Japan. *P. ginseng* has been reported to possess neuroprotective, antioxidant, antiapoptotic and immunostimulatory properties and extracts have been reported to protect against neurotoxicity in vitro and in vivo models of PD. *P. ginseng* extract G115 blocked the loss of tyrosine hydroxylase (TH) (+) cells in substantia nigra and reduced locomotor dysfunction in MPTP induced PD model C57BL/6 mice and rats.<sup>[47]</sup> In an in vitro study, ginseng saponins have shown positive effects of enhancing neurite growth of dopaminergic SK-N-SH neuroblastoma cells. Ginsenosides (Rb1 and Rg1) have been demonstrated to increase neuritic growth in 1-methyl-4-phenylpyridinium (MPP+) or glutamate stressed primary cultured mesencephalic dopaminergic cells in vitro.<sup>[48]</sup> Ginseng has been suggested to inhibit both N-methyl-D-aspartate (NMDA) and non-NMDA glutamate receptors resulting in reduction of Ca<sup>2+</sup> over-influx into neurons and thus protecting the cells from neurodegeneration mediated via Ca<sup>2+</sup> overload.<sup>[49]</sup> Ginseng has also been reported to attenuate dopamine-induced apoptotic cell death through suppression of intracellular oxidative stress and therefore, rescue neurons against apoptosis induced by exogenous dopamine in PC12 cells.<sup>[50]</sup> Ginseng exerted neuroprotective effect in experimental PD model in vitro in which SH-SY5Y cells were injured by MPP+ and in MPTP induced in vivo model of mouse.<sup>[51]</sup>

**11. *Peltophorum africanum* (Fabaceae) (Weeping wattle).**It is a tree with a dense rounded to spreading crown, greyish stem, fern-like stipules and clustered fruits and is widespread in south Africa and most tropical areas. Bizimanyera et. al.<sup>[52]</sup> reported neuroprotective and

antioxidant properties from the bark and root extracts of *P. africanum*.

**12. *Polygonum cuspidatum*** (Polygonaceae) inhibited staurosporin induced apoptosis in dose dependent manner.<sup>[36]</sup>

**13. *Schizandra chinensis*** (Magnoliaceae). Extracts of *S. chinensis* inhibited staurosporin induced apoptosis by 30-50% in a dose dependent manner. The extract showed 1. neuroprotective, antioxidant & free radical scavenging activities.<sup>[36]</sup> Schisantherin A, a dibenzocyclooctadiene lignan from the fruit of *S. chinensis* protects 6-OHDA-induced dopaminergic neuron damage in zebrafish and cytotoxicity in human neuroblastoma SH-SY5Y cells through the ROS/NO and AKT/GSK3 $\beta$  pathways.<sup>[53]</sup>

**14. *Smilacis chiniae*** rhizome exhibited a neuroprotective effect in an in vitro model of N-methyl D aspartate (NMDA) induced neurotoxicity. It showed a similar effect in an in vivo model of focal cerebral ischemia.<sup>[54]</sup>

**15. *Scutellaria baicalensis*** Geogri (Lamiaceae). *Scutellaria baicalensis* Geogri (SBG) is native of China and the dried roots are rich in flavones such as baicalin, baicalein and wogonin. Baicalin and baicalein flavones from *S. baicalensis* have been shown to possess antioxidant, anti-inflammatory and cognitive enhancing properties.<sup>[55]</sup> Marked reduction in tremors, mitigation of astroglial response and increased tyrosine hydroxylase positive neurons in substantia nigra have been reported in 6-OHDA lesioned rat models in vivo and in vitro.<sup>[56]</sup> Mu et al 2011<sup>[57]</sup> demonstrated similar results in MPTP mice models of PD along with inhibition of dopamine turnover. Further, baicalein has also been reported to increase dopamine and 5HT levels in the striatum in MPTP mice models of PD.<sup>[58]</sup> Baicalein has been shown to prevent 6-OHDA-induced oxidative damage in PC12 cells by activating Keap1/Nrf2/HO-1 channels, as well as PKC $\alpha$  and PI3K/AKT signal channels.<sup>[59]</sup>

**16. *Withania somnifera*** (L) Dunal Eng- Winter cherry, Indian ginseng, Hindi- Ashwagandha. Erect tomentose shrub, 30-150 cm high. It is found throughout the drier parts of India in waste places and on bunds in areas of upper Gangetic plain, West Bengal, Bihar, Orissa, Gujarat, Konkan, Karnataka & Coimbatore. *W. somnifera* fruit extract has been shown to possess antiaging, antioxidant, free radical scavenging, adaptogenic and immunomodulatory properties.<sup>[60]</sup> Antiparkinsonian effect of *W. somnifera* extract was evaluated in 6-OHDA induced parkinsonism in rats. Treatment with *W. somnifera* extract reversed some symptoms of PD such as decreased striatal dopamine level, increased lipid peroxidation, increased dopaminergic D<sub>2</sub> receptors, reduced nigral glutathione level, reduced activity of SOD, Catalase and lessened tyrosine hydroxylase expression.<sup>[61]</sup> Neuroprotective effects of *W. somnifera* has also been reported in maneb paraquat induced parkinsonism in mice.<sup>[62]</sup>

