



**CHEMICAL AND POTENTIAL BIOLOGICAL PERSPECTIVES OF GENUS *SENECIO*
[ASTERACEAE]**

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Article Received on 05/09/2017

Article Revised on 26/09/2017

Article Accepted on 17/10/2017

ABSTRACT

Ethnopharmacological Relevance: The plant of the genus *Senecio* [Astereaceae] is widely distributed ,and have long been used in folk medicine for the treatment of various ailments particularly treatment of dysentery, conjunctivitis, infections, rheumatism, cancer, cough suppressant, asthma, bronchitis, eczema inflammation, and muscular pain. **Aim of the Review:** In present review we emphasized the recent progress in the chemistry and biologyog this genus as well as its traditional uses.This database may provide guidance for researchers, chemists and herbologists for further investigation in the field. **Materials and Methods:** All literature available on the genus *Senecio* was collected via electronic search [using Scifinder, Google scholar, Scirus and Web of science], books, thesis and journals. **Results:** Ethnobotanical uses of different species of genus *Senecio* have been reported from China, Pakistan, India, Malasiya,Australia and Nepal for their different types of ailments,Genus *Senecio* possesses different chemical constituents including alkaloids,glycosides, sugar, sesquiterpenoids, monoterpenoids, diterpenoids, triterpenoids, flavonoids,coumarins, and some other compounds.Crude extract, fraction, and isolated secondary metabolities of genus *Senecio* have shown a wide range of pharmacological activities including antibacterial, antifungal, antifeedant,antioxidant and cholinesterase inhibiting activities. **Conclusion:** The leaves and roots of *Senecio* plants have been used for the treatment of dysentery, conjunctivitis, infections, rheumatism, fever in the folk medicine system for years without any adverse effect.However there is a need to search for individual secondary metabolities responsible for these actions and study their mode of actions, and physiological pathways in sufficient details.

KEYWORDS: The genus *Senecio*, which belongs to the tribe Senecioneae.

1. INTRODUCTION

The genus *Senecio*, which belongs to the tribe Senecioneae, is the largest and most complex genus in the family of the Asteraceae [Compositae] and includes more than 1500 species with a worldwide distribution [B. Nordenstam et al 1977] that have been extensively investigated for their secondary metabolities. pyrrolizidine alkaloids [Pas], eremophilolides and cacularides are particulary characteristic for species of this genus [E.Burgueno et al 2004].

The plants of this genus have been studied extensively because of the traditional medicinal uses associated with them. The leaves, stems and flowers are used mostly in folk medicine for the treatment of various ailments [G. B. Hammond et al 1998 E. Uzun, et al 2004]. These plants are the major sources of bioactive pyrrolizidine alkaloids [Pas] and the furanoeremophilane sesquiterpenoids and these are the phytoconstituents which are responsible for almost all of pharmacological activities of this genus [F. Bohlmann, et al 1986,W. H. Heywood et al 1977, D. J. Robins et al 1982]. The chemical constituents of the genus *Senecio* include

notably sesquiterpenoids, monoterpenoids [F. Bohlmann, et al 1985,S. Dupre et al 1991], diterpenoids [C. Dong-Liang et al 1992], triterpenoids [P. Torres et al 1998], essential oils [Y. Zhao et al 2011], pyrrolizidine alkaloids, phenolic and flavonoid compounds [B. E. Juarez et al 1995, P. Torres et al 1997,E. M. Suleimenov et al 2000, H. Zhong-Mei et al 2010,T. Dao-peng et al 2010]. Most of the aquatic plants of this genus are known for containing furanoeremophilanes [A. B. Pomilio et al 1997, E. A. Jares et al 1990], steroids [M. C. Tettamanzi et al 1992] and the pyrrolizidine alkaloids senecionine and platiphylline [J. A. Paiva, et al 2004,C. M. Silva, et al 2006] with toxic properties.Chemically investigated parts of the *Senecio* species include roots, leaves, stems, flowers, aerial parts, and whole plants. The PAs and furanoeremophilanes are the most important constituents of this genus. Somecompounds isolated from the genus *Senecio* and even crude extracts are known antimicrobial activity, including antibacterial, antifungal, and antitubercular activities.

2. Traditional Uses

The plants of the genus *Senecio* are used traditionally for medicinal purposes that include treatment of dysentery, conjunctivitis, infections, rheumatism, cancer, cough suppressant, asthma, bronchitis, eczema and inflammation. *Senecioaryunensis* is used in traditional Chinese medicine in northwestern China to treat dysentery, conjunctivitis and tumefaction [The Encyclopedia et al 1977].

