

ORGANOCHLORINE TOXICITY ON LIVER OF SWISS ALBINO RATS

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ABSTRACT

Endosulfan, an organochlorine insecticide, is being used extensively and its broad-spectrum activity in commercial agriculture, poultry, livestock, home and garden pest control were administered with endosulfan. In the present study, the histological changes in the liver of Endosulphan-treated rats were significantly different from that of the normal control rats. Few hepatocytes appeared swollen and empty with indistinct cell membranes. Their, nuclei were also enlarged. The nuclear membrane of a few of these cells was lost. The size of the nucleus is an indicator of functional activity of the cell. Therefore, the observed increase in the size of nucleus suggests that these cells are over actively involved in the metabolism of endosulphan. The disrupted pattern of hepatocytic cords, capsular fibrosis, subcapsular inflammatory cells, enlarged hepatocytes, evidence of increased cellular metabolism coexistent with ballooning degeneration, microvesicular and macrovesicular fatty changes, cytoplasmic basophilia, fibrosis and inflammatory infiltrate around the portal triads along with the dilatation and congestion of the blood vessels and proliferation of bile ducts and areas of hemorrhage are suggestive of toxic hepatitis.

KEYWORDS: Hepatocytes, fibrosis, hepatitis.

INTRODUCTION

Organochlorine pesticides are being extensively used in agriculture and disease control purposes for the last more than fifty years worldwide and their long persistence in soil have been reported. This pesticide residue is capable to affect the soil fertility, crop productivity, ecological imbalance and caused human health problems (Verma *et al.*, 2011). In developing countries such as India where the economy depends largely on agricultural products, one cannot afford to loose the harvest to pests. In India, fifteen to twenty percentage of the total harvest is destroyed by pests resulting in uncontrolled use of pesticides by the Indian cultivators. Endosulfan is a widely used insecticide in India to maintain crops and food productions, fight against infecting pests, safeguard humans from vector borne diseases and related epidemics (Ayub *et al.*, 2003).

However, this pesticide has significant toxicity to non-target organisms including humans. The major routes of exposure to these chemicals are through food chain series, dermal contact, respiratory tract etc. The workers in the field and those involved in the application of these insecticides are at high risk of toxicity manifestations by these chemicals (Ayub, 2003). Various experimental studies reported congenital malformation in chicken and duck embryos with carbaryl.

Chemical nature of endosulfan

Endosulfan is an organochlorine insecticide belonging to the cyclodiene group that is extensively applied in agriculture to protect crops (Li *et al.*, 2009) It is sold under the trade name of Thiodan® which is a mixture of 70% endosulfan- α (endosulfan I) and 30% endosulfan- β (endosulfan II) (Butlar *et al.*, 1993). The technical grade endosulfan (1,2,3,4,7,7-hexachlorobicyclo-2,2,1-heptene-2,3-bishydroxy methane-5,6-sulfite) is a mixture of two stereoisomers, a \square - and \square -endosulfan, in the ratio of 7:3. It is extensively used throughout the world to control the pests on different crops.

Endosulfan and its application

Endosulfan is a broad-spectrum cyclodiene insecticide that has been used extensively for a longer period of time on a variety of crops. This chemical has been used in many countries. This pesticide has been banned in many countries; however, it is still used in the United States (Parbhu *et al.*, 2009). It has its effect on flora and fauna. Throughout the world in different countries, endosulfan is used as a pesticide and in so many countries it is banned because of its ill effects. It was a cause of panic in British Columbia, where it has been sprayed since years. Some other issues were reported in countries like Cuba, Benin and India where it is still in use and has produced very dangerous effects such as congenital birth defects, cancer, loss of immunity, neurological and mental disorders, reproductive health problems, etc.

Along with this, in many countries its toxicity was reported (Balgiar *et al.*, 2005).

The changes were seen in target organ of Albino rats liver on dermal exposure to endosulphan for 15,30 and 45 days. An increase in the activities of LDH and Acid Phosphatase suggesting hepatocellular damage was also recorded. Inhibition of liver enzymes with Endosulphan was also reported. This study was conducted to develop insights about the role of Endosulphan on liver histopathology of Albino rat.

