



AN EFFECTIVE APPROACH TO REDUCE THE PENETRATION POTENTIAL OF SARS-COV-2 AND OTHER VIRUSES BY SPIKE PROTEIN: THROUGH SURFACE PARTICLE ELECTROSTATIC CHARGE NEGOTIATION

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ABSTRACT

The objective of this paper is to provide a mathematical model to construct a barrier that may be useful to prevent the penetration of different viruses (Eg. SARS-COV-2) as well as charged aerosols through the concept of electrostatic charge negotiation. (Fusion for the opposite types of charges and repulsion for the similar types of charges). Reviewing the works of different authors, regarding charges, surface charge densities (σ), charge mobility (μ) and electrostatic potentials of different aerosols under varied experimental conditions, a similar intensive study has also been carried out to investigate the electron donating and accepting (hole donating) properties of the spike proteins (S-proteins) of different RNA and DNA viruses, including SARS-COV-2. Based upon the above transport properties of electrons of different particles having different dimensions, a mathematical model has been established to find out the penetration potential of those particles under different electrostatic fields. An intensive study have been carried out to find out the generation of electrostatic charges due to the surface emission of electrons (SEE), when a conducting material like silk, nylon or wool makes a friction with the Gr IV elements like Germanium or Silicon, it creates an opposite layer of charges in the outer conducting surface and the inner semiconducting surface separated by a dielectric materials. This opposite charge barriers may be considered as Inversion layers (IL). The electrostatic charges accumulated in the layers between the Gr IV Ge is sufficient enough to either fuse or repel the charges of the spike proteins of the RNA, DNA viruses including SARS-Cov-2 (RNA virus) or the aerosols.

KEYWORDS: SEE, Transport properties, Inversion Layer, surface charge density, SARS-Cov-2.

1) Theoretical Background, Journal study and Investigation

1.1 Calculation of Charges of Different Aerosols and Graphical Analysis

A study by M.V.Rodrigues^[1] et al on the values of charges accumulated on different aerosol particles found, that the accumulation of the charges and charge densities(σ) largely depend upon the Stokes diameter of the particle. The variation of charges with the Stokes diameter has been found most likely to be following a linear graph with a definite slope. The results have been used extensively in our study.

However, it has also been observed that for a larger particle having diameter more than $d > 4 \mu\text{m}$, abruptly different levels of charge density are present.

In the experimental setup consisting of an ECC (Schematics in adjoining figure), a set of values for the

charge accumulation on dry egg dust particles and their respective Stokes radius was found. It is referred below in Table 1.

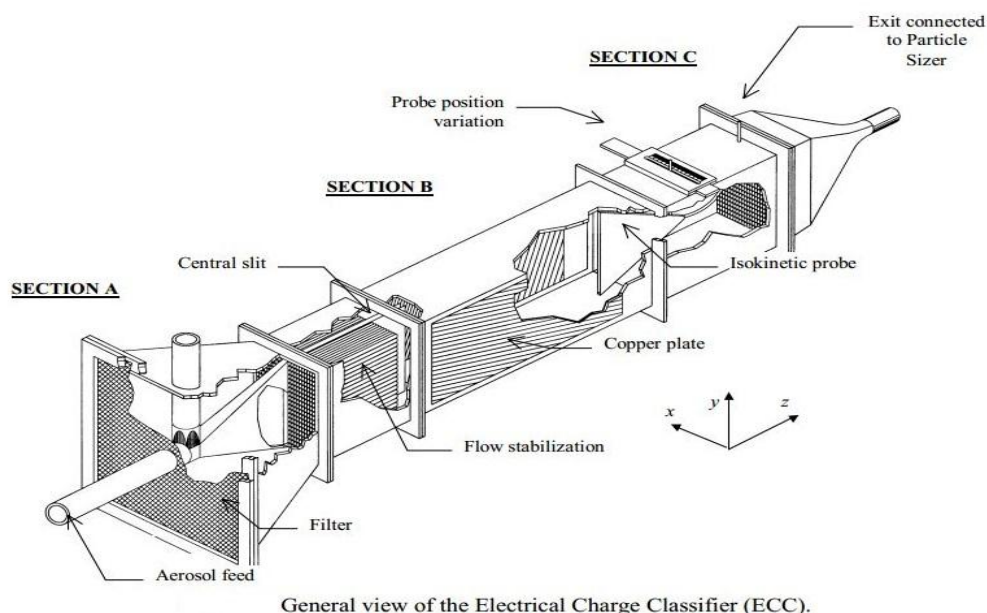


Table 1.

Stokes Diameter [10^{-6} m]	Charge[10^{-17} C]
1.7	0.5
2.1	0.5
2.5	1.2
3.2	1.6
4.8	3.0

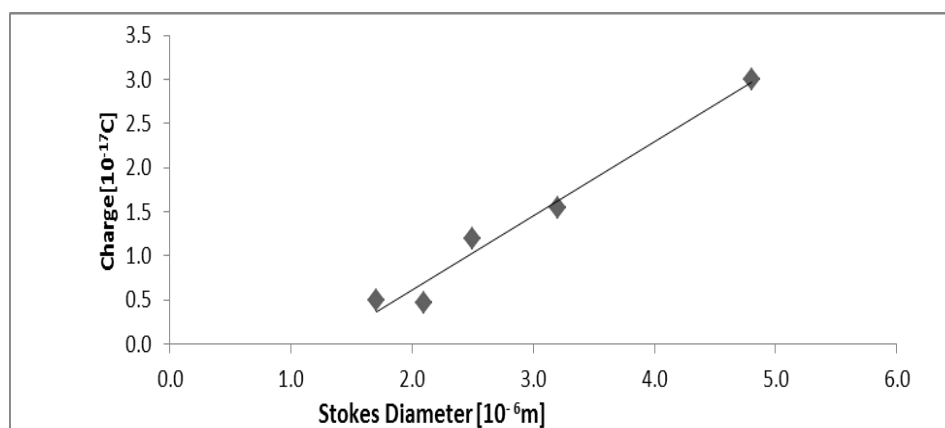


Figure I: Particle charge as a function of Stokes diameter for aerosol (Concentrated dry egg dust particle - Data set in Table 1).

