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EFFECT OF ETHANOLIC EXTRACT OF TINOSPORA CORDIFOLIA ON OXIDATIVE STRESS INDUCED BY CEREBRAL ISCHEMIA-REPERFUSION IN RATS

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ABSTRACT

Restoration of blood flow to ischemic brain is linked with generation of reactive oxygen species. In Ayurveda, the medicinal properties of Tinospora cordifolia have been attributed to its anti-stress, antioxidant, neuroprotective, adaptogeneic and nootropic properties. The present study investigates the effect of ethanolic extract of T. cordifolia on acute cerebral ischemia-reperfusion in rats. Acute cerebral ischemia-reperfusion (30 min occlusion of bilateral common carotid arteries followed by 45 min reperfusion) in Charles Foster (C.F.) strain rats was produced following standard technique. Effect of Tinospora cordifolia on lipid peroxidation, superoxide dismutase (SOD) activity, ascorbic acid, cyclic AMP level and total tissue sulfhydryl (T-SH) group in for brain region in acute cerebral ischemia-reperfusion were evaluated. T. cordifolia pre-treatment (100 mg/kg p.o. for 7 days) attenuated the reperfusion induced biochemical alterations. The results suggest protective role of T. cordifolia in cerebral ischemia reperfusion injury.

KEYWORDS: Ischemia, Reperfusion injury, Oxidative stress, Neuroprotection, Tinospor.

INTRODUCTION

A number of herbal drugs have been evaluated for their possible role in neurodegenerative disorders and cognitive functions. Tinospora cordifolia (TC) or Amrita or Giloya in Hindi and Guduchi in Sanskrit in Ayurveda (the classical Indian system of medicine), has been used for centuries, for a variety of diseases. [1] Tinospora cordifolia (Guduchi) is a large glabrous, deciduous, climbing shrub of Menispermaceae family found throughout tropical India. [2] It is classed as Medhya (Learning and memory enhancer).^[3] Neuroprotective and ameliorative properties are due to their antioxidant and trace element contents. [4] Tinospora cordifolia is known to be a rich source of trace elements (Zinc and Copper) which act as antioxidants and protects cells from the damaging effects of oxygen radicals generated during immune activation. Tinospora cordifolia has been claimed to possess learning and memory enhancing, [6] antioxidant, [7,8] and anti-stress activity. [9] Tinospora cordifolia enhanced the cognition in normal and cognition deficits animals. [10] Mechanism of cognitive enhancement is by immunostimulation and increasing the synthesis of acetylcholine, supplementation of choline enhances the cognition.^[11] Guduchi may also be attributed to its immunomodulatory properties.[12]

Chemical constituents' classes are alkaloids, diterpenoid lactones, glycosides, steroids, sesquiterpenoid, phenolics, aliphatic compounds, lignans and polysaccharides. [13] The polyphenols have the ability of penetrate the blood brain barrier and act as potential neuroprotective agent.

A majority of the present day disease are reported to be due to shift in the balance of pro-oxidant and antioxidant homeostatic phenomenon in the body. Pro-oxidant conditions dominate either due to the increased generation of free radicals caused by excessive oxidative stress or due to their poor scavenging in the body caused by gradual decline in antioxidant defence mechanism. [14] Oxidative free radicals play an important role in cerebral ischemia as well as reperfusion injury which is a distinct entity from the primary ischemia injury. This study was designed to assess the neuroprotective activity of ethanolic extract of T. cordifolia on acute cerebral ischemia-reperfusion.

MATERIALS AND METHODS

Drug and reagents

1, 1, 3, 3-Tetraethoxypropane (TEP), (Merck, Germany), Thiobarbituric acid (TBA), NADH, nitroblue tetrazolium (NBT) and phenazine methosulfate (PMS) (Sigma, USA) were used. All other chemicals and reagents were of the highest analytical grades available.