Administration of *W. somnifera* root extract in MPTP mice model of PD caused increase in the level of dopamine, 3, 4- dihydroxy phenylacetic acid (DOPAC), homovanillic acid (HVA), glutathione, glutamine peroxidase and normalized levels of lipid peroxidation marker i.e. thiobarbituric acid reactive substance (TBARS).<sup>[63]</sup>

### Neuroprotective phytochemicals

**1. Curcumin** (diferuloylmethane), the well-known component of yellow curry spice derived from the rhizome of *Curcuma longa* L (Zingiberaceae) Eng-Turmeric Hindi- Haldi, has been used as a food preservative and herbal medicine in India for hundreds of years. Rhizome of *Curcuma longa* L contains curcuminoids (curcumin, demethoxy curcumin & bis demethoxy curcumin) as its active phytochemical. Curcuminoids possess anti-inflammatory, antioxidant, proapoptotic, antiproliferative, wound healing and antiparkinsonian effects. Several studies in cellular and animal models indicated that curcumin is a neuroprotective agent in neurodegenerative disorders such as PD.<sup>[64, 65]</sup> Curcumin has been reported to protect substantia nigra (SN) neurons, improves striatal dopamine levels and chelates Fe<sup>2+</sup>, in 6-OHDA administered rats.<sup>[66]</sup> Curcumin produces increase in the density of dopaminergic neurons in the SN.<sup>[67]</sup> Mythri et al. 2011<sup>[68]</sup> has observed that chronic dietary consumption of turmeric caused an increase in the tyrosine hydroxylase (TH) positive neurons in the SN, in consistent with earlier reports of Kim et al. 2008,<sup>[69]</sup> and Xu et al. 2007<sup>[70]</sup> who reported that curcumin contributes to neurogenesis. Apart from MPTP model, curcumin has been found to be neuroprotective in 6-OHDA-induced hemiparkinsonian mice model where 6-OHDA-induced loss of striatal TH fibers and nigral TH-immunoreactive neurons had decreased.<sup>[71]</sup>

**2. Gastrodin**, the predominant constituent of the rhizome of *Gastrodia elata* Blume (Orchidaceae), a Chinese herbal medicine, has long been used for treating vertigo, general paralysis, epilepsy, tetanus, stroke, dementia and Parkinson's disease. Methanol extract of *Gastrodiae Rhizoma* or the pure compound vanillyl alcohol reduced oxidative stress and cell apoptosis in both SH-SY5Y and MN9D dopaminergic cell lines in response to the damage induced by MPP+.<sup>[72-73]</sup> Gastrodin was shown to reduce the loss of TH-positive cells in a rotenone-induced model of PD by protecting dopaminergic neurons, down-regulating nigral IL-1 $\beta$  expression resulting in a restraining of neuroinflammation during the damage and improved muscle rigidity and endurance in PD rats.<sup>[74]</sup> It has been reported to mobilizes neuro-protective capacities used and is used often for the treatment of headache, convulsions, hypertension and neurodegenerative diseases.<sup>[75]</sup> Gastrodin has been reported to protect dopaminergic neurons in SH-SY5Y cells stressed with MPP+ through regulating free radicals, Bax/Bcl- 2 mRNA, caspase-3, and cleaved poly(ADP-ribose)

polymerase. It also showed neuroprotective effects in MPTP induced mouse PD model by ameliorating bradykinesia and motor impairment in the pole and rotarod tests, respectively.<sup>[76]</sup> Gastrodin prevented motor deficits and oxidative stress in the MPTP mouse model of PD via interrupting extracellular signal regulated protein kinases (ERK) 1/2-nuclear factor erythroid 2-related factor 2 (Nrf2) signaling pathway.<sup>[77]</sup>

**3. Paeoniflorin** (Paeoniaceae) a major active component of Chinese herb *Paeoniae alba* Radix has shown neuroprotective effect in MPTP mouse model of PD (73). Subchronic treatment with Paeoniflorin had shown an alleviating effect on 6-OHDA induced neurological impairment in rats.<sup>[78]</sup>

**4. Resveratrol**, a polyphenolic compound naturally present in red wine, grapes, peanuts, berries & Pine has been shown to possess anti-inflammatory, antiapoptotic, antioxidant and neuroprotective properties.<sup>[79]</sup> Resveratrol has shown neuroprotective effect in Balb/c mice by alleviating HPTP induced motor incoordination, oxidative stress and TH positive neuronal cell loss.<sup>[80]</sup> Reduced oxidative stress, microglial activation, neuroinflammation besides increasing number of TH positive cells and dopamine content have been reported in Pakquat and Maneb models of PD.<sup>[81]</sup> Neuroprotective effects of resveratrol has also been reported in rotenone and 6-OHDA induced dopaminergic cell death.<sup>[82,83]</sup> Therefore, resveratrol protects dopaminergic neurons by reducing inflammation, oxidative stress, diminishing dopamine apoptosis and by altering the expression of CYP2d22 as well as paraquat accumulation.<sup>[81]</sup>