The data from Table 1 is fitted to the equation 1.1.1 (by Jonhston et al. 1987). $|Q/e|=Ad_p^B$ [1.1.1]

Where e is the elementary charge of an electron and A is the median number of elementary charges of magnitude e present on a particle of diameter $1\text{ }\mu\text{m}$. It is found that, the value of B is nearly equal to 2 and also as per as suggested by Jonhston. (Range between 1 and 2).

This expected range of B is also applicable for calculating the charge of any small particles.

1.2 Theoretical investigation for the factors, influencing the charge on SARS-COV-2 surfaces.

The accumulation of charges on the surface of SARS-COV-2 can be attributed by the following four factors.

- **The size and shape of the virus:** Literature study [Angeletti et al⁴] suggests, SARS-COV-2 has a spherical or ellipsoidal shape having average diameter in the range of 60 nm to 140 nm. This makes a prediction that the pattern of charge on the surface of SARS-COV-2 is similar to that of aerosols as established in 1.1.

- **The (pKa)s of the protein on the surface:** At the isoelectric point of a protein, there is an equivalent distribution of negative and positive charges, leading to a neutral response to a potential difference. However, if the pH be lowered from the isoelectric point, it is found that, a bias is created towards the centre of positive charge of the protein. The pKa's could be derived from the pH values of the titration curve for the S(spike)-protein of the SARS-CoV-2 using the Henderson-Hasselbalch equation (from titration analysis).

$$\text{pH} = \text{pK}_a + \log \left(\frac{[\text{conjugate base}]}{[\text{weak acid}]} \right) \quad [1.2.1]$$

To establish this concept of the influence of surrounding pH on the surface charge of a virus, similar experiments had already been carried out also over the other virus strains.

Relevant examples

Poliovirus 1 (strain LSc) was established to be isoelectric in nature at pH level of 6.6. [Zerda *et al*⁵]. A second strain of poliovirus 1 (strain Brunhilde) had been found isoelectric at the level of pH 7.1. Phage MS-2 was isoelectric at pH 3.9. Also, reovirus type 3 has been reported to be isoelectric at pH 3.9 by column focusing techniques [Zerda *et al*⁵]. Hence, it can be concluded that, there is a tendency of viruses to accumulate different charges on the surface area at different pH levels.

- **The protein size of the outermost spikes of the virus:** Also considering that the 1255 long amino acid sequence of the S-protein has been found and the corresponding possible oxidizing state of the contributing amino acids can be visualized, one may actually determine exactly how much charge is accumulated on each protein subunit at a particular pH level.

As stated earlier, actual fractional dissociation (α_i) of any group which can be ionized is related to pH by the Henderson-Hasselbalch equation.

$$\text{pH} = \text{pK}_i + \log \left[\frac{\alpha_i}{1 - \alpha_i} \right] \quad [1.2.2]$$

Where, pK_i may be calculated if α_i at corresponding pHs are known. The pK_i depicts the negative logarithm of the effective dissociation constant. In the usual charge determination procedure [Jakubke and Jeschkeit, 1977; Skoog and Wichman, 1986], the pK_i for each type of ionizable group is assigned a magnitude. The verified knowledge of the amino acid sequence of a protein is used to calculate the net charge, z_p , as $-(z_p)_i$, where the charge of protein arising from n_i ionizable groups of type i , $(z_p)_i$, is given by.

$$(z_p)_i = n_i z_i \alpha_i \quad [1.2.3a]$$

where the ionizable group, i , is anionic (z_i is negative)

$$\text{and, } (z_p)_i = n_i z_i (1 - \alpha_i) \quad [1.2.3b]$$

where, the ionizable group, i , is cationic (since, z_i is positive).

However, pK_i cannot be assigned a fixed magnitude because of its dependence on the overall charges of the protein. It is more difficult to dissociate a proton from a negatively charged molecule than from one with a net positive charge. This variation of pK_i can be taken into account [Compton and O'Grady, 1991] by means of the theoretical expression for proton dissociation that has been in existence for over 80 years [Linderstrøm-Lang 1924; Linderstrøm-Lang and Nielsen, 1959]. Specifically, pK_i can be expressed by.

$$\text{pK}_i = (\text{pK}_{\text{int}})_i - 0.868 w z_p \quad [1.2.4]$$

The Spike protein (S Protein) is a large type of transmembrane protein ranging from 1160 amino acid for Avian infectious Bronchitis virus (IBV) and upto 1400 amino acids for Feline Corona virus (FCo) [Taylor Heald-Sargent, *et al*²¹]. It also has been revealed from different studies of Donald J. Winzor⁷ and Harold P. Erickson⁸ that, approximate numbers of electrons can effectively be calculated from the ratio of the mass of the spike proteins and that of the amino acids since it may be considered that each of the amino acid can carry maximum a single donating or accepting charge. (depending upon their shapes and sizes). The ultimate structural understanding of a protein comes from an atomic-level structure obtained by X-ray crystallography or nuclear magnetic resonance (NMR) spectroscopy. However, structural information at the nanometer level is frequently invaluable. Hydrodynamics, in particular sedimentation and gel filtration, can provide this structural information, and it becomes even more powerful when combined with electron microscopy (EM).

