



ANTIBACTERIAL POTENTIAL AND GC-MS PROFILING OF METHANOLIC LEAF EXTRACT OF *VITEX NEGUNDO* LINN.

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ABSTRACT

The current study was designed to catalyze the primary metabolites and their authentication by using GC-MS analysis and antibacterial power of methanolic leaf extract of *Vitex negundo*. The antibacterial potential of the methanol leaf extract of *Vitex negundo* was tested against *Bacillus cereus*, *Proteus vulgaris* and *Proteus mirabilis* using by agar well diffusion method. The higher zone of inhibition (55.03mm) was observed against the bacterium *P.vulgaris* at 20μl and 25μl concentration of methanol leaf extract. GC-MS analysis confirms the occurrence of more than 16 different compounds in the methanol leaf extract of *Vitex negundo*.

KEYWORDS: *Vitex negundo*, Antibacterial activity, Disc diffusion assay, GC-MS.

INTRODUCTION

Systematic investigations of medicinal plants have started in many countries because of their contributions to health care. The primary benefits of using plant based medicines are relatively safer than synthetic alternatives, offering marked therapeutic benefits and more reasonable treatment. (Ahmad et al). Thus, over 50% of these modern drugs are of natural product origin and these natural products play an important role in the drug development in the pharmaceutical industry. (Jeyachandran et al). In general, bacteria have the genetic ability to transfer and gain resistance to drugs used as therapeutic agents. The only way to prevent antibiotic resistance is by using new compounds which have not based on the existing synthetic anti-microbial agents. (Shah et al). To promote the use of medicinal plants as potential sources of anti-microbial compounds, it is important to thoroughly find out their composition and activity and thus confirm their use. (Nair et al) Plants and their active constituents play an important role in the prevention of a variety of ailments. *Vitex negundo* Linn., Verbenaceae, known as Nirgundi in Hindi, grows gregariously in wastelands and is also planted as a hedge-plant. It is an erect, 2–5 m in height, slender tree with quadrangular branchlets distributed throughout India. The leaves have five leaflets in a palmately arrangement, which are lanceolate, 4–10 cm long, hairy beneath and pointed at both ends. The bluish purple flowers are numerous. The fruit is succulent, black when ripe, rounded and about 4 mm in diameter (Chopra, Nayar, & Chopra, 1956; Wealth of India, 1976).

Although all parts *V. negundo* are used as medicine in the indigenous system of medicine, the leaves are the most potent for medicinal use. It is used for treatment of eye-disease, toothache, inflammation, leucoderma, enlargement of the spleen, skin-ulcers, in catarrhal fever, rheumatoid arthritis, gonorrhoea, and bronchitis. They are also used as tonics, vermifuge, lactagogue, emmenagogue, antibacterial, antipyretic and antihistaminic agents (Chaturvedi & Singh, 1965). Recently, attention has been focused on the medicinal properties of the extracts. The present study was aimed to evaluate the antibacterial potential of methanol leaf extract of *Vitex negundo* against bacterial pathogens and identify the secondary metabolites **through GCMS study**. The result also indicated that scientific studies carried out on medicinal plant having traditional claims of effectiveness might warrant fruitful results.

MATERIALS AND METHODS

Materials

The plant *Vitex negundo* Linn. was collected from Adhiparasakthi Agricultural College, Kalavai, Tamil Nadu. Methanol and required chemicals (HPLC grade) were procured from Merck, India. The plant was identified by the scientist of the Institute.

Methods

The leaves of *Vitex negundo* Linn was collected, washed, air dried in shadow and grinded by mixer grinder. After grinding, 5 gm of plant material was extracted in 50 ml of methanol solvent separately at

room temperature for 6 hours. The organic solvent was filtered by whatman filter paper till clear solution was obtained. Solvent was evaporated in a rotatory evaporator (Buchi, Switzerland) under reduced pressure (vacuum) at 40-50°C till complete dryness.

Extract yield percentage: The methanolic extract was transferred in preweighed beaker and was dried. Further the final weight of the beaker with extract was measured. Then the extract yield was calculated. Noted in Table 1. Extract yield % = ((Extract + Beaker weight) - (Empty beaker weight))/Sample weight *100.

Sample Preparation

6mg of methanolic extract of VN was suspended in 1 ml of methanol and from that 0.005, 0.010, 0.015, 0.020 ml and 0.025 ml was used for the test to give a concentration 30, 60, 90, 120 and 150 mcg each.

Microorganisms used and growth conditions

The following organisms were used in this study and they consist of both Gram positive and Gram negative bacteria: *Bacillus cereus*, *Proteus vulgaris* and *Proteus mirabilis*. The bacterial strains were maintained at 40°C and their stock was kept in 10% glycerine saline at – 20°C. Antibacterial activity Sensitivity of different test bacterial strains to various extracts of *Vitex negundo Linn* was measured by agar well diffusion method.(Raman BV et al., 2005, raman BV et al 2009). Zone of inhibition was determined using Himedia zone of inhibition scale and results are expressed in millimeters (mm). For each combination of extract and the bacterial strain, the experiment was performed in triplicate. The bacteria with a clear zone of inhibition of more than 8 mm were considered to be sensitive.

GC-MS analysis

The chromatographic procedure was performed using GC-MS model MSQP2010 (Shimadzu, Kyoto, Japan) with auto sampler. 1000 ppm solution in methanol was prepared from methanolic extracts and 1 µL of methanolic extract was injected for analysis using DB - 5MS column (30 meter × 0.25 mm, film thickness 0.25µm). Helium gas was used at flow rate 1ml/min. as a carrier gas. The analysis was carried out using oven programming of initial temperature 50°C for 2 minutes followed by ramp rate of 20°C/minute up to 130°C followed by ramp of 12°C/min. to a temperature of 180°C, finally raised temperature to 280°C at 3°C per minute and hold for 15 minutes. The ion source temperature was set at 250°C. The injection port temperature was set as a 250°C and the total run time was 58.5 minute. The instrument was operated in electron impact (EI) mode with electron energy 70ev. Confirmation of analytes was done by SIM (selective ion mode) mode.(abirami et al 2012, prajapathi R et al., 2014).

RESULT AND DISCUSSION

Antibacterial activity of methanolic leaf extract of *Vitex nigundo linn*.

The antibacterial activity of methanolic leaf extracts was evaluated against three opportunistic bacterial pathogens. The results of antibacterial activity of different concentrations of *Vitex nigundo linn leaf extract* were summarized in Table 2. The results revealed that methanolic leaf extract of *Vitex nigundo linn* showed the maximum activity against *P.vulgaris* at 20 and 25 µl concentration. The antibacterial activity was distinctly varied against the bacterial test pathogens. Kanamycin (30 µg) was used as a reference standard. The results of well diffusion method and minimum inhibitory concentration shown in figure 6, 7 and in Table 2. From the results, it is identified that the methanolic leaf extract of *Vitex nigundo linn* exhibited significant antibacterial activity when compared with the standard antibiotics tested.

Assessment of phytocomponents in methanolic leaf extract of *Vitex nigundo linn*

P.vulgaris showed the significant antibacterial activity. Consequently, it was subjected to GC-MS analysis. The mass spectrum of methanol extract showed more than 16 prominent peaks. The major phytoconstituents were phosphonic acid, ledol, benzene methanol 1 - t - butyl - 4 (adamantly-1) benzene, Z-5,17 - octadecadiene - 1-ol acetate, 6, 11 - Undecadiene, 1-acetoxy - 3,7-dimethyl, 4,(2,2,6-trimethyl - bicycle(4.1.0) hept - 1-yl)-butane - 2-one, heptacosone, dotriacontane, hydroxyl dehydro steric acid showed in figures 8.

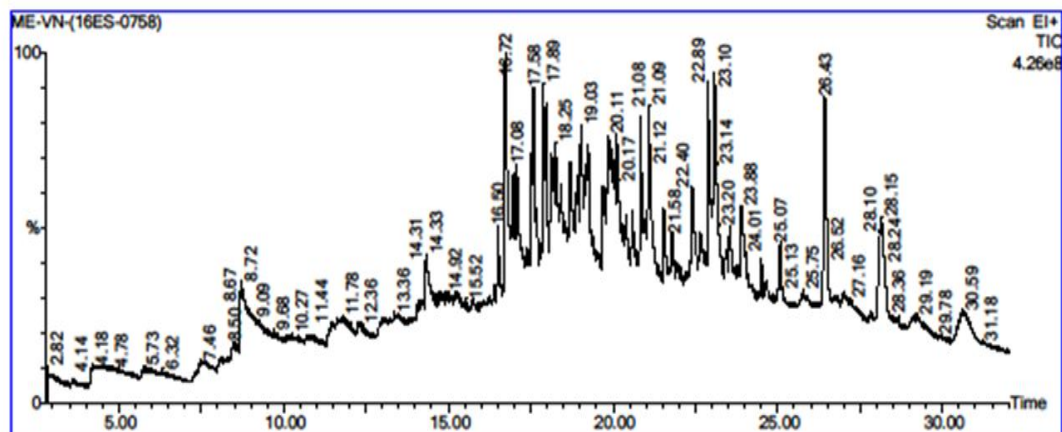
Qualitative Report

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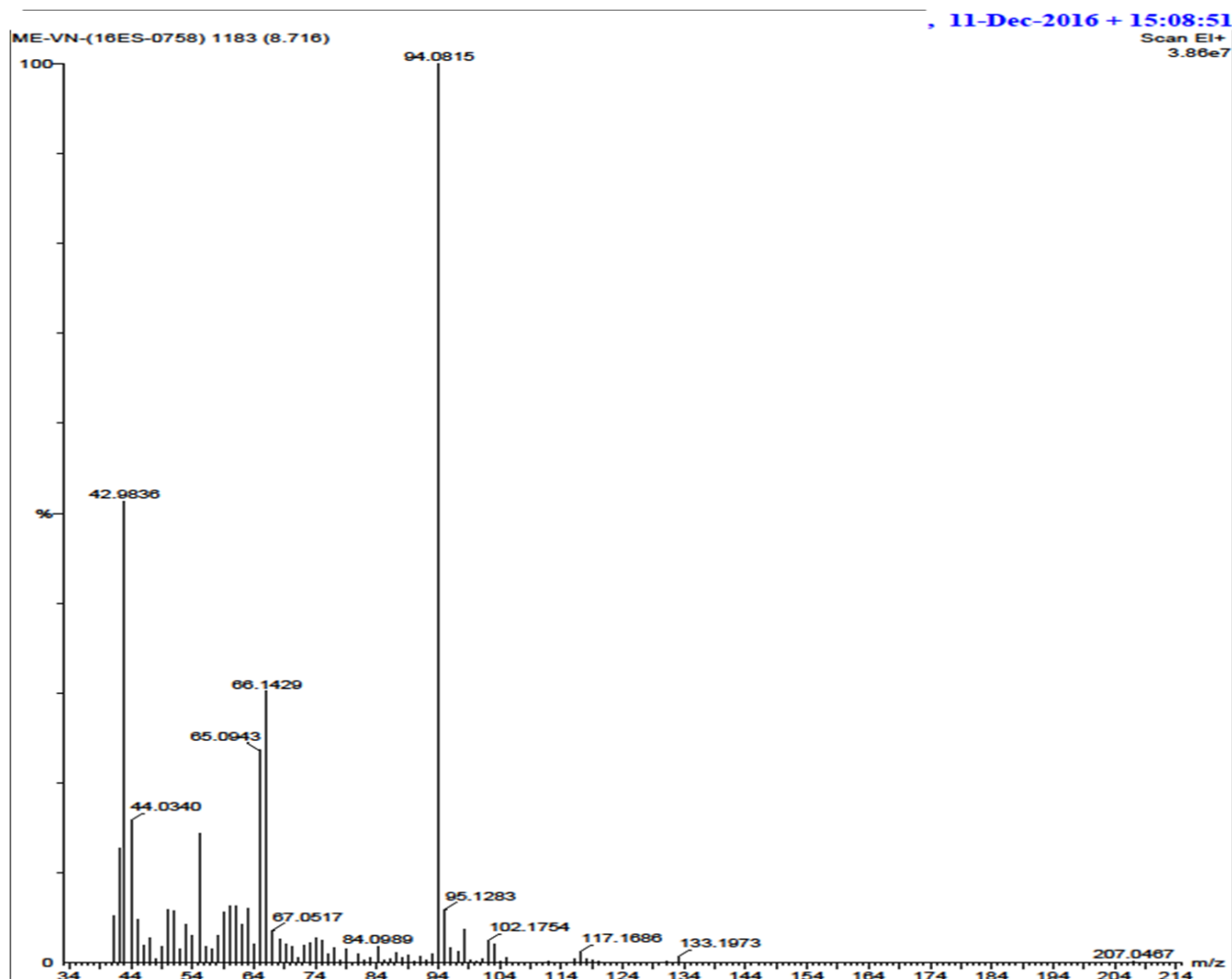
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#	RT	Scan	Height	Area	Area %	Norm %
1	8.716	1183	84,031,136	38,545,300.0	13.607	100.00
2	14.333	2306	63,814,992	8,131,971.5	2.871	21.10
3	16.724	2784	291,537,376	26,909,288.0	9.499	69.81
4	16.974	2834	131,878,488	12,450,566.0	4.395	32.30
5	17.084	2856	136,212,400	12,510,705.0	4.417	32.46
6	19.250	3289	104,280,000	10,334,188.0	3.648	26.81
7	19.855	3410	108,226,008	21,277,508.0	7.511	55.20
8	20.105	3460	116,934,088	9,349,389.0	3.301	24.26
9	20.831	3605	172,088,352	15,010,450.0	5.299	38.94
10	21.086	3656	185,753,408	16,920,506.0	5.973	43.90
11	22.416	3922	104,642,848	10,684,571.0	3.772	27.72
12	22.886	4016	219,656,608	16,577,576.0	5.852	43.01
13	23.072	4053	233,424,944	27,418,176.0	9.679	71.13
14	23.882	4215	84,661,112	9,883,107.0	3.489	25.64
15	26.428	4724	250,164,976	23,759,578.0	8.388	61.64
16	28.154	5069	124,024,704	23,508,688.0	8.299	60.99

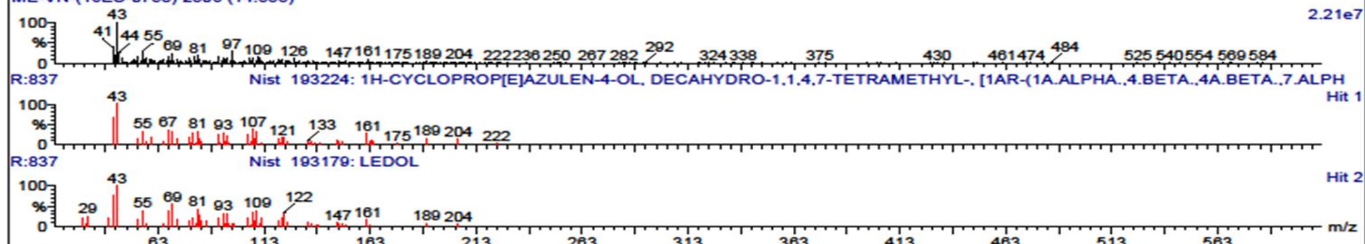
Inst() ACQUISITION PARAMETERS

Oven: Initial temp 60°C for 2 min, ramp 10°C/min to 300°C, hold 6 min, InjAauto=260°C, Volume=0 µL, Split=10:1, Carrier Gas=He, Solvent Delay=2.80 min, Transfer Temp=240°C, Source Temp=240°C, Scan: 40 to 600Da, Column 30.0m x 250µm