**5. Tenuigenin**, Root of *Polygala tenuifolia* (polygalaceae) a traditional Korean medicine rich in tenuigenin, which is used for treating various cognitive problems associated with ageing and PD.<sup>[84]</sup> Neuroprotective effect of tenuigenin has been demonstrated against 6-OHDA induced cytotoxicity in SHSY54 cells by protecting mitochondrial damage by increasing glutathione, SOD levels and thereby increasing cell viability.<sup>[85]</sup> Tenuigenin protects against lipopolysaccharide induced neuroinflammatory damage in rats by improving TH immunoreactive neurons and dopamine levels in the striatum. Furthermore, lipopolysaccharide induced upgradation of TNF $\alpha$  and IL-1 $\beta$  was also overturned by tenuigenin.<sup>[86]</sup>

#### Miscellaneous agents

**Chrysanthemum morifolium** inhibited MPTP-induced cytotoxicity and maintained cell viability of SH-SY5Y cell line by preventing ROS formation, decreasing Bax/Bcl2 ratio and caspase-3 activation.<sup>[87]</sup>

**Echinoid**, an active compound of *Cistanche salsa* in a dose of 20mg/kg maintained striatal dopamine levels, reduced cell death, significantly increased the tyrosine hydroxylase enzyme expression, and reduced the

activation of caspase-3 and caspase-8 expression in MPTP mouse model of PD.<sup>[88]</sup>

**Silymarin** preserved dopamine levels, diminished the number of apoptotic cells and preserved dopaminergic neurons in SN of MPTP- and 6-OHDA-intoxicated mice.<sup>[89]</sup>

**Anemopaegma mirandum**, a Brazilian tree, the extract of *Anemopaegma mirandum* produced protective effect on Rotenone-induced apoptosis in human neuroblastomas SHSY5Y cells.<sup>[90]</sup>

**Valeriana officinalis** increased the viability of SH-SY5Y cells in rotenone induced in vitro experimental model of PD.<sup>[91]</sup>

**Tripterygium regelii** reduced oxidative stress-induced cell death through the inhibition of apoptotic cascades, preserved mitochondrial function, and promoted tyrosine hydroxylase expression and brain-derived neurotrophic factor (BDNF) production in H<sub>2</sub>O<sub>2</sub> treated SH-SY5Y cells.<sup>[92]</sup>

**Uncaria rhynchophylla** decreased cell death and ROS production; increased GSH levels in cultured PC12 cells, while 6-OHDA-induced caspase-3 activation was attenuated preventing cell death. Rotational behavior was significantly reduced in the 6-OHDA PD model.<sup>[93]</sup>

**Isoflavones daidzin, daidzein and genistein** contained in *Pueraria thomsonii* protected PC12 cells stimulated with 6-OHDA through the inhibition of the caspase-3 activation.<sup>[94]</sup> Genistein also protected neurons from substantia nigra pars compact and attenuated the rotational behavior in a hemiparkinsonian 6-OHDA model.<sup>[95]</sup>

**Psoralea corylifolia** protected SK-N-SH cells from MPP+ intoxication and prevented the dopaminergic neurons loss in MPTP intoxicated mice by inhibition of the monoamine transporter.<sup>[96]</sup>

**Rosmarinus officinalis** protected dopaminergic neurons in different degenerative disease models.<sup>[97]</sup>

#### REFERENCES

1. Eriksen JL, Petrucelli L. Parkinson's disease—molecular mechanisms of disease. *Drug Discov Today Dis Mech*, 2004; 1: 399–405.
2. Singh N, Pillay V, Choonara YE. Advances in the treatment of Parkinson's disease. *Prog Neurobiol*, 2009; 81: 29–44.
3. Langston JW. The Parkinson's complex: parkinsonism is just the tip of the iceberg. *Ann Neurol*, 2006; 59: 591–6.
4. Richard IH, Frank S, McDermott MP, Wang H, Justus AW, LaDonna KA, Kurlan R. The ups and downs of parkinson disease, a prospective study of

