

ASSAY METHOD DEVELOPMENT AND VALIDATION FOR SIMULTANEOUS ESTIMATION OF PAROXETINE AND CLONAZEPAM BY RP- HPLC

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Article Received on 13/05/2017

Article Revised on 02/06/2017

Article Accepted on 22/06/2017

ABSTRACT

A simple, precise, rapid, specific and accurate reverse phase high performance liquid chromatography method was developed for simultaneous estimation of Paroxetine and Clonazepam in pharmaceutical dosage form. Chromatographic separation was performed on INERTSIL ODS 3V column, C18(250x4.6 ID) column, with mobile phase comprising of mixture of buffer (pH7, adjusted with ammonium acetate), methanol, acetonitrile in the ratio of 3:2:5, at the flow rate 1.0 ml/min. The detection was carried out at 224 nm. The retention times of paroxetine and clonazepam were found to be 2.367 and 4.867 mins respectively with a run time of 6 mins, theoretical levels for paroxetine and clonazepam were 2320 and 3211 respectively, with a resolution of 8.249. As per ICH guidelines the method was validated for linearity, accuracy, precision, limit of detection and limit of quantitation, robustness and ruggedness. Linearity of paroxetine was found in the range of 60-140µg/mL and that for clonazepam was found to be 2.4-5.6µg/mL.

KEYWORDS: Paroxetine, Clonazepam, RP-HPLC, Method development, Validation.

INTRODUCTION

Paroxetine is chemically a (3S,4R)-3-[(2H-1,3-benzodioxol-5-yl)oxy]methyl-4-(4-fluorophenyl)piperidine. It is an antidepressant in a group of drugs called selective serotonin reuptake inhibitors (SSRIs).^[1-5] SSRIs are primarily classified as antidepressants. The mechanism of action of Paroxetine affects neurotransmitters, the chemicals that nerves within the brain use to communicate with each other. Neurotransmitters are manufactured and released by nerves and then travel and attach to nearby nerves.^[6-10] Paroxetine works by preventing the reuptake of one neurotransmitter, serotonin, by nerve cells after it has been released. Since reuptake is an important mechanism for removing released neurotransmitters and terminating their actions on adjacent nerves, the reduced uptake caused by paroxetine increases free serotonin that stimulates nerve cells in the brain.^[11-16]

Clonazepam is a benzodiazepine drug having anxiolytic, anticonvulsant. Clonazepam is chemically 5-(2-chlorophenyl)-1,3-dihydro-7-nitro-2H-1,4-benzodiazepin-2-one. Clonazepam is approved by the Food and Drug Administration for treatment of epilepsy and Panic Disorder. Clonazepam is a chlorinated derivative of nitrazepam and therefore a chloro-nitrobenzodiazepine. Clonazepam's primary mechanism of action is the modulation of GABA function in the brain, by the

benzodiazepine receptor, located on GABA receptors, which, in turn, leads to enhanced GABAergic inhibition of neuronal firing. Clonazepam exerts its action by binding to the benzodiazepine site of the GABA receptors, which causes an enhancement of the electric effect of GABA binding on neurons, resulting in an increased influx of chloride ions into the neurons. This results in an inhibition of synaptic transmission across the central nervous system.^[16]

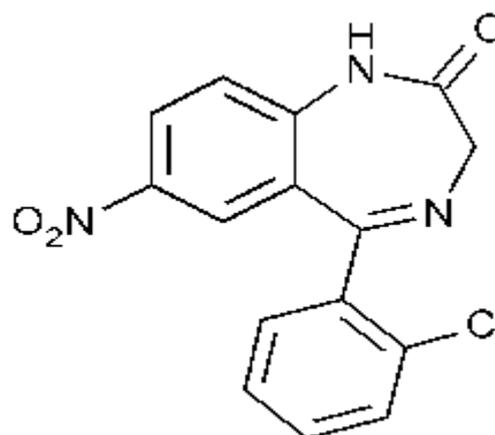


Figure 1: Molecular Structure of paroxetine.