The raw material of test formulation was procured from Ayurvedic Garden; Banaras Hindu University campus in the month of January. It was authenticated in Department of Dravya Guna, Faculty of Ayurveda, IMS, BHU. The specimen copy has been kept in department for future references. Dried and powdered stem of T. Cordifolia was defatted with petroleum ether to remove lipids and fats, and then the residue was extracted with ethanol using soxhlet apparatus. The ethanolic extract was evaporated under reduced pressure at 40°C, using a rotary vapour evaporator. The extract was kept in 4°C for further analysis and experimental studies. The dose selection for the experimental studies was made according to our initial pilot experimental results.

Animals

After approval of Institutional Ethical Committee, the present study was conducted on inbred CF male albino rats weighing 250-300g, obtained from the central animal house of the Institute of Medical Sciences, Banaras Hindu University, Varanasi. They were kept in the departmental animal house in colony cages at an ambient temperature of 25±2°C and 45-55% relative humidity with 10:14 h light: dark cycles. They had free access to standard rodent pellet diet and drinking water. The food was withdrawn 18-24h before the surgical procedure, however, water was allowed ad libitum. Principles of laboratory animal care (NIH Publication No. 86-23, revised 1985) guidelines were followed throughout the experiments.

Experimental Procedure Surgical Procedure

Surgical technique for induction of cerebral ischemia by bilateral common carotid artery occlusion (BCCAO) was adapted from earlier published method of Iwasaki et al. [15] Rats were anaesthetized by ketamine (100 mg kg⁻¹, i.p). After a midline skin incision in the neck, both common carotid arteries were identified and isolated carefully from accompanying vagosympathetic nerve.

Acute ischemia-reperfusion injury was produced by blocking bilateral common carotid arteries (BCCA) for 30 min (lifting arteries with the help of thread) and reperfusion for 45 min was allowed by releasing the thread. Body temperature was maintained at about 37°C. This protocol was adopted on the basis of earlier reports from our laboratory and elsewhere. [16,17]

Study Design

The animals were divided into four groups of six animals each. First group served as sham-operated control (underwent all surgical procedure except BCCAO). In second group, ethanolic extract of T.cordifolia was administered to sham-operated animals to determine effect of drug per se. Third group of animals underwent 30 min BCCAO and 45 min reperfusion. In the fourth group (treatment) T.cordifolia 100 mg/kg/day, p.o. for 7 days, was administered before subjecting animals to ischemia-reperfusion.

Biochemical analysis

At the end of experiments animals were sacrificed by decapitation and frontoparietal part of cerebral cortex from both the hemispheres were separated. After rinsing with ice-cold normal saline the brain tissue were transferred to the appropriate homogenizing medium and analyzed for the biochemical parameters of the oxidant-antioxidant status i.e. thiobarbituric acid reactive substances (TBARS), superoxide dismutase (SOD) activity, tissue total sulfhydryl (T-SH) level, ascorbic acid and cyclic AMP. All the procedures on the brain samples were performed on ice or ice bath and sample were kept at -20°C. For all biochemical parameter studies, frontoparietal part of cerebral cortex of both the hemispheres was analysed.

Lipid peroxidation

Estimation of lipid peroxidation was done by measuring the lipid peroxidation product TBARS (Thio Barbituric Acid Reactive Substances) following the method of Ohkawa et al. [18] TEP was used as an external standard, and the level of lipid peroxidation was expressed as nanomoles TBARS mg⁻¹ of protein.

Superoxide dismutase (SOD)

SOD was estimated by adopting the procedure of Kakkar et al and results are expressed in milliunits mg⁻¹ of protein.^[19]

Total tissue sulfhydryl groups (T-SH)

Total T-SH in brain was measured according to the method of Sedlack and Lindsay. The level of T-SH was expressed as moles of SH 100⁻¹ g of wet tissue weight. [20]

Ascorbic acid

Ascorbic acid levels were determined by the method of Omaye et al and the results are expressed in terms of mg/100g wet weight. [21]

Estimation of brain total protein

The protein content of brain tissue was estimated using the method of Lowry et al. $^{[22]}$

Cyclic AMP estimation

Cyclic AMP estimation of frontoparietal part of forebrain was done by ELISA using EIATM cyclic AMP kit (Assay Designs Inc., USA). This kit uses a polyclonal antibody to cyclic AMP which binds, in a competitive manner with the cyclic AMP. Results were expressed as nmol of cyclic AMP per g (wet weight) of tissue.