- mood and anxiety fluctuations. *Cogn Behav Neurol*, 2004; 17: 201-7.
5. Naimark D, Jackson E, Rockwell E. Psychotic symptoms in Parkinson's disease patients with dementia. *J Am Geriatr Soc*, 1996; 44: 296-99.
  6. Buzhor E, Leshansky L, Blumenthal J, Barash H, Warshawsky D, Mazor Y, Shtrichman R. Cell-based therapy approaches: the hope for incurable diseases. *Regen Med*, 2014; 9: 649-72.
  7. Birkmayer W, Hornykiewicz O. The effect of L-3,4-dihydroxy phenylalanine (L-DOPA) on akinesia in Parkinsonism, *Parkinsonism relat Disorder*, 1998; 4: 59-60.
  8. Ho YS, So KF, Chang RC. Anti-aging herbal medicine--how and why can they be used in aging-associated neurodegenerative diseases? *Ageing Res Rev*, 2010; 9: 354-62.
  9. Park HJ, Shim HS, Choi WK, Kim KS, Shim I. Neuroprotective effect of *Lycium chinense* fruit on Trimethyltin-Induced learning and memory deficits in the rats. *Exp Neurobiol*, 2011; 20: 137-43.
  10. Singh HK, Dhawan BN. Neuropsychopharmacological affects of the Ayurvedic nootropic *Bacopa monnieri* Linn (Brahmi). *Indian J Pharmacol*, 1997; 29: 5359-65.
  11. Mohanty IR, Maheshwari U, Joseph D, Deshmukh Y. *Bacopa monniera* protects rat heart against ischaemia-reperfusion injury, role of key apoptotic regulatory proteins and enzymes. *J Pharm Pharmacol*, 2010; 62: 1175-84.
  12. Bhattacharya SK, Bhattacharya A, Kumar A, Ghosal S. Antioxidant activity of *Bacopa monniera* in rat frontal cortex, striatum and hippocampus. *Phytother Res*, 2000; 14: 174-79.
  13. Hosamani R, Muralidhara K. Neuroprotective efficacy of *Bacopa monnieri* against rotenone induced oxidative stress and neurotoxicity in *Drosophila melanogaster*. *Neurotoxicology*, 2009; 30: 977-85.
  14. Jadiya P, Khan A, Sammi SR, Kaur S, Mir SS, Nazir A. Anti-Parkinsonian effects of *Bacopa monnieri*, insights from transgenic and pharmacological *Caenorhabditis elegans* models of Parkinson's disease. *Biochem Biophys Res Comm*, 2011; 413: 605-10.
  15. Renaud J, Nabavi SF, Daglia M, Nabavi SM, Martinoli M-G. Epigallocatechin-3-Gallate, a Promising Molecule for Parkinson's Disease? *Rejuvenation Res*, 2015; 18: 257-69.
  16. Saha P, Das S. Regulation of hazardous exposure by protective exposure: modulation of phase II detoxification and lipid peroxidation by *Camellia sinensis* and *swertia chirata*. *Teratog Carcinog Mutagen*, 2003; suppl. I: 313-22.
  17. Choi JY, Park CS, Kim DJ, Cho MH, Jin BK, Pie JE, Chung WG. Prevention of nitric oxide-mediated 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinson's disease in mice by tea phenolic epigallocatechin 3-gallate. *Neurotoxicol*, 2002; 23(3): 367-74.
  18. Mandel S, Maor G, Youdim MB. Iron and alpha-synuclein in the substantia nigra of MPTP-treated mice: effect of neuroprotective drugs R-apomorphine and green tea polyphenol (-)-epigallocatechin-3-gallate. *J Mol Neurosci*, 2004; 24: 401-16.
  19. Pinto NB, Alexandre BDS, Neves KRT, Silva AH, Leal LKAM, Viana GSB. Neuroprotective properties of the standardized extract from *Camellia sinensis* (green tea) and its main bioactive components, epicatechin and epigallocatechin gallate, in the 6-OHDA model of parkinson's disease. *Evidence-Based Complem Alter Med*, 2015; Article ID 161092, 12 pages.
  20. Al-amri JS, Hagraas MM, Mujallid MI. Effect of Epigallocatechin-3-gallate on inflammatory mediators release in LPS-induced Parkinson's disease in rats. *Indian J Exp Bio*, 2013; 52: 357-62.
  21. Appa Rao MVR, Srinivadan K, Rao TK. The effect of mandookaparni (*Centella asiatica*) on the general mental ability on mentally retarded children. *Indian J Psychiatry*, 1977; 19: 54-9.
  22. Shinomol GK, Muralidhara K. Effect of *Centella asiatica* leaf powder on oxidative markers in brain regions of prepubertal mice in vivo and it's in vitro efficacy to ameliorate 3-NPA-induced oxidative stress in mitochondria. *Phytomedicine*, 2008; 15: 971-84.
  23. Hussin M, Abdul-Hamid A, Mohamad S, Saari N, Ismail M, Bejo MH. Protective effect of *Centella asiatica* extract and powder on oxidative stress in rats, *Food Chemistry*, 2007; 100: 535-41.
  24. Haleagrahara N, Ponnusamy K. Neuroprotective effect of *Centella asiatica* extract (CAE) on experimentally induced Parkinsonism in aged Sprague-Dawley rats, *J Toxicol Sci*, 2010; 35: 41-47.
  25. Lopez V, Calvo MI. White tea (*Camellia sinensis* Kuntze) exerts neuroprotection against hydrogen peroxide-induced toxicity in PC12 cells. *Plant Foods Hum Nutr*, 2011; 66: 22-6.
  26. Voss P, Horakova L, Jakstadt M, Kiekebusch D, Grune T. Ferritin oxidation and proteasomal degradation: protection by antioxidants. *Free Radic Res*, 2006; 40: 673-83.
  27. Augustin S, Rimbach G, Augustin K, Schliebs R, Wolfram S, Cermak R. Effect of a short- and long-term treatment with *Ginkgo biloba* extract on amyloid precursor protein levels in a transgenic mouse model relevant to Alzheimer's disease. *Arch Biochem Biophys*, 2009; 481: 177-82.
  28. Chandrasekaran K, Mehrabian Z, Spinnewyn B, Chinopoulos C, Drieu K, Fiskum G. Neuroprotective effects of bilobalide, a component of *Ginkgo biloba* extract (EGb 761) in global brain ischemia and in excitotoxicity induced neuronal death. *Pharmacopsychiatry*, 2003; 36(Suppl 1): 89-94.
  29. Li LY, Zhao XL, Fei XF, Gu ZL, Qin ZH, Liang ZQ. Bilobalide inhibits 6-OHDA-induced activation

