

**MOLECULAR BASIS OF FOCAL SEGMENTAL GLOMERULOSCLEROSIS
ASSOCIATED WITH STEROID RESISTANCE NEPHROTIC SYNDROME**

Bhoomi B. Joshi¹, Kinnari N. Mistry^{1*}, Prakash G. Koringa², Sishir Gang³ and Chaitanya G. Joshi²

¹Ashok and Rita Patel Institute of Integrated Study and Research in Biotechnology and Allied Sciences (ARIBAS),
Affiliated to Sardar Patel University, Vallabh Vidyanagar, Gujarat, India, 388120.

²College of Veterinary Sciences and Animal Husbandary, Anand Agricultural University.

³Muljibhai Patel Urological Hospital, Dr. V.V. Desai Road, Nadiad-387 001, Gujarat.

***Author for Correspondence: Dr. Kinnari N. Mistry**

Ashok and Rita Patel Institute of Integrated Study and Research in Biotechnology and Allied Sciences (ARIBAS), Affiliated to Sardar Patel University, Vallabh Vidyanagar, Gujarat, India, 388120.

Article Received on 21/06/2017

Article Revised on 12/07/2017

Article Accepted on 01/08/2017

ABSTRACT

According to the classical description, focal segmental glomerulosclerosis (FSGS) is characterized by the presence of scarring lesion in some (i.e. focal) segmental portion of glomeruli. FSGS patients generally do not respond to steroid treatment, thus despite the availability of a number of agents with variable efficacy in inducing remission, the optimal treatment of patients with steroid resistant nephrotic syndrome (SRNS) is unclear. Genetic causes can be identified in nearly 10% of affected children with highly heterogeneous disorder. Therefore it is important to know about genetic components of underlying cause of SRNS so that patients can be treated effectively before they develop end stage renal disease. This review provides potential clinical application of polymorphisms in candidate genes involved in FSGS-SRNS which may serve as markers in disease prediction. The mutant variants showing genotypic-phenotypic association can be translated to clinical practice through genetic testing. Rather, the most significant outcome of this review will be a better understanding of disease pathogenesis, which will hopefully lead in turn to novel and better treatments and more tailored drug therapy.

KEYWORDS: Nephrotic syndrome, Focal Segmental Glomerulosclerosis, Steroid Resistance Nephrotic Syndrome, SNPs.

Abbreviations: Nephrotic syndrome: NS; Focal Segmental Glomerulosclerosis: FSGS; Steroid Resistance Nephrotic Syndrome: SRNS; Steroid Sensitive Nephrotic Syndrome: SRSS; End Stage Renal Disease: ESRD; Single Nucleotide Polymorphism: SNPs; non-synonymous Single Nucleotide Polymorphism: nsSNP.

INTRODUCTION

FSGS (Focal Segmental Glomerulosclerosis), focal stands for damage in some of the filters of glomeruli leading to sclerosis where the kidney is no more able to filter the blood properly. Focal segmental glomerulosclerosis (FSGS) is one of the most complicated and mysterious histological abrasion, rather than a disease in nephrology (Fig. 1) (Stokes et al., 2006; D'Agati, 2008b). It can happen as a primary disorder without any known cause or as a sickness, secondary to diverse problems. On the basis of the response to standard to corticosteroid therapy NS has been characterized as (1) steroid-resistant NS (SRNS) and (2) steroid-sensitive NS (SSNS). Recognition of the genetics

of FSGS started with understanding the molecular composition of glomerular filtration barrier. It consists of podocytes, basement membrane and fenestrated endothelium (Klahr and Morrissey, 2003). This barrier isolates urinary space from blood under proper physiologic conditions, preventing the unnecessary escape of large molecules having molecular weight higher than 40 kDa, such as albumin and other clotting factors. In the case of NS, including FSGS this glomerular filtration barrier becomes useless and permeable (Caulfield and Farquhar, 1974; Klahr and Morrissey, 2003). Podocytes have been reported as the chief cells in the progression of FSGS (Pardon et al., 2006; Wiggins, 2007). Effacement of the podocyte foot processes is mainly caused by disrupted podocytes, this transformation in podocyte shape requires rearrangement of the actin cytoskeleton but, this is permanent and progressive process in FSGS (Zenker et al., 2009). For decades, there have been multiple broad-based studies going on to interpret probable threats that are influencing FSGS vulnerability, healing response, and development of the NS.