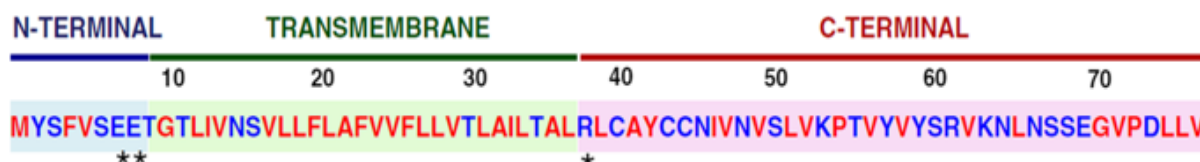
A hypothetical visualization is that if we consider one charged site per 'n' amino acids at a particular pH, the number of free electrons present per nano meter length of spike proteins remain in the order of (1400/n) and the value of the charges per nano meter length of the spike may be of the order of $1.6 \times 10^{-19} \times (1400/n) = (2240 \times 10^{-19}/n)$ C. (Experimental determination of the value of n is beyond the scope of this paper and is strongly recommended by the Authors). Therefore, at a particular pH.

$$\text{Charge per nm length of spike protein} = (2240 \times 10^{-19}/n) \text{ C} \quad [1.2.5]$$

Considering the above values, a mathematical model has been established in 1.3 to find out the charges of the spike proteins of different lengths ranging from 8.0 nm to 10.0 nm for SARS-CoV-2. It has been revealed by [Zerda *et al*⁵]. Therefore, it can be predicted that, the spike proteins of SARS-CoV-2 shows measurable electron donating or accepting capabilities.

• Maintenance of Membrane Potential across Enveloping protein.

If we look at the enveloping proteins of the SARS-CoV, refereeing to the following research by **Dewald Schoeman *et al*⁴⁵**, we have found that, the Envelope Proteins (E-proteins) have a capability to maintain a neutrality through the membrane potential of their own, till a considerable amount of charge difference is created when the structure of whole envelope collapses.



Amino Acid Sequence and Domains of the SARS-CoV E Protein. Amino acid properties are indicated: hydrophobic (red), hydrophilic (blue), polar, charged (asterisks).

Comparative and phylogenetic analysis of SARS-COV E revealed that, a substantial portion of the TMD consists of the two nonpolar, neutral amino acids, valine and leucine, lending a strong hydrophobicity to the E protein. The peptide exhibits an overall net charge of zero, the middle region being uncharged and flanked on one side by the negatively charged amino (N)-terminus, and, on the other side, the carboxy (C)-terminus of variable charge. The C-terminus also exhibits some hydrophobicity but less than the TMD due to the presence of a cluster of basic, positively charged amino acids.

An investigation regarding maintenance of this membrane potential had been done in case of Semiliki Forest Virus [Andre V. Samsonov *et al*⁶] which is also an RNA virus, where it was established that, at a relatively positive charge, the potential barrier of the E1 and E2 envelope had been collapsed and the virus was spilled out by its contents. Hence, it might be abstracted that coronavirus will also show similar results (Experimental determination of which is beyond scope of this theoretical work.) and that, there is a tendency among RNA viruses to develop (by any process) a general neutral Hydrophobic nature until the potential difference is very high, when the envelope system collapses.

1.3 Calculation of the charges of the S-proteins of different coronaviruses viruses including SARS-COV-2 based on similarity to aerosols and their Graphical analysis.

The virus responsible for COVID-19 is SARS-CoV-2 which belongs to the category of β corona virus. The structure of these viruses are found mostly spherical or ellipsoidal in nature having diameter in the range of 60nm to 140 nm, additionally having spikes in the range

The COV E protein is a short, integral membrane protein of 76–109 amino acids, ranging from 8.4 to 12.0 kDa in size. The primary and secondary structure reveals that, E protein has a short, hydrophilic amino terminus consisting of 7–12 amino acids, followed by a large hydrophobic Trans Membrane Domain (TMD) of 25 amino acids, and ends with a long, hydrophilic carboxyl terminus, which comprises the majority of the protein. The hydrophobic region of the TMD contains at least one predicted amphipathic α -helix that oligomerizes to form an ion-conductive pore in membranes.

of length 8 nm to 10 nm. [Point to be noted: The aerosol particles referred to in 1.1 also have a diameter in the range of 50 nm to 100 nm.].

It should be considered here that the S-protein in the spikes of the coronavirus is essentially a complex folded structure of a chain formed from the the S gene that comprises an ORF encoding a protein of 1353 amino acid residues, with a predicted molecular weight of 149,918 (*S Mounir¹, P Labonté, P J Talbot*). It tends to have different charge accumulations and configurations in different pH levels of the surroundings, unlike simple inorganic particles of aerosols.

Now, reaction to pH level is actually generated due to the difference of electric potential energy level of an electron from the energy level of the 1s orbital of the H^+ . Similar possible different energy state of electron is obtained from bringing a molecule near to a charged surface. Hence this similarity has been exploited in this investigation. According to the recent research work^[5,7], mutation in the spike protein is probably responsible for jumping this virus (after mutation) from species to human and [Angeletti *et al*⁴] proposed that the ORF1ab is the largest gene in SARS-CoV-2 which encodes the pp1ab protein and 15 nsps. This means that a structure forming such a major bulk of the virus is expected to have significant effects on the charge accumulation on the virus surface, behaving in closed spaces not unlikely to that in certain pH's.