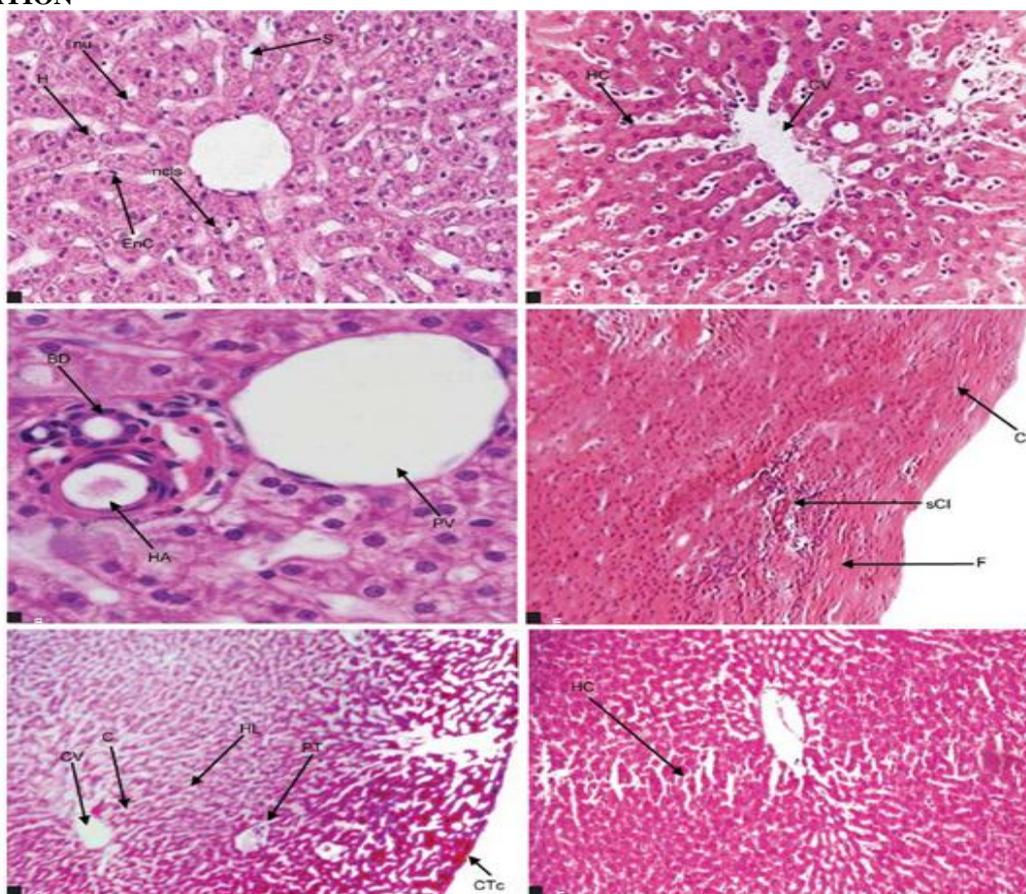
MATERIALS AND METHODS

The toxicity experiments were conducted using the chosen concentration of endosulfan LC_{50} was determined using simple graphic (% Mortality Vs. Log Concentration), probit graphic Probit value Vs. Log concentration and regression analysis methods. Sub-lethal concentration of endosulphan was 12.84. ppm. In

the present work the histopathological effects have been undertaken. Albino rats were sacrificed after 15, 30 and 45 days of exposure. The liver was dissected out. Tissue must undergo preparatory treatment before microtomy, the slides were stained with eosin for half a minute and then place in 95% alcohol again. The stain sections were place in xylene for 20 minutes, then clean and mounted in D.P.X. The liver of the rat exposed to endosulphan at 30° C temperature showed drastic changes, the result of present work showed the pathological symptoms.

