



YEASTS COLONIZATION OF LOWER RESPIRATORY TRACT INFECTION IN A TERTIARY HOSPITAL IN BENIN CITY, NIGERIA

Omua Evelyn Oladugba^{1,2,*}, Helen Oroboghae Ogefere¹ and Edirin Omonigho Yusuf³

¹Department of Medical Laboratory Science, School of Basic Medical Sciences, College of Medical Sciences, University of Benin, Benin City, Nigeria.

²Department of Medical Laboratory Science, School of Health Technology, Benin City, Nigeria.

³Department of Medical Microbiology, University of Benin Teaching Hospital, Benin City, Nigeria.

***Corresponding Author: Omua Evelyn Oladugba**

Department of Medical Laboratory Science, School of Basic Medical Sciences, College of Medical Sciences, University of Benin, Benin City, Nigeria.

Article Received on 05/05/2022

Article Revised on 25/05/2022

Article Accepted on 15/06/2022

ABSTRACT

Yeast colonization of the lower respiratory tract is on the increase. This study aimed to determine prevalence of yeast lower respiratory tract colonization, speciate the recovered yeast isolates and determine their susceptibility profile. A total of 248 sputum specimens were collected from patients with signs and symptoms of lower respiratory tract infection attending the University of Benin Teaching Hospital, Benin City. The sputum specimens were processed to recover yeast isolates. The yeast isolates were identified and speciated using morphological, biochemical techniques as well as growth on CHROMagar™ candida. Susceptibility test was performed using agar diffusion method. The prevalence of yeasts colonization was 32.66%. Age and gender did not significantly affect the prevalence ($P = 0.765$) and ($P = 0.1478$) respectively. In – patients were significantly more at risk of yeast colonization (OR 9.127, 95% CI = 2.498, 33.350; $P = 0.0003$). The most predominant yeast species recovered was *Candida albicans* with a prevalence of 78.31%. *Saccharomyces cerevisiae* (2.41%) was recovered from both male and female patients. The polyenes were the most active antifungal agents, while the azoles were the least active. Routine surveillance for the colonization of yeast in lower respiratory tract and prudent use of antifungal agents is advocated.

KEYWORDS: Yeast, lower respiratory tract.

INTRODUCTION

Lower respiratory tract infections (LRTI) are infections that affects the trachea, bronchi and lungs (Latha *et al.*, 2006). They range from tuberculosis, whooping cough to pneumonia (Latha *et al.*, 2006). Most pneumonia infections are caused by bacteria and opportunistic fungi (Abdulla and Yehia, 2012). In time past, *Aspergillus* species and *Histoplasma* species were the leading causes of fungal pneumonia followed by *Cryptococcus neoformans* and rarely *Candida* species (Jha *et al.*, 2006). Currently, *Candida* species are the most common causes of opportunistic mycosis worldwide. *Candida* pneumonia is known to be the most challenging of all candida infections (Tamai *et al.*, 2012). *Candida* pneumonia have two classical cases, candida primary pneumonia which is restricted to the lungs and the secondary candida pneumonia that involves the movement of invasive *candida* to other areas of the body (Sol, 2002). The increase in the rate of immunocompromised persons in the last two decades and the unregulated use of antimicrobial agents has increase the number of opportunistic infections caused

by candida and other yeast species (Banesh and Kalyani, 2011). *Candida* species are normal commensal of human. They are capable of producing a wide variety of yeast infections. The ability to distinguish between colonization and infection can sometimes be challenging (Barenfanger *et al.*, 2003). *Candida* species are generally known to be the most common causes of opportunistic mycosis worldwide, is a major cause of most endogenous infections due to its commensal nature (Abdulla and Yehia, 2012). Although, candida pneumonia is rare, it is associated with high mortality rate, high morbidity and poor patient outcome (Liu *et al.*, 2003). There is lack of robust clinical studies evaluating treatments and severe underlying disease in the patients. (Romani, 2008). Identifying candida to species level from sputum specimens has become mandatory to aid the clinicians in the selection of antifungal agents in the treatment of both opportunistic and invasive fungal infections. This is because most of the non-albicans species usually exhibit reduced susceptibility to the common antifungal agents (Liu *et al.*, 2003; Elena *et al.*, 2006). Due to paucity of data in our environment, this study aimed to determine

the prevalence of yeasts colonization of the lower respiratory tract, speciation of the recovered yeast isolates and their susceptibility profiles.

