



**ANTIBACTERIAL EFFECT OF BACTERIOCIN FROM *LEUCONOSTOC
MESENTEROIDES* SSP. *CREMORIS* AGAINST DIARRHEAL CAUSATIVE BACTERIA**

Raghad A. Aziz¹, Jehan Abdul Sattar Salman^{2*} and Omar A. H. Hachim³

^{1,3}Department of Science, College of Basic Education, Al-Mustansiriyah University, Baghdad-Iraq.

²Department of Biology, College of Science, Al-Mustansiriyah University, Baghdad-Iraq.

***Author for Correspondence: Dr. Jehan Abdul Sattar Salman**

Department of Biology, College of Science, Al-Mustansiriyah University, Baghdad-Iraq.

Article Received on 16/09/2016

Article Revised on 06/10/2016

Article Accepted on 27/10/2016

ABSTRACT

Antibacterial activity of crude bacteriocin of *Leuconostoc mesenteroides* ssp. *cremoris* included non-concentrated and concentrated supernatant (bacteriocin) was tested against diarrheal causative bacteria *Escherichia coli*, *Salmonella ser. paratyphi A*, *Shigella* group. and *Vibrio cholerae*. Results showed that non-concentrated crude supernatant (bacteriocin) from 2 isolates [*L. mesenteroides* ssp. *cremoris* (Lc3) and *L. mesenteroides* ssp. *cremoris* (Lc4)] had inhibitory effect with inhibition diameter ranged between (11-13) mm against *E.coli*, *Shigella* group, *Salmonella ser. paratyphi A* and *V. cholera*, while other isolates were unable to inhibit the growth of pathogenic bacteria. The concentrated crude supernatant bacteriocin showed inhibitory activity against *E.coli*, *Shigella* group, *Salmonella ser. paratyphi A* and *V. cholerae* with diameter of 19, 18, 15 and 25 mm respectively for for *Leuconostoc mesenteroides* ssp. *cremoris* (Lc3) isolate and inhibition diameter of 17, 18, 16 and 20 mm for *Leuconostoc mesenteroides* ssp. *cremoris* (Lc4) isolate.

KEYWORDS: *Leuconostoc mesenteroides*, Bacteriocin, Diarrheal causative bacteria.

INTRODUCTION

Leuconostoc mesenteroides ssp. *cremoris* characterized as gram positive coccobacilli bacteria, they are non-motile, catalase- negative and non- spore former bacteria, they are found in foods of animal origin which include raw milk, dairy products, poultry, meat and fish as well as found in vegetable products, despite the resistance to vancomycin it is sensitive to erythromycin and clindamycin (Usta-Atmaca *et al.*, 2015).

It is a part of the lactic acid bacteria, it's able to produce anti molecules such as organic acids, hydrogen peroxide, a dual acetylcholine, and bacteriocins (Ringø *et al.*, 2010).

Bacteriocins are antimicrobial peptides (AMPs) which contains molecules acts as a defense system against many organisms, such as bacteria, fungi, parasites and viruses. These components become very important due to the increase in microorganisms resistant to the usual vital antibiotics (Balciunas *et al.*, 2013).

Diarrhea is a major cause of morbidity among children in developing countries. transmitted microbes responsible for the majority of injuries through the feces to the mouth through water and food (Fung *et al.*, 2011). It is caused by a wide range of pathogens, including bacteria,

viruses, etc, only a small number of organisms are the cause of most cases of acute childhood diarrhea such as adenovirus and rotavirus reasons leading to acute diarrhea, The biggest section contains pathogenic bacteria such as *E.coli*, *Shigella*, *Campylobacter*, *Salmonella*, *V.cholera* *Clostridium difficili* caused diarrhea during epidemics (Moreno *et al.*, 2000 ;Hera *et al.*, 2016).

The mechanism of diarrhea occurrence are adhesion of pathogenic bacteria to the small intestine and the production of toxins when they connected to particularly receptors located on the cell membrane layer of the gut where it lead to activate the enzyme adenylate cyclase where increasing its concentration leads to increasing Cycli Adenosin Mono phosphate (cAMP) and thus lead to increased fluid loss and salts from the intestine, causing diarrhea (Ateia, 2013).

This study was aimed to production of bacteriocin from the *Leuconostoc mesenteroides* ssp. *cremoris* isolated from raw cow milk and study their antibacterial effect against some bacteria causing diarrhea.