The charges of the spike proteins have been measured in different laboratories by different methods but all of them have established a concrete correlation with the length and diameter of the spike proteins. However, it has been found that in most of the cases, the level of accumulation of charges show saturation beyond a certain length of the S-proteins. A possible explanation would be due to the destructive field interference of similar charges.

Presented below are some sets of data furnished from equation [1.2.5] with minimal standard deviation to extrapolate viable charge concentrations on coronaviruses.

Accumulated charges per nm length of S-protein spikes on Coronavirus at a particular pH (S-protein in the range of 8 nm to 10 nm)

Spike length (nm)	(Charge X n) (X 10 ⁻¹⁹ C)
8.1	1.296
8.2	1.311
8.3	1.318
8.4	1.324
8.5	1.334

8.6	1.366
8.7	1.372
8.8	1.391
8.9	1.401
9.0	1.412
9.1	1.443
9.2	1.461
9.3	1.472
9.4	1.481
9.5	1.501
9.6	1.512
9.7	1.532
9.8	1.544
9.9	1.564
10.0	1.589

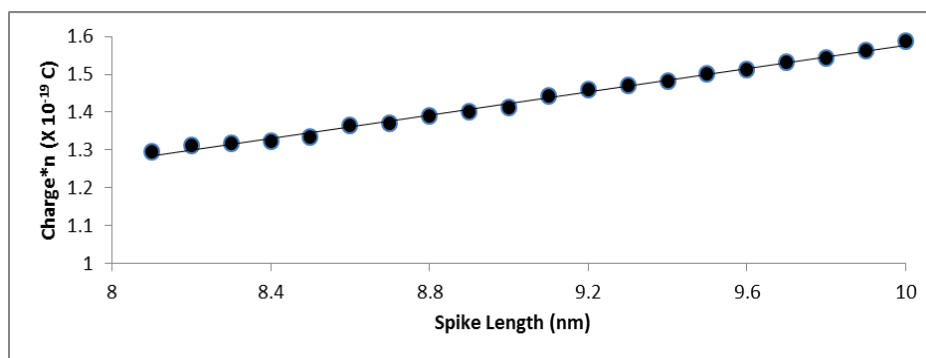


Figure II: Particle charge as a function of Spike Length of SARS-COV-2.

Here, it can be seen that, the charge of the S-protein is predicted to vary linearly with spike length at a particular pH.

2) Mathematical model developed for calculating net charge on the surface of a single virus:

2.1 The factors affecting charge formation.

The effects due to various factors affecting the charge are listed below in the form of equations stated beforehand.

i. Due to size of virus: (From 1.1.1)

$$|Q/e| = A d_p^B$$

ii. Due to interaction of S-protein with surrounding pH (from 1.2.3a):

$$(z_p)_i = n_i z_i \alpha_i$$

And from [1.2.3b]

$$(z_p)_i = n_i z_i (1 - \alpha_i)$$

$$\text{From [1.2.2]} \quad \text{pH} = \text{pK}_i + \log \left[\frac{\alpha_i}{1 - \alpha_i} \right]$$

$$\text{From [1.2.4]} \quad \text{pK}_i = (\text{pK}_{\text{int}})_i - 0.868 \text{wz}_p$$

From the above four equations, we may derive a relation between the net charges on protein with the surrounding pH if pK_i is known:

$$\text{pH} = [(\text{pK}_{\text{int}})_i - 0.868 \text{wz}_p] + \log \left[\frac{((z_p)_i / n_i z_i)}{1 - ((z_p)_i / n_i z_i)} \right] \text{ for anionic ionisable group [2.1.1a]}$$

$$\text{pH} = [(\text{pK}_{\text{int}})_i - 0.868 \text{wz}_p] - \log \left[\frac{((z_p)_i / n_i z_i)}{1 - ((z_p)_i / n_i z_i)} \right] \text{ for cationic ionisable group [2.1.1b]}$$

2.2 Derivation of probable formula to predict charge formation on virus

At the isoelectric point of the S-protein of the virus, Equation 1.1.1 is the only factor affecting the charge formation. However, when the virus is in a different pH level from that of the isoelectric points of S-protein, Equation 2.1.1a and equation 2.1.1b are also applicable which affects the formation of charges. Hence at a particular pH, incorporating both the equations, the following mathematical model has been established.

$$\text{From 1.1.1,} \quad Q = A e d_p^B$$

$$\text{From 2.1, } \text{pH} = [(\text{pK}_{\text{int}})_i - 0.868 \text{wz}_p] \pm \log$$

$$\left[\frac{((z_p)_i / n_i z_i)}{1 - ((z_p)_i / n_i z_i)} \right]$$

$$\Rightarrow (\text{pH} - (\text{pK}_{\text{int}})_i + 0.868 \text{wz}_p) = \pm \log \left[\frac{((z_p)_i / n_i z_i)}{1 - ((z_p)_i / n_i z_i)} \right]$$

$$\Rightarrow e^{\{\pm (\text{pH} - (\text{pK}_{\text{int}})_i + 0.868 \text{wz}_p)\}} = \left[\frac{((z_p)_i / n_i z_i)}{1 - ((z_p)_i / n_i z_i)} \right]^{-1} - 1$$

$$\Rightarrow (z_p)_i = (n_i z_i) [1 + e^{\{\pm (\text{pH} - (\text{pK}_{\text{int}})_i + 0.868 \text{wz}_p)\}}]^{(-1)} \quad [2.2.1]$$

Here an assumption is made that, the charge formation on the S-protein due to pH change shall not affect the inherent tendency of the virus.