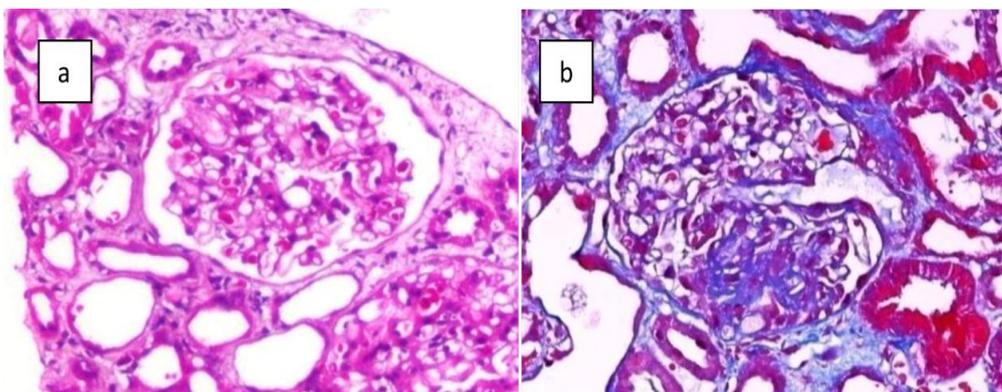


Fig. 1: (a) Biopsy of normal glomeruli under simple light microscopy showing no sclerosis with clear cytoplasmic region. (b) Biopsy of FSGS patients stained with masson trichrome X400, this dye typically stains sclerosis to blue colour indicating expansion in cytoplasmic region of glomeruli (i.e. collapsing glomerulonephropathy).

Epidemiology of FSGS-SRNS

The occurrence of FSGS has been growing worldwide in nearly all racial and age groups over last 20-30 years with a frequency of FSGS is 8-9 per million cases, which is 2-3 folds higher in a rate of disease findings. With the high levels of proteinuria, 50% of FSGS cases reach towards ESRD within 3 to 8 years with the recurrence rate of 20-25% even after kidney transplantation (KT) (Wiggins, 2007; D'Agati, 2008b; Kiffel *et al.*, 2011). In USA, around 35% of renal biopsies proved to have FSGS in adults, among this, 40-50% of patients stopped responding towards steroids treatment whereas in children 63-73% SRNS were found to have FSGS. A UK study estimated the incidence of pediatric SRNS to be 0.3 per 100,000 (i.e. about 20-30 children every year). Over all there were 5-20% cases of SRNS in children leading towards ESRD (Gbadegesin *et al.*, 2007). The male-female ratio for steroid-resistant patients is 1.2:1,

which means male are more prone to this disease (Ruf *et al.*, 2004b). The conventional studies showed that yearly occurrence rates of FSGS in African Americans were considerably higher than Caucasians (0.4 to 1.9 cases per annum, respectively), with high risk of FSGS in black persons (50%) than that of white (35%) (D'Agati, 2008b). There is 1-2 folds increase in the occurrence of FSGS-SRNS in Asian population.

Pathogenesis of FSGS-SRNS

In patients with FSGS-SRNS increase in protein permeability of glomerular basement membrane is caused due to various circulating agents such as the soluble form of the urokinase-type plasminogen activator receptor (uPAR), cardiotrophin like cytokines of immune cells beside the injury to podocytes due to oxidative stress (Fig. 2).