MATERIALS AND METHODS

Bacterial Isolates

Isolation and Identification of *Leuconostoc mesenteroides*

For *Leuconostoc mesenteroides* isolation, each sample of raw cow milk was inoculated in MRS broth at (1%) and cultivated at 30°C for 24 h under anaerobic condition. A series of dilutions (10^{-1} , 10^{-2} , 10^{-3} , 10^{-4}) were made for each sample with phosphate buffer saline, then spread (0.1) ml of the fourth dilution on solid MRS Vancomycin agar plates (Fatma and Benmechernene, 2013). MRS Vancomycin agar plates were incubated at 30°C for 24 h anaerobically. Sub culturing of isolated colonies on MRS agar for further purification, incubated under anaerobic condition at 30°C for 24 h, then the individual colonies were transported to MRS broth and incubated under the same conditions. The purified isolates were identified through cultural, microscopically and biochemical tests (Gravie, 1986) and Vitek2 system.

Isolation and Identification of diarrheal causative bacteria

The diarrheal causative bacteria was isolated from feces of children under five years old and it was identified through cultural, microscopically and biochemical tests and Vitek2 system according to (Baron and Finegold, 1994).

Detection of bacteriocin production by *Leuconostoc mesenteroides* ssp. *cremoris*

The bacterial isolates were inoculated in MRS broth with pH 6.2 for 24h at 37°C. The crude supernatant (bacteriocin) was obtained by centrifugation at 10,000 rpm for 15 min, sterilized by Millipore filter (0.02 m), the supernatant was neutralized to pH 6.2 by

adding 1N NaOH to remove the influence of organic acid (Salman, 2014).

Concentration of crude supernatant (bacteriocin)

The crude bacteriocin was concentrated using Dialysis bags and poly ethylene glycol 20000 (Janson and Ryden., 1998).

Antibacterial effect of crude bacteriocin from *L. mesenteroides* ssp. *cremoris*

Leuconostoc mesenteroides ssp. *cremoris* non concentrated and concentrated crude supernatants (bacteriocin) were screened for their antibacterial activity against *Escherichia coli*, *Salmonella* spp., *Shigella* spp. and *Vibrio cholerae* isolates, using agar well diffusion method. The plates were spread with 10^5 cfu/ml culture broth of pathogenic isolates on the surface of nutrient agar, then the plates were left for 10 min dispensing 50µl of each samples into the wells. The plates were incubated at 37°C for 24h. Inhibition zone were measured in millimeter diameter (Gupta, 1996).

RESULTS AND DISCUSSION

Isolation and Identification of *Leuconostoc mesenteroides*

Twelve isolates of *L. mesenteroides* were obtained from 25 bacterial isolates. All isolates were Gram positive, ovoid shape associated in pairs and/or short chains, negative catalase reaction, able to produce carbon dioxide from glucose and unable to hydrolyze Arginine. All isolates were grown at 45°C. After identification by biochemical tests and Vitek 2 system, the results showed that 9 out of 12 isolates belonged to *Leuconostoc mesenteroides* ssp. *cremoris*, while 2 isolates of *Leuconostoc mesenteroides* ssp. *mesenteroides* and one isolate *Leuconostoc mesenteroides* ssp. *dextranicum* (Table-1).

Table(1): Incidence of *Leuconostoc* isolates from raw milk.

Type of isolates	Number of isolates
<i>Leuconostoc mesenteroides</i> ssp. <i>cremoris</i>	9
<i>Leuconostoc mesenteroides</i> ssp. <i>mesenteroides</i>	2
<i>Leuconostoc mesenteroides</i> ssp. <i>dextranicum</i>	1
Total	12

From the result above, 9 of 12 isolates were identified as *L. mesenteroides* ssp. *cremoris* which isolated from raw milk, Abbas (2015) reported that the majority of *Leuconostoc* spp. isolated from raw milk were identified as *L. mesenteroides* ssp. *cremoris*, while Fatma and Benmechernene (2013) reported the possibility to isolate *Leuconostoc* spp. from raw camel milk. Bao *et al.* (2012) mentioned that 40.8% of lactic acid bacteria isolated from raw milk product was belonged to *Leuconostoc* spp.