- of NF-kappaB and loss of dopaminergic neurons in rat substantia nigra. *Acta Pharmacol Sin*, 2008; 29: 539-47.
30. Rojas, P, Serrano-García N, Mares-Sámano JJ, Medina-Campos ON, Pedraza-Chaverri J, Ogren SO. EGb761 protects against nigrostriatal dopaminergic neurotoxicity in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinsonism in mice: role of oxidative stress. *Eur J Neurosci*, 2008; 28: 41-50.
  31. Ahmad M, Saleem S, Ahmad AS, Yousuf S, Ansari MA, Badruzzaman M. *Ginkgo biloba* affords dose dependent protection against 6-hydroxydopamine induced parkinsonism in rats: neurobehavioural, neurochemical and immunohistochemical evidences. *J Neurochem*, 2005; 93: 94-104.
  32. Cao F, Sun S, Tong ET. Experimental study on inhibition of neuronal toxic effect of levodopa by *Ginkgo biloba* extract on Parkinson disease in rats. *J Huazhong Univ Sci Technolog Med Sci*, 2003, 23: 151-3.
  33. Herrschaft H, Nacu A, Likhachev S, Sholomov I, Hoerr R, Schlaefke S. *Ginkgo biloba* extract EGb 761® in dementia with neuropsychiatric features: A randomised, placebo-controlled trial to confirm the efficacy and safety of a daily dose of 240 mg. *J Psychia Res*, 2012; 46: 716-23.
  34. Zhu WW, Liu ZL, Xu HW, Chu WZ, Ye QY, Xie AM, Chen L, Li JR. Effect of the oil from *Ganoderma lucidum* spores on pathological changes in the substantia nigra and behaviors of MPTP-treated mice. *Di Yi Jun Yi Da Xue Xue Bao*, 2005; 25: 667-71.
  35. Ding H, Zhou M, Zhang RP, Xu SL. *Ganoderma lucidum* extract protects dopaminergic neurons through inhibiting the production of inflammatory mediators by activated microglia. *Acta Physiologica Sinica*, 2010; 62: 547-54.
  36. Shen B, Truong J, Halliwell R, Govindaraghvm, Sucher NJ. An in vitro study of neuroprotective properties of traditional Chinese herbal medicines thought to prevent ageing and longevity. *BMC Comple Alter Med*, 2013; 13: 373-9.
  37. Yu XQ, Xue CC, Zhou ZW, Li CG, Du YM, Liang J, Zhou SF. In vitro and in vivo neuroprotective effect and mechanisms of glabridin, a major active isoflavone form *Glycyrrhiza glabra* (Liquorice). *Life Sci*, 2008; 82: 68-78.
  38. Lin R.D., Hou W.C., Yen K.Y. & Lee M.H. Inhibition of monoamine oxidase B (MAO-B) by Chinese herbal medicines. *Phytomedicine*, 2003; 10: 650-56.
  39. Im AR, Kim YH, Uddin MR, Chae S, Lee HW, Kim YS, Lee MY. Neuroprotective Effects of *Lycium chinense* Miller against rotenone-Induced neurotoxicity in PC12 Cells. *Am J Chin Med*, 2013; 41: 1343-59.
  40. Patwardhan B, Vaidya ABV. Natural products drug discovery: accelerating the clinical candidate development using reverse pharmacology approaches. *Indian J Exp Bio*, 2010; 48: 220-27.