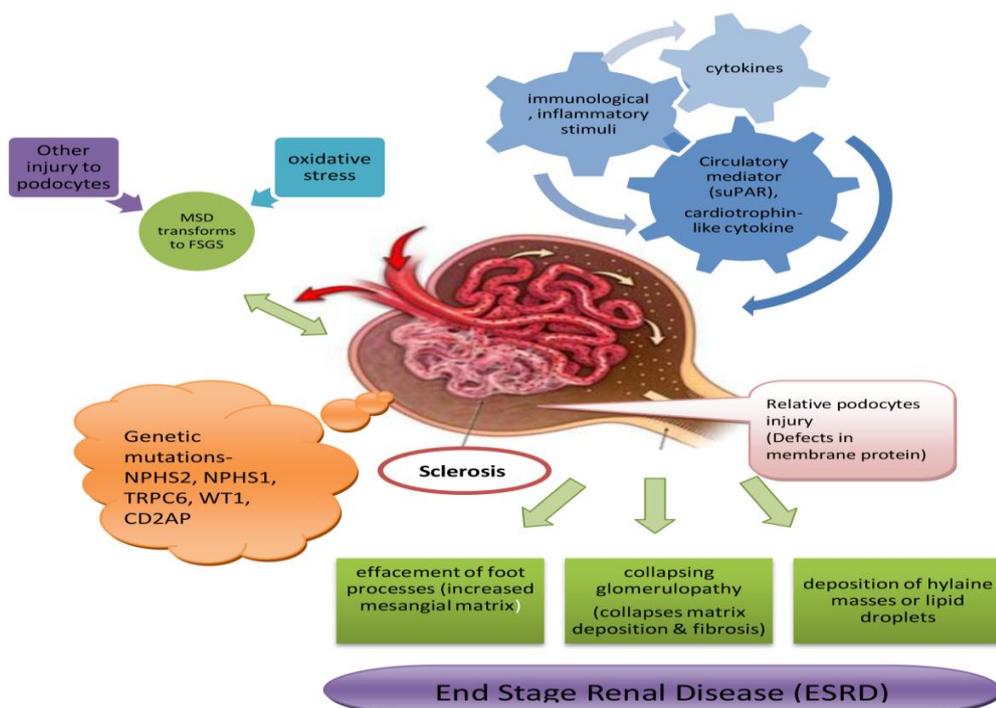


Fig. 2: Pathogenesis of focal segmental glomerulosclerosis.

The principal pathogenesis of FSGS-SRNS is still unidentified, but proof robustly relates the importance of genetic factors (i.e. 10%) (D'Agati, 2008b). A rising number of mutated genes have been recognized that can lead to inherited forms of idiopathic NS. These genes helps in guiding different structural proteins or enzymes that work in harmony to manage the glomerular membrane permeability and take part in various signaling events of regulating podocyte enlargement, segregation, and communications among cell-cell, cell-matrix interactions. Proteinuria results from the damage caused by these transformations in glomerular filtration barrier and in this event, podocytes require their specific epithelial cell markers such as fibroblast specific protein, nephrin, desmin, actin, collagen, and fibronectin (D'Agati, 2008a). The findings of these new podocyte proteins and their mutation study have shed light on the pathogenesis of proteinuria linked with NS and FSGS lesions.

SNPs as biological marker

A new type of marker, named SNPs, enlightening polymorphisms at the DNA level have recently appeared on the picture in dominating the molecular genetics field for human and animal genome studies (Cooper and Krawczak, 1989). SNP is biallelic co-dominant markers with only a single alter base present in a DNA sequence, with a common substitute of two probable nucleotides at

a given location with frequency of 1% or greater within a given population (Wang *et al.*, 1998). The probable chance of SNPs to occur is hardly one in every 1000 to 2000 base pairs, currently released SNP data projected total 160,508,575 SNPs in the human genome (dbSNP web query for build 144: Jun 08, 2015). Non-synonymous SNP (nsSNP) is a single nucleotide replacement occurring within the coding region of a gene which causes an amino acid change in the subsequent protein product which ultimately results in a structural or functional change in the protein product whose consequences may be minor or major phenotypic change accounting for the pathology of disease. Around 50-60% of induced mutations in concerned with inherited genetic disorders are due to nsSNPs (Wang *et al.*, 1998).