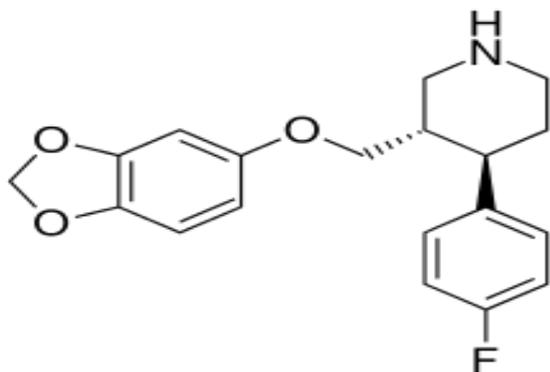


Figure 2: Molecular structure of clonazepam.

Literature survey revealed that few analytical techniques are available for estimation of paroxetine alone as well as in combine dosage form such as UV, HPLC.^[4-8] Similarly few analytical methods are available for estimation of clonazepam alone and its combination with drugs such as UV and HPLC, colorimetry^[1-8] but there is few methods for their simultaneous estimation using RP-HPLC. The present study was designed to develop a simple, precise, and rapid analytical RP-HPLC procedure, which can be used for the analysis of assay method for simultaneous estimation of paroxetine and clonazepam as there was only individual methods reported for both drugs. The combination of these two drugs is not official in any pharmacopoeia; hence no official method is available for the simultaneous estimation of these two drugs in their combined dosage forms. Various validation aspects of the analysis accuracy, precision, recovery, the limits of detection and quantification etc have been measured as per ICH guidelines.^[15]

MATERIALS AND METHODS

Equipment

Chromatographic separation was performed on HPLC system-Water's, PDA Detector 2998 module, equipped with a solvent delivery pump, sample injector and column thermostats. CLASSVP Chromatographic system software was applied for data collecting and processing.

Chemicals and reagents

Methanol, Acetonitrile (HPLC grade) was used. Buffer used was pH-7 (pH adjusted with ammonium acetate). (Reference standards Paroxetine and Clonazepam were obtained from chandra Lab. paroxetine (100mg) and Clonazepam (4mg) manufactured by Johnson and Johnson Ltd, India. were procured from local market.

Preparation of Standard Solution

Standard stock solutions of Paroxetine and Clonazepam ($\mu\text{g/ml}$) were prepared by dissolving 100 mg of Paroxetine and 4 mg of Clonazepam dissolved in sufficient mobile phase. After that filtered the solution using 0.45-micron syringe filter and Sonicated for 5 min and dilute to 100 ml with mobile phase. Further dilutions are prepared in 5 replicates of 100 $\mu\text{g/ml}$ of Paroxetine

and 4 $\mu\text{g/ml}$ of Clonazepam was made by adding 1 ml of stock solution to 10 ml of mobile phase.

Preparation of Sample Solution

20 tablets (each tablet contains 100 mg of Paroxetine and 4 mg of Clonazepam) were weighed and taken into a mortar and crushed to fine powder and uniformly mixed. Tablet stock solutions of Paroxetine and Clonazepam ($\mu\text{g/ml}$) were prepared by dissolving weight equivalent to 100 mg of Paroxetine and 4 mg of Clonazepam and dissolved in sufficient mobile phase. After that filtered the solution using 0.45-micron syringe filter and sonicated for 5 min and dilute to 100 ml with mobile phase. Further dilutions are prepared in 5 replicates of 100 $\mu\text{g/ml}$ of Paroxetine and 4 $\mu\text{g/ml}$ of Clonazepam was made by adding 1 ml of stock solution to 10 ml of mobile phase.

Preparation of Buffer

6.8 gm of potassium di hydrogen phosphate (KH_2PO_4) was weighed and dissolved in 100 ml of water and volume was made up to 1000 ml with water. Adjust the pH to 3.5 using ortho phosphoric acid. The buffer was filtered through 0.45 μ filters to remove all fine particles and gases.

Preparation of Mobile Phase

A mixture of 30 volumes of 50 mM Phosphate buffer (KH_2PO_4) pH 4.0, 20 volumes of Methanol, and 50 volumes of Acetonitrile. The mobile phase was sonicated for 10 min to remove gases.