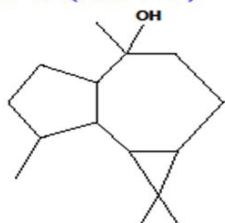
Figure 8: GC-MS profiling along with chromatogram of methanolic leaf extract of *Vitex negundo linn*

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ME-VN-(16ES-0758) 2306 (14.333)



ME-VN-(16ES-0758)

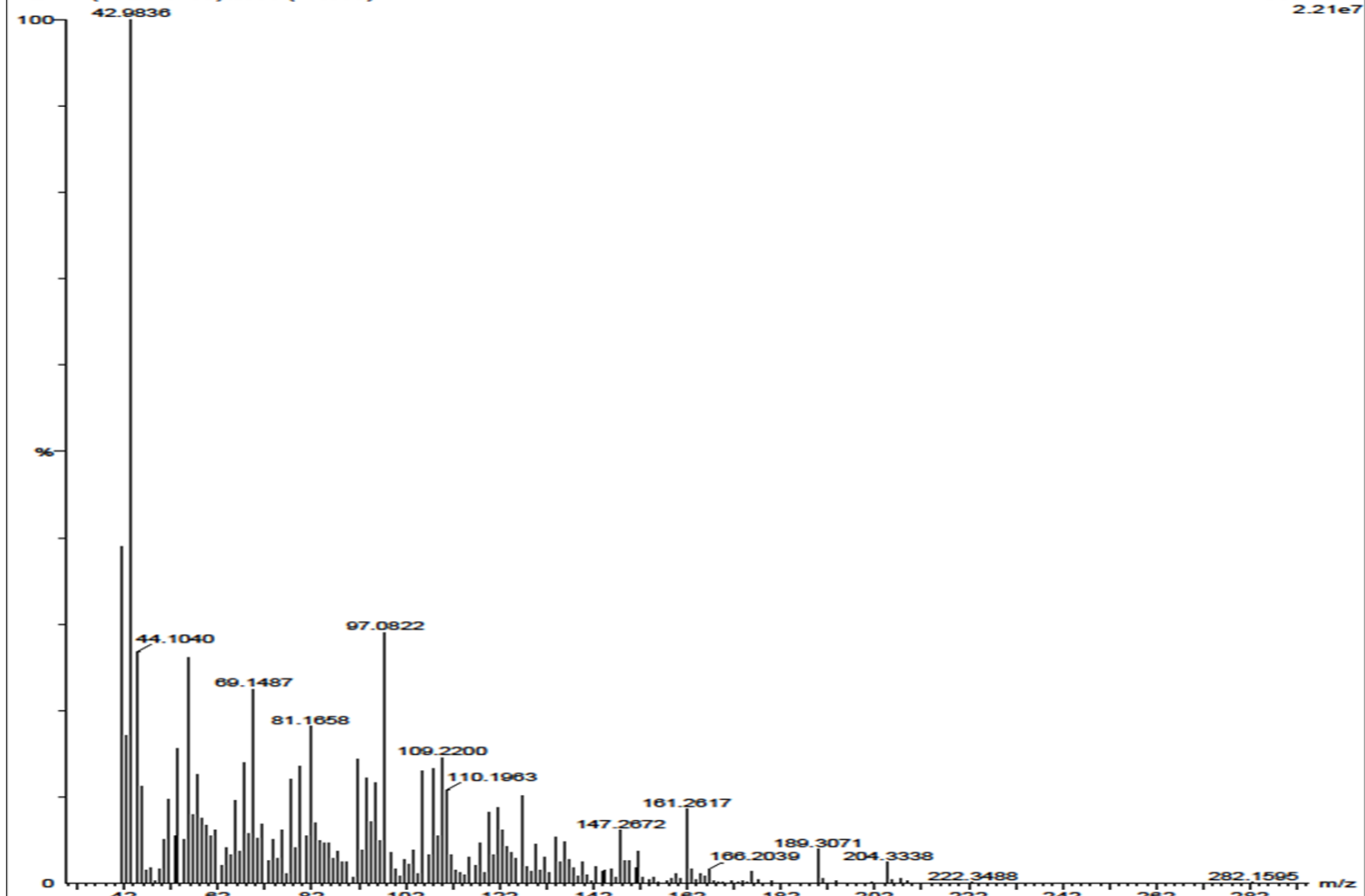


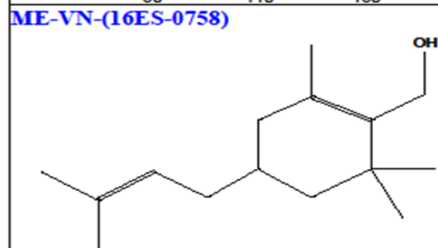
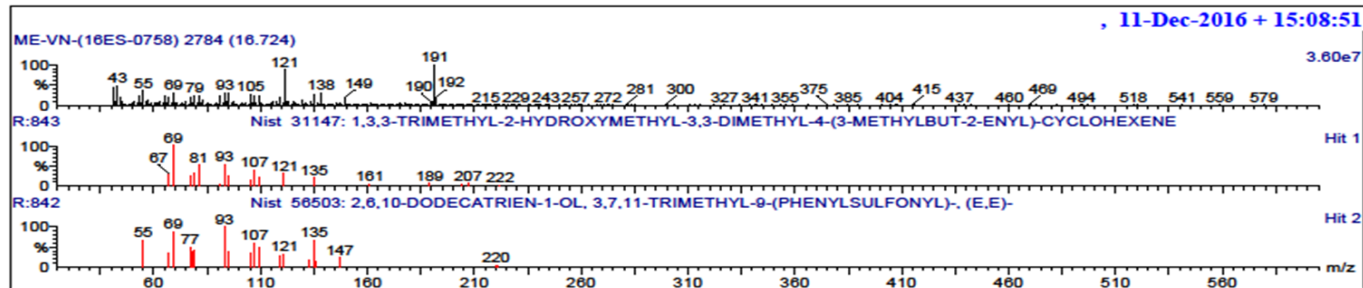
Hit	REV	for	Compound Name	M.W.	Formula	CAS
1	837	632	1H-CYCLOPROP[E]AZULEN-4-OL, DECAHYDRO-1,1,4,7-TETRAMETHYL-, [1AR-(1A.A	222	C15H26O	552-02-3
2	837	643	LEDOL	222	C15H26O	577-27-5
3	829	602	LEDOL	222	C15H26O	577-27-5
4	829	602	VERIDIFLOROL	222	C15H26O	900122-17-3
5	827	649	3-BUTEN-2-ONE, 4-(3-HYDROXY-6,8-DIMETHYL-2-METHYLENOCYCLOHEXYL)-	208	C13H20O2	900142-31-3
6	820	619	1H-CYCLOPROP[E]AZULEN-4-OL, DECAHYDRO-1,1,4,7-TETRAMETHYL-, [1AR-(1A.A	222	C15H26O	552-02-3
7	819	600	1H-CYCLOPROP[E]AZULEN-4-OL, DECAHYDRO-1,1,4,7-TETRAMETHYL-, [1AR-(1A.A	222	C15H26O	552-02-3
8	804	594	GLOBULOL	222	C15H26O	51371-47-2
9	786	571	7-DEHYDROCHOLESTERYL ISOCAPROATE	482	C33H54O2	900251-07-6
10	780	534	CYCLOARTANOL	428	C30H52O	4657-58-3
11	771	519	9,19-CYCLOLANOST-24-EN-3-OL, ACETATE, (3.BETA.)-	468	C32H52O2	1259-10-5
12	769	566	LEDENE OXIDE-(I)	220	C15H24O	900151-93-3
13	765	602	5,8-DECADIEN-2-ONE, 5,9-DIMETHYL-, (E)-	180	C12H20O	130876-99-2
14	763	584	3,5,9-TRIMETHYL-DECA-2,4,8-TRIEN-1-OL	194	C13H22O	900188-00-3
15	757	566	1H-BENZOCYCLOHEPTEN-7-OL, 2,3,4,4A,5,6,7,8-OCTAHYDRO-1,1,4A,7-TETRAMET	222	C15H26O	6892-80-4
16	754	512	9,19-CYCLOERGOST-24(28)-EN-3-OL, 4,14-DIMETHYL-, ACETATE, (3.BETA., 4.ALPH	468	C32H52O2	10376-42-8
17	753	568	1,6-DIMETHYL-9-(1-METHYLETHYLIDENE)-5,12-DIOXATRICYCLO[9.1.0.0(4,6)]DODE	250	C15H22O3	900072-83-2

ME-VN-(16ES-0758)

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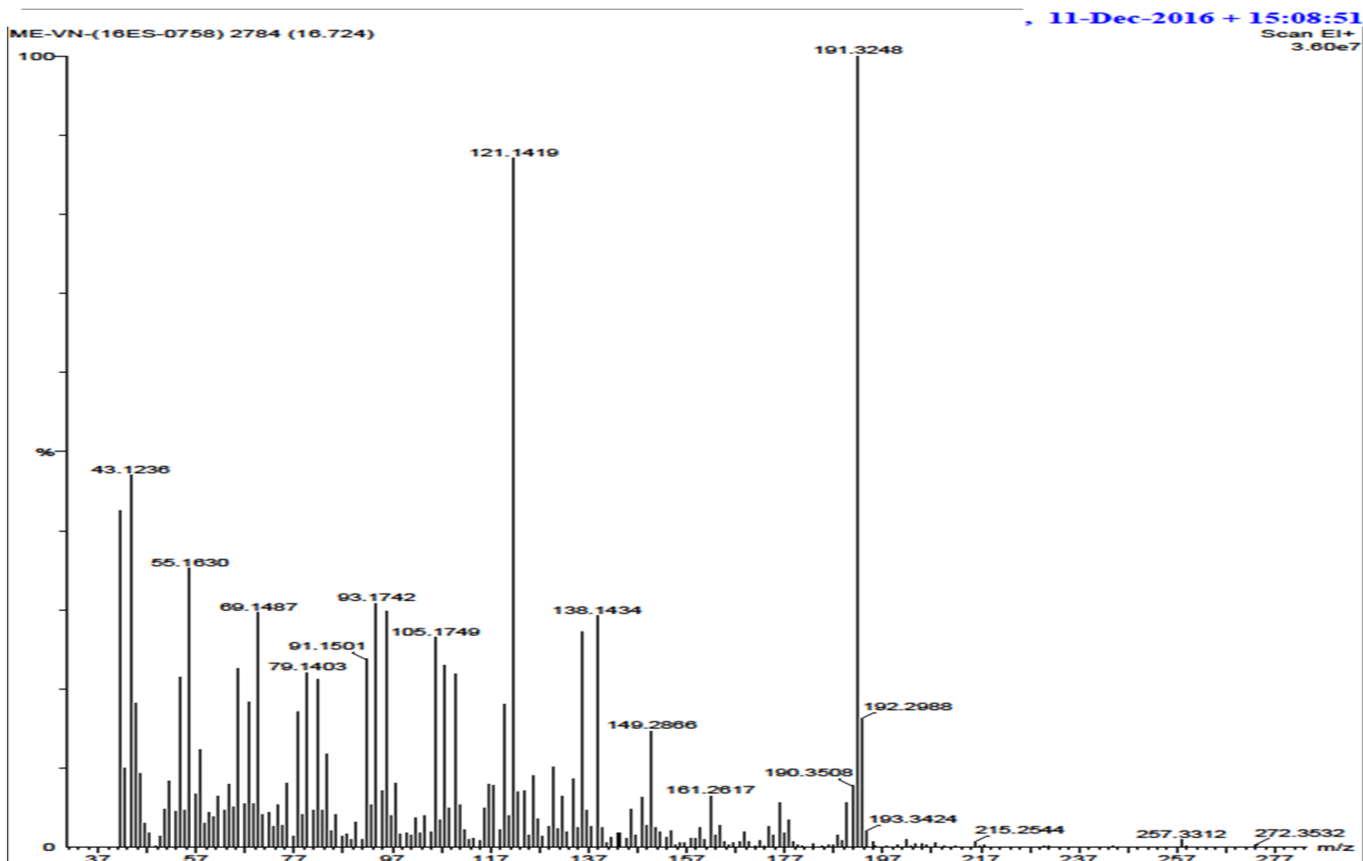
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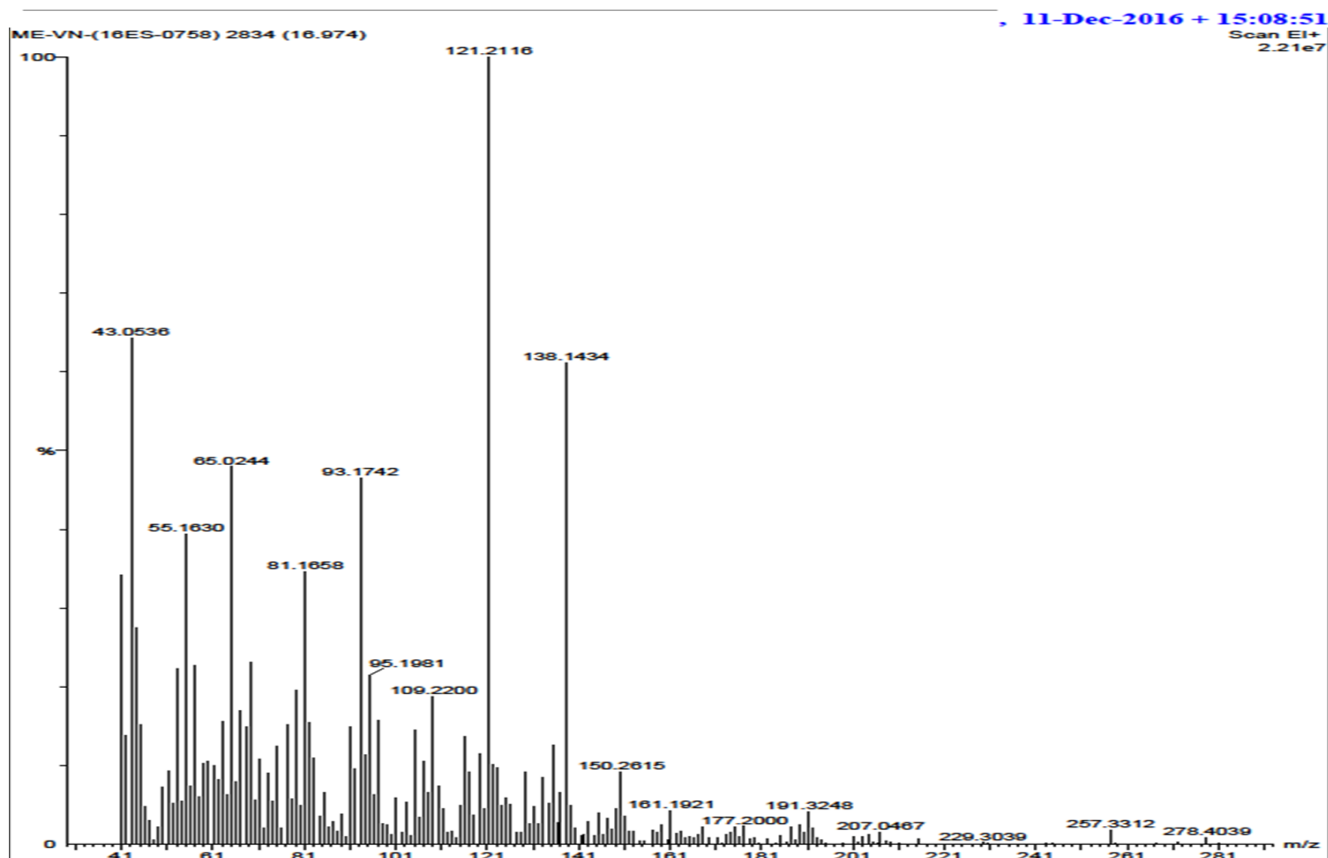
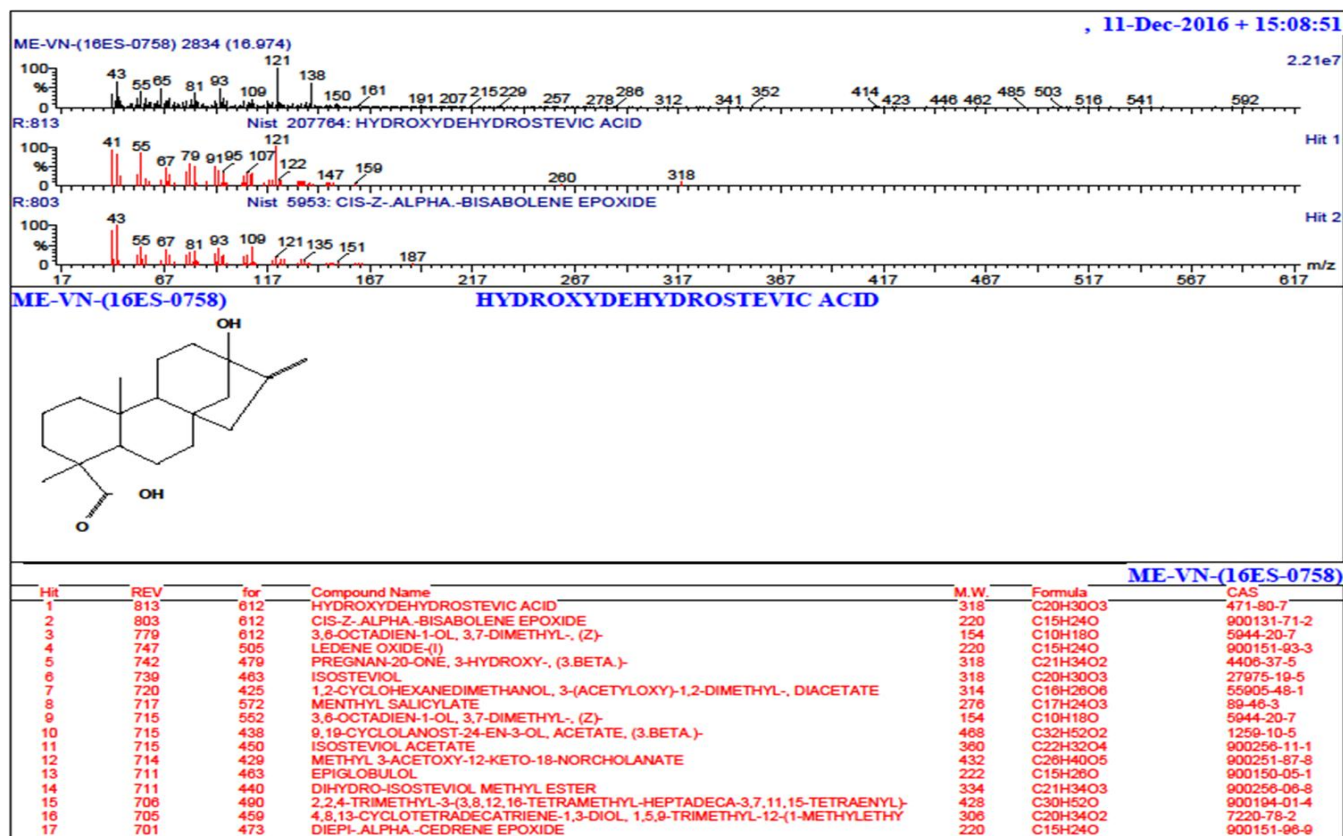