MATERIALS AND METHODS

Study population: This study was carried out in the University of Benin Teaching Hospital Benin City, Nigeria. A total of two hundred and forty-eight (248) patients comprising of one hundred and twelve (112) males and one hundred and thirty-six (136) females with signs and symptoms of lower respiratory tract infection were recruited for this study. Informed consent was obtained from each patient prior to specimen collection. The Ethics and Research Committee of the University of Benin Teaching Hospital, Benin City, approved the protocol for the study.

Collection and Processing of specimens

Deep coughed out early morning sputum was collected into a sterile wide mouth container. A modification of the method described by Chessbrough, (2004) was used to process the sputum specimens. Briefly, the mucopurulent parts of the sputum were obtained with the aid of a sterile wire loop and inoculated onto sabourand dextrose agar containing 20 µg/ml of gentamicin and incubated at 37⁰ C for 24 – 48 hours.

Wet mount: For direct microscopic examination, the sputum specimens was placed on the glass slide and a drop of 10% (KOH) potassium hydroxide was added and the preparation was covered with a coverslip and examined under X10 and X40 objective lens for yeast cells.

Gram stain: Smears were made from the sputum specimens on a clean grease free slide and then heat fixed by just passing the glass slide over the flame. The smears were stained by Grams staining method and then examined under X100 objective lens for the presence of Gram-positive oval yeast cells.

Identification and Speciation of yeast isolates

The yeast isolates were further identified by Germ tube test, Chlamydiospore formation on corn meal agar, sugar

fermentation and assimilation tests as previously described by Amar *et al.*, (2013) as well as CHROMagarTM candida (CHROMagar Paris, France) (Paritpokee *et al.*, 2005).

Antifungal susceptibility testing

Disc antifungal testing was performed on the yeast isolates using agar diffusion method as described by the Clinical Laboratory Standard Institute (CLSI) guidelines (CLSI, 2009). The antifungal agents used were fluconazole (25 µg), ketoconazole (10 µg), amphotericin B (25 µg) and nystatin (100 units).

Statistical analysis

The data obtained were analysed with chi square (X^2) and odd ratio analysis test using the statistical software INSTAT^(R) (Graph pad software Inc. La Jolla Ca, USA).

RESULTS

The prevalence of yeast colonization in the lower respiratory tract in this study was 32.66% with the male and female patients having a prevalence of 33.82% and 31.25% respectively. The prevalence of yeast colonization of the lower respiratory tract between male and female did not differ significantly ($P = 0.765$) as well as in relation to age $P = (0.1478)$. In- patients were more infected than out - patients with a significant difference ($P = 0.0003$). Among the sputum specimens, 2 (1.79%) yielded mixed growth of *Candida albicans* and *Candida lusitanea* respectively from male patients (Table1). *Candida albicans* was the most predominant species recovered generally from both male and female patients with a prevalence of 65(78.31%) *Candida parapsiplosis* and *Candida krusei* were the least recovered isolates with a prevalence of 1.2% each (Table 2). The antifungal susceptibility profile showed that the polyenes (amphotericin B and nystatin) were the most active while the azoles (fluconazole and ketoconazole) were the least active against the non-*Candida albicans* (Table 3).

Table 1: Prevalence of yeasts infection in relation to Gender, Age and source of patients.

Characteristics	No. tested	No. infected (%)	OR	95% CI	P value
Gender					
Male	112	35(31.25)	0.889	0.521, 1.518	0.7687
Female	136	46(33.82)			
Mixed infections					
Male	112	2(1.79)	6.176	0.293,130.09	0.3946
Female	136	0(0.00)			
Age (years)					
≤ 1 – 10	14	6(42.86)			0.1478
11 – 20	18	8(44.40)			
21 – 30	35	15(42.86)			
31 – 40	45	19(42.20)			
41 – 50	29	7(24.14)			
≥ 51	107	28(26.17)			
Source of patients					

In – patients	15	12(80.00)	9.127	2,498,33.350	0.0003
Out – patients	233	71(30.47)			

OR = Odd ratio; CI = confidence interval

Table 2: Distribution of yeast isolates recovered from patients.