  41. HP-200 PD Study Group. An alternative medicine treatment for Parkinson's disease: results of a multicenter clinical trial. HP-200 in Parkinson's Disease Study Group. *J Altern Complement Med*, 1995; 1: 249-55.
  42. Kasture S, Pontis S, Pinna A, Schintu N, Spina L, Longoni R, Simola N, Ballero M, Morelli M. Assessment of symptomatic and neuroprotective efficacy of *Mucuna pruriens* seed extract in rodent model of Parkinson's disease. *Neurotox Res*, 2009; 15: 111-22.
  43. Lieu CA, Venkiteswaran K, Gilmour TP, Rao AN, Petticoffer AC, Gilbert EV, Deogaonkar M, Manyam BV, Subramanian T. The antiParkinsonian and antidyskinetic mechanisms of *Mucuna pruriens* in the MPTP-treated nonhuman primate. *Evid Based Complement Alternat Med*, 2012; ID 840247: 10 pages.
  44. Lieu CA, Kunselman AR, Manyam BV, Venkiteswaran K, Subramanian T. A water extract of *Mucuna pruriens* provides long-term amelioration of parkinsonism with reduced risk for dyskinesias. *Parkinsonism Relat Disord*, 2010; 16: 458-65.
  45. Katzenschlager R, Evans A, Manson A, Patsalos P, Ratnaraj N, Watt H, Timmermann L, Giessen RV, Less A. *Mucuna pruriens* in Parkinson's disease: a double blind clinical and pharmacological study. *J Neurol Neurosurg Psychiatry*, 2004; 75: 1672-77.
  46. Vaidya AB, Rajagopalan TG, Mankodi NA, Antarkar DS, Tathed PS, Purohit AV, Wadia NH. Treatment of Parkinson's disease with the cowhage plant-*Mucuna pruriens* Bak. *Neurol India*, 1978; 26: 171-76.
  47. Van Kampen J, Robertson H, Hagg T, Drobitch R. Neuroprotective actions of the ginseng extracts G115 in two rodent models of Parkinsons disease. *Exp Neurol*, 2003; 184: 521-29.
  48. Radad K, Gille G, Moldzio R, Saito H, Rausch WD. Ginsenosides Rb1 and Rg1 effects on mesencephalic dopaminergic cells stressed with glutamate. *Brain Res*, 2004; 17: 41-53.
  49. Liu M, Zhang J. Effects of ginsenoside Rb1 and Rg1 on synaptosomal free calcium level, ATPase and calmodulin in rat hippocampus. *Chin Med J*, 1995; 108: 544-47.
  50. Chen XC, Zhu YG, Zhu LA, Huang C, Chen Y, Chen LM, Fang F, Zhou YC, Zhao CH. Ginsenoside Rg1 attenuates dopamine-induced apoptosis in PC12 cells by suppressing oxidative stress. *Eur J Pharmacol*, 2003; 473: 1-7.
  51. Liu Y, Zhang RY, Zhao J, Dong Z, Feng DY, Wu R, Shi M, Zhao G. Ginsenoside Rd protects SH-SY5Y cells against 1-Methyl-4-phenylpyridinium induced injury. *Int J Mol Sci*, 2015; 16: 14395-408.
  52. Bizimenyera ES, Aderogba MA, Eloff JN, Swan GE. Potential of neuroprotective antioxidant-based therapeutics from *Peltophorum africanum* sond. (Fabaceae). *Afr J Trad CAM*, 2007; 4: 99-106.