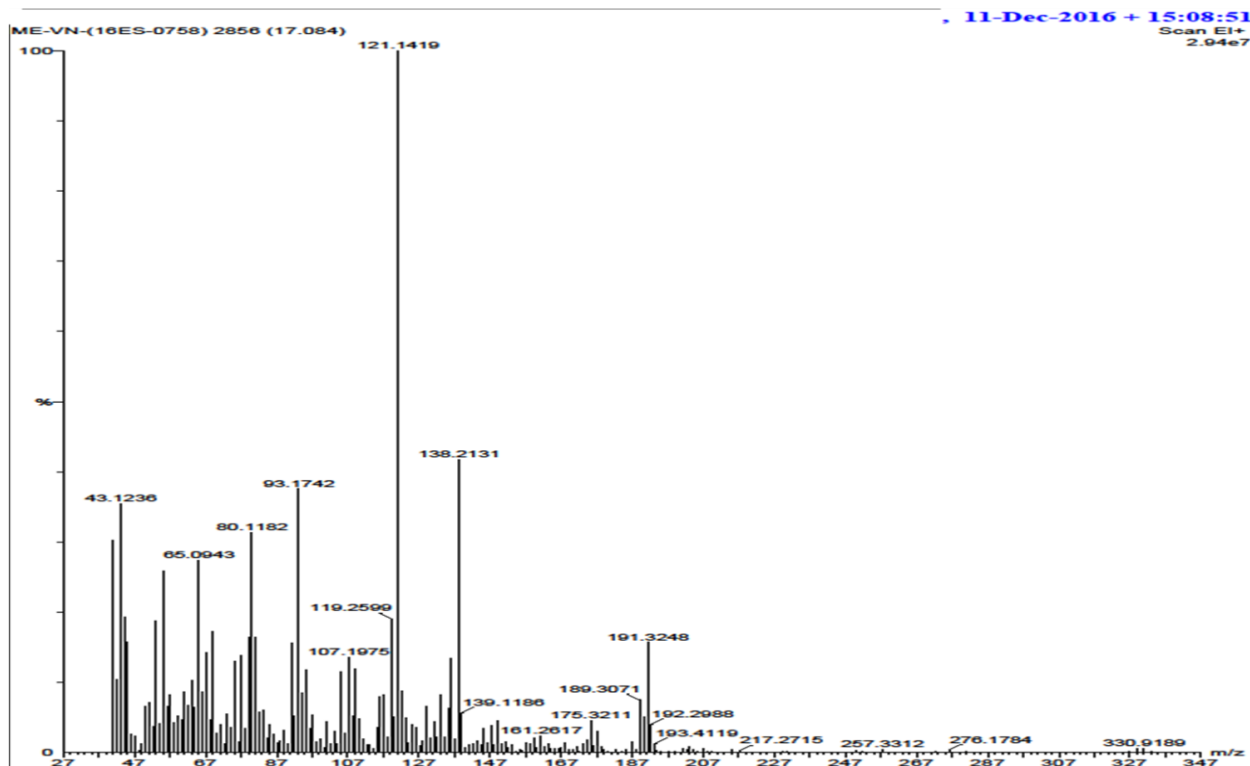
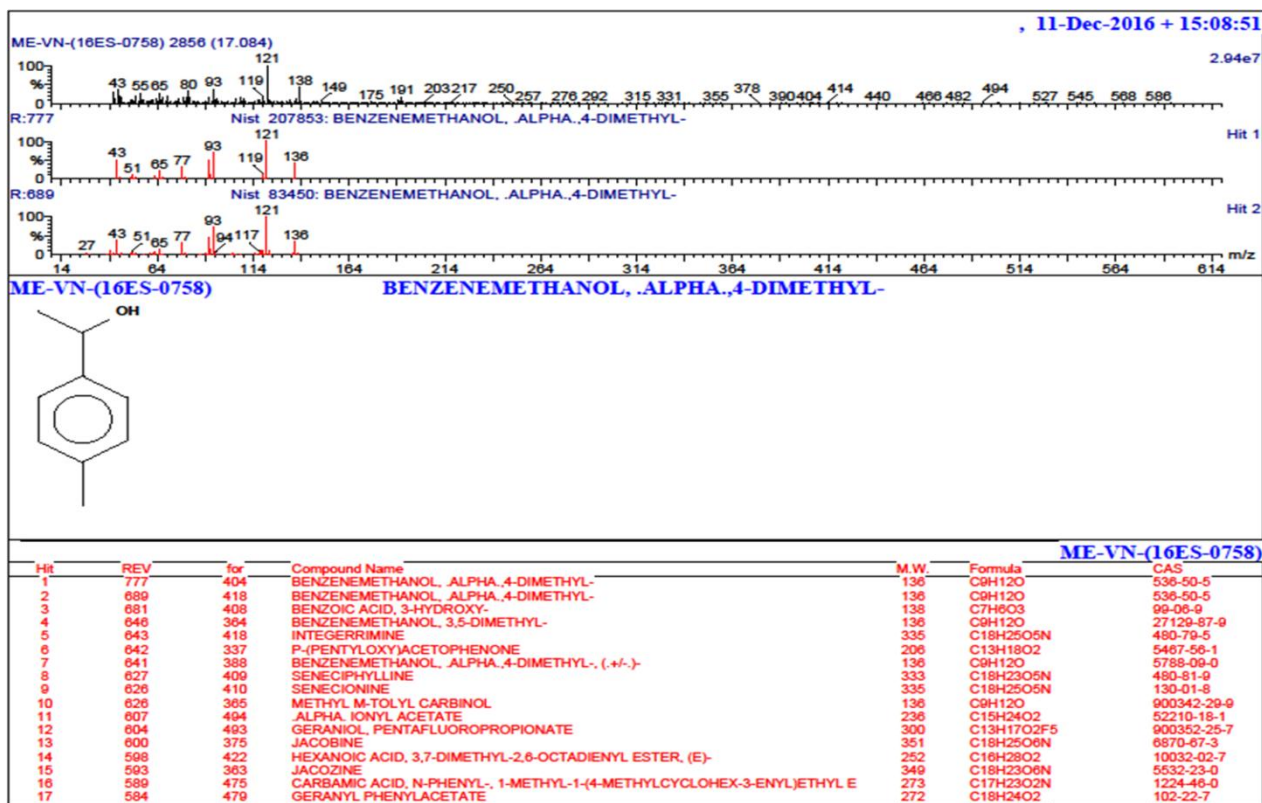