In this research 15 isolates of diarrheal causative bacteria were identified as (9) *E. coli*, (2) *Salmonella* ser. *paratyphi A*, (3) *Shigella* group and one isolate of *Vibrio cholerae*,

Detection of bacteriocin production by *Leuconostoc mesenteroides* ssp. *cremoris*

Screening of crude supernatant (bacteriocin) of *L. mesenteroides* ssp. *cremoris* isolates to select the more efficient producing bacteriocin isolates, the results showed that *L. mesenteroides* ssp. *cremoris* (Lc3) and *L. mesenteroides* ssp. *cremoris* (Lc4) isolates gave the highest inhibition activity against *E. coli* and *Shigella* group with inhibition diameter (10-13) mm, against *Salmonella* ser. *paratyphi A* (10-12) mm, while against *Vibrio cholerae* was (11-12) mm (Table-2-). Gupta *et al.*, (1998) observed that the supernatant of *Lactobacillus* bacteria grown on MRS broth gave inhibitory activity against gram positive and negative bacteria, Salman *et al.*, (2008) noticed the inhibitory activity of of the

supernatant of lactic acid bacteria against bacteria causing UTI urinary tract infection, Karthikeyan and Santho, (2009) showed that MRS broth is the best medium for the production of Bacteriocin, O'Shea *et al.*, (2011) mentioned that the inhibitory activity of

supernatant of *Lactobacillus salivarius* was limited effect, Salman, (2014) showed that *Staphylococcus aureus* (PSM producer isolates) showed resistance to the neutralized concentrated cell free supernatant (CSF) and bacteriocin of *Lactobacillus salivarius*.

Table(2): Antibacterial activity of *Leuconostoc mesenteroides* ssp. *cremoris* crude supernatant (bacteriocin) against diarrheal causative bacteria.

Pathogenic bacteria isolates	Inhibition zone (mm)								
	Lc1	Lc2	Lc3	Lc4	Lc5	Lc6	Lc7	Lc8	Lc9
<i>E.coli</i> (E1)	-	-	10	10	-	-	-	-	-
<i>E.coli</i> (E2)	-	-	11	13	-	-	-	-	-
<i>E.coli</i> (E3)	-	-	12	12	-	-	-	-	-
<i>E.coli</i> (E4)	-	-	11	11	-	-	-	-	-
<i>E.coli</i> (E5)	-	-	13	12	-	-	-	-	-
<i>E.coli</i> (E6)	-	-	11	10	-	-	-	-	-
<i>E.coli</i> (E7)	-	-	12	13	-	-	-	-	-
<i>E.coli</i> (E8)	-	-	12	12	-	-	-	-	-
<i>E.coli</i> (E9)	-	-	11	11	-	-	-	-	-
<i>Shigella group</i> (Sh1)	-	-	10	12	-	-	-	-	-
<i>Shigella group</i> (Sh2)	-	-	12	11	-	-	-	-	-
<i>Shigella group</i> (Sh3)	-	-	11	12	-	-	-	-	-
<i>Salmonella ser. paratyphi A</i> (Sa1)	-	-	12	12	-	-	-	-	-
<i>Salmonella ser. paratyphi A</i> (Sa2)	-	-	11	12	-	-	-	-	-
<i>Vibrio cholerae</i> (V1)	-	-	12	11	-	-	-	-	-

Lc: *Leuconostoc mesenteroides* ssp. *cremoris* , - : no inhibitory effect

Inhibitory effect of concentrated crude supernatant (bacteriocin) against diarrheal causative bacteria.

The concentrated crude supernatant (bacteriocin) produced by *L.mesenteroides* ssp. *cremoris* showed inhibitory effect against all pathogenic bacteria with inhibition zone ranged from (12-19) mm against *E.coli*, (14-18) mm against *Shigella group*, (14-15) mm *Salmonella ser. paratyphi A* and (22) mm against *Vibrio cholera* for *L. mesenteroides* ssp. *cremoris* (Lc3) isolate and from (11-17) mm against *E.coli*, (16-18) mm against *Shigella group*, (15-16) mm against *Salmonella ser. paratyphi A* and (20) mm against *Vibrio cholera* for *L. mesenteroides* ssp. *cremoris* (Lc4) isolate. (Table-3-) (Figure 1, 2).

The reason for the effectiveness of inhibitory activity of concentrated crude supernatant against many gram positive and negative bacteria due to the ability of

bacteria to produce many inhibitory substances, especially bacteriocins (Brashear *et al.*, 2003), Salman (2014) observed that the concentrated supernatant of lactic acid bacteria showed inhibitory activity against *S. aureus*.

The impact of bacteriocin in bacterial cells can be either fatal bactericidal (Todorov, 2009; Al-Charrakh *et al.*, 2011) or it can be a disincentive to the growth bacteriostatic (Serika *et al.*, 2010) and similar mechanisms of action of bacteriocins on its impact on the target cell with the mechanisms of action of antibiotics it has been acting on cell wall or plasma membrane (Han *et al.*, 2011). Sarika *et al.*, (2010) have showed that bacteriocins produced from *lactobacillus rhamnosus* had an effective inhibitory against *E.coli* , *S. aureus*, *Pseudomonas aeruginosa* and *Vibrio harveyi*.