Optimized chromatographic conditions are as follows

- Column : INERTSIL ODS 3V column, C18 (4.6 mm \times 250 mm \times 5 μ)
- Mobile Phase : phosphate buffer: methanol: acetonitrile (3:2:5)
- Flow Rate : 1.0 ml/min
- Volume : 5 μ l
- Column Temperature : 30 $^\circ\text{C}$
- Sample temperature : 25 $^\circ\text{C}$
- Detector wavelength : 224 nm (PDA)
- Run Mode : Isocratic
- Instrument : Shimadzu with CLASSVP Software

METHOD VALIDATION

Linearity

Standard stock solutions of Paroxetine and Clonazepam ($\mu\text{g/ml}$) were prepared by dissolving 100 mg of Paroxetine and 4 mg of Clonazepam in 100 ml of mobile phase. After that filtered the solution using 0.45-micron syringe filter and Sonicated for 5 min. and dilute 100 ml with mobile phase and further dilutions.

Accuracy

Accuracy of the method was determined by Recovery studies. To the formulation (preanalyzed sample), the reference standards of the drugs were added at the level of 80%, 100%, 120%. The recovery studies were carried out three times and the percentage recovery and

percentage mean recovery were calculated for drug is shown in table. To check the accuracy of the method, recovery studies were carried out by addition of standard drug solution to pre-analyzed sample solution at three different levels 80%, 100%, 120%.

Precision

Intraday and interday variations were determined by using six replicate injections of one concentration and analyzed on the same day and different days. Precision of An analytical method is usually expressed as the standard deviation correlative standard deviation (coefficient of variation) of seriesof measurements.

Robustness

The robustness was evaluated by assaying test solutions after slight but deliberate changes in the analytical conditions. The factors chosen for this study were the flow rate (± 0.1 ml/min), temperature.

Limit of detection (LOD) and Limit of quantification (LOQ) LOD and LOQ was calculated from linear curve using formulae.

$$\text{LOD} = 3.3 * \sigma / \text{slope}, \text{LOQ} = 10 * \sigma / \text{slope}$$

(Where σ =the standard deviation of the response and S= Slope of calibration curve).

Specificity

Specificity was checked for the interference of impurities in the analysis of blank solution and injecting sample

solution under optimized chromatographic conditions to demonstrate separation of both paroxetine and clonazepam from impurities.

RESULT AND DISCUSSION

Several mobile phase compositions were tried to resolve the peak of paroxetine and clonazepam. The mobile phase containing buffer: Ammonium acetate: Acetonitrile in proportion of 82:18v/v was found ideal to resolve the peak of paroxetine and clonazepam. Retention time of paroxetine and clonazepam were 2.1 and 3.3 min respectively (Figure 3). Result of assay is shown in Table- 2. The proposed method was found to be linear in concentration range 100-300 μ g/ml for paroxetine and 2- 6 μ g/ml for clonazepam. The data was shown in Table-3 and Figure-4&5. System suitability parameters were evaluated and results shown in (Table-4), which were within acceptance criteria. The mean percentage recovery for paroxetine and clonazepam was found to be 99.9 % and 100% respectively, which are well within the limit and hence the method was found to be accurate (Table-5). LOD and LOQ values were 2.34 μ g/mL and 7.10 μ g/mL for paroxetine and 0.03 μ g/mL and 0.09 μ g/mL for clonazepam (Table-6). Results of intraday and interday precision were shown in the (Table-7a&7b). The robustness of the method was investigated by varying experimental conditions such as changes in flow rate and temperature. The result obtained implies method is robust for routine qualitative analysis (Table-8).