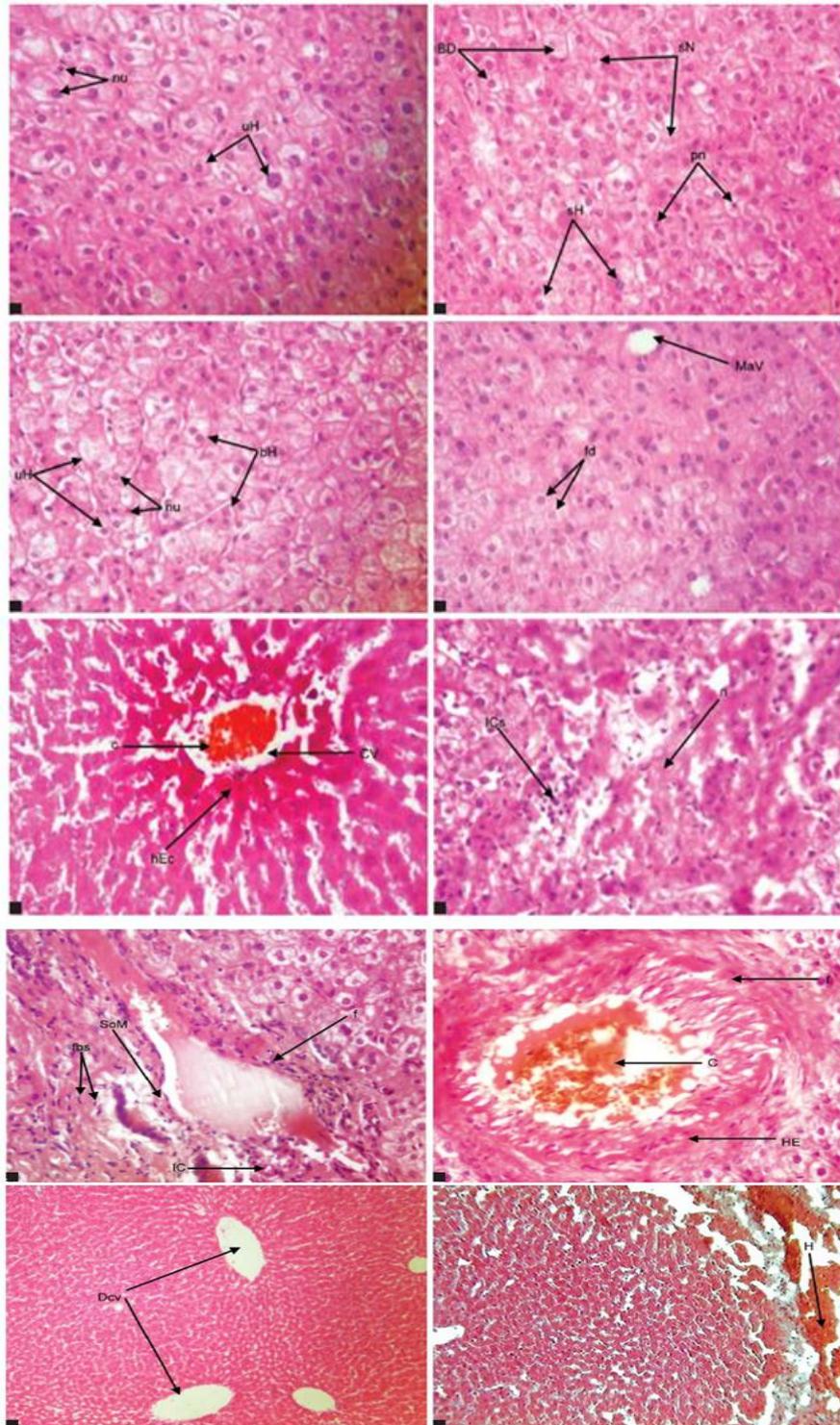
The body weights were recorded before the onset of experiment and prior to the sacrifice of animals. The liver were cut into smaller pieces (5 mm) and immediately fixed in 10% formalin. The blocks were prepared for section cutting with a microtome by paraffin wax embedding method. Sections of 6 μ thickness were cut and stained with hematoxylin and eosin (H&E) stain.13.