Thus, if the total charge at a particular pH on the virus be Q_{tot} , then from Equation 1.1.1 and Equation 2.2.1, it's value may be expressed as,

$$Q_{\text{tot}} = Q + [(z_p)_i] \{(\pi d_p^2) k\}$$

(Where k is the number of spikes per unit surface area of the virus. However, this value is not determined and the authors strongly suggest an experimental determination of this value)

$$Q_{\text{tot}} = Ae d_p^B + [(n_i \cdot z_i) [1 + e^{\pm (pH - (pK_{\text{int}})I + 0.868wzP)}]]^{(-1)}$$

$$\Rightarrow Q_{\text{tot}} = (Ae) d_p^B + \{[(n_i z_i) [1 + e^{\pm (pH - (pK_{\text{int}})I + 0.868wzP)}]]^{(-1)}\} (\pi k) d_p^2 \quad [2.2.2]$$

Here it can be observed that, at a particular pH, the Q_{tot} is still an exponential function of the diameter of the virus. Now, if we relate the charge of the virus to the dry egg dust aerosol, it is expected to show similar types of properties as the virus is also behave like a dry egg dust particle, which is a rich in protein. Thus a value of B nearly equal to 2 can be expected here. Assuming B to be almost equal to 2, in this case the equation may be simplified for relatively smaller values of d_p as follows.

At a particular pH level,

$$Q_{\text{tot}} = (Ae + K) d_p^2$$

$$\text{Where, } K = [(n_i z_i) (1 + e^{\pm (pH - (pK_{\text{int}})I + 0.868wzP)})^{(-1)}] (\pi k)$$

$$\Rightarrow Q_{\text{tot}} = \chi d_p^2 \quad (2.2.3)$$

$$\text{Where, } \chi = Ae + [(n_i z_i) [1 + e^{\pm (pH - (pK_{\text{int}})I + 0.868wzP)}]]^{(-1)}$$

(πk)

This parameter ' χ ' may be experimentally determined through suitable procedure. Hence, it would provide a fairly accurate prediction of the net charge on the surface of a virus at a particular pH level following the lines of this mathematical model.

Now, a particular potential difference may show very similar effect on ionizing proteins at a particular pH. Hence, this model may be extended for the calculation of charges on a virus surface under the influence of a particular potential gradient. In that case, let the parameter be ' C ' (To be experimentally determined) and the equation be as follows.

$$Q_{\text{tot}} = C d_p^2 \quad (2.2.4)$$

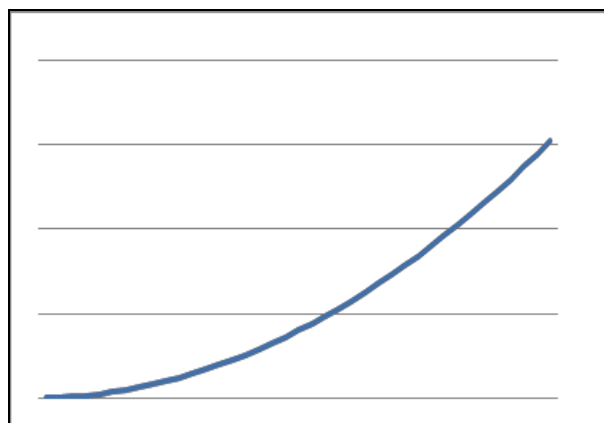


Fig III: This is a demonstration of nature of increase of Q_{tot} (depicted in vertical axis) with respect to d_p (depicted in horizontal axis).

3) Prospective model of the potential barrier to stop viruses

3.1 Formation of surface charge density (σ) over IL as a function of time

The electrostatic charges form due to the surface emission owing to the frictions between the layers of semiconducting materials like Ge with the conducting materials like nylon, wool.

When the conducting materials like wools make friction with a semiconducting Gr IV materials, it shows a surface emission of electrons and the charges remain accumulated at the outer conducting layers since the middle layer of the mask is a semiconductor and is separated by an effective dielectric medium.

The charge densities per square cm area (σ) follow empirically the growth of charges equation in a capacitor.

$$\sigma = [\epsilon \mu N_f / 4\pi r^2] [(1 - e^{-t/CR}) dt] [(dS/dx)] \quad [3.1.1]$$

Where, ϵ is the permittivity of the medium, μ is the coefficient of friction between the two layers, N_f is the normal force acting over the surface depending upon the rate of movement, $4\pi r^2$ is the surface area and dx is the separation distance between the conducting and semi conducting surface separated by the dielectric materials. Given below is a graphical representation of 3.1.1