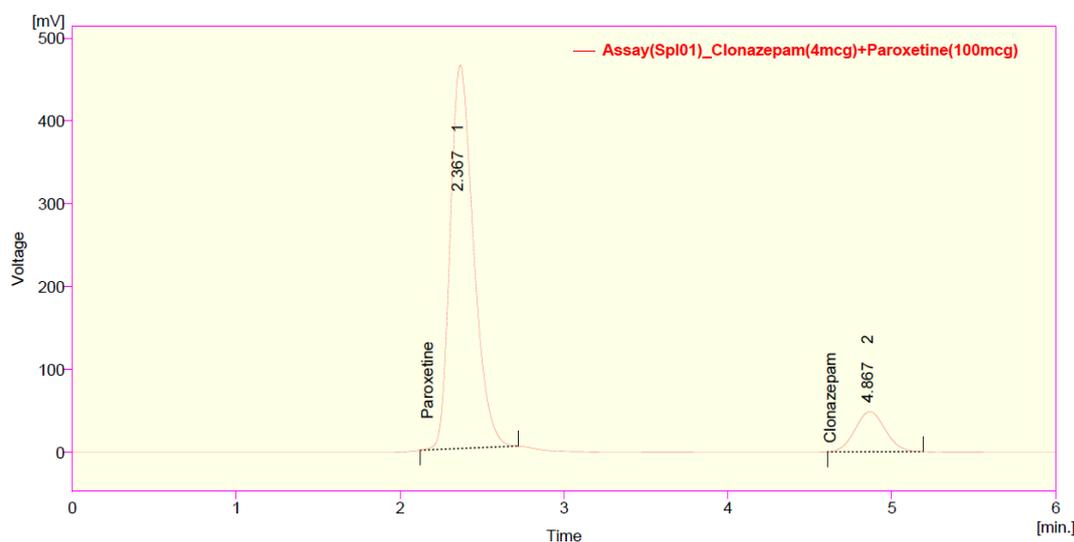


Figure 3: Chromatogram of sample.

Table 1. Observations of sample Chromatogram.

| Sl.No | Name | Retention Time | Area | USP Resolution | USP Tailing | USP Plate Count |
|-------|------------|----------------|---------|----------------|-------------|-----------------|
| 1 | Paroxetine | 2.367 | 2360112 | | 1.20 | 2320 |
| 2 | Clonazepam | 4.867 | 6610849 | 8.247 | 1.34 | 3211 |

Table 2. Analysis data of formulation.

| Injection | Label claim(mg) | Assay (%) |
|------------|-----------------|-----------|
| Paroxetine | 100 | 99.5 |
| Clonazepam | 4 | 100 |

Table 3: Linearity of Paroxetine and Clonazepam.

| S.No. | Paroxetine | | Clonazepam | |
|-------|--------------|----------|--------------|---------|
| | Conc.(µg/ml) | Area | Conc.(µg/ml) | Area |
| 1 | 60 | 2844.012 | 2.4 | 383.121 |
| 2 | 80 | 3607.702 | 3.2 | 482.379 |
| 3 | 100 | 4668.147 | 4 | 620.664 |
| 4 | 120 | 5523.646 | 4.8 | 726.780 |
| 5 | 140 | 6337.242 | 5.6 | 834.914 |

Table 4: System suitability parameters.

| Parameters | Paroxetine | Clonazepam | Acceptance criteria |
|--------------------|------------|------------|---------------------|
| Theoretical plates | 2320 | 3211 | Not less than 2000 |
| Tailing factor | 1.20 | 1.34 | Not more than 2 |
| Resolution | - | 8.247 | Not less than 2 |

Table 5. Recovery studies for Paroxetine and Clonazepam

| Drug | Spiked | Amount taken | Amount found | Percent recovery | Mean recovery |
|------------|--------|--------------|--------------|------------------|---------------|
| | level% | (µg/ml) | (µg/ml) | | |
| Paroxetine | 80 | 100 | 98.98 | 98.98 | 100 |
| | 100 | 120 | 121.61 | 101.34 | |
| | 120 | 140 | 139.88 | 99.92 | |
| Clonazepam | 80 | 4 | 3.93 | 98.15 | 99.89 |
| | 100 | 4.8 | 4.93 | 102.80 | |
| | 120 | 5.6 | 5.53 | 98.70 | |

Table 6. LOD and LOQ for Paroxetine and Clonazepam.

| Drug | LOD (µg/ml) | LOQ (µg/ml) |
|------------|-------------|-------------|
| Paroxetine | 2.34 | 7.10 |
| Clonazepam | 0.03 | 0.09 |

Table 7(a). Result of Precision.

| S.No | Sample Weight | Sample Area-1 | Sample Area-2 | % Assay |
|----------------|---------------|---------------|---------------|---------|
| 1 | 434.00 | 4583.034 | 629.697 | 99 |
| 2 | 434.00 | 4584.532 | 627.874 | 99 |
| 3 | 434.00 | 4598.577 | 632.997 | 99 |
| 4 | 434.00 | 4570.390 | 627.37 | 99 |
| 5 | 434.00 | 4546.120 | 629.795 | 99 |
| 6 | 434.00 | 4530.777 | 628.228 | 99 |
| Average Assay: | | | | 99 |
| STD | | | | 0.12 |
| %RSD | | | | 0.13 |