Different podocyte genes responsible for FSGS-SRNS

All the identified genetic defects affects gene transcription and assembly of podocyte structure together with actin based cytoskeleton, and adhesion complexes (Hinkes *et al.*, 2007). Cloning techniques were used in the identification of various genes such as NPHS1, NPHS2, TRPC6, WT1 and CD2AP which were involved in podocytes damage and were further confirmed by knockout or transgenic models (Caridi *et al.*, 2001; Rood *et al.*, 2012). The SNP data generated using dbSNP in different podocyte genes is graphically (Fig. 3)

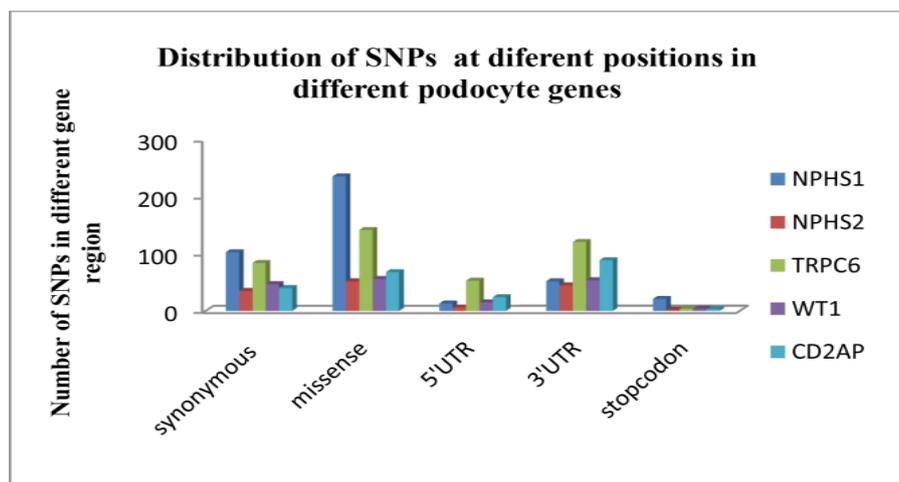


Fig. 3: SNP data generated using NCBI-dbSNP and plotted in a form of bar graph. Targeted genes are showing distribution of SNPs in different regions excluding intronic SNPs. Highest number of SNPs was found to be missense and present in NPHS1 gene than compared to other genes.

NPHS1 gene

The NPHS1 gene is 26 kb in size, contains 29 exons located on chromosome 19. The NPHS1 gene product is nephrin, made up of 1,241 amino acid residues and a member of cell adhesion molecules of immunoglobulin (Ig) family (Lahdenkari *et al.*, 2004). Nephrin contains a transmembrane domain as well as eight Ig like repeated sequence, and one fibronectin III module (Beltcheva *et al.*, 2001). Nephrin is only expressed in visceral cell of glomerular epithelial that form a network of 30 nm globular cross strands possessing elongated pores in zipper fashion (Koziell *et al.*, 2002). Mutation in NPHS1

gene is autosomal recessive type causing congenital NS having massive uteroproproteinuria from birth. Many patients with FSGS are showing mutations in overlapping genes of NPHS1/NPHS2 (Koziell *et al.*, 2002). Reports showed 5 out of 30 patients having joint mutations in the NPHS1/NPHS2 genes, but proper phenotype/genotype correlation evidence was not yet found (Koziell *et al.*, 2002). In 2010, overall 37 different mutations were reported from which 19 were novel in NPHS1 gene, out of those 58% of mutations were missense, whereas others included splice-site and nonsense mutations in 44 unrelated patients (Schoeb *et*

al., 2010). Generally majority of cases are heterozygous compared to homozygote, but the homozygous or compound heterozygous states are more lethal or linked with more severe phenotype (Beltcheva *et al.*, 2001).

NPHS2 gene

The NPHS2 gene size is 25 kb, made up of 8 exons that are located on chromosome 1. The gene contains 383 a.a. residues that encodes for podocin. It is an essential membrane protein that very precisely organizes and regulates structure of glomerular membrane by interacting with NPHS1, CD2AP, TRPC6 and various other genes (Reiterova *et al.*, 2012). Podocin facilitates membrane transport of nephrin and directs podocytes intracellular signaling pathways. Boute *et al.* in 2000 identified NPHS2 gene as causative agent for early onset autosomal recessive SRNS. Overall 6.4-30% of sporadic SRNS were found to have mutations in NPHS2 gene in different parts of the world (Boute *et al.*, 2000; Reiterova *et al.*, 2012).