53. Zhang LQ, Sa F, Chong CM, Wang Y, Zhou ZY, Chang RCC, Chan SW, Hoi PM, Lee SMY. Schisantherin A protects against 6-OHDA-induced dopaminergic neuron damage in zebrafish and cytotoxicity in SH-SY5Y cells through the ROS/NO and AKT/GSK3 $\beta$  pathways. *J Ethnopharmacol*, 2015; 170: 2015 Pages 8-15.
54. Uddin R, Kim HH, Lee JH, Park SU. Neuroprotective effects of medicinal plants. *EXCLI J*, 2013; 12: 541-45.
55. Huang WH, Lee AR, Yang CH. Antioxidative and anti-inflammatory activities of polyhydroxyflavonoids of *Scutellaria baicalensis* GEORGI. *Biosci Biotechnol Biochem*, 2006; 70: 2371-80.
56. Mu X, He G, Cheng Y, Li X, Xu B, Du G. Baicalein exerts neuroprotective effects in 6-hydroxydopamine-induced experimental parkinsonism in vivo and in vitro. *Pharmacol Biochem Behav*, 2009; 92: 642-48.
57. Mu X, He GR, Yuan X, Li X.-X, Du GH. Baicalein protects the brain against neuron impairments induced by MPTP in C57BL/6 mice. *Pharmacol Biochem Behav*, 2011; 98: 286-91.
58. Cheng Y, He G, Mu X, Zhang T, Li X, Hu J, Xu B, Du G. Neuroprotective effect of baicalein against MPTP neurotoxicity, behavioral, biochemical and immunohistochemical profile. *Neurosci Lett*, 2008; 441: 16-20.
59. Zhang J, Cui W, Li G, Yuan S, Xu D, Hoi MP, Lin Z, Dou J, Han Y, Lee SM. Baicalein protects against 6-OHDA-induced neurotoxicity through activation of Keap1/Nrf2/HO-1 and involving PKC $\alpha$  and PI3K/AKT signaling pathways. *J Agric Food Chem*, 2012; 60: 8171-82.
60. Sharma PC, Yelne MB, Dennis TJ. In "Database on medicinal plants used in Ayurveda" Vol 3. Central Council for Research in Ayurveda & Sidha, Deptt. of ISM & H. Ministry of Health & Family Welfare, Govt. of India, New Delhi, 2005; 88-128.
61. Ahmad M, Saleem S, Ahmad AS, Ansari MA, Yousuf S, Hoda MN, Islam F. Neuroprotective effects of *Withania somnifera* on 6-hydroxydopamine induced parkinsonism in rats. *Hum Exp Toxicol*, 2005; 24: 137-47.
62. Prakash J, Yadav SK, Chouhan S, Singh SP. Neuroprotective role of *Withania somnifera* root extract in maneb-paraquat induced mouse model of parkinsonism. *Neurochem Res*, 2013; 38: 972-80.
63. RajaSankar S, Manivasagam T, Sankar V, Prakash S, Muthusamy R, Krishnamurti A, Surendran S. *Withania somnifera* root extract improves catecholamines and physiological abnormalities seen in a Parkinsons disease model mouse. *J Ethnopharmacol*, 2009; 125: 369-73.
64. Cole, GM, Teter B, Frautschy SA. Neuroprotective effects of curcumin. *Adv Exp Med Biol*, 2007; 595: 197-212.
65. Rajeswari A, Sabesan M. Inhibition of monoamine oxidase-B by the polyphenolic compound, curcumin and its metabolite tetrahydrocurcumin, in a model of Parkinson's disease induced by MPTP neurodegeneration in mice. *Inflammopharmacology*, 2008; 16: 96-9.
66. Zbarsky V, Datla KP, Parkar S, Rai DK, Aruoma OI, Dexter DT. Neuroprotective properties of the natural phenolic antioxidants curcumin and naringenin but not quercetin and fisetin in a 6-OHDA model of Parkinson's disease. *Free Radic Res*, 2005; 39: 1119-25.
67. Vajragupta O, Boonchoong P, Watanabe H, Tohda M, Kummasud N, Sumanont Y. Manganese complexes of curcumin and its derivatives: Evaluation for the radical scavenging ability and neuroprotective activity. *Free Radic Biol Med*, 2003; 35: 1632-44.
68. Mythri RB, Harish G, Dubey SK, Misra K, Bharath MM. Glutamoyl diester of the dietary polyphenol curcumin offers improved protection against peroxynitrite-mediated nitrosative stress and damage of brain mitochondria in vitro: implications for Parkinson's disease. *Mol Cell Biochem*, 2011; 347: 135-43.
69. Kim SJ, Son TG, Park HR, Park M, Kim MS, Kim HS, Chung HY, Mattson MP, Lee J. Curcumin stimulates proliferation of embryonic neural progenitor cells and neurogenesis in the adult hippocampus. *J Biol Chem*, 2008; 283: 14497-505.
70. Xu Y, Ku B, Cui L, Li X, Barish PA, Foster TC, Ogle WO. Curcumin reverses impaired hippocampal neurogenesis and increases serotonin receptor 1A mRNA and brain-derived neurotrophic factor expression in chronically stressed rats. *Brain Res*, 2007; 1162: 9-18.
71. Tripanichkul W, Jaroensuppaperch EO. Curcumin protects nigrostriatal dopaminergic neurons and reduces glial activation in 6-hydroxydopamine hemiparkinsonian mice model. *Inter J Neurosci*, 2012; 122: 263-70.
72. An H, Kim IS, Koppula S, Kim BW, Park PJ, Lim BO, Choi WS, Lee KH, Choi DK. Protective effects of *Gastrodia elata* Blume on MPP $^{+}$ -induced cytotoxicity in human dopaminergic SH-SY5Y cells. *J Ethnopharmacol*, 2010; 130: 290-8.
73. Kim IS, Choi DK, Jung HJ. Neuroprotective effects of vanillyl alcohol in *Gastrodia elata* Blume through suppression of oxidative stress and anti-apoptotic activity in toxin-induced dopaminergic MN9D cells. *Molecules*, 2011; 16: 5349-61.
74. Li C, Chen X, Zhang N, Song W, Mu Y. Gastrodin inhibits neuroinflammation in rotenone-induced Parkinson's disease model rats. *Neural Regen Res*, 2012; 7: 325-331.
75. Manavalan A, Ramachandran U, Sundaramurthi H, Mishra M, Sze SK, Hu JM, Feng ZW, Heese K. *Gastrodia elata* Blume (tianma) mobilizes neuroprotective capacities. *Int J Biochem Mol Biol*, 2012; 3: 219-41.
76. Kumar H, Kim IS, More SV, Kim BW, Bahk YY, Choi DK. Gastrodin protects apoptotic dopaminergic