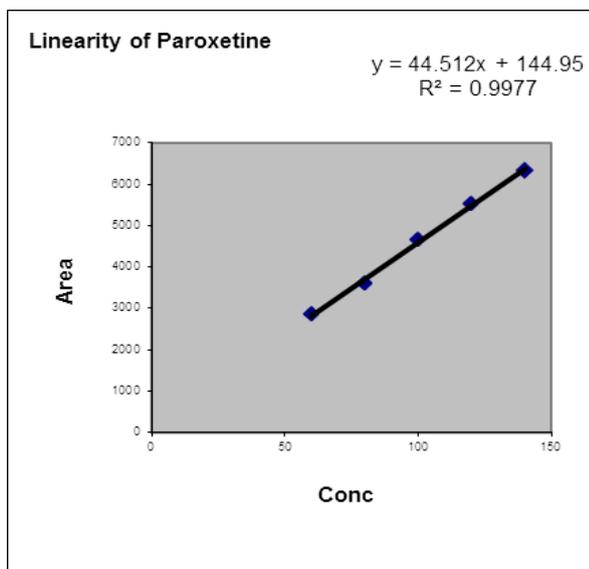
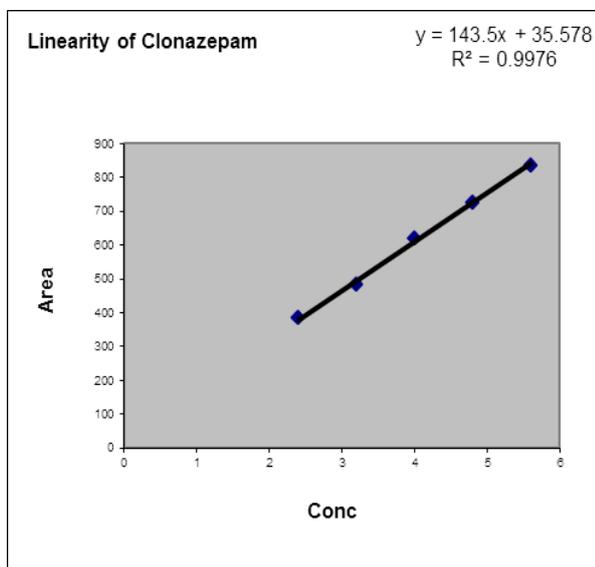
n- Number of replicate injections.

Table 7(b). Results of precision.

| Drug | Conc. (µg/ml) | Peak area (n=6) | % RSD |
|------------|---------------|-----------------|-------|
| Paroxetine | 100 | 4568.905 | 0.56 |
| Clonazepam | 4 | 629.327 | 0.33 |

Table 8: Results of Robustness study

| S.no | Parameter | Condition | Theoretical levels | | Tailing factor | | Retention time | |
|------|-------------|------------|--------------------|------|----------------|------------|----------------|-------|
| | | | PARA | CNZ | paroxetine | clonazepam | PARA | CNZ |
| 1. | Flow rate1 | 0.8 ml/min | 2335 | 3212 | 1.529 | 0.982 | 3.253 | 5.927 |
| | Flow rate2 | 1.0ml/min | 2025 | 3115 | 1.432 | 1.134 | 2.413 | 5.054 |
| | Flow rate3 | 1.2 ml/min | 2425 | 3215 | 1.438 | 1.540 | 2.157 | 4.765 |
| 2. | Wave length | 222 nM | 2334 | 3214 | 1.441 | 1.038 | 2.400 | 4.753 |
| | | 224 nM | 2025 | 3213 | 1.440 | 1.031 | 2.413 | 5.041 |
| | | 226 nM | 2435 | 3222 | 1.545 | 1.137 | 2.350 | 4.743 |

**Fig. 4: Linearity graph of Paroxetine.****Fig 5: Linearity graph of Clonazepzm.****CONCLUSION**

The proposed RP-HPLC method was used for the simultaneous estimation of paroxetine and clonazepam was found to be sensitive, accurate, precise, simple, and rapid. Hence the present RP-HPLC method may be used for routine analysis of the raw materials, in vitro dissolution study of combinational dosage formulations containing paroxetine and clonazepam.

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