Yeast isolates	Male n (%)	Female n (%)	Total n (%)
<i>Candida albicans</i>	25(73.53)	40(81.63)	65(78.31)
<i>Candia parapsilosis</i>	1(2.94)	0(0.00)	1(1.20)
<i>Candida tropicalis</i>	5(14.71)	5(10.20)	10(12.04)
<i>Candida lusitanae</i>	2(5.88)	1(2.04)	3(3.61)
<i>Candida krusei</i>	0(0)	1(2.04)	1(1.20)
<i>Saccharomyces cerevisiae</i>	1(2.94)	1(2.04)	2(2.41)
Total	34(40.96)	49(59.04)	83(100)

Table 3: Susceptibility profiles of the yeast isolates.

Yeast isolates	FCN n (%)	KCN n (%)	AMP n (%)	NYS n (%)
<i>Candida albicans</i> (n=66)	28(42.42)	33(50.00)	61(92.42)	63(95.45)
<i>Candida krusei</i> (n=1)	0(0.00)	0(0.00)	1(100.00)	1(100.00)
<i>Candida lusitanae</i> (n=3)	0(0.00)	1(33.33)	3(100.00)	3(100.00)
<i>Candia parapsilosis</i> (n=1)	(0.00)	(0.00)	1(100.00)	1(100.00)
<i>Candida tropicalis</i> (n=10)	(0.00)	(0.00)	10(100.00)	10(100.00)
<i>Saccharomyces cerevisiae</i> (n=2)	1(100.00)	1(100.00)	1(100.00)	1(100.00)

Key

FCN = Fluconazole

KCN = Ketoconazole

AMP = Amphoterin B

NYS = Nystatin

DISCUSSION

The prevalence and identification of candida in lower respiratory tract infection is rare (Romani, 2008). Due to the increase in the number of immunocompromised persons, unregulated use of antimicrobial agents, increasing population of terminally ill persons and debilitated patients, there has been phenomenal rise in the occurrence of fungal lung infection over the last two decades (Chen *et al.*, 2001). However, there is paucity of data on candida lower respiratory tract infection in our environment. Against this background this study was conducted.

The prevalence of yeast infection in this study was 32.66%. This is higher than the 14.7% and 12.1% reported by Ogba *et al.*, (2013) and Jha *et al.*, (2006) respectively. The high prevalence reported in this study could be attributed to the fact that the yeast isolates may only represent colonization and not an infection. The criteria for the diagnosis of pulmonary candidiasis are still controversial, since candida is known to be a commensal of the lower respiratory tract, the definite diagnosis of pulmonary candidiasis still rest on histological demonstration of yeast in lung tissue with associated inflammatory changes (Diaz-fuentes *et al.*, 2007). The difference may be due to geographical location. Among children with diarrhoea, prevalence of infection varies with geographical location, regions within the same countries and even overtime in the same

location and population (Petri *et al.*, 2008). The study of Jha *et al.*, (2006) was carried out in India while that of Ogba *et al.*, (2013) was carried out in Calabar, Nigeria among HIV/AIDS patients. Ours was carried out in Benin City, Nigeria among patients whose HIV/AIDS status were not known.

Mixed infections were observed in two of the sputum specimens 1.79% each. It is believed that most people usually have a single strain of candida in different parts of the body for a long period of time. However, some individuals have more than one species at the same time (Klotz *et al.*, 2007).