Table(3): Antibacterial activity of *Leuconostoc mesenteroides* ssp. *cremoris* concentrated crude supernatant (bacteriocin) against diarrheal causative bacteria.

Pathogenic bacteria isolates	Inhibition zone (mm)	
	<i>Leuconostoc mesenteroides</i> ssp. <i>cremoris</i> (Lc3)	<i>Leuconostoc mesenteroides</i> ssp. <i>cremoris</i> (Lc4)
<i>E.coli</i> (E1)	12	11
<i>E.coli</i> (E2)	13	16
<i>E.coli</i> (E3)	18	16
<i>E.coli</i> (E4)	18	17
<i>E.coli</i> (E5)	19	16
<i>E.coli</i> (E6)	12	12
<i>E.coli</i> (E7)	18	15
<i>E.coli</i> (E8)	19	16

<i>E.coli</i> (E9)	14	14
<i>Shigella group</i> (Sh1)	14	17
<i>Shigella group</i> (Sh2)	18	18
<i>Shigella group</i> (Sh3)	14	16
<i>Salmonella ser. paratyphi A</i> (Sa1)	15	15
<i>Salmonella ser. paratyphi A</i> (Sa2)	14	16
<i>Vibrio cholerae</i> (V1)	25	20

Lc: *Leuconostoc mesenteroides* ssp. *cremoris* , - : no inhibitory effect



Figure (1): Antibacterial activity of *Leuconostoc mesenteroides* ssp. *cremoris* concentrated crude supernant (bacteriocin) against *E.coli*.

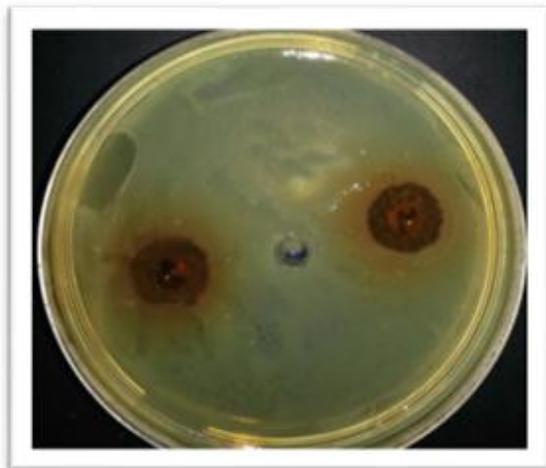


Figure (2): Antibacterial activity of *Leuconostoc mesenteroides* ssp. *cremoris* concentrated crude supernant (bacteriocin) against *Shigella group*.

CONCLUSION

In conclusion, some of *Leuconostoc mesenteroides* ssp. *cremoris* isolated from local raw cow milk had ability to produce bacteriocin that inhibit diarrheal causative bacteria.

REFERENCES

1. Abbas, A. Y. (2015). Effect of Biosurfactant Produced by Locally *Leuconostoc mesenteroides* ssp. *cremoris* in Pathogenic Bacteria Isolated from Catheters and Urinary Tract Infections.(Master's Thesis). AL-Mustansiriyah University. Iraq. Baghdad.
2. Al-Charrakha, A .H.; Yousif, S.Y. and Al- Janabi, H.S.(2011). Antimicrobial spectrum of the action of bacteriocins from *Klebsiella* isolates from Hilla/Iraq. *J.Microbial.*, 2(5): 1-11.
3. Ateia, A. M. (2013). Activity of some Proteinous Inhibitors Produced by *Saccharomyces cerevisiae* Against some enteric bacteria. Master`s Thesis. AL-Mustansiriyah University. Iraq. Baghdad.
4. Balciunas, E.M.; Martinez, F.A.C.; Todorov, S.D.; De Melo Franco, B.D.G.; Converti, A.and De Souza Oliveira, R.P. (2013).Novel biotechnological applications of bacteriocins: A review. *Food Control.*, 32: 134-142.
5. Bao, Q.; Yu, J.; Liu, W.; Qing, M.; Wang, W.; Chen, X. and Zhang, H. (2012). Predominant lactic acid bacteria in traditional fermented yak milk products in the Sichuan Province of China. *Dairy science and technology.*, 92(3): 309-319.
6. Baron, E. J. and Finegold, S. M. (1994). Microorganism Encountered in Urinary Tract. In Baily and Scott's Diagnostic Microbiology, 9th ed., Mosby Company, USA.
7. Brashears, M.M.; Galyean, M.L.; Loneragan, G.H.; Mann, J.E. and Mann, K.K. (2003). Prevalence of *Escherichia coli* O157: H7 and performance by beef feedlot cattle given *Lactobacillus* direct – fed Microbials. *J. Food. Prot.* 66(5): 748 –754.
8. Fatma, C. H. and Benmechernene, Z. (2013). Isolation and identification of *Leuconostoc mesenteroides* producing bacteriocin isolated from Algerian raw camel milk. *African Journal of Microbiology Research.*, 7(23): 2961-2969.
9. Fung, W.; Lye, H.; Lim, T.; Kuan, C. and Liong, M. (2011). Roles of Probiotic on Gut Health. Springer-Verlag Berlin Heidelberg., 21.3: 139- 142.
10. Gupta, S. (1996). The Short Text Book of Pediatrics. 7th ed., Jaypee Brothers Medical Publishers (P) Ltd.