Statistical Analysis

Statistical analysis was performed by applying one-way Analysis of Variance (ANOVA) followed by post hoc Tukey Test for biochemical parameters. A p-value of <0.05 was considered statistically significant.

RESULTS

Acute BCCAO for 30 min followed by 45 min reperfusion induced increase in lipid peroxidation

(TBARS), superoxide dismutase (SOD), activity and fall in T-SH levels. Ethanolic extract of T.cordifolia pretreatment attenuated enhanced TBARS level (p < 0.01) and SOD activity (p < 0.01) as well as prevented the consumption of T-SH significantly (p < 0.01) following cerebral ischemia reperfusion injury. T.cordifolia per se had no significant effect on any of these biochemical parameters (Table 1). Ischemia followed by reperfusion increased cyclic AMP level significantly as compared to

that in sham-operated animals (p < 0.05). T.cordifolia pre-treatment of ischemia reperfused animals led to a significant rise in cyclic AMP level compared to ischemia reperfusion group (p < 0.01) (Table 2). Ascoric acid levels, however, did not show any change after reperfusion injury and/or T.cordifolia pre-treatment. Thus total ascorbic acid levels appear unaffected during reperfusion injury (Table-1).

Table 1: Effect of T.cordifolia (100 mg/kg p.o. x 7 days) on biochemical parameters of oxidative stress in rat forebrain following cerebral ischemia-reperfusion injury (30 min BCCAO followed by 45 min reperfusion).

Groups	TBARS (nmol/mg protein)	SOD (MU/mg protein)	T-SH (x 10 ⁻⁵ M/mg protein)	Ascorbic Acid(mg/100g wet weight)
Sham-operated control	2.01±0.51	312.18±85.00	4.78±0.45	10.16±3.01
Per Se	1.95±0.27	338.01 ±79.66	3.91 ±0.40	9.33±1.95
Ischemia-reperfusion	3.95±0.67	646.80±115.56	2.46±0.20	7.80±2.13
T.cordifolia	2.67 ± 0.54	405.52±80.19	3.42±0.37	7.20±2.46

All data is expressed as mean \pm SD, n=6 in each group. Sham-operated control and treatment groups are compared with ischemia-reperfusion group. T.cordifolia per se is compared with sham-operated control group. Superscript indicates p-value <0.01. Statistical analysis was done by one-way ANOVA followed by Tukey test.

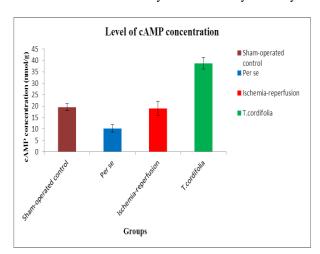


Figure 1: Effect of T.cordifolia (100 mg/kg p.o. x 7 days) on level of cyclic AMP in frontoparietal region of rat brain following cerebral ischemia-reperfusion injury (30 min BCCAO followed by 45 min reperfusion). Each bar represents Mean \pm SEM of six rats in each group.* and** indicates p-value <0.05 and < 0.01 respectively. Statistical analysis was done by one-way ANOVA followed by Tukey test.