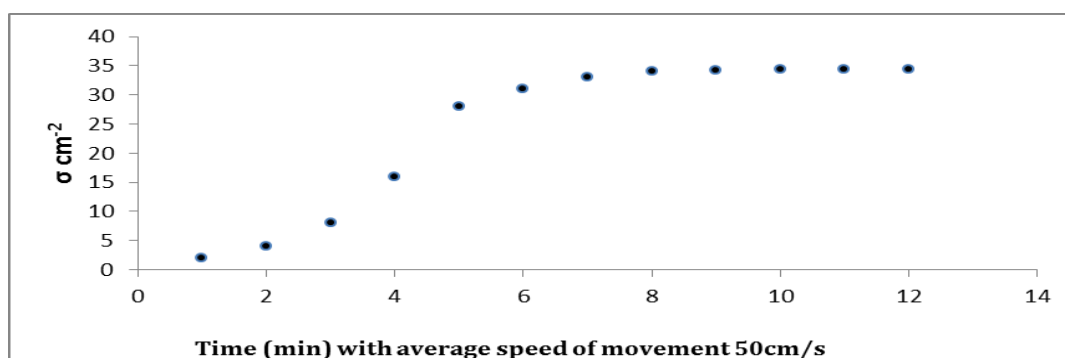


Fig III: Variation of accumulation of charge densities (σ) per cm^{-2} with time in the IL due to average movement of 50 cm/s.

Clearly, the density of accumulated charge is a time dependent, area dependent and movement dependent function. The produced charges will form an inversion layer (IL) at the outer and inner surface due to the induction. However the charge accumulation attains a finite quantity after a certain period of time, as can be seen from Fig III.

3.2 Comparison of static charges produced in the layers of mask (IL) and the surface of the viruses and/or the air-borne aerosols.

From the above studies it is clear that, the accumulation of charges per square cm area of a conducting bi layer due to the surface emission of electrons in normal movement is at least 10^6 times more than the charges on surface of viruses including SARS-CoV-2 and also the charge density is much higher than the charges accumulated in the concentrated dust particles of normal ranges.

Therefore, it may be concluded that, due to the surface emission of electrons, the IL of a mask can effectively trap, neutralize or repel the the viruses and/or the charged aerosols which might be carrying air borne viruses.

4) CONCLUSION

The charge accumulation on the surface of a virus is established to be in accordance with the Equation 2.2.2:

$$Q_{\text{tot}} = (Ae)d_p^B + \left[\left((n_i z_i) \left[1 + e^{\left\{ \pm \frac{(pH - (pK_{\text{int}})I + 0.868wzP)}{1} \right\}} \right] \right)^{(-1)} \right] \{\pi k\} d_p^2$$

Which has been simplified with some basic assumption to Equation 2.2.3:

$$Q_{\text{tot}} = \chi d_p^2$$

(Where, $\chi = [Ae + \left((n_i z_i) \left[1 + e^{\left\{ \pm \frac{(pH - (pK_{\text{int}})I + 0.868wzP)}{1} \right\}} \right)^{(-1)} \right] \{\pi k\}]$)

Using a derivation from this mathematical model, the model of a protective barrier has been established as follows.

A three layer mask can be produced to prevent the immediate infections of DNA, RNA viruses including SARS-COV-2 and others because the electric charge accumulation of the RNA viruses is much less than the electrostatic charges accumulated in the layers of the mask within few minutes. In the inner surface, a cotton type non conducting material can be used which will work basically as a nonconductor so that the electrostatic charges produced inside the two layers does not drain out through surface of the (human)body. Additionally, the cotton layer may be effective to protect the skin from electrostatic thermal radiation.

Frictions between the middle and the outer layers, where the hydrophobic conducting and semiconducting materials are used, effectively lead to accumulation of the static charges.

The rate of accumulation of charges increase with the increase of friction but come to a saturation level following the model of growth of charges in a capacitor per square cm of area.

5) Suggestions for further experimental determinations.

From the above studies, authors strongly recommend the following investigations:

- Experimental determination of the value of 'n' as specified in Equation 1.2.5.
- Experimental determination of the value of 'k' as specified in Equation 2.2.2.
- Experimental determination of parameters 'χ' and 'C' as specified in this article.
- Conducting of further experiments to find out the applications of the same for the preparation of PPE or gloves.

REFERENCES

1. M.V.Rodrigues, W.D. Marra Jr, R.G. Almeida and J.R.Coury, Measurement of the electrostatic charge in airborne particles: II-particle charge distribution of different aerosols, Brazilian Journal of Engineering, January- March, 2006; 23(01): 125-133, ISSN: 0104-6632.
2. Yun-Haeng Joe, Joonmik Shim, Il-Kyoung Shine, Se-Jin Yook, Hyun-Seol Park, A study on electrical charge distribution of aerosol using gerdien ion counter, Aerosol and Air Quality Research, 2018; 18: 2922-2928, ISSN: 1680-8584 print/ 2071-1409 online, doi: 10.4209/aaqr.2018.08.0309
3. Bruce Forsyth, Benjamin Y.H.Liu & Francisco J. Romy, Particle charge measurement for commonly generated laboratory aerosols, Aerosol Science and Technology, 28: 6, 489-501, ISSN: 0278-6826, doi: 10.1080/02786829808965540.
4. Angeletti S, Benvenuto D, Bianchi M, Giovanetti M, Pascarella S, Ciccozzi M, COVID-2019: The role of the nsp2 and nsp3 in its pathogenesis, J. Med. Virol, 2020 Feb 2.
5. KATHERINE S. ZERDA,lt CHARLES P. GERBA,l* KENNETH C. HOU, AND SAGAR M. GOYAL: Adsorption of Viruses to Charge-Modified Silica Departments of Microbiology and Immunology and Nutrition and Food Science, University of Arizona, Tucson, Arizona 857211; AMFICUNO, Meriden, Connecticut 065402; and Department of Veterinary Diagnostic Investigation, College of Veterinary Medicine, University of Minnesota, St. Paul, Minnesota 55083 Received 20 April 1984/Accepted 5 October 1984 APPLIED AND ENVIRONMENTAL MICROBIOLOGY, Jan. 1985; P. 91-95 0099-2240/85/010091-05\$02.00/0 Copyright C) 1985, American Society for Microbiology
<https://aem.asm.org/content/49/1/91.full.pdf>
6. Andrey V. Samsonov, Prodyot K. Chatterjee, Vladimir I. Razinkov, Christina H. Eng, Margaret Kielian, Fredric S. Cohen, Effects of