Even though initial reports showed recessive mutation in NPHS2 as a source of familial SRNS along with ESRD in children happening between the ages of 3 months to 5 years of age, but current data presented its association with a wide range of clinical spectrum (Franceschini *et al.*, 2006). In many patients with 2 mutations of NPHS2 gene showing pathogenic effect develops FSGS in the early age of six years, most of these patients are not responding immunosuppressive treatment and reach towards ESRD (Hinkes *et al.*, 2007). From the time when the NPHS2 gene got identified, various researchers in Europe, the North America and Middle East confirmed NPHS2 mutation to be a common source of sporadic SRNS, taking place in 10-30% of children with sporadic SRNS (Caridi *et al.*, 2001). Podocin variation R229Q is one of most frequently reported one with marginally higher frequency of around 5% in SRNS as compared to healthy individuals (Ruf *et al.*, 2004a).

TRPC6 gene

The TRPC6 gene is made up of 13 exons that are located on chromosome 11, having 931 amino acids encoding for the short transient receptor potential channel with a size of 106325Da (Dietrich *et al.*, 2005). TRPC6 is a part of TRP family, expressed in many tissues that regulate intracellular Ca²⁺ concentration via G protein- coupled receptors (Bach, 2001; Winn *et al.*, 2005b). Reports demonstrated 12 different mutation with familial FSGS and 4 with late onset of sporadic cases (age of 15-55 years, with a few exception at 1-9 years of age) resulting towards unpredictable rate of development to ESRD (Winn *et al.*, 2005a).

Almost all the reported mutations were missense, except two K874X and 89fsX8 mutations. Among all missense mutations 8 (i.e. H218L, P112Q, N125S, E897K, M132T, R895L, Q889K and R895C) were gain-of-function that cause increase in Ca²⁺ current amplitudes where as the rest may probably showing pathogenic

effect on the basis of biochemical and biophysical variations. Majority of TRPC6 mutations were dispersed all throughout N and C terminal cytosolic domains while no mutation has been observed in transmembrane domains. In European and African families, 6 families were recognized having autosomal dominant FSGS with a distinct missense variant (Gudermann, 2005; Reiser *et al.*, 2005).

WT1 gene

The WT1 gene is made up of 10 exons that are located on chromosome 11p13, spanning 48 kb of genomic DNA. A number of case-control studies reported renal phenotypes related to WT1 mutations as a cause of FSGS-SRNS or related with urogenital malformations, these variants are characteristically heterozygous and germline or *de novo* in nature (Gessler *et al.*, 1990; Kaltenis *et al.*, 2004). Till now most of WT1 mutations are observed in 3,6,7,8 and 9 exons, but the majority of these variants were present in zinc finger domains, correspondingly coded in exon 8 and 9, thus significantly affecting NPHS1, mRNA expression levels. Bettina, *et al.* carried out sequencing studies taking 115 sporadic SRNS and 110 SSNS patients having WT1 mutation (exon 6-9), 6-8% SRNS patients were found with mutation in exon 8 and 9 (Hu *et al.*, 2004; Kaltenis *et al.*, 2004; Mucha *et al.*, 2006). Orloff *et al.* demonstrated WT1 involvement with FSGS in African America population where 218 were FSGS cases and the genotyping results showed total 8 SNPs in different regions of WT1 (Orloff *et al.*, 2005).

CD2AP gene

The CD2AP represents CD2-associated protein, which is a cytoplasmic ligand molecule for T-cell adhesion protein CD2 with a size of 80 kDa is generally expressed in almost all tissues excluding the brain. In kidneys, CD2AP plays a significant role in ultrafiltration processes of slit-diaphragm by interacting with nephrin and podocin (Schwarz *et al.*, 2001; Shih *et al.*, 2001). In animals, CD2AP mutations associated with FSGS gave rise to mesangial cell proliferation leading to glomerulosclerosis, whereas such association is less seen in humans (Shih *et al.*, 2001). In African Americans, there has been a report on heterozygous nucleotide variant that causes CD2AP splicing resulting to FSGS (Gigante *et al.*, 2009). In recent studies, homozygous mutation in CD2AP resulted in premature stop codon to some extent forming a truncated protein. The protein formed down regulated CD2AP expression by lymphocytes and the binding with F-actin (Shih *et al.*, 2001; Gigante *et al.*, 2009).