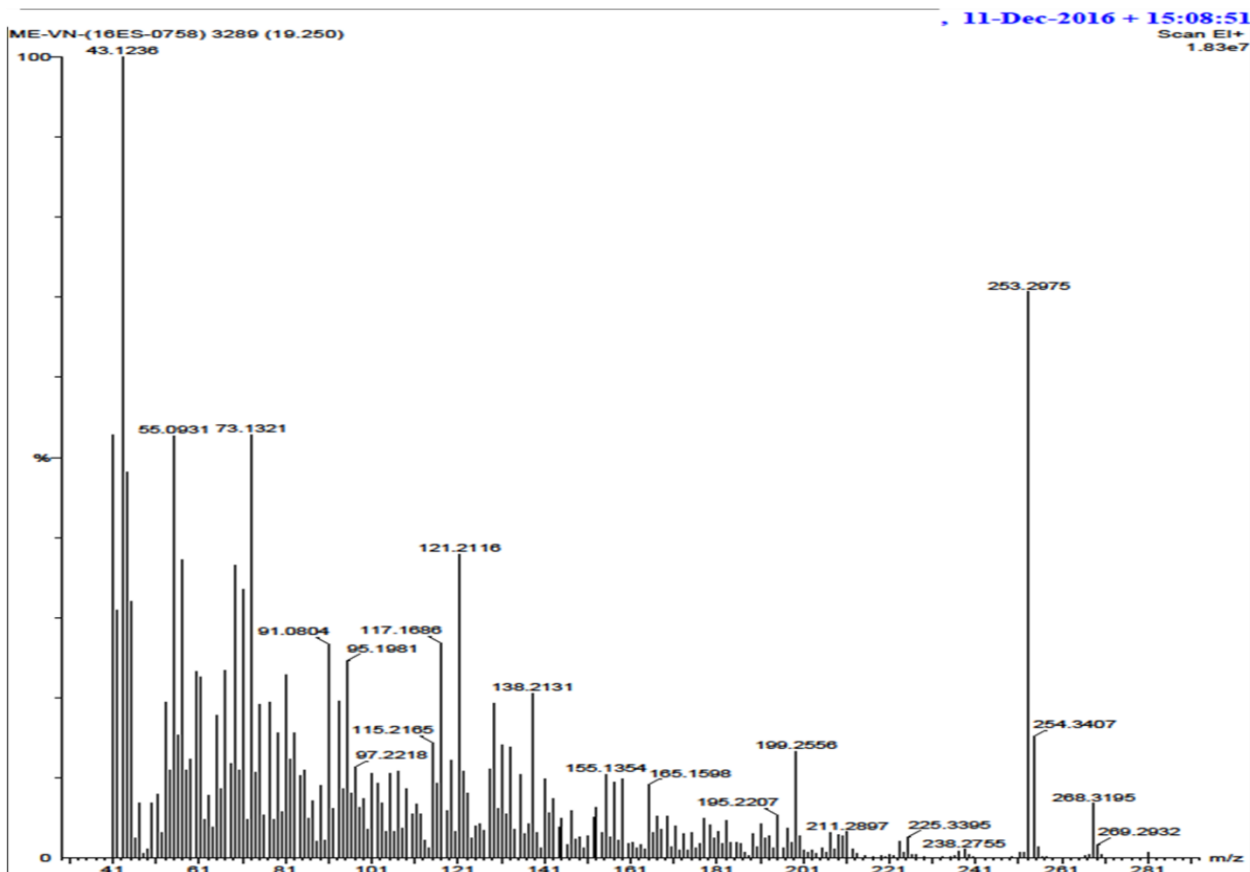
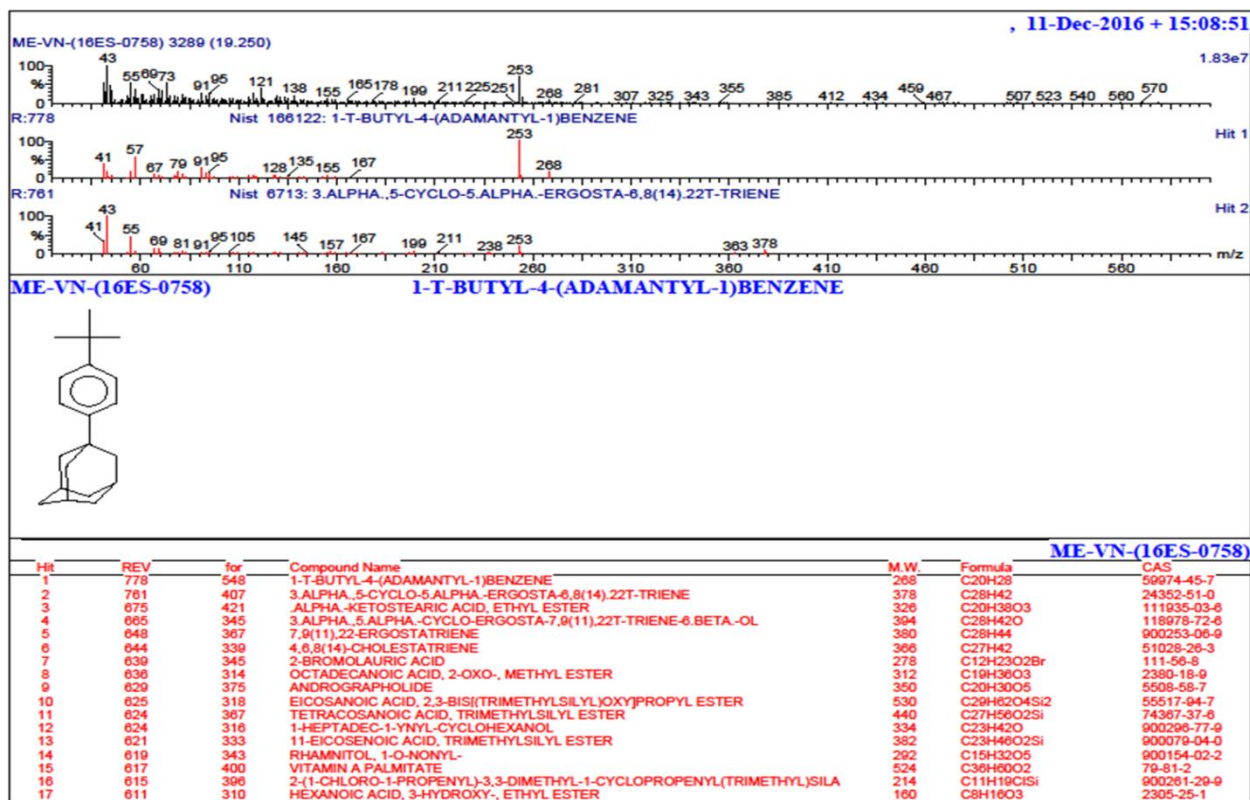
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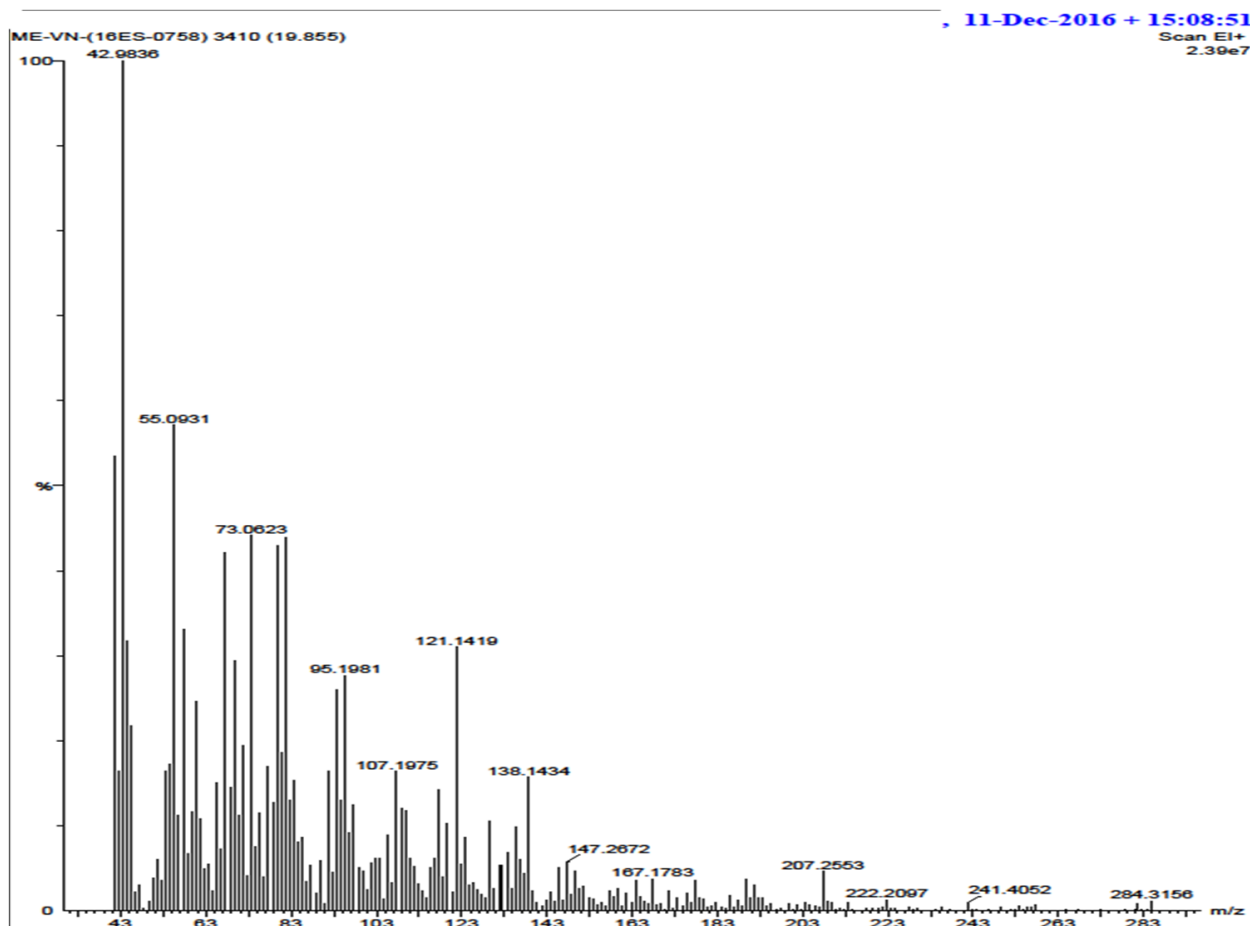
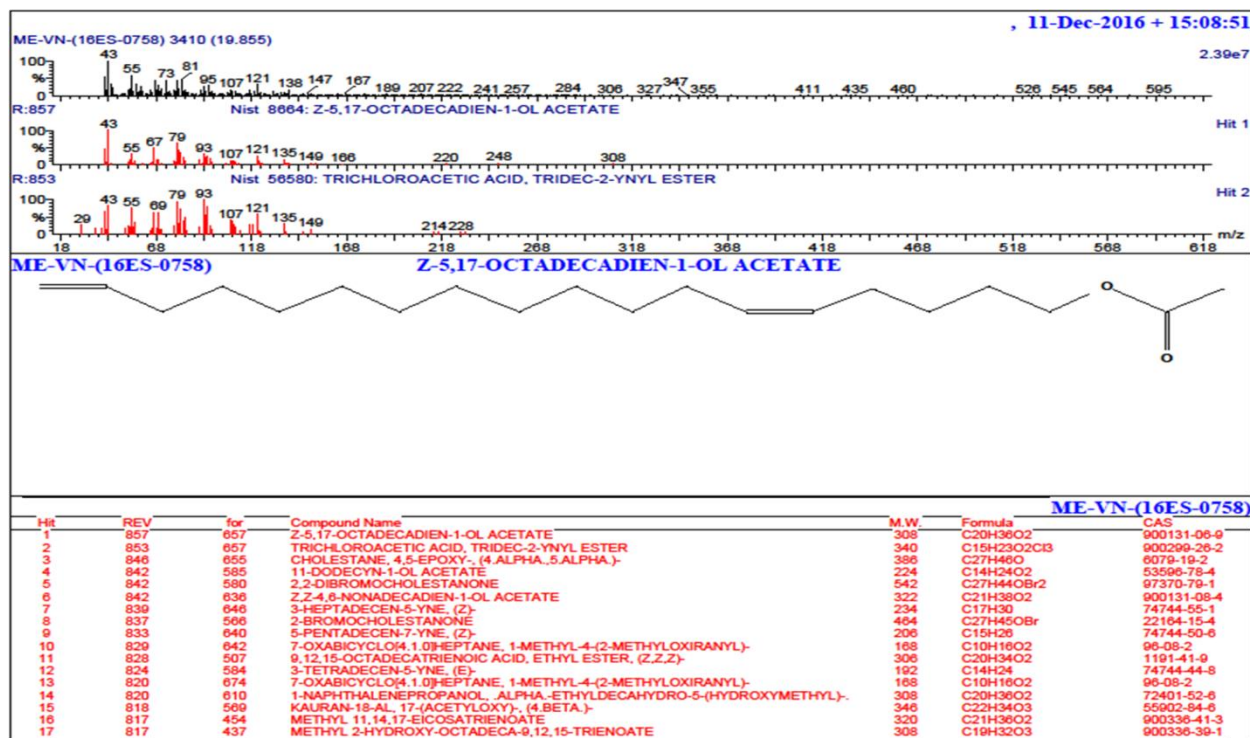
Hit	REV	for	Compound Name	M.W.	Formula	CAS
1	843	402	1,3,3-TRIMETHYL-2-HYDROXYMETHYL-3,3-DIMETHYL-4-(3-METHYLBUT-2-ENYL)-C	222	C15H26O	900144-10-7
2	842	412	2,6,10-DODECATRIEN-1-OL, 3,7,11-TRIMETHYL-9-(PHENYLSULFONYL)-, (E,E)-	362	C21H30O3S	57683-67-7
3	818	641	(7A-ISOPROPENYL-4,5-DIMETHYLOCTAHYDROINDEN-4-YL)METHANOL, (E,E)-	222	C15H26O	900187-02-9
4	790	533	9,19-CYCLOLANOST-24-EN-3-OL, ACETATE, (3.BETA.)-	468	C32H52O2	1259-10-5
5	786	488	2,4,4-TRIMETHYL-3-HYDROXYMETHYL-5A-(3-METHYLBUT-2-ENYL)-CYCLOHEXEN	222	C15H26O	900144-10-5
6	761	639	7A-ISOPROPENYL-4,5-DIMETHYLOCTAHYDROINDENE-4-CARBOXYLIC ACID	236	C15H24O2	900192-14-7
7	754	481	DIHYDROTACHYSTEROL	368	C28H46O	67-96-9
8	754	596	5-(7A-ISOPROPENYL-4,5-DIMETHYL-OCTAHYDROINDEN-4-YL)-3-METHYL-PENT-2-	290	C20H34O	900193-54-0
9	748	334	(E,E,E)-(5-PHENYLSULFONYLGERANYL)GERANIOL	430	C28H38O3S	67428-43-7
10	746	582	5-(1-ISOPROPENYL-4,5-DIMETHYLBICYCLO[4.3.0]NONAN-5-YL)-3-METHYL-2-PENT	332	C22H36O2	900195-69-8
11	741	561	5-(7A-ISOPROPENYL-4,5-DIMETHYL-OCTAHYDROINDEN-4-YL)-3-METHYL-PENT-2-	288	C20H32O	900193-54-1
12	741	655	4-(2,2-DIMETHYL-6-METHYLENECYCLOHEXYLIDENE)-3-METHYLBUTAN-2-ONE	206	C14H22O	93175-74-7
13	741	613	TRICYCLO[3.1.0.0(2,4)]HEXANE, 2,2,5,5,8,8-HEXAMETHYL-, (1ALPHA.,6BETA.,7 AL	206	C15H26	54832-82-5
14	736	553	TRICYCLO[3.1.0.0(2,4)]HEXANE, 3,3,6,6-TETRAETHYL-, TRANS-	192	C14H24	900150-14-4
15	726	420	1,4,5,6-TETRAHYDROCYCLOPENTAPYRAZOLE-3-CARBOXYLIC ACID, (1-ADAMANT	326	C19H26ON4	900265-46-6
16	719	571	2-(4A,8-DIMETHYL-6-OXO-1,2,3,4,4A,5,6,8A-OCTAHYDRO-NAPHTHALEN-2-YL)-PRO	234	C15H22O2	900190-21-8
17	714	438	3-O-ACETYL-6-METHOXY-CYCLOARTENOL	498	C33H54O3	900286-40-9