- Membrane Potential and Sphingolipid Structures on Fusion of Semliki Forest Virus
DOI: 10.1128/JVI.76.24.12691-12702.2002
<https://jvi.asm.org/content/76/24/12691>
7. Donald J. Winzor (Current Protocols in Protein Science (2005) 2.10.1-2.10.8 Copyright © 2005 by John Wiley & Sons, Inc,
 8. Harold P. Erickson, Size and Shape of Protein Molecules at the Nanometer Level Determined by Sedimentation, Gel Filtration, and Electron Microscopy: Shulin Li (ed.), Biological Procedures Online, Volume 11, Number 1 © to the author(s) 2009 DOI: 10.1007/s12575-009-9008-x URL: springerprotocols.com; springerlink.com.
 9. Marina Macchiagodena, Marco Pagliai, Piero Procacci, Identification of potential binders of the main protease 3CLpro of the COVID-19 via structure-based ligand design and molecular modeling, Chemical Physics Letters, 2020; 750: 137489. doi.org/10.1016/j.cplett.2020.137489.
 10. Alexandra C. Walls, Young-Jun Park, M. Alejandra Tortorici, Abigail Wall, Andrew T. McGuire, David Veasler, Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein, Cell, April 16, 2020; 180: 281–292, Elsevier Inc., <https://doi.org/10.1016/j.cell.2020.02.058>.
 11. Daniel Wrapp, Nianshuang Wang, Kizzmekia S. Corbett, Jory A. Goldsmith, Ching-Lin Hsieh, Olubukola Abiona, Barney S. Graham, Jason S. McLellan, Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation, Science, 2020; 367: 1260–1263.
 12. Vladimir A. Belyi and M. Muthukumar, Electrostatic origin of the genome packing in viruses, PNAS, November 14, 2006; 103(46): 17174–17178, doi.10.1073/pnas.0608311103.
 13. Cascella M, Rajnik M, Cuomo A, Scott C, Napoli Di, Features, evaluation and treatment coronavirus (COVID-19), NCBI Bookshelf, StatPearls Publishing, 2020.
 14. Majid Monajjemi, Fatemeh Mollaamin, Shahrokh Shojaei, An overview on Coronaviruses family from past to Covid-19: introduce some inhibitors as antiviruses from Gillan's plants, Biointerface Research in Applied Chemistry, 2020; 10(3): 5575–5585 ISSN 2069-5837.
 15. Fang Li, Structure, function, and evolution of coronavirus spike proteins, Annu. Rev. Virol, 2016; 3: 237–61.
 16. Berend Jan Bosch, Ruurd van der Zee, Cornelis A. M. de Haan, and Peter J. M. Rottier, The coronavirus spike protein is a class I virus fusion protein: structural and functional characterization of the fusion core complex, Journal of Virology, 16 Aug. 2003; 77: 8801–8811, doi: 10.1128/JVI.77.16.8801–8811.2003.
 17. Christopher Forrey and M. Muthukumar, Electrostatics of capsid-induced viral RNA organization, The Journal of Chemical Physics, 2009; 131: 105101.
 18. Mushtaq Hussain, Nusrat Jabeen, Fozia Raza, Sanya Shabbir, Ayesha A. Baig, Anusha Amanullah, Basma Aziz, Structural variations in human ACE2 may influence its binding with SARS-CoV-2 spike protein, J Med Virol, 2020; 1–7, DOI: 10.1002/jmv.25832.
 19. Yanning Lu, Tuan Ling Neo, Ding Xiang Liu, James P. Tam, Importance of SARS-CoV spike protein Trp-rich region in viral infectivity, Biochemical and Biophysical Research Communications, 2008; 371: 356–360, doi:10.1016/j.bbrc.2008.04.044.
 20. Patrick C. Y. Woo, Yi Huang, Susanna K. P. Lau, and Kwok-Yung Yuen, Coronavirus genomics and bioinformatics analysis, Viruses, 2010; 2: 1804–1820; ISSN: 1999-4915 doi:10.3390/v2081803.
 21. Taylor Heald-Sargent, et al. Ready, Set, Fuse! The Coronavirus Spike Protein and Acquisition of Fusion Competence. Viruses, 2012; 4(4): 557–580.
 22. Hui DS, Azhar EI, Kim YJ, Memish ZA, Oh MD, Zumla A. Middle East respiratory syndrome coronavirus: risk factors and determinants of primary, household, and nosocomial transmission. Lancet Infect Dis 2018; 18(8): e217–27.
 23. Qianqian Yao, Paul S. Masters, and Rong Ye, Negatively charged residues in the endodomain are critical for specific assembly of spike protein into murine coronavirus, Virology. 2013 Jul 20; 442(1): 74–81, Published online 2013 Apr 28. doi: 10.1016/j.virol.2013.04.001
 24. Walls, A.C.; Tortorici, M.A.; Bosch, B.J.; Frenz, B.; Rottier, P.J.M.; DiMaio, F.; Fey, F.A.; Veasler, D. Cryo-electron microscopy structure of a coronavirus spike glycoprotein trimer. Nature, 2016; 531: 114–117, <http://dx.doi.org/10.1038/nature16988>.
 25. Kirchdoerfer, R.N.; Cottrell, C.A.; Wang, N.; Pallesen, J.; Yassine, H.M.; Turner, H.L.; Corbett, K.S.; Graham, B.S.; McLellan, J.S.; Ward, A.B. Pre-fusion structure of a human coronavirus spike protein. Nature, 2016; 531: 118–121, <http://dx.doi.org/10.1038/nature17200>.
 26. Z.-L. Shi, P. Zhou, X.-L. Yang, et al., Discovery of a novel coronavirus associated with the recent pneumonia outbreak in humans and its potential bat origin, bioRxiv, 2020; doi: 10.1101/2020.01.22.914952.
 27. F. Wu, S. Zhao, B. Yu, et al., A new coronavirus associated with human respiratory disease in China, Nature, 2020; 579: 265–269.
 28. Muhammad Tahir ul Qamar, Safar M. Alqahtani, Mubarak A. Alamri, Ling-Ling Chen, Structural basis of SARS-CoV-2 3CLpro and anti-COVID-19 drug discovery from medicinal plants, Journal of Pharmaceutical Analysis, 2020, <https://doi.org/10.1016/j.jpha.2020.03.009>
 29. N. Zhu et al., A Novel Coronavirus from Patients with Pneumonia in China, 2019, The New England Journal of Medicine, DOI: 10.1056/NEJMoa2001017, January 24, 2020.