Benefits and applications of genetic studies

A lack of controlled studies has hindered development of effective treatment. Understanding the interaction between various factors is critical to developing new strategies for treating patients with disease. The discovery of mutation could benefit the patient by avoiding exposure to prolonged treatment with steroids

and/ or the use of alkylating agent. Genetic screening might help in making treatment decisions, patients care and counseling during their follow-up, counseling of the family. Identification of genetic mutation in a child can help the parents in their decision to plan new pregnancies and also, the results can be used for prenatal genetic testing.

ACKNOWLEDGEMENT

Authors are grateful to Charutar Vidya Mandal (CVM) and Anand Agriculture University VallabhVidyanagar, Gujarat for providing platform for this research work.

REFERENCES

- Bach G. Mucopolidosis type IV. *Mol Genet Metab*, 2001; 73: 197-203.
- Beltcheva O, Martin P, Lenkkeri U and Tryggvason K. Mutation spectrum in the nephrin gene (NPHS1) in congenital nephrotic syndrome. *Hum Mutat*, 2001; 17: 368-73.
- Boute N, Gribouval O, Roselli S, Benessy F, Lee H, Fuchshuber A, Dahan K, Gubler MC, Niaudet P and Antignac C. NPHS2, encoding the glomerular protein podocin, is mutated in autosomal recessive steroid-resistant nephrotic syndrome. *Nat Genet*, 2000; 24: 349-54.
- Caridi G, Bertelli R, Carrea A, Di Duca M, Catarsi P, Artero M, Carraro M, Zennaro C, Candiano G, Musante L, Seri M, Ginevri F, Perfumo F and Ghiggeri GM. Prevalence, genetics, and clinical features of patients carrying podocin mutations in steroid-resistant nonfamilial focal segmental glomerulosclerosis. *J Am Soc Nephrol*, 2001; 12: 2742-6.
- Caulfield JP. and Farquhar MG. The permeability of glomerular capillaries to graded dextrans. Identification of the basement membrane as the primary filtration barrier. *J Cell Biol*, 1974; 63: 883-903.
- Cooper DN and Krawczak M. Cytosine methylation and the fate of CpG dinucleotides in vertebrate genomes. *Hum Genet*, 1989; 83: 181-8.
- D'Agati VD. Podocyte injury in focal segmental glomerulosclerosis: Lessons from animal models (a play in five acts). *Kidney Int*, 2008a; 73: 399-406.
- D'Agati VD. The spectrum of focal segmental glomerulosclerosis: new insights. *Curr Opin Nephrol Hypertens*, 2008b; 17: 271-81.
- Dietrich A, Mederos YSM, Gollasch M, Gross V, Storch U, Dubrovskaja G, Obst M, Yildirim E, Salanova B, Kalwa H, Essin K, Pinkenburg O, Luft FC, Gudermann T and Birnbaumer L. Increased vascular smooth muscle contractility in TRPC6-/- mice. *Mol Cell Biol*, 2005; 25: 6980-9.
- Franceschini N, North KE, Kopp JB, McKenzie L and Winkler C. NPHS2 gene, nephrotic syndrome and focal segmental glomerulosclerosis: a HuGE review. *Genet Med*, 2006; 8: 63-75.