The recovery of yeast isolates from sputum specimens was found to be highest among the age group of 11 – 20 years followed by the age groups of $\leq 1 - 10$ years and 21 – 30 years the least was among the age group of 41 – 5 years. This does not agree with Jha *et al.*, (2006) and Phino *et al.*, (2002) who found the highest isolates of *Candida* species among the age group of 70 – 80 years. The finding that yeasts colonization of the lower respiratory tract's colonization was significantly associated with in – patients (OR 9.127, 95% C.I. = 2.498, 33.350; P = 0.0003) agrees with the norm that hospitalized patients consume a lot of antibiotics which may increase the chances of opportunistic yeast infections (Panwar and Faujder, 2016).

The predominant yeast in this study isolate recovered in this study is *Candida albicans* (78.31%). The non-albicans species of candida recovered have been previously recovered from sputum specimens (St. German *et al.*, 2001). The prevalence of *Saccharomyces cerevisiae* recovered in this study was 2.41% from both male and female patients. The consideration of *Saccharomyces cerevisiae* as a health related or hospital acquired infection has been reported in literature (Perapoch, *et al.*, 2000; Salonen *et al.*, 2000; Lherm *et al.*, 2002; Cassone *et al.*, 2003 and Enache - Angoulvant and Hennequin, 2005).

Generally, the yeast isolates recovered in this study were more susceptible to polyenes (amphotericin B and nystatin) while the non – albicans species of candida were resistant to azoles (fluconazole and ketoconazole). This may be due to increased antifungal resistance of the non – albicans species of candida Liu *et al.*, 2003; and Elena *et al.*, 2006; Achkar and Fries, 2010).

Conclusively, the study reveals high prevalence of yeast colonization of the lower respiratory tract. In – patients were more at risk of yeasts colonization. *Candida albicans* was the most predominant yeast isolate recovered and the polyenes were the most active antifungal agents. Routine surveillance for the colonization of yeast in lower respiratory tract infections and prudent use of antifungal agent is advocated.