11. Gupta, S. S.; DeWitt, N. D.; Allen, K. E. and Slayman, C.W. (1998). Evidence for a salt bridge between transmembrane segments 5 and 6 of the yeast plasma-membrane H⁺-ATPase. *J Biol Chem.*, 273(51): 34328-34.
12. Han, B.; Yu, Z.; Liu, B.; Ma, Q. and Zhang, R. M.(2011). Optimization of bacteriocin production by *Lactobacillus plantarum* YJG, isolated from the mucosa of the gut of healthy chickens. *J. Microbiology Research.*, 5(10): 1147-1155.
13. Hera, N.; Tri, W.; Abu Tholib, A.; Abdul, W. and Yati, S. (2016). Detection of group A rotavirus strains circulating among children with acute diarrhea in Indonesia. *J. Springer Plus.*, 97(5): 137-140.
14. Janson, J. C. and Rydén, L. (1998). Protein Purification: principles, high resolution methods and applications. New York: John Wiley and Sons.
15. Karthikeyan, V. and Santhosh, S.W.(2009). Study of Bacteriocin as a Food Preservative and the *L. acidophilus* Strain as Probiotic .Department of Biotechnology, School of Bioengineering, SRM University, Kattankulathur-603203, Tamilnadu, India. *Pakista Journal of Nutrition*, 8(4): 335-340.
16. Moreno, A.R.; Filho, A.F.; Gomes, T.A.T.; Ramos, S.T.S.; Montemor, L.P.G.; Tavares, V.C.; Filho, L.S.; Irino, K. and Martinez, M.B. (2000). Etiology of children diarrhea in the Northeast of Brazil: significant emergent diarrhea pathogens. *Dia. Microbiol. Infec. Dis.*, 66(1): 50-57.
17. O'shea, E.F.; O'connor, P.m\M.; Raftis, E.J.; O'tooie, P.W.; Stanton, C.; Cotter, P.D.R. Oss, R.P. and Hill, C. (2011). Production of multiple Bacteriocins from a single locus by gastrointestinal strains of *Lactobacillus salivarius*. *J. Bacterial.*, 193(24): 6973-6982.
18. Ringø, E.; Løvmo, L.; Kristiansen, M.; Bakke, Y. and Salinas, I. (2010). Lactic acid bacteria vs. pathogens in the gastrointestinal tract of fish. *Aquac Res.*, 41: 451-467.
19. Salman, J. A. S. (2014). Effect of *Lactobacillus* metabolites against *Staphylococcus aureus* producing Phenol-Soluble Modulins (PSMs). *Int. J. Curr. Microbial. App. Sci.*, 3(8): 178-184.
20. Salman, J. A. S. and Majeed, H. Z. and Abdul Razzaq, H. A. (2008). Inhibitory effect of the bacteria *Lactobacillus gasseri* boosts vital to the growth and virulence factors of some bacteria that cause urinary tract infections. *AL- Mustansiriyah Journal of Science.*, 19(4): 23-13.
21. Sarika, A.R.; Lipton, A.P. and Aishwarya, M.S. (2010). Bacteriocin Production by a New Isolate of *Lactobacillus rhamnosus* GP1 under Different Culture Conditions. *J. Food Science and Technology.*, 2(5): 291-297.
22. Todorov, S.D. (2009). Bacteriocins from *Lactobacillus plantarum* – Production Genetic Organization and Mode of Action. *J. Brazilian Journal of Microbiology.*, 40: 209-221.
23. Usta- Atmaca, H.; akbas, f.; karagoz, y. and piskinpasa, M.E. (2015). A rarely seen cause for empyema: *Leuconostoc mesenteroides*. *J infect dev ctries.*, 9(4): 425-427.