- Gbadegesin R, Hinkes B, Vlangos C, Mucha B, Liu J, Hopcian J and Hildebrandt F. Mutational analysis of NPHS2 and WT1 in frequently relapsing and steroid-dependent nephrotic syndrome. *Pediatr Nephrol*, 2007; 22: 509-13.
- Gessler M, Poustka A, Cavenee W, Neve RL, Orkin SH and Bruns GA. Homozygous deletion in Wilms tumours of a zinc-finger gene identified by chromosome jumping. *Nature*, 1990; 343: 774-8.
- Gigante M, Pontrelli P, Montemurno E, Roca L, Aucella F, Penza R, Caridi G, Ranieri E, Ghiggeri GM and Gesualdo L. CD2AP mutations are associated with sporadic nephrotic syndrome and focal segmental glomerulosclerosis (FSGS). *Nephrol Dial Transplant*, 2009; 24: 1858-64.
- Gudermann T. A new TRP to kidney disease. *Nat Genet*, 2005; 37: 663-4.
- Hinkes BG, Mucha B, Vlangos CN, Gbadegesin R, Liu J, Hasselbacher K, Hangan D, Ozaltin F, Zenker M and Hildebrandt F. Nephrotic syndrome in the first year of life: two thirds of cases are caused by mutations in 4 genes (NPHS1, NPHS2, WT1, and LAMB2). *Pediatrics*, 2007; 119: e907-19.
- Hu M, Craig J, Howard N, Kan A, Chaitow J, Little D and Alexander SI. A novel mutation of WT1 exon 9 in a patient with Denys-Drash syndrome and pyloric stenosis. *Pediatr Nephrol*, 2004; 19: 1160-3.
- Kaltenis P, Schumacher V, Jankauskiene A, Laurinavicius A and Royer-Pokora B. Slow progressive FSGS associated with an F392L WT1 mutation. *Pediatr Nephrol*, 2004; 19: 353-6.
- Kiffel J, Rahimzada Y and Trachtman H. Focal segmental glomerulosclerosis and chronic kidney disease in pediatric patients. *Adv Chronic Kidney Dis*, 2011; 18: 332-8.
- Klahr S and Morrissey J. Progression of chronic renal disease. *Am J Kidney Dis*, 2003; 41: S3-7.
- Koziell A, Grech V, Hussain S, Lee G, Lenkkeri U, Tryggvason K and Scambler P. Genotype/phenotype correlations of NPHS1 and NPHS2 mutations in nephrotic syndrome advocate a functional inter-relationship in glomerular filtration. *Hum Mol Genet*, 2002; 11: 379-88.
- Lahdenkari AT, Kestila M, Holmberg C, Koskimies O and Jalanko H. Nephrin gene (NPHS1) in patients with minimal change nephrotic syndrome (MCNS). *Kidney Int*, 2004; 65: 1856-63.
- Mucha B, Ozaltin F, Hinkes BG, Hasselbacher K, Ruf RG, Schultheiss M, Hangan D, Hoskins BE, Everding AS, Bogdanovic R, Seeman T, Hoppe B, Hildebrandt F and Members of the, A.P.N.S.G. Mutations in the Wilms' tumor 1 gene cause isolated steroid resistant nephrotic syndrome and occur in exons 8 and 9. *Pediatr Res*, 2006; 59: 325-31.
- Orloff MS, Iyengar SK, Winkler CA, Goddard KA, Dart RA, Ahuja TS, Mokrzycki M, Briggs WA, Korbet SM, Kimmel PL, Simon EE, Trachtman H, Vlahov D, Michel DM, Berns JS, Smith MC, Schelling JR, Sedor JR and Kopp JB. Variants in the Wilms' tumor gene are associated with focal segmental glomerulosclerosis in the African