REFERENCES

1. Abdulla AZ, Yehia MM. Opportunistic fungi in lower respiratory tract infection among immunocompromised and immunocompetent patients. *Annals of the College of Medicine, Mosul*, 2012; 28, 38(1): 59-67.
2. Achkar JM, Fries BC. Candida infections of the genitourinary tract. *Clin Microbiol Rev*, 2010; 23(2): 253-73.
3. Amar CS, Ashisu J, Hajare V, Sreekantha YB. Study of prevalence and antifungal susceptibility of candida. *Int J Pharmaceut Biosci*, 2013; 4(2): 361–381.
4. Banesh LY, Kalyani M. Phenotypic characterization of Candida species and their antifungal susceptibility from a tertiary care centre. *J Pharmaceut Biomed Sci*, 2004; 11(2): 110-115.
5. Barenfanger J, Arakere P, Cruz RD, Imran A, Drake C, Lawhorn J, Verhulst SJ, Khardori N. Improved outcomes associated with limiting identification of Candida spp. in respiratory secretions. *J Clin Microbiol*, 2003; 41(12): 5645-9.
6. Cassone M, Serra P, Mondello F, Girolamo A, Scafetti S, Pistella E, Venditti M. Outbreak of *Saccharomyces cerevisiae* subtype *boulardii* fungemia in patients neighboring those treated with a probiotic preparation of the organism. *Journal of clinical microbiology*, 2003; 41(11): 5340-3.
7. Chen WY, Tseng HI, Wu MT, Hung HC, Wu HT, Chen HL, Lu CC. Synergistic effect of multiple indoor allergen sources on atopic symptoms in primary school children. *Environmental research*, 2003; 93(1): 1-8.
8. Chessbrough M. *District Laboratory Practice in Tropical Countries. Part 2 (Low price edition)* Cambridge University Press, 2004; 479.
9. Clinical Laboratory Standards Institute. *Method of antifungal disc susceptibility testing of yeast; Approved guidelines – second edition. C.L.S.I. document M 44 – A2 (ISBNI – 56238 – 703 – 0).* Clinical Laboratory Standard Institute, Wayne: Pennsylvania, 2009.
10. Diaz-Fuentes G, Shin C, Sy E, Niazi M, Menon L. Pulmonary fungal involvement in HIV-positive patients in an inner-city hospital in New York. *Internet J Pulmonary Med*, 2007; 7: 1-6.
11. Enache-Angoulvant A, Hennequin C. Invasive *Saccharomyces* infection: a comprehensive review. *Clinical Infectious Diseases*, 2005; 41(11): 1559-68.
12. Jha BJ, Dey S, Tamang MD, Joshy ME, Shivananda PG, Brahmadata KN. Characterization of Candida species isolated from cases of lower respiratory tract infection. *Kathmandu University medical journal (KUMJ)*, 2006; 4(3): 290-4.
13. Klotz SA, Chasin BS, Powell B, Gaur NK, Lipke PN. Polymicrobial bloodstream infections involving Candida species: analysis of patients and review of the literature. *Diagnostic microbiology and infectious disease*, 2007; 59(4): 401-6.
14. Latha R, Sasikala R, Muruganandam N, Babu RV. Study on the shifting patterns of Non-Candida albicans Candida in lower respiratory tract infections and evaluation of the CHROMagar in identification of the Candida species. *J Microbiol Biotech Res*, 2011; 1(3): 113-9.
15. Lherm T, Monet C, Nougère B, Soulier M, Larbi D, Le Gall C, Caen D, Malbrunot C. Seven cases of fungemia with *Saccharomyces boulardii* in critically ill patients. *Intensive care medicine*, 2002; 28(6): 797-801.
16. Liu XY, Sheng RY, Li XL, Li TS, Wang AX. Nosocomial fungal infections analysis of 149 case. *zhonghua Yi, Xue Za, Zhi*, 2003; 83(5): 399 – 402.
17. Ogba OM, Abia-Bassey LN, Epoke J, Mandor BI, Iwatt GD. Characterization of Candida species isolated from cases of lower respiratory tract infection among HIV/AIDS patients in Calabar, Nigeria. *World J AIDS*, 2013; 3(03): 201.
18. Panwar S, Faujdar SS. Prevalence, distribution, risk factors and antifungal susceptibility profiles of Candida species in a tertiary care hospital. *Int J Curr Microbiol Appl Sci*, 2016; 5: 78-94.
19. Paritpokee SI, Hall GE, Procop GA. Rapid identification of yeast isolates using BD BBL™ CHROMagar™ Candida. In 105th General Meeting of the American Society for Microbiology, 2005; 1-2.
20. Perapoch J, Planes AM, Querol A, Lopez V, Martinez-Bendayan I, Tormo R, Fernández F, Peguero G, Salcedo S. Fungemia with

Saccharomyces cerevisiae in two newborns, only one of whom had been treated with ultra-levura. *Europ J Clin Microbiol Infect Dis*, 2000; 19(6): 468-70.

21. Petri WA, Miller M, Binder HJ, Levine MM, Dillingham R, Guerrant RL. Enteric infections, diarrhea, and their impact on function and development. *The Journal of clinical investigation*, 2008; 118(4): 1277-90.
22. Tamai K, Tachikawa R, Tomii K, Imai Y. Fatal community-acquired primary *Candida* pneumonia in an alcoholic patient. *Int Med*, 2012; 51(22): 3159-61.
23. Romani L. Cell mediated immunity to fungi: a reassessment. *Sabouraudia*, 2008; 46(6): 515-29.
24. Salonen JH, Richardson MD, Gallacher K, Issakainen J, Helenius H, Lehtonen OP, Nikoskelainen J. Fungal colonization of haematological patients receiving cytotoxic chemotherapy: emergence of azole-resistant *Saccharomyces cerevisiae*. *Journal of Hospital Infection*, 2000; 45(4): 293-301.
25. Sol DR. *Candida* and virulence. The evolution of phenotypic plasticity. *Acta Tropical*, 2002; 81(2): 101 – 105.
26. St-Germain G, Laverdiere M, Pelletier R, Bourgault AM, Libman M, Lemieux C, Noel G. Prevalence and antifungal susceptibility of 442 *Candida* isolates from blood and other normally sterile sites: results of a 2-year (1996 to 1998) multicenter surveillance study in Quebec, Canada. *Journal of Clinical Microbiology*, 2001; 39(3): 949-53.