- neurons in a Toxin-Induced Parkinson's Disease Model. Evidence-Based Complementary and Alternative Medicine. Volume 2013, Article ID 514095, 13 pages.
77. Wang XL, Xing GH, Hong B, Li XM, Zou Y, Zhang XJ, Dong MX. Gastrodin prevents motor deficits and oxidative stress in the MPTP mouse model of Parkinson's disease: Involvement of ERK1/2–Nrf2 signaling pathway. *Life Sci*, 2014; 114: 77–85.
  78. Liu DZ, Zhu J, Jin DZ, Zhang LM, Ji XQ, Ye Y, Tang CP, Zhu XZ. Behavioral recovery following sub-chronic paeoniflorin administration in the striatal 6-OHDA lesion rodent model of Parkinsons disease. *J Ethnopharmacol*, 2007; 112: 327–32.
  79. Renaud J, Bournival J, Zottig X, Martinoli MG. Resveratrol protects DAergic PC12 cells from high glucose-induced oxidative stress and apoptosis: effect on p53 and GRP75 localization. *Neurotox Res*, 2014; 25: 110–23.
  80. Lu KT, Ko MC, Chen BY, Huang JC, Hsieh CW, Lee MC, Chiou RY, Wung BS, Peng CH, Yang YL. Neuroprotective effects of resveratrol on MPTP-induced neuron loss mediated by free radical scavenging. *J Agric Food Chem*, 2008; 56: 6910–3.
  81. Srivastava G, Dixit A, Yadav S, Patel DK, Prakash O, Singh MP. Resveratrol potentiates cytochrome P450 2 d22-mediated neuroprotection in maneb- and paraquat-induced parkinsonism in the mouse. *Free Radic Biol Med*, 2012; 52: 1294–306.
  82. Chang CY, Choi DK, Lee DK, Hong YJ, Park EJ. Resveratrol confers protection against rotenone-induced neurotoxicity by modulating myeloperoxidase levels in glial cells. *PLoS ONE*, 2013; 8: Article ID e60654.
  83. Jin F, Wu Q, Lu YF, Gong QH, Shi JS. Neuroprotective effect of resveratrol on 6-OHDA-induced Parkinsons disease in rats. *Euro J Pharmacol*, 2008; 600: 78–82.
  84. Zhang H, Han T, Zhang L, Yu CH, Wan DG, Rahman K, Qin LP, Peng C. Effects of tenuifolin extracted from radix polygalae on learning and memory, a behavioral and biochemical study on aged and amnesic mice. *Phytomedicine*, 2008; 15: 587–94.
  85. Liang Z, Shi F, Wang Y, Lu L, Zhang Z, Wang X, Wang X. Neuroprotective effects of tenuigenin in a SH-SY5Y cell model with 6-OHDA-induced injury. *Neurosci Lett*, 2011; 497: 104–9.
  86. Yuan HL, Li B, Xu J, Wang Y, He Y, Zheng Y, Wang XM. Tenuigenin protects dopaminergic neurons from inflammation-mediated damage induced by the lipopolysaccharide. *CNS Neurosci Therap*, 2012; 18: 584–90.
  87. Kim IS, Koppula S, Park PJ, Kim EH, Kim CG, Choi WS, Lee KH, Choi DK. “*Chrysanthemum morifolium* Ramat (CM) extract protects human neuroblastoma SH-SY5Y cells against MPP+-induced cytotoxicity,” *J Ethnopharmacol*, 2009; 126: 447–54.
  88. Geng X, Tian X, Tu P, Pu X, “Neuroprotective effects of echinacoside in the mouse MPTP model of Parkinson's disease,” *Eur J Pharmacol*, 2007; 564: 66–74.
  89. Perez-H J, Carrillo-S C, Garcia E, Ruiz-Mar G, Perez-Tamayo R, Chavarria A, “Neuroprotective effect of silymarin in a MPTP mouse model of Parkinson's disease,” *Toxicology*, 2014; 319: 38–43.
  90. de Andrade DVG, de Oliveria DM, Barreto G. “Effects of the extract of *Anemopaegma mirandum* (Catuba) on rotenone-induced apoptosis in human neuroblastomas SHSY5Y cells,” *Brain Res*, 2008; 1198: 188–96.
  91. Oliveria DMD, Barreto G, Andrade DVGD, Saraceno E, Aon-Bertolino L, Capani F, Bacha RDSEI, Giraldez LD. “Cytoprotective effect of *Valeriana officinalis* extract on an in vitro experimental model of parkinson disease,” *Neurochem Res*, 2009; 34: 215–20.
  92. Choi BS, Sapkota K, Kim S, Lee HJ, Choi HS, Kim SJ, “Antioxidant activity and protective effects of *Tripterygium regelii* extract on hydrogen peroxide-induced injury in human dopaminergic cells, SH-SY5Y.” *Neurochem Res*, 2010; 35: 1269–80.
  93. Shim JS, Kim HG, Ju MS, Choi JG, Jeong SY, Oh MS. “Effects of the hook of *Uncaria rhynchophylla* on neurotoxicity in the 6-hydroxydopamine model of Parkinson's disease,” *J Ethnopharmacol*, 2009; 126: 361–5.
  94. Lin CM, Lin RD, Chen ST, Lin YP, Chiu WT, Lin JW, Hsu FL, Lee MH. “Neurocytoprotective effects of the bioactive constituents of *Pueraria thomsonii* in 6-hydroxydopamine (6-OHDA)-treated nerve growth factor (NGF)-differentiated PC12 cells,” *Phytochemistry*, 2010; 71: 2147–56.
  95. Baluchnejadmojarad T, Roghani M, Nadoushan MRJ, Bagheri M. “Neuroprotective effect of genistein in 6-hydroxydopamine hemi-parkinsonian rat model,” *Phytother Res*, 2009; 23: 132–5.
  96. Zhao G, Zheng XW, Qin GW, Gai Y, Jiang ZH, Guo LH, “In vitro dopaminergic neuroprotective and in vivo antiparkinsonian-like effects of  $\Delta$  3,2-hydroxybakuchiol isolated from *Psoralea corylifolia* (L.),” *Cell Mol Life Sci*, 2009; 66: 1617–29.
  97. Park SE, Kim S, Sapkota K, Kim SJ, “Neuroprotective effect of *Rosmarinus officinalis* extract on human dopaminergic cell line, SH-SY5Y,” *Cell Mol Neurobio*, 2010; 30: 759–67.