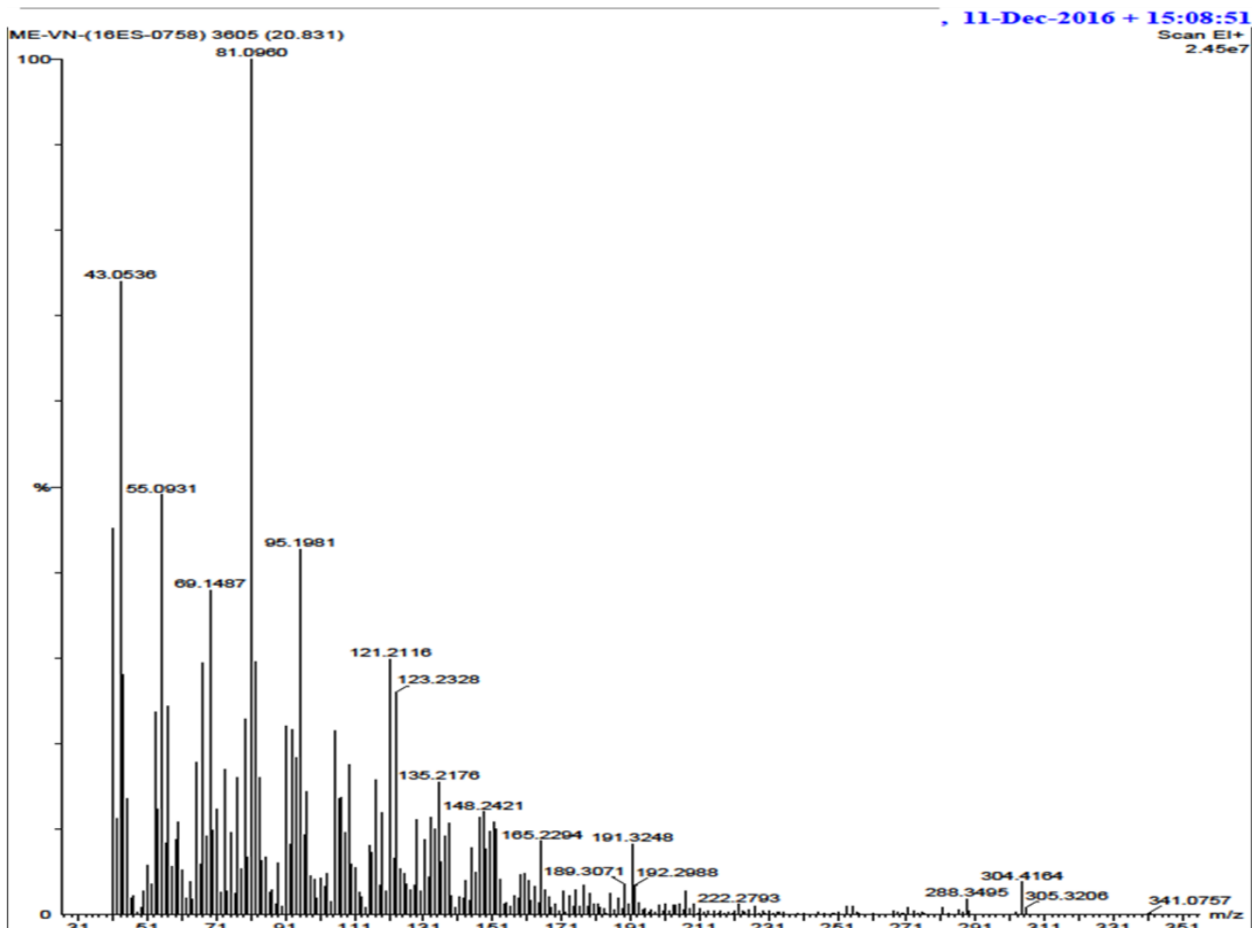
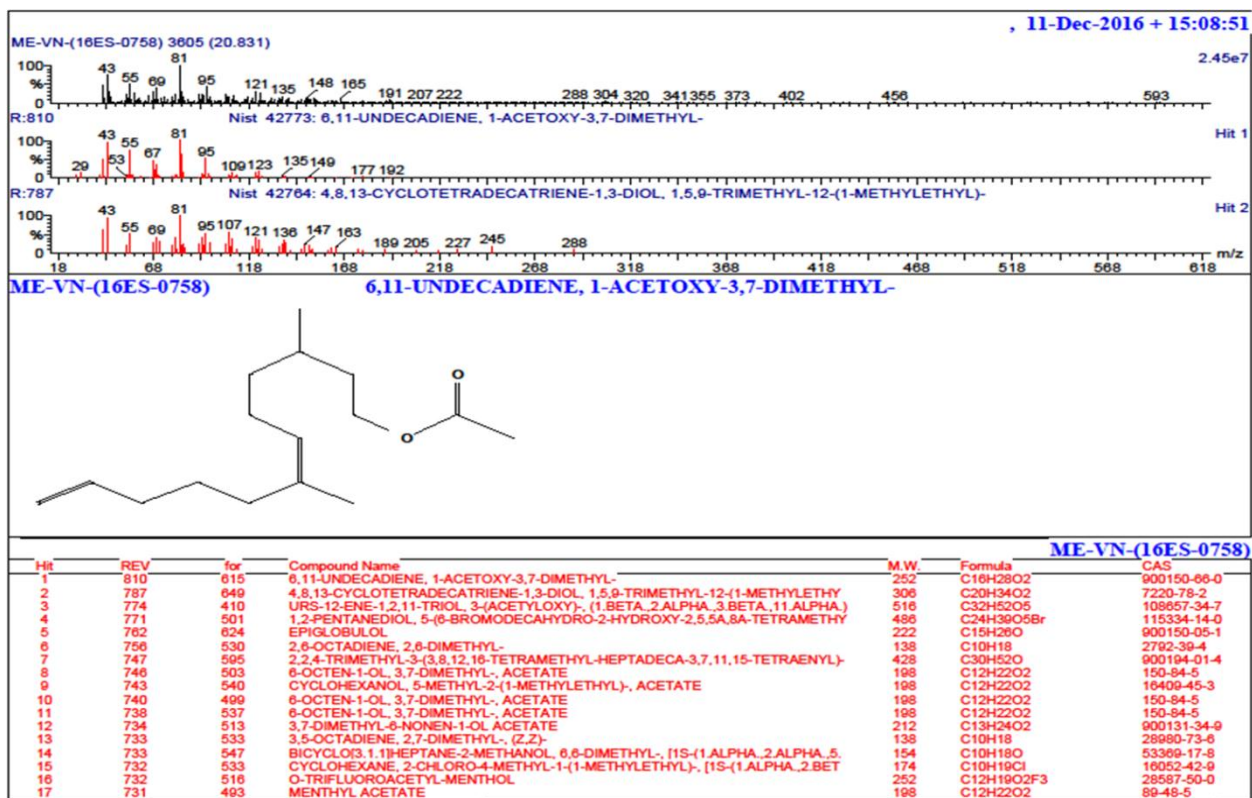


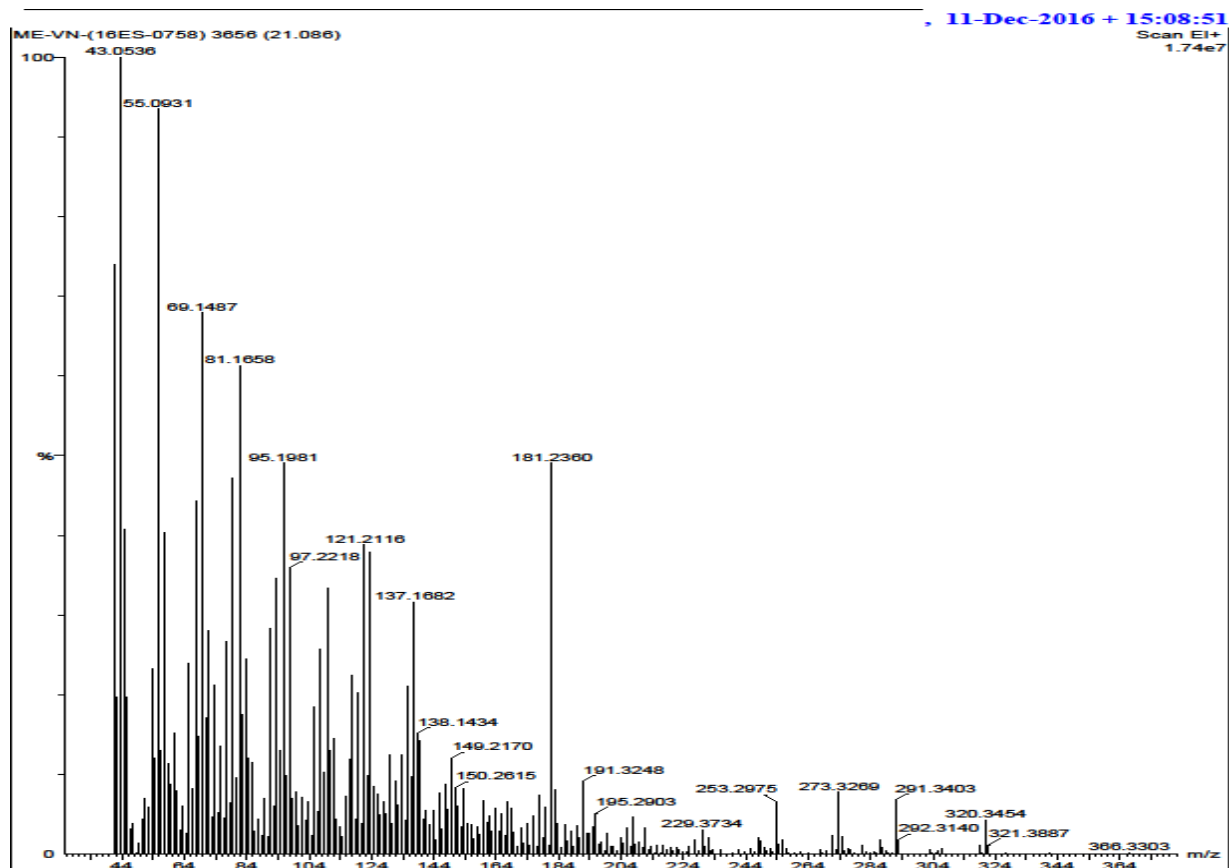
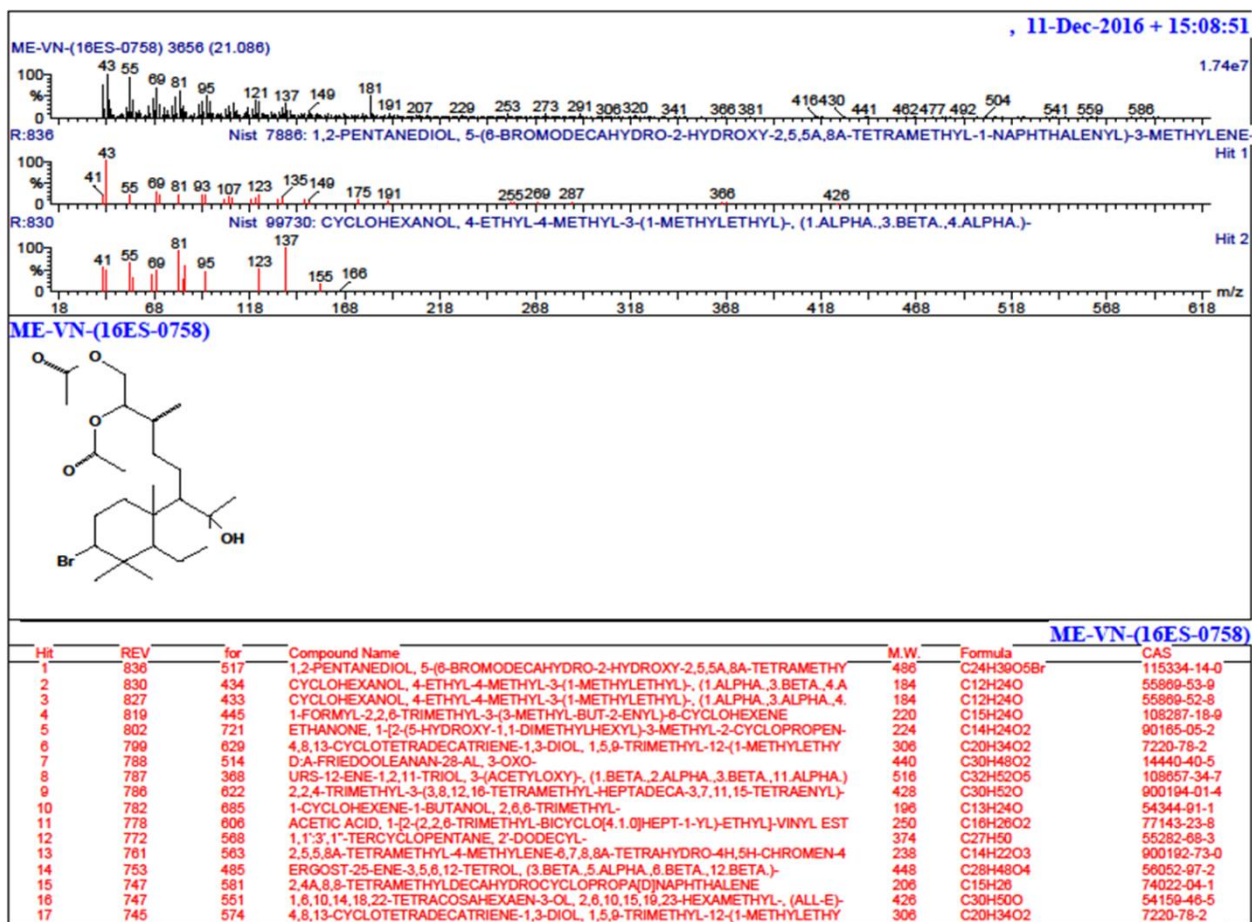


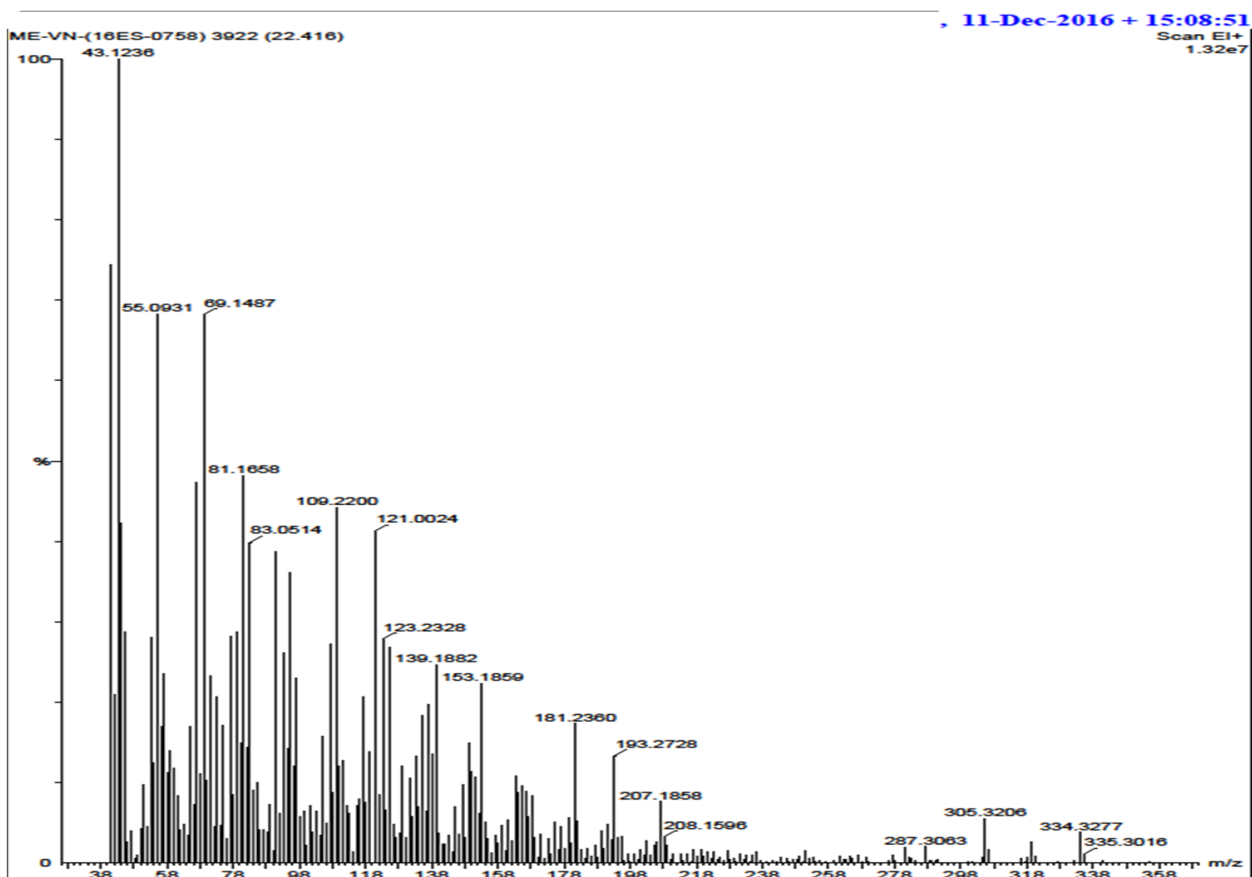
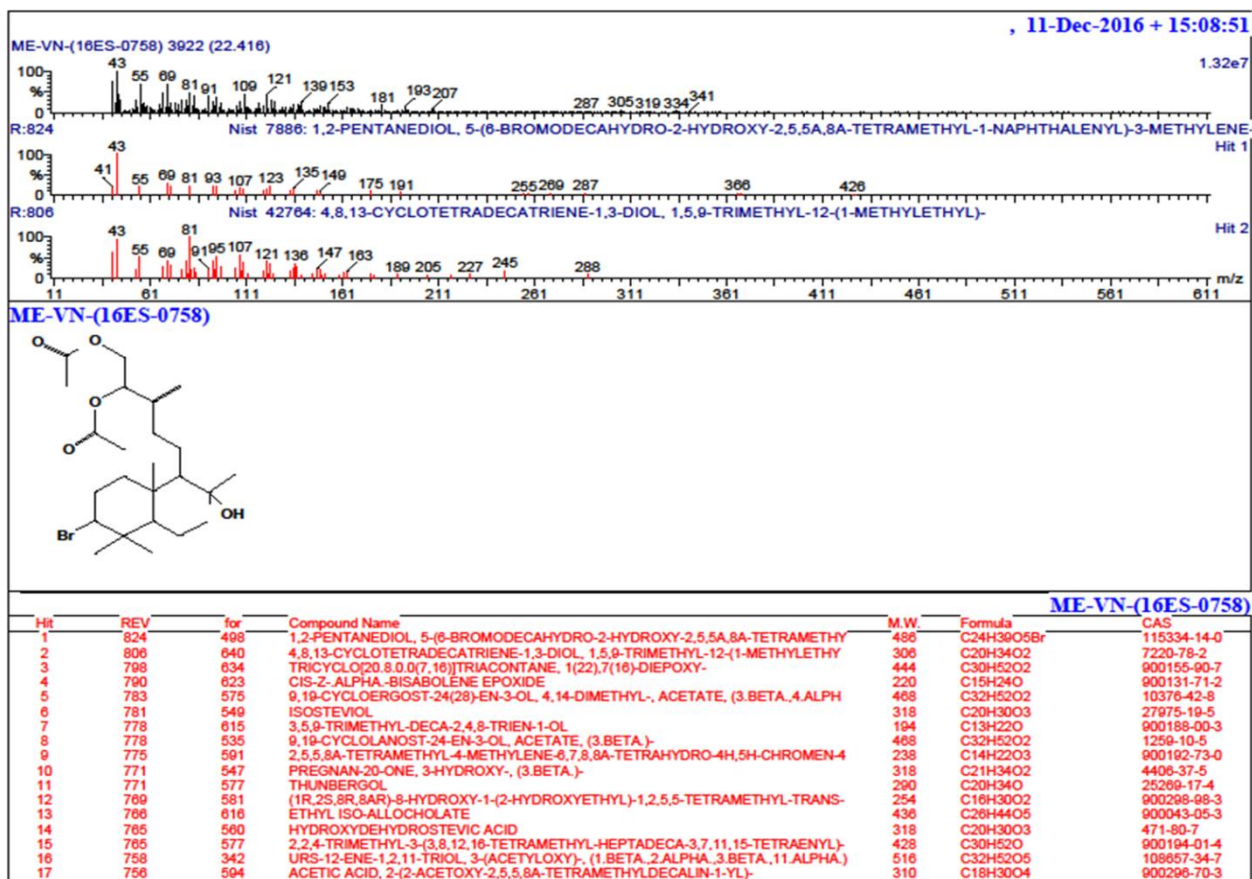