OBSERVATION



Figs 1A to F: Photomicrograph of transverse section of liver of group I rat showing hepatocytes (H) which are polyhedral in shape placed rounded euchromatic nucleus (nu) and a prominent nucleolus (ncls) with endothelial cells (EnC) lining the sinusoids (S). H&Estain (400 \times), (B) transverse section of liver of group I rat showing liver parenchyma with branches of hepatic with eccentrically artery(HA), portal vein (PV) and bile duct (BD) forming the portal triad. H&E stain (800 \times), rat showing unequal size of hepatocytes (uH) (C) Photomicrograph of transverse section of liver of group II rat showing connective tissue capsule (CTc) and radial arrangement of cords (C) around central vein (CV) with portal triad(PT) at the periphery of hepatic lobule (HL). and their nuclei (nu) showing(pleomorphism)with number of binucleate hepatocytes (bH). H&E stain (400 \times), (D) transverse section of liver of group II rat demonstrating the radial arrangement of hepatocyte cords around central vein. H&E stain (800 \times), (E) transverse section of liver of group III rat Showing thickened capsule (C)

with fibrosis (F) and subcapsular inflammatory cells (sCI). H&E stain (100×), transverse section of liver of group III showing disrupted hepatocytic cords. H&E stain (100×) showing necrosis (n) and inflammatory cells (ICs).



Figs 2A to J: (A) Transverse section of liver of group III rat showing unequal size of hepatocytes and their nuclei (pleomorphism). H&E stain (400×), (B) transverse section of liver of group III rat showing unequal size of hepatocytes (uH) and their nuclei (nu) (pleomorphism) with number of binucleate hepatocytes (bH). H&E stain (400×), (C) transverse section of liver of group III rat showing degenerating swollen and empty hepatocytes (sH) with central vein (CV). H&E stain (400×), (D) transverse section of liver of group III rat showing few hyper eosinophilic cells (hEc) and congestion (C) in indistinct cell membrane, few hepatocytes with swollen and partially lysed nuclei (sN), i.e. ballooning degeneration (BD) of hepatocytes and some with dense and pyknotic nuclei (pn). H&E stain (400×), (E) TS of liver of group III rat showing areas of microvesicular (foamy)

degeneration) (fd) and macrovesicular (MaV) fatty change H&E stain (400×), (F) transverse section of liver of group III rat showing necrosis (n) and inflammatory cells (ICs), predominantly lymphocytes in the liver parenchyma. H&E stain (400×), (G) transverse section of liver of group III rat showing of fibroblasts (fbs) in the space of Mall (SoM) along with inflammatory cells (IC). H&E stain (400×), (H) TS of liver of group III rat showing fibrosis (f) in the number central veins (Dcv). H&E stain (100×), (I) transverse section of liver of group III rat showing Congestion© and fibrosis (f) around the hepatic artery (HE). H&E stain (400×), (J) transverse section of liver of group III rat showing hemorrhage, (H) in the liver parenchyma. H&E stain (100×).

RESULT AND DISSUCION

Groups I and II did not show any physical signs while group III showed physical signs in the form of irritability, sneezing, lacrimation, shivering and tremors for about 1 to 1.5 hours after administration of dose. A notable clinically significant reduction in the body weight and decrease in appetite of the experimental animals was observed after Endosulphan administration, the liver in control groups was dark, reddish maroon colored large organ suspended under diaphragm by perito- neal ligaments while the liver in experimental group was reddish brown in color with some pin-point subcapsular hemorrhages over the surface. In the liver of experimental rats group, the connective tissue capsule was thickened at places, showed fibrotic changes and inflammatory cells. The one cell thick, orderly arranged pattern of the hepatocyte cords was disrupted in many areas. Most of the hepatocytes of group III were enlarged as compared to groups I and II. Many areas showed hepatocytes with dense and pyknotic nuclei. At sites, few of the hepatocytes were binucleated. There were areas of micro vesicular and macro vesicular fatty changes. The areas around the central vein showed hepatocytes that had highly eosinophilic cytoplasm with inflammatory infiltration around the portal triads due to hepatocellular degeneration. Carrol et al (1956) Many liver cells in areas away from central vein showed increased cytoplasmic basophilia due to higher metabolic activity. There was also proliferation of bile ductules in the portal triads and fibrosis was seen around many portal triads. The sinusoids, the central veins and branches of portal vein appeared dilated and congestion was seen in the central vein and branches of hepatic artery. At places, there were areas of hemorrhage where the normal parenchyma was replaced by large blood-filled spaces.