DISCUSSION

The study confirms the previous reports that cerebral post-ischemic reperfusion is associated with generation of free radicals. The analysis of biochemical parameters show that BCCAO for 30 min followed by 45 min reperfusion causes ischemia-reperfusion injury. Increased generation of free radicals initiates lipid peroxidation and this reflected as increased level of TBARS. Polymorphonuclear leukocytes are known to be involved in cerebral reperfusion injury. Leukocyte accumulation has been noted in brain after cerebral

ischemia. [26] These activated neutrophils are a source of free radicals, especially superoxide anion. [27] The increased SOD activity is an indication that brain's antioxidant machinery is activated in response to excessive generation of free radicals. [28] Enhanced SOD activity catalyzes the conversion of superoxide anion to hydrogen peroxide and molecular oxygen. Hydrogen peroxide, the product of this reaction, is more toxic than the oxygen derived free radicals and requires to be scavenged further by tissue thiols (glutathione redox pathway) and catalase. [29] A fall in GSH (a non-protein sulfhydryl) during cerebral reperfusion injury is well reported and reduced level of T-SH reflects consumption of tissue thiols.^[30] Sulfhydryl compounds are among the most important endogenous antioxidants. They have role in maintenance of cellular proteins and lipids in their functional states. When these are consumed, the toxic effects of oxidative insult are exacerbated resulting in increased membrane and cell damage. [31]

The data reveals that T.cordifolia could antagonize ischemia-reperfusion injury induces rise in TBARS level. Similarly, T.cordifolia reverses ischemia reperfusion induced change in SOD and T-SH. These findings are in agreement with earlier reported antioxidant and of neuroprotective properties T.cordifolia. Neuroprotective and ameliorative properties are due to their antioxidant and trace element contents. [4] Tinospora cordifolia has been claimed to possess learning and memory enhancing, antioxidant, and anti-stress activity. [6,7,8,9] Reperfusion injury did not produce any significant change in ascorbic acid level. Possibly, reperfusion injury increases the ascorbate levels (reduced form of ascorbic acid) without altering the total ascorbic acid levels. This finding receives direct support from an earlier investigation that also suggests lack of change in

total ascorbic acid levels with a decrease in ascorbate levels secondary to cerebral reperfusion injury. [32]

The study also revealed significant increase in cyclic AMP level in brain (frontoparietal region). Following ischemia-reperfusion injury, cyclic AMP is known to increase in striatum, neocortex and hippocampus and in cerebral cortex. The increase in cyclic AMP levels following such injury has been implicated in reversing stroke induced vasospasm in central vessles. Increased level of cycle AMP is known to inhibit release of excitatory amino acid like glutamate through modulation of adenosine. The concentration in ischemia-reperfused animals. It is rather alluring to hypothesize that part of the therapeutically beneficial effect of T.cordifolia might be due to its effect on camp.

Several studies have identified and isolated the active principle of T.cordifolia in which lignans, such as 3(a, 4-dihydroxy-3-methoxybenzyl)-4-(4-hydroxy-3-methoxybenzyl) and alkaloids (jatrorrhizine, palmatine, beberine, tembeterine39, choline40), has gained attention, as it represents the major constituents responsible for modulation in neurotransmitter levels in the brain of rodents. [38] Free radical scavenging effect of T.cordifolia ethanolic extract could be related to an active constituent.

The data of present studies thus suggested protective role of ethanolic extract of T.cordifolia in cerebral ischemia reperfusion induced injury in rats. The present finding, apart from supporting previously reported therapeutic effects of T. cordifolia in various animal models of oxidative stress, is suggestive of its efficacy in the treatment of cerebral reperfusion like cerebrovascular insults.

CONCLUSION

Concluding the effects of T.cordifolia we demonstrated that T.cordifolia prevents the oxidative stress during the cerebral reperfusion injury. In conclusion, the present study provides experimental evidences for T.cordifolia as a neuroprotective drug and emphasizes the necessity to appreciate the neuropharmacological effects of T.cordifolia.