- American population. *Physiol Genomics*, 2005; 21: 212-21.
24. Pardon A, Audard V, Caillard S, Moulin B, Desvaux D, Bentaarit B, Remy P, Sahali D, Roudot-Thoraval F, Lang P and Grimbert P. Risk factors and outcome of focal and segmental glomerulosclerosis recurrence in adult renal transplant recipients. *Nephrol Dial Transplant*, 2006; 21: 1053-9.
 25. Reiser J, Polu KR, Moller CC, Kenlan P, Altintas MM, Wei C, Faul C, Herbert S, Villegas I, Avila-Casado C, McGee M, Sugimoto H, Brown D, Kalluri R, Mundel P, Smith PL, Clapham DE and Pollak MR. TRPC6 is a glomerular slit diaphragm-associated channel required for normal renal function. *Nat Genet*, 2006; 37: 739-44.
 26. Reiterova J, Safrankova H, Obeidova L, Stekrova J, Maixnerova D, Merta M and Tesar V. Mutational analysis of the NPHS2 gene in Czech patients with idiopathic nephrotic syndrome. *Folia Biol (Praha)*, 2012; 58: 64-8.
 27. Rood IM, Deegens JK and Wetzels JF. Genetic causes of focal segmental glomerulosclerosis: implications for clinical practice. *Nephrol Dial Transplant*, 2012; 27: 882-90.
 28. Ruf RG, Lichtenberger A, Karle SM, Haas JP, Anacleto FE, Schultheiss M, Zalewski I, Imm A, Ruf EM, Mucha B, Bagga A, Neuhaus T, Fuchshuber A, Bakkaloglu A, Hildebrandt F and Arbeitsgemeinschaft Fur Padiatrische Nephrologie Study G. Patients with mutations in NPHS2 (podocin) do not respond to standard steroid treatment of nephrotic syndrome. *J Am Soc Nephrol*, 2004a; 15: 722-32.
 29. Ruf RG, Schultheiss M, Lichtenberger A, Karle SM, Zalewski I, Mucha B, Everding AS, Neuhaus T, Patzer L, Plank C, Haas JP, Ozaltin F, Imm A, Fuchshuber A, Bakkaloglu A, Hildebrandt F and Group A.P.N.S. Prevalence of WT1 mutations in a large cohort of patients with steroid-resistant and steroid-sensitive nephrotic syndrome. *Kidney Int*, 2004b; 66: 564-70.
 30. Schoeb DS, Chernin G, Heeringa SF, Matejas V, Held S, Vega-Warner V, Bockenhauer D, Vlangos CN, Moorani KN, Neuhaus TJ, Kari JA, MacDonald J, Saisawat P, Ashraf S, Ovunc B, Zenker M, Hildebrandt F and Gessellschaft fur Paediatrische Nephrologie Study G. Nineteen novel NPHS1 mutations in a worldwide cohort of patients with congenital nephrotic syndrome (CNS). *Nephrol Dial Transplant*, 2010; 25: 2970-6.
 31. Schwarz K, Simons M, Reiser J, Saleem MA, Faul C, Kriz W, Shaw AS, Holzman LB and Mundel P. Podocin, a raft-associated component of the glomerular slit diaphragm, interacts with CD2AP and nephrin. *J Clin Invest*, 2001; 108: 1621-9.
 32. Shih NY, Li J, Cotran R, Mundel P, Miner JH and Shaw AS. CD2AP localizes to the slit diaphragm and binds to nephrin via a novel C-terminal domain. *Am J Pathol*, 2001; 159: 2303-8.
 33. Stokes MB, Valeri AM, Markowitz GS and D'Agati VD. Cellular focal segmental glomerulosclerosis: Clinical and pathologic features. *Kidney Int*, 2006; 70: 1783-92.
 34. Wang DG, Fan J.B, Siao CJ, Berno A, Young P, Sapolsky R, Ghandour G, Perkins N, Winchester E, Spencer J, Kruglyak L, Stein L, Hsie L, Topaloglou T, Hubbell E, Robinson E, Mittmann M, Morris MS, Shen N, Kilburn D, Rioux J, Nusbaum C, Rozen S, Hudson TJ, Lipshutz R, Chee M and Lander ES. Large-scale identification, mapping, and genotyping of single-nucleotide polymorphisms in the human genome. *Science*, 1998; 280: 1077-82.
 35. Wiggins RC. The spectrum of podocytopathies: a unifying view of glomerular diseases. *Kidney Int*, 2007; 71: 1205-14.
 36. Winn MP, Conlon PJ, Lynn KL, Farrington MK, Creazzo T, Hawkins AF, Daskalakis N., Kwan SY, Ebersviller S, Burchette JL, Pericak-Vance MA, Howell DN, Vance JM and Rosenberg PB. A mutation in the TRPC6 cation channel causes familial focal segmental glomerulosclerosis. *Science*, 2005a; 308: 1801-4.
 37. Winn MP, Conlon PJ, Lynn KL, Farrington MK, Creazzo T, Hawkins AF, Daskalakis N, Kwan SY, Ebersviller S, Burchette JL, Pericak-Vance MA, Howell DN, Vance JM and Rosenberg PB. A mutation in the TRPC6 cation channel causes familial focal segmental glomerulosclerosis. *Science*, 2005b; 308: 1801-1804.
 38. Zenker M, Machuca E and Antignac C. Genetics of nephrotic syndrome: new insights into molecules acting at the glomerular filtration barrier. *J Mol Med (Berl)*, 2009; 87: 849-57.