In the present study, the histomorphological changes in the liver of Endosulphan-treated rats were significantly different from that of the normal control rats. The disrupted pattern of hepatocytic cords, capsular fibrosis, subcapsular inflammatory cells, enlarged hepatocytes, evidence of increased cellular metabolism coexistent with ballooning degeneration, microvesicular and macrovesicular fatty change, cytoplasmic basophilia, fibrosis and inflammatory infiltrate around the portal triads along with the dilatation and congestion of the blood vessels and proliferation of bile ductules and areas of hemorrhage are suggestive of toxic hepatitis (Figs 2A to J).

The liver makes up 4.15% of the total body weight. Since liver is the organ where most of the substances

undergo metabolism, it becomes an organ of extreme importance to study the effect of various substances. Ton et al (2012) administered a single oral dose of C14 carbaryl (24 µCi/kg or 0.9 mg/kg) to normal mice and detected higher amounts of radioactivity in the liver and in the blood as compared to the other organs in the body at various postadministration time intervals. The accumulation of C14 carbaryl in the liver of rat and mice fetuses. In the present study, the rats became very active and irritable immediately after receiving the first dose of endosulphan. This was accompanied by sneezing, shivering and tremors for half an hour. These findings are in accordance with the reports of Adhakari, et al (2011) where endosulphan by a single oral or dermal route produced symptoms typical of cholinergic poisoning such as muscle fasciculations, tremors, excessive salivation and lacrimation, diarrhea and involuntary urination. Similar cholinergic effects were also noted.

A statistically significant decrease in the body weight ($p < 0.0001$) was observed in the -treated rats as compared to the normal control rats. It is quite obvious that endosulphan toxicity causes metabolic and structural derangements which in turn lead to wasting of the muscle mass and loss of body weight. Pant et al (2011) noted a significant decrease in the absolute weights of testes, epididymis, seminal vesicles, ventral prostate at a dose of 100 mg/kg in young rats as compared to adult rats. The lesser weight gain in the young rats was probably a direct effect on the somatic cells or an indirect influence through central nervous system and appetite. observed decrease in the weight in an elderly retired coal miner who was unknowingly exposed to carbaryl dust (10%) for a period of 8 months. The difference in the observations could probably be due to a sustained exposure to higher amounts of carbaryl over a longer duration. In the present study, many hepatocytes showed an increase in size in response to endosulphan administration. Increase in cell size following administration of carbaryl was also noted in the hypophysis and in the adrenal glands, which was due to an increase in the activity of the cells. Thus, in addition to liver, other metabolically active organs also show hyperactivity. Babu et al (1990).

In the present study, at some sites, few hepatocytes appeared swollen and empty with indistinct cell membranes. Their, nuclei were also enlarged. The nuclear membrane of a few of these cells was lost. The size of the nucleus is an indicator of functional activity of the cell. Therefore, the observed increase in the size of

nucleus suggests that these cells are over actively involved in the metabolism of endosulfan.

These findings are suggestive of an ongoing ballooning degeneration of the hepatocytes. According to observations made by -Luty et al.(2013) after dermal application of pesticide in rats, there was edema and an increase in vacuolation in the cells of the epidermis and swelling of the mitochondria. Similar changes were seen in the endothelium of the capillary vessels in the lungs and the heart. The cytoplasm of many hepatocytes appeared to contain several tiny vacuoles, giving it a foamy appearance. This is indicative of a fatty change in the liver cells. Chaudhary et al (2003) These ultrastructural changes seen suggest that the cytoplasm might be participating in the metabolism of endosulphan and the overactivity progressively exhausted the cell leading to degeneration. It also suggests that endosulphan affects many organs of the body by transdermal route in addition to the intraperitoneal route used in the present study.