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REFERENCES

1. Czygan, F.C., Kulturgeschichte and Mystik des Johanniskrutes. Zeitschrift fur Phytotherapie, 1993; 14: 276-82.

- Sharma, P.C., Yelne, M.B., Dennis, T.J., Database on Medicinal plants used in Ayurveda and Sidha. New Delhi: CCRAS, Dept. of AYUSH, Ministry of Health and Family Welfare, Govt. Of India, 2005; 3: 256-81.
- 3. Acharya, Y.T., Agnivesha. Caraka Samhita with Chakrapani's Ayurveda Deepika Teeka., Varanasi: ChoukhambaSamskrita Samsthana, 1994; 385.
- Avinash, K.R., Manohar, G.M., Saibal, K.B., Rubia cordifolia, Fagonia cretica Linn. And Tinospora cordifolia exert neuroprotection by modulating the antioxidant system in rat hippocampal slices subjected to oxygen glucose deprivation. BMC Complementary Alternative Medicine, 2004; 4: 11.
- 5. Chulet, R., Pradhan, P., A review on Rasayana. Pharmacognosy Reviews, 2009; 3: 229-234.
- 6. Agarwal, A., Malini, S., Bairy, K.L., Rao, M.S., Effect of Tinospora cordifolia on Learning and Memory in normal and memory deficit rats. Indian Journal of Pharmacology, 2002; 34: 339-49.
- Singh, R.P., Banergee, S., Kumar, P.V., Raveesha, K.A., Rao, A.R., Tinospora cordifolia induces enzymes of carcinogen/ drug metabolism and antioxidant System, and inhibits lipid peroxidation in mice. Phytomedicine, 2006; 13: 74-84.
- 8. Stanely, M., Prince, P., Menon, V.P., Antioxidant action of Tinospora cordifolia root extract in alloxan diabetic rats. Phytotherapy Research, 2001; 15: 213-218
- 9. Patil, M., Patki, P., Kamath, H.V., Patwardhan, B., Antistress activity of Tinospora cordifolia (Willd) Meirs. Indian drugs, 1997; 34: 211-215.
- Reddy, Y.Y., Mohana, L.S., Saravana K.A., Review on Effect of Natural Memory Enhancing Drugs On Dementia. International Journal of Phytopharmacology, 2010; 1: 1-7.
- 11. Asuthosh, A., Malini, S., Bairy, K.L., Muddanna, S.R., Effect of Tinospora cordifolia on learning and memory in normal and memory deficits rats. Indian Journal of Pharmacology, 2000; 34: 339-349.
- 12. Manjrekar, P.N., Jolly, C.I., Narayanan, S., Comparative studies of the immunomodulatory activity of Tinospora cordifolia and Tinospora sinensis. Fitoterapia, 2000; 71: 254-257.
- 13. Singh, S.S., Pandey, S.C., Srivastava, S., Gupta, V.S., Patro, B., Ghosh, A.C., Chemistry and Medicinal properties of Tinospora cordifolia (Guduchi). Indian Journal of Pharmacology, 2003; 35: 83-91.
- Schulz, J.B., Lindnau, J., Seyfriend, J., et al. Glutathione oxitative stress and neuro-degeneration. European Journal of Biochemistry, 2000; 167: 4904-4911.
- Iwasaki, Y., Ito, S., Suzuki, M., et al. Forebrain ischemia induced by temporary bilateral common carotid artery occlusion in normotensive rats. Journal of Neurological Sciences, 1989; 90: 155-165.
- 16. Yanpallewar, S.U., Hota, D., Rai, S., et al. Nimodipine attenuates biochemical, behavioural and