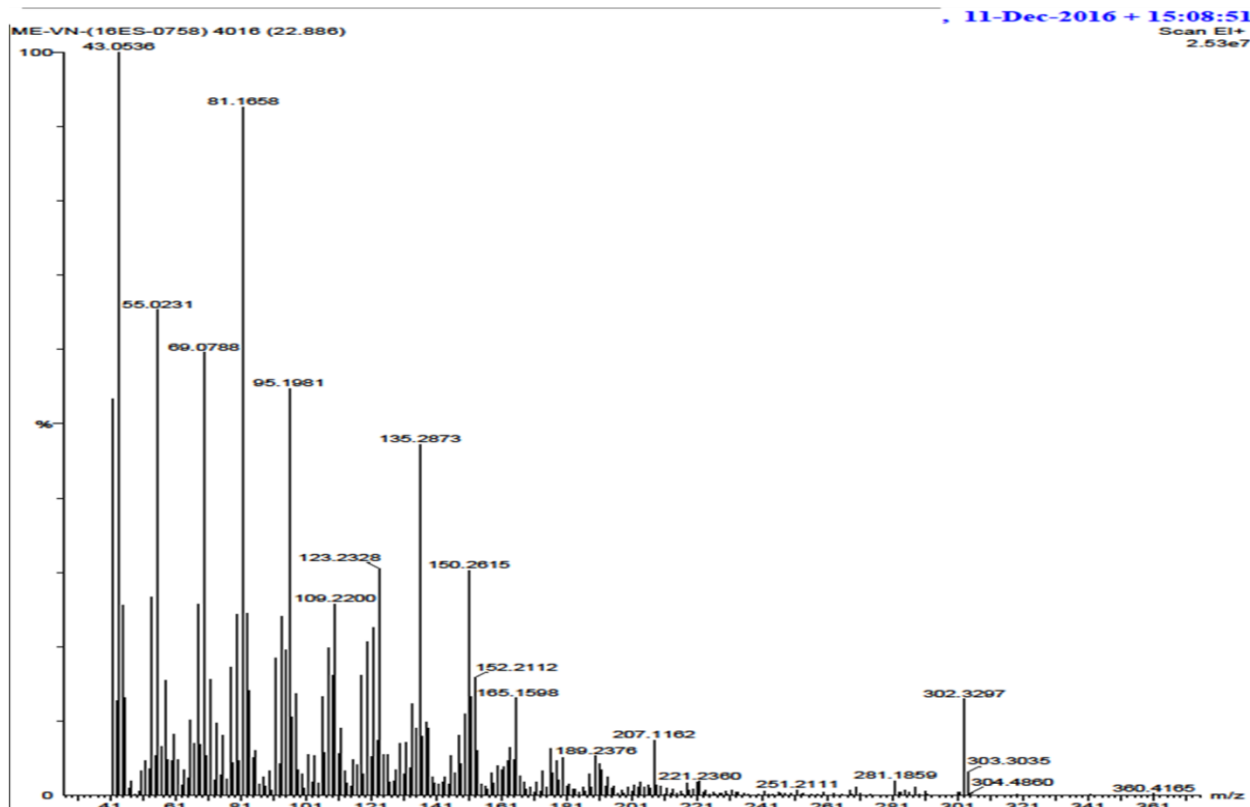
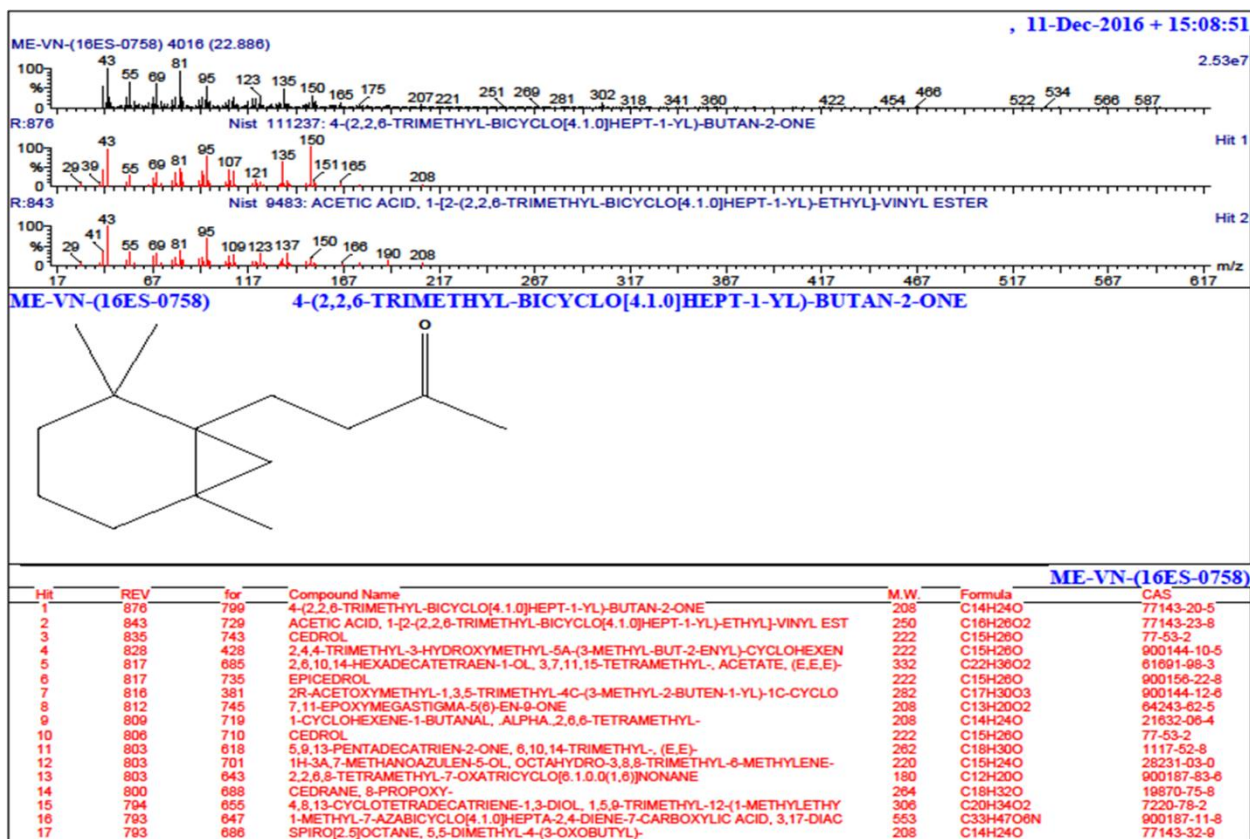


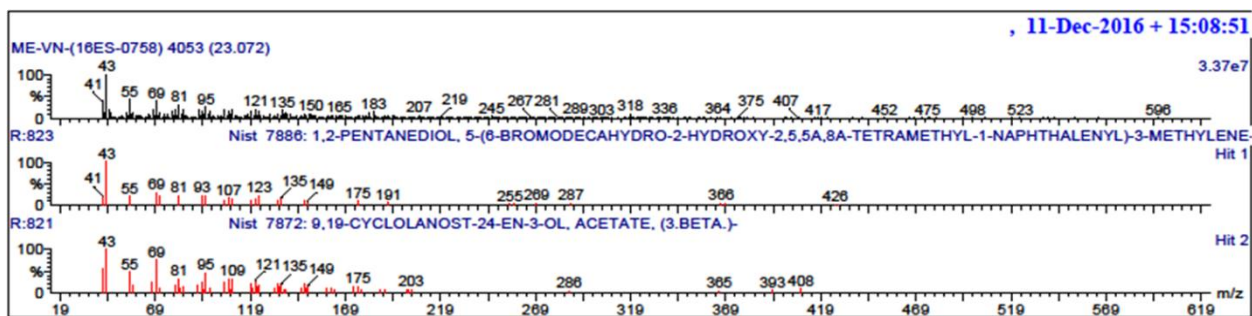




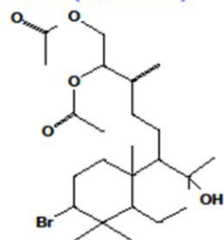








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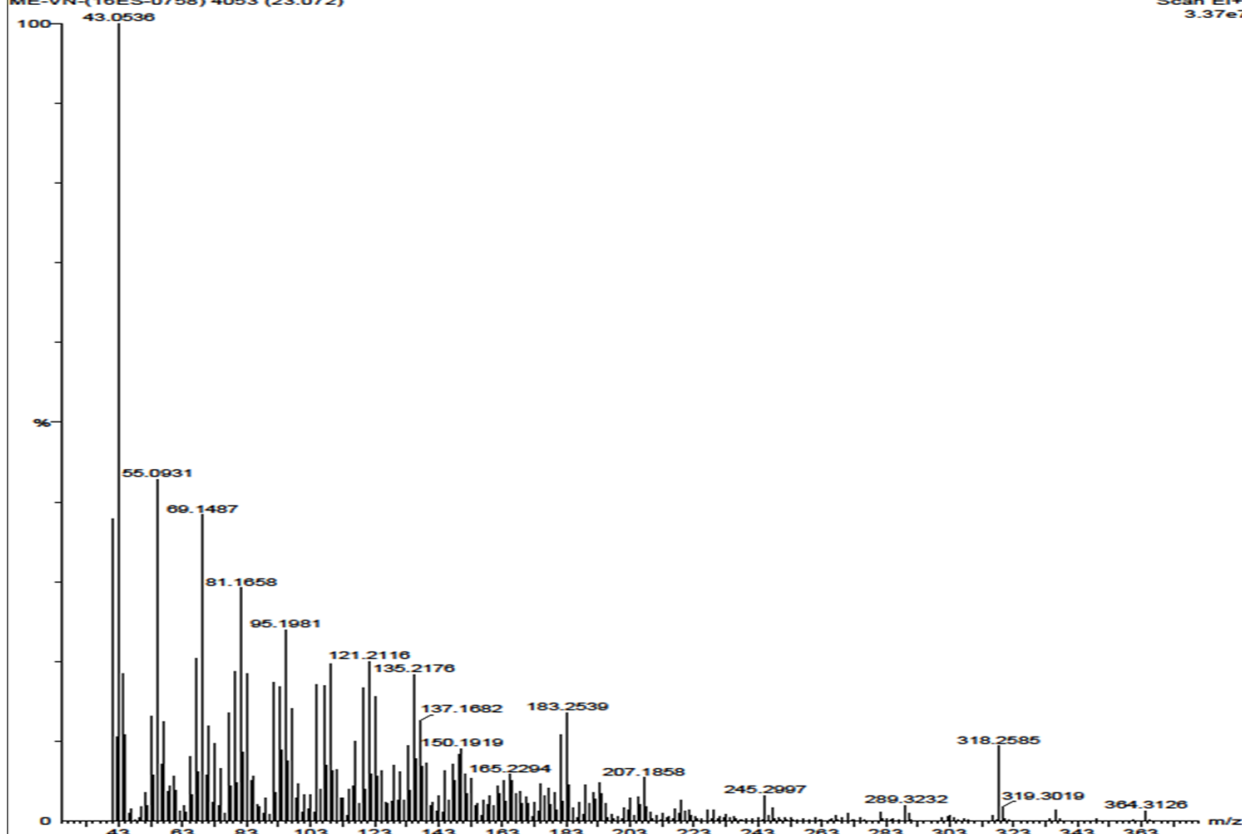