Areas around the portal triads and central vein showed hepatocytes that become shrunken and had a highly eosinophilic cytoplasm. Their nucleus was dense and pyknotic. These findings are suggestive of hepatocellular degeneration and are in accordance with the findings of Fukuto et al.(1953) who also noted degenerative changes in the hepatocytes, pyknotic cells in the stratum spinosum of the epidermis and in the Purkinje cells of the cerebellum following dermal application of chlorpyrifos. In the present study, the histomorphological changes, in the liver of endosulphan treated rats was significantly different from that of the control rats. The disrupted pattern of the one cell thick orderly arrangement of hepatocytic cords, is evidence of increased cellular metabolism coexistent with ballooning degeneration, inflammatory infiltrate around the portal triads along with the dilatation of the blood vessels and the bile canaliculi, are suggestive of toxic hepatitis induced by endosulphan. Khan- et al.(2013).

CONCLUSION

Endosulfan, though used for betterment of mankind, has ultimately resulted in being dangerous to environment. The toxic residues of it impairs the development and normal functioning of the hormone-dependent processes in flora, fauna and has lead to severe and chronic human poisoning. It has been banned in many developed countries. Bane of endosulfan in India is in consideration, as it leads to many drastic effects, resulting in mortality. It is the requirement of the present time to reduce the ill effect of this pesticide. Government must come forward with an alternative to reduce the hazardous effect of this pesticide strictly or stop its future use completely.

REFERENCES

1. Ayub, M., Brodie .B. P. and Kendal. F. E (2003) Simplified method for estimation of total cholesterol

- in serum and demonstration of its specificity. *J. Biol. Chem.*; 195: 357- 361.
2. Adhikari, N. and Grover, I. S (1988) Genotoxic effects of some systemic pesticides: in vivo chromosomal aberrations in bone marrow cells in rats. *Environ. Mol. Mutagen.*, 12: 235-242.
3. Babu, G. R. V., Reddy, G. R., Reddy, A. T.V., Rajendra, W. and Chetty, C. S(1990) Effect of in vitro of propoxur on calcium dependent ATP hydrolysis and calcium uptake by the rat brain synaptosomes. *Biochem. Intrnl.*, 21: 1089-1094.
4. Baligar, P. N. and Kaliwal, B. B., Malviya, A. N(2001). Survey among workers in Pesticide factories. *Toxicol. Lett.* 10: 129- 132.
5. Butler, A.M., Murray, M(1993) Inhibition and inactivation of constitutive cytochromes P450 in rat liver by parathion. *Mol. Pharmacol.*, 43: 902-908.
6. Carrol, N.V., Langely, R.W. and Row, R.H(1956) Glycogen determination in liver and mode by use of anthrone reagent. *J.Biol. Chem.*, 20: 583-593.
7. Choudhary, N., Sharma, M, Verma, P., Joshi, S.C(2003) Hepato and nephrotoxicity in rat exposed to endosulfan. *J. Environ. Biol.*, 24: 305-308.
8. Lie, T., Desai, D. (2010) Mechanisms of chlordecone⁷⁴ *Effects of Carbosulfan on Albino Mice* neurotoxicity A Review. *Neurotoxicity*, 3: 103-110.
9. Prabhu, B., Dikshith, T. S. S., Raizada, R. B., Datta, K.K(2009) Interaction of phosphomidon and Benzene in female rabbits. *Ind. J. Exptl. Biol.*, 18: 1273-1277.
10. Pan, W. F., Wolfe, H. R. (2011) Measurement of the exposure of workers to pesticides, *Bull. Wld. Hlth. Org.*, 26: 75-91.
11. Fukuto, R. T. (1983) Structure-activity relationship in derivatives of anticholinesterase insecticides. In: Pesticide chemistry human welfare and the environment. Pergmen Press Ltd., Headington Hill Hall, Oxford OX3 OBW, England, pp. 203- 212.
12. Khan, S. and Sharma, N. 2010. Effect of organophosphate Insecticide on protein and glycogen content in liver and kidney of *Gambusia affinis*. *International Journal of Ultra Scientist of Physical Sciences*, 22(3): 625-631.
13. Ton, L., Klassen, C. P. and Plaa, G. L. (2013) Relative effects of various chlorinated hydrocarbon on liver and kidney function in mice. *Toxicol. & Appl. Pharmacol.*, 9: 139.
14. Verma, S., Kostka, G., Palut, D., Kopec-Szlezak, J., Kudwicki, J. K. (2012) Early hepatic changes in rats induced by permethrin in comparison with DDT. *Toxicology*, 142: 135-143.