- hisotopathological alterations induced by acute transient and long-term bilateral common carotid occlusion in rats. Pharmacological Research, 2004; 49: 143-150.
- 17. Shukla, R., Tyagi, E., Kumar, R., Protective effect of COX and NOS inhibitors on Ips induced oxidative stress in rat. Annals of Neurosciences, 2008; 15(1): 6-10.
- Ohkawa, H., Ohishi, N., Yagi, K., Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Analytical Biochemistry, 1979; 95: 351-358.
- Kakkar, P., Das, B., Viswanathan, P.N., A modified spectrophotometric assay of superoxide dismutase. Indian Journal of Biochemistry and Biophysics, 1984; 21: 130-132.
- 20. Sedlack, J., Lindsay, R.H., Estimation of total, protein bound and non-protein bound sulfhydryl groups in tissue with Ellman's reagent. Analytical Biochemistry, 1968; 25: 192-205.
- 21. Omaye, S.T., Turnball, D.J., Samberlich, H.E., Selected methods for the determination of ascorbic acid in animal cells, tissues and fluids. Methods in Enzymology, 1979; 62: 3-11.
- 22. Lowry, O.H., Rosenborough, N.J., Farr, A.L., Randall, R.J., Protein measurement with folin phenol reagent. Journal biological Chemistry, 1951; 193: 265-275.
- 23. Nakashima, M., Niwa, M., Iwai, T., Uematsu, T., Involvement of free radicals in cerebral vascular reperfusion injury evaluated in a transient focal cerebral ischemia model of rats. Free Radical Biological Medicine, 1999; 26: 722 729.
- Sorrenti, V., Di Giaxomo, C., Renis, M., et al. Lipid peroxidation and survival in rats following cerebral post-ischemic reperfusion: effect of drugs with different molecular mechanisms. Drugs Experimental Clinical Research, 1994; 20: 185-189.
- 25. Halliwell, B., Reactive oxygen species in living system: source. Biochemistry and role in human disease. American Journal of Medicine, 1991; 99: 145-225.
- Barone, F.C., Hillegans, L.M., Price, W.J., Polymorphonuclear leukocytes in cerebral focal ischemic tissue: myeloperoxidase activity assay and histologic verification. Journal of Neurosciences Research, 1991; 29: 331–345.
- 27. Gringo, J.M., Reperfusion injury. Transplantation Proceedings, 1997; 29: 59-61.
- Bannister, J.V., Bannister, W.H., Rotillio, G., Aspects of structure, function and application of superoxide dismutase. CRC. Critical Reviews in Biochemistry, 1987; 22: 111-180.
- Fridovich, I., Superoxide radicals and superoxide dismutase. Annual Review in Biochemistry, 1995; 44: 147–59
- Panigrahi, M., Sadguna, Y., Bangalere. R., Alphalipoic acid protects against reperfusion injury following cerebral ischemia in rats. Brain Research, 1996; 217: 184-88.

- 31. Thomas, J.A., Poland, B., Honzatko, R., Protein Sulfhydryls and their role in antioxidant function of protein-S-thiolation. Archives of Biochemistry and Biophysics, 1995; 319: 1-9.
- 32. Cooper, O.J.L., Pulsinelli, W.O., Duffy, T.E., Glutathione and ascorbate during ischemia and postischemic reperfusion in rat brain. Journal of Neurochemistry, 1980; 35: 1242-1245.
- 33. Prado, R., Busto, R., Mordecai, Y.T., et al. Ischemia induced changes in extracellular levels of striatal cyclic AMP: Role of dopamine neurotransmitter. Journal of Neurochemistry, 1992; 59: 1581-1584.
- 34. Blomquist, P., Lindvall, O., Stenevi, U., et al. Cyclic AMP concentrations in rat neocortex and hippocampus during and following incomplete ischemia: Effects of central noradrenergic neurons, prostaglandins, and adenosine. Journal of Nuerochemistry, 1985; 44: 1345-1353.
- 35. Kobayashi, M., Lust, W.D., Passonneau, J.V., Concentrations of energy metabolites and nucleotides during and after bilateral ischemia in the gerbil cerebral cortex. Journal of Neurochemistry, 1977; 29: 53-59.
- 36. Palmer, G.C., Cyclic nucleotides in stroke and related cerebrovascular disorders. Life Sciences, 1985; 36: 1995-2006.
- 37. Phillis, J.W., Smith-Barbour, M., Oreagam, M.H., et al. Amino acid and purine release in rat brain following temporary middle cerebral artery occlusion. Neurochemical Research, 1994; 19: 1125-1130.
- 38. Serdarevic, N., Eckert, G.P., Nuller, W.E., The effects of extracts from St. John's Wort and Kava kava on brain neurotransmitter levels in the mouse. Pharmacopsychiatry, 2001; 34: 134-136.