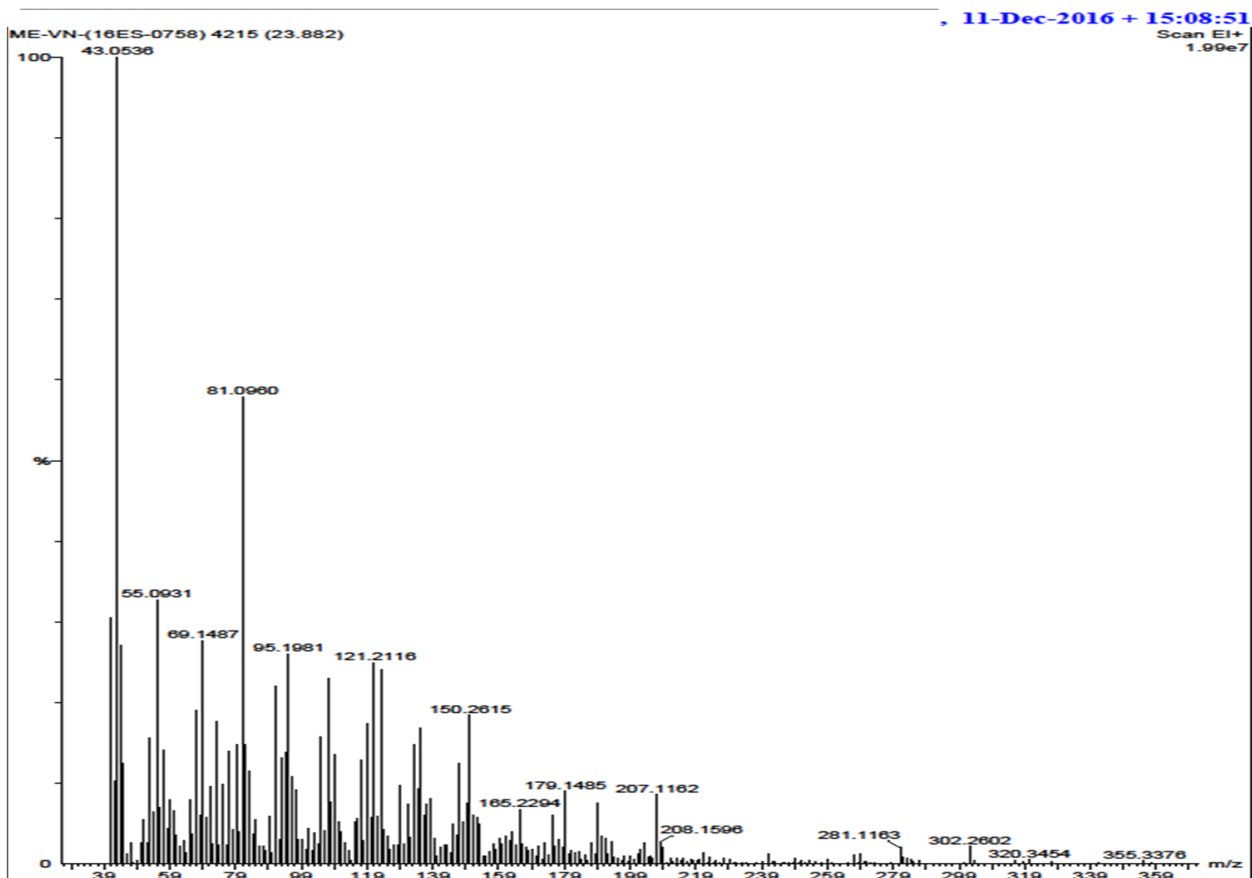
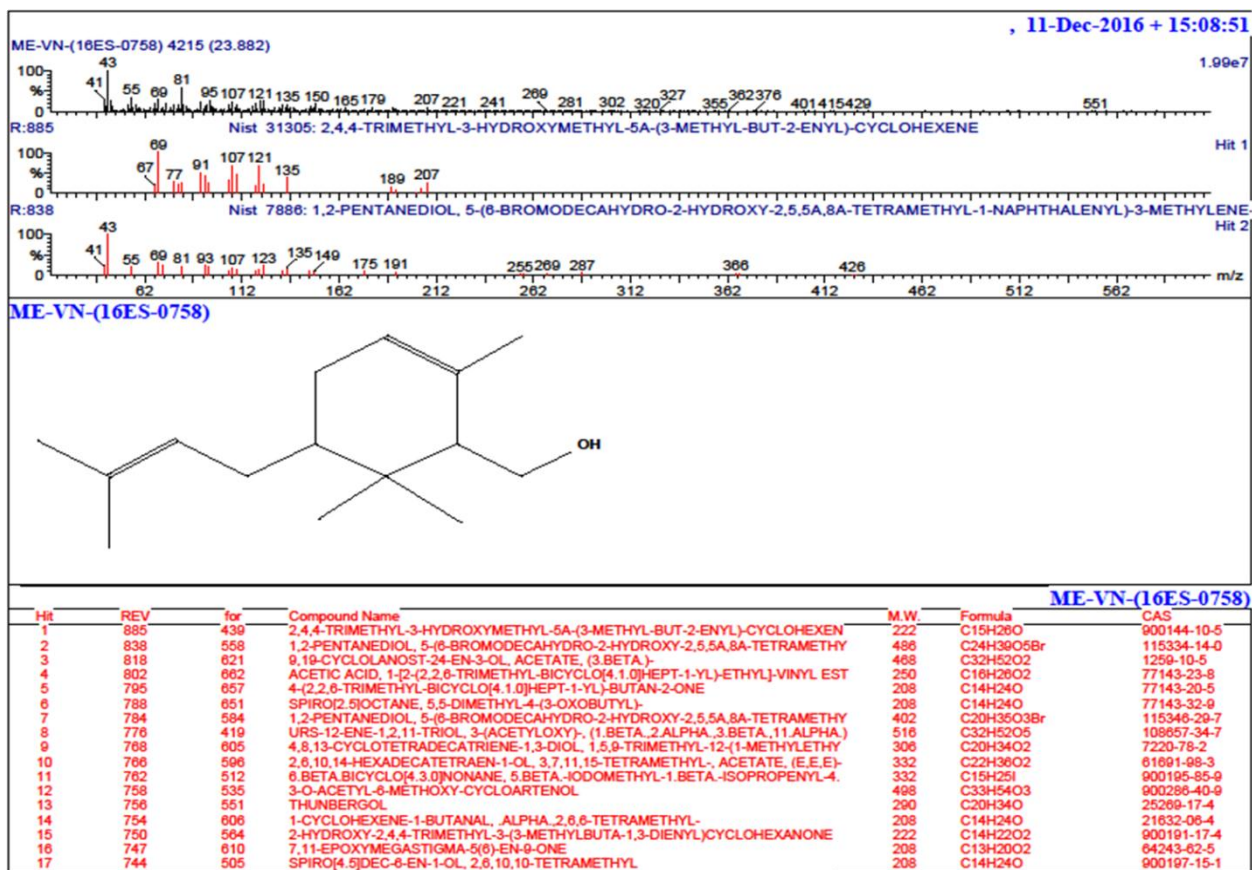
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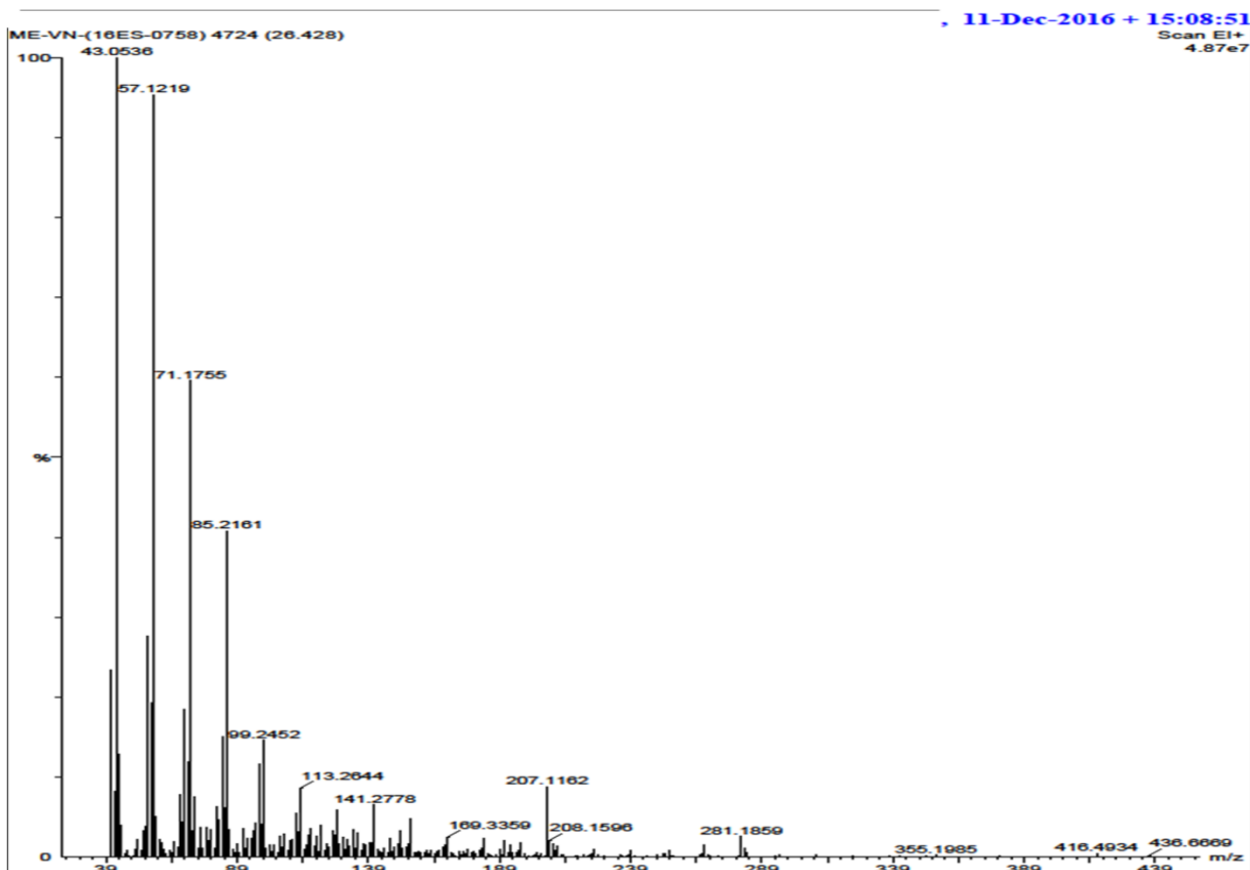
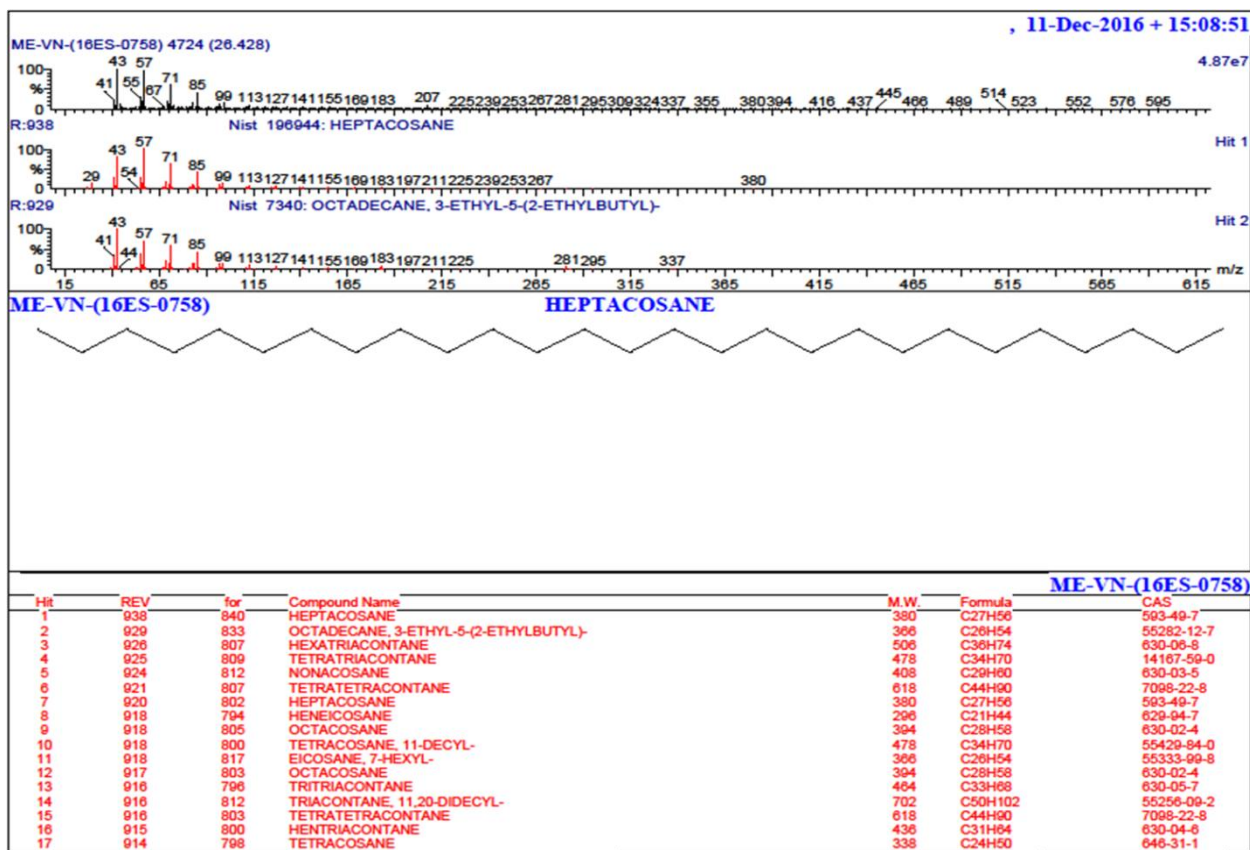
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2	821	637	9,19-CYCLOANOST-24-EN-3-OL, ACETATE, (3.BETA.)-	468	C32H52O2	1259-10-5
3	818	698	ACETIC ACID, 1-[2-(2,6-TRIMETHYL-BICYCLO[4.1.0]HEPT-1-YL)-ETHYL]-VINYL EST	250	C18H26O2	77143-23-8
4	814	371	2-ISOPROPYL-5-METHYLCYCLOHEXYL 3-(1-(4-CHLOROPHENYL)-3-OXOBUTYL)-C	524	C30H33O6Cl	900143-59-5
5	812	654	9,19-CYCLOERGOST-24(28)-EN-3-OL, 4,14-DIMETHYL-, ACETATE, (3.BETA.,4.ALPH	468	C32H52O2	10376-42-8
6	808	685	4,8,13-CYCLOTETRADECATRIENE-1,3-DIOL, 1,5,9-TRIMETHYL-12-(1-METHYLETHY	306	C20H34O2	7220-78-2
7	800	613	METHYL 3-ACETOXY-12-KETO-18-NORCHOLANATE	432	C26H40O5	900251-87-8
8	793	428	URS-12-ENE-1,2,11-TRIOL, 3-(ACETOXY)-, (1.BETA.,2.ALPHA.,3.BETA.,11.ALPHA.)	516	C32H52O5	108657-34-7
9	791	649	THUNBERGOL	290	C20H34O	25269-17-4
10	786	627	2,6,10,14-HEXADECATETRAEN-1-OL, 3,7,11,15-TETRAMETHYL-, ACETATE, (E,E,E)-	332	C22H36O2	61691-98-3
11	785	618	1,2-PENTANEDIOL, 5-(6-BROMODECAHYDRO-2-HYDROXY-2,5,5A,8A-TETRAMETHYL-1-NAPHTHALENYL)-3-METHYLENE-	402	C20H35O3Br	115346-29-7
12	782	587	9,19-CYCLOANOSTAN-3-OL, ACETATE, (3.BETA.)-	470	C32H54O2	4575-74-0
13	781	593	CHOLESTA-22,24-DIEN-5-OL, 4,4-DIMETHYL-	412	C29H48O	900128-66-1
14	781	608	2,2-DIBROMOCHOLESTANONE	542	C27H44OBr2	97370-79-1
15	771	633	ANDROSTAN-3-ONE, 17-HYDROXY-2,4-DIMETHYL-, (2.ALPHA.,4.ALPHA.,5.ALPHA.,1	318	C21H34O2	54550-08-2
16	769	542	LANOSTA-8,24-DIEN-3-OL, ACETATE, (3.BETA.)-	468	C32H52O2	2671-68-3
17	762	581	CYCLOARTANOL	428	C30H52O	4657-58-3