30. Wallace Woon Fong Leung, Qiangqiang Sun, Electrostatic charged nanofiber filter for filtering airborne novel coronavirus (COVID-19) and nano-aerosols, Separation and Purification Technology, April 2020, doi: <https://doi.org/10.1016/j.seppur.2020.116886>.
31. Muhammad Adnan Shereen, Suliman Khan, Abeer Kazmi, Nadia Bashir, Rabeea Siddique, COVID-19 infection: Origin, transmission, and characteristics of human coronaviruses, Journal of Advanced Research, 2020; 24: 91–98.
32. Cui J, Li F, Shi Z-L, Origin and evolution of pathogenic coronaviruses, Nat Rev Microbiol, 2019; 17(3): 181–92.
33. Lai C-C, Shih T-P, Ko W-C, Tang H-J, Hsueh P-R. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and corona virus disease-2019 (COVID-19): the epidemic and the challenges, Int. J. Antimicrob Agents, 2020; 105.
34. S.Chatterjee, S.K.Biswas, A simple theoretical analysis of the effective electron mass in heavily doped III-V semiconductor in the presence of band-tails, Physica Scripta, 2003; 68: 368.
35. S.Chatterjee, S.K.Biswas, The simplified analysis of the Burstein-Moss shift in degenerate n-type semiconductors, Physica B, 2003.
36. S.Chatterjee, S.K.Biswas, S. Ghosal, On the magnetic susceptibilities in quantum wires of II-VI materials” J. Wqve Material Interaction, 2003.
37. S. Chatterjee, P. Kinget, A 0.5-V 1-Msps track-and-hold circuit with 60-dB SNDR, IEEE Journal of Solid State Circuits, Apr. 2007; 42(4): 722-729.
38. K.P. Pun, S. Chatterjee, P. Kinget, A 0.5-V 74-dB SNDR 25-kHz continuous-time delta-sigma modulator with a return-to-open DAC, IEEE Journal of Solid State Circuits, Mar. 2007; 42(3): 496-507.
39. S. Chatterjee, Y. Tsividis, P. Kinget, Ultra-low voltage analog integrated circuits, IEICE Transactions on Electronics, Vol. E89-C, Jun. 2006; 6: 673-680.
40. S. Chatterjee, Y. Tsividis, P. Kinget, 0.5-V analog circuit techniques and their application in OTA and filter design, IEEE Journal of Solid State Circuits, Dec. 2005; 40(12): 2373-2387.
41. P.K.Chattopadhyay, S.Chatterjee, Studies on novel dual filler based epoxidized natural rubber nanocomposite, Polymer composites 2009 DOI10.1002/pc.20866.
42. T.K.Chaki, S. Chatterjee, Thermal and mechanical properties of polymer nanocomposites based on ethylene methyl acrylate and multiwalled carbon nanotube, Polymar Composites 2009 DOI 10.1002/pc.20903.
43. T.K.Chaki, S.Chatterjee, Influence of acrylate content on the properties of ethylene methyl acrylate multiwalled carbon nano tube composites, Advanced Science letters, 2010; 3: 1-10.
44. S.Chatterjee, Studies on the filler microstructures in thermal and dynamic mechanical property development in ENR based ternary nanocomposites, Composites part A, 2009.
45. S.Chatterjee, D.K.Pahari, The study of Einstein relation in quantum wires of biomaterials: simplified theory and suggestions for further experimental determination, ETHENCE, 2015.
46. Sudip Chatterjee, Durjoy Bandyopadhyay, Kausik Rakshit, The Study of Einstein relation in quantumdots superlattices (QDSL) of nonparabolic semiconductors: simplified theory and suggestions for future experimental determination in biomaterials, INC, June 2020 communicated and accepted.
47. Schoeman, D., Fielding, B.C. Coronavirus envelope protein: current knowledge. Virol J, **16**, 69 (2019). <https://doi.org/10.1186/s12985-019-1182-0>,