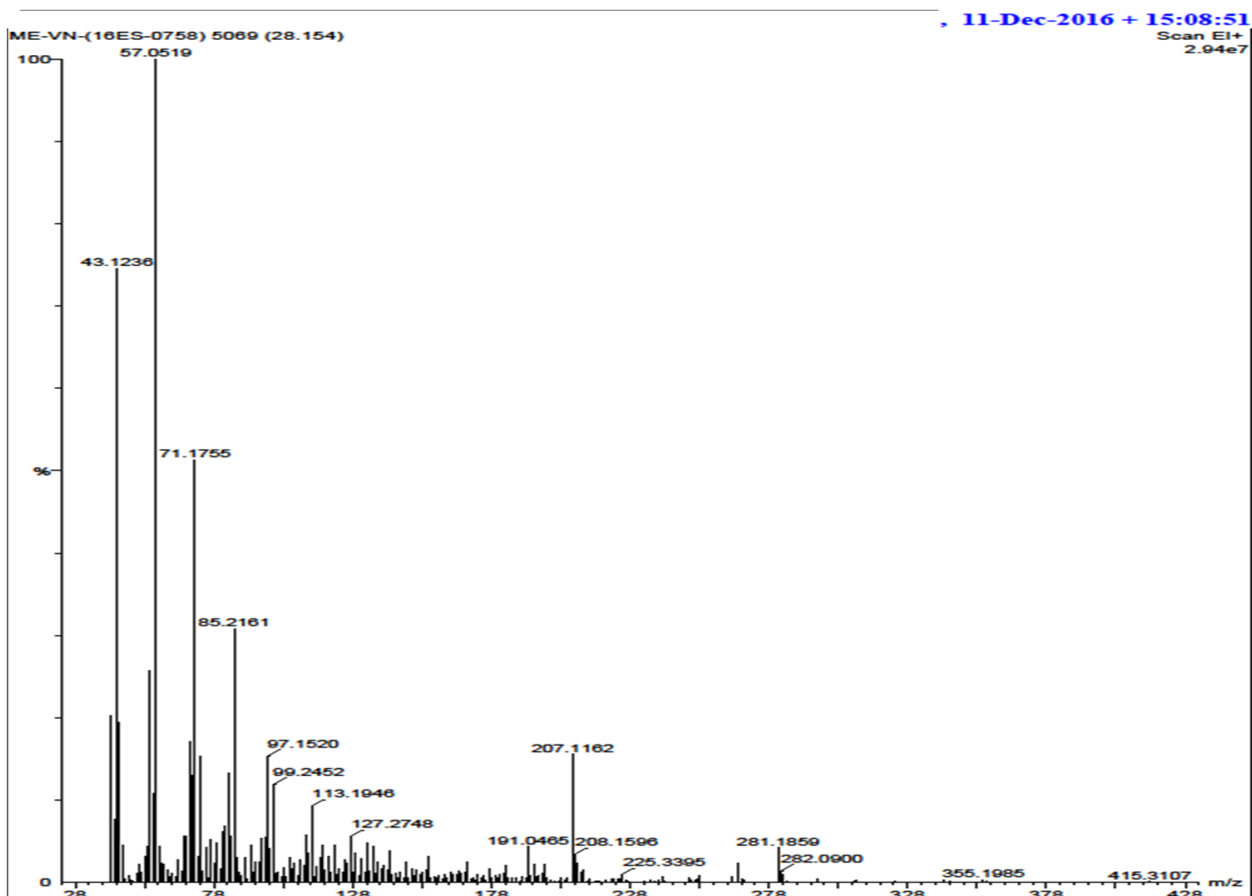
ME-VN-(16ES-0758) 4053 (23.072)

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Scan EI+
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Result

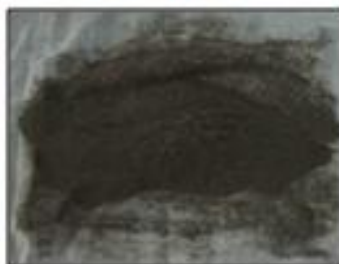
Figure 1: *Vitex negundo*Figure 2: Dried leaf powder of *Vitex negundo*

Figure 3: Soxhlet extraction using methanol



Figure 4: Rotary vacuum evaporator



Figure 5: Methanolic extract of VN (After drying)

Antibacterial activity for the methanolic extract of VN

i. Well diffusion method:

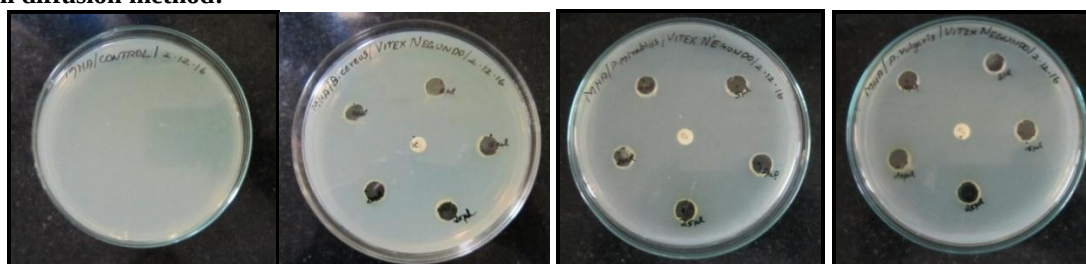
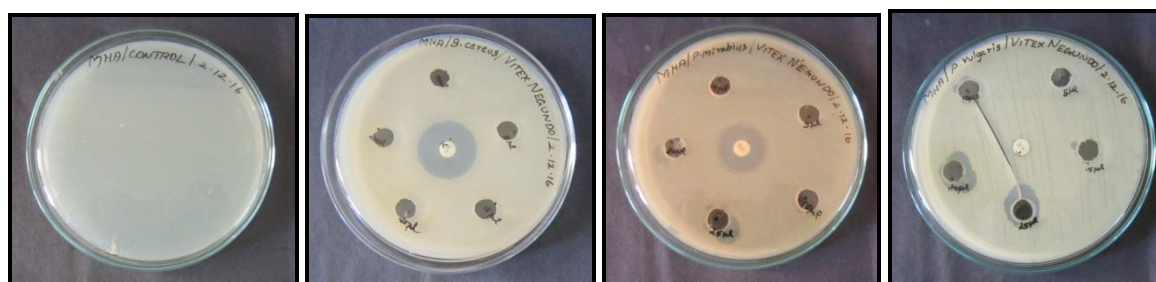
Figure 6: Antibacterial activity for VN (methanolic extract) *B.cereus*, *P.mirabilis*, *P.vulgaris* (before inoculation)Figure 7: Antibacterial activity for VN (methanolic extract) *B.cereus*, *P.mirabilis*, *P.vulgaris* (after inoculation)

Table 1: Extractive yield (%) of VN

Name of the extract	Empty beaker wt (g)	Sample + beaker wt (g)	Sample wt (g)	Extract yield % (25g)	Extract yield % (100g)
Methanol	30.1662	34.0001	25	3.8339	15.34

Table 2: Antibacterial activity – Zone of Inhibition (mm)

INHIBITION ZONE(mm)							
Bacteria	Methanolic extract (VN)					R	MIC (µg/ml)
	5 (µl)	10 (µl)	15 (µl)	20 (µl)	25 (µl)		
<i>B.cereus</i>	0	0	0	0	0	24	0
<i>P.mirabilis</i>	0	0	0	0	0	18	0
<i>P.vulgaris</i>	0	0	0	12	13	7	55.03

DISCUSSION

In recent years, the interest for the study of the organic compounds found in plants and their activity has increased. The GC–MS is an ideal technique for qualitative and quantitative analysis of active components in plant. The aim of the present study was to confirm the phytochemicals present in the plant extracts and to evaluate their antibacterial activity. Diversity of medicinal plants and herbs containing various phytochemicals with biological activity can be of valuable therapeutic key. Different phytochemicals have been found to have a broad range of activities, which may help in protection against chronic diseases.(Liu RH., 2003)

The discovery of novel antimicrobial metabolites from medicinal plants is an important alternative to overcome the increasing levels of drug resistance by human pathogens. Due to the world's urgent need for new antibiotics and chemotherapeutic agents, growing interest is taken into the research on the chemistry of medicinal plants. Antimicrobial activity of medicinal plants and their phytoconstituents have been deliberated in the late 19th century [L.L. Zaika., 1998]. In the present study, leaf extract of *Vitex nigundo linn* was evaluated for antibacterial activity against opportunistic bacterial pathogens *Bacillus cereus*, *Proteus vulgaris* and *Proteus mirabilis*. The results revealed that the methanolic leaf extract of *Vitex nigundo linn* showed considerable antibacterial activity against *P.vulgaris*.

GC–MS analysis plays a key role in the analysis of components of plant origin. Generally, the plant materials are highly complexes, which make GC–MS well suited for their analysis because of its high sensitivity and selectivity. It is considered to be the gold standard in scientific analysis (M. Sajewicz, ETAL., 2009, K. Balamurugan ETAL., 2012). Several phytochemical screening studies have been carried out in different parts of the world by using GC–MS [D. Dubey ETAL., 2014, G.M. Doshi ETAL., 2015). The methanolic leaf extract of *Vitex nigundo linn* used for bioactive compounds investigation using the GC–MS analysis.

In this study, we reported for the first time a high resolution GC–MS method for the evaluation of the chemical constituents of methanolic leaf extract of *Vitex nigundo linn*. This accurate and sensitive analysis of methanolic leaf extract of *Vitex nigundo linn* revealed the various phytoconstituents present in the extracts. The major bioactive compounds in methanolic leaf extract of

Vitex nigundo linn is **phosphonic acid**. It is the major compound present in this plant extract. Ledol, benzene methanol 1 – t – butyl – 4 (adamantly-1) benzene, Z-5,17 – octadecadiene – 1-ol acetate, 6, 11 – Undecadiene, 1-acetoxy – 3,7-dimethyl, 4,(2,2,6-trimethyl – bicycle(4.1.0) hept – 1-yl)-butane – 2-one, heptacosone, dotriacontane, hydroxyl dehydro steric acid another major compounds in the studied plant extracts.

The results of the present investigation also complement the ethnobotanical usage of the studied plants which possess several phytoconstituents with biological activity. Based on the present investigation, it is concluded that methanolic leaf extract of *Vitex nigundo linn* have potential source of bioactive compounds with great pharmaceutical value. The study can be extended for separation and *in vitro*, *in vivo* evaluation of bioactive compounds in novel drug discovery.

CONCLUSION

In the present study, more than sixteen chemical constituents have been confirmed from the methanolic leaf extract of *Vitex nigundo linn* using GC-MS analysis. The antibacterial study result indicated that the lowest MIC against *B.cereus* and *P.mirabilis* was observed in methanol extract, against kanamycin. The results also revealed that the methanolic leaf extract of *Vitex nigundo linn* showed considerable antibacterial activity against *P.vulgaris*. The presence of various phytochemical constituents justifies the use of plant for various ailments by traditional practitioners. Further studies are needed on these extracts in order to isolate, identify, characterize and elucidate the structure of these compounds.

REFERENCES

1. Abirami P, Rajendran A. GC-MS analysis of methanol extracts of Vernonia cinerea. Euro. J. Exp. Bio. 2012; 2(1): 9-12.
2. Ahmad I. and Beg A.Z. Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens. J. of Ethnopharmacol. 2001; 74: 113-123.
3. Bauer AW, Kirby WMM, Sherris JC & Turck M, Antibiotic susceptibility testing by a standardized single disk method, Am J Clin Pathol 1966; 43: 493.
4. Chaturvedi GN, Singh RH. Indian Journal of Medical Research. 1965; 53: 71.
5. Chopra R N, Nayar SL, & Chopra C. Glossary of Indian medicinal plants. New Delhi: CISR; 1956; 256.

6. D. Dubey, R. Patnaik, G. Ghosh, R.N. Padhy *In vitro* antibacterial activity, gas chromatography-mass spectrometry analysis of *Woodfordia fruticosa* Kurz. leaf extract and host toxicity testing with *in vitro* cultured lymphocytes from human umbilical cord blood *Osong Public Health Res Perspect*, 2014; 5(5): 298-312.
7. G.M. Doshi, V.V. Nalawade, A.S. Mukadam, P.K. Chaskar, S.P. Zine, R.R. Somani, *et al.* Structural elucidation of chemical constituents from *Benincasa hispida* seeds and *Carissa congesta* roots by gas chromatography: mass spectroscopy *Pharmacogn Res*, 2015; 7(3): 282-293.
8. Jeyachandran R. and Mahesh A. Antimicrobial evaluation of *Kigelia africana* (Lam). *Res. J. of Microbiol.* 2007; 2(8): 645-649.
9. Kiruthika KA, Jaisheeba A, Sornaraj R. Evaluation of antibacterial activity of some selected Angiosperm flower extract. *International Journal of Chem. Tech. Research.* 2011; 3(4): 1945-51.
10. K. Balamurugan, A. Nishanthini, V.R. Mohan GC-MS analysis of *Polycarpaea corymbosa* (L.) Lam whole plant. *Asian Pac J Trop Biomed*, 2012; 2(Suppl 3): S1289-S1292.
11. Liu RH. Health benefits of fruits and vegetables are from additive and synergic combinations of phytochemicals. *Am. J. Clin. Nutr.* 2003; 78(3): 517-20.
12. L.L. Zaika Spices and herbs: their antimicrobial activity and its determination *J Food Saf*, 1988; 9: 97-118.
13. M. Sajewicz, J. Rzepa, M. Hajnos, Ł. Wojtal, D. Staszek, T. Kowalska, *et al.* GC-MS study of the performance of different techniques for isolating the volatile fraction from sage (*Salvia* L.) species, and comparison of seasonal difference in the composition of this fraction. *Acta Chromatogr*, 2009; 21(3): 453-471.
14. Nair R. and Chanda S. Activity of some medicinal plants against certain pathogenic bacterial strains. *Ind. J. Pharmacol.* 2006; 38: 142- 144.
15. NCCLS (2000) Methods for dilution Antimicrobial susceptibility test for bacteria that grow aerobically: Approved standard. Fifth edition, NCCLS M7-A5, NCCLS: Wayne, PA. USA.
16. Prajapati R, Roy S, Mishra S, Raza SK, Thakur LK. Formulation development, standardization and antimicrobial activity of *ageratum conyzoides* extracts and their formulation *Int J Pharm Pharm Sci.* 2014; 6(2): 369-74.
17. Raman BV, Radhakrishnan TM & Rajagopal SV, Antibacterial and immunomodulatory studies on selected brown algal species of Visakhapatnam seacoast, *Indian J Microbiol* 2005; 45: 245.
18. Raman BV, Sai Ramkishore A, Uma Maheswari M & Radhakrishnan TM, Antibacterial activities of some folk medicinal plants of Eastern Ghats, *J Pure & Appl Microbiol* 2009; 3: 187.
19. Shah P.M. The need for new therapeutic agents: what is in pipeline? *Clin. Microbiol. Infect.* 2005; 11: 36-42.