The aerial parts of *Senecio canescens* are used for the treatment of infections the leaves are especially used topically for treatment of rheumatism [A. White, et al 1976]. *Senecio integrifolius* is used in traditional Chinese medicine and has been investigated for its cytotoxic effect [The Encyclopedia et al 1977]. *Senecio graveolensis* characterized by a typical smell and is used as a popular medicine to counteract mountain sickness and as an emenagogue, digestive and cough suppressant [J. Bautista et al 1991]. South American traditional system of medicine reported the use of *Senecio* species as a remedy for altitude sickness ['soroche'] and to relieve stomach pain [S. Abdo et al 1992, L. A. Loyola et al 1985].

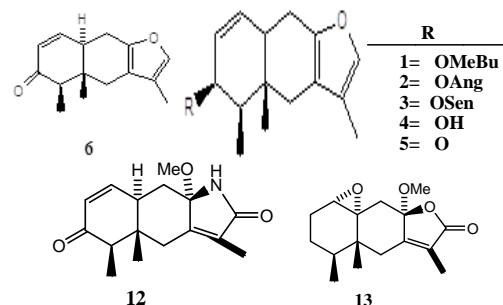
In traditional medicine, the use of *Senecio* species for treatment of asthma, coughs, bronchitis, eczema and wound healing have also been reported [E. Burgueno et al 2004, G. B. Hammond et al 1998, E. Uzun, et al 2004]. This species is used in traditional medicine for the treatment of skin, respiratory and osteoarticular diseases [A. A. Bolzan et al 2007]. *Senecio tenuifolius* Burm.f. is poisonous to livestock, but the leaves of the plant are used topically as remedy for skin diseases to reduce swelling and pain [C. K. Parikh et al 1999, D. S. Bhakuni et al 1982].

3. Chemical constituents

The chemical constituents of the genus *Senecio* include alkaloids, sesquiterpenoids, monoterpenoids, diterpenoids, triterpenoids, flavonoids, coumarins, and some other compounds. The chemicals reported from some of the plants of this genus are as follows.

3.1. *Senecio flavus*

3 β -Methylbutyryloxyeuryopsin,^[1] 3 β -angeloyloxyeuryopsin,^[2] and 3 β -senecioyloxyeuryopsin,^[3] 3 β -Hydroxyeuryopsin,^[4] Euryopsin-3-one,^[5] Furanoligularenone,^[6] 3-Oxo-8 α ,12 α -dimethoxy-8,12-dihydro-10 α H-furanoeremophil-1-ene,^[7] 3-Oxo-8 α -methoxy-10 α H-eremophila-1,7[11]-dien-12,8 β -olide^[8] 3-Oxo-8 α -hydroxy-10 α H-eremophila-1,7[11]-dien-12,8 β -olide,^[10] 3-Oxoeremophila-1,7[11]-dien-12,8 β -olide,^[11] 3-Oxo-8 α -methoxy-10 α H-eremophila-1,7[11]-dien-12,8 β -lactam,^[12] 1 β ,10 β -Epoxy-8 α -methoxyeremophil-7[11]-en-12,8 β -olide^[13] [P. Torresa et al 1999].

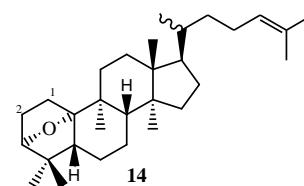


3.2. *Senecio ruwenzoriensis*

Bisline, Isoline, Ruwenine, Ruzorine, Isolinecic acid, Pyrrolizidine alkaloids [E. M. Suleimenov et al 2000].

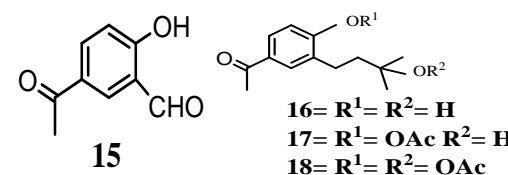
3.3. *Senecio selloi*

[20R]-3 α ,10 α -Epoxy-9-epi-cucurbita-24-ene,^[14] [20 S]-3 α ,10 α -Epoxy-9-epi-cucurbita-24-ene[2][G. R. Ckera et al 1999].



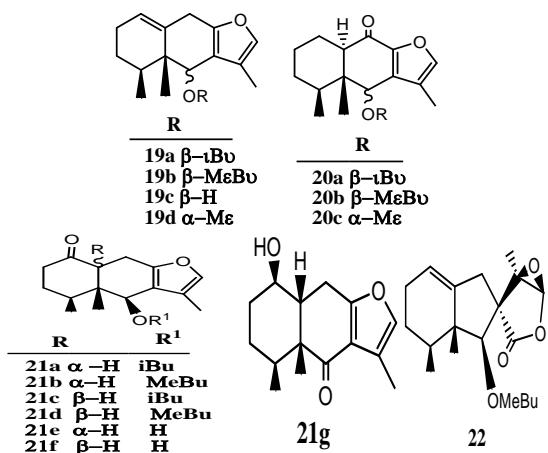
3.4. *Senecio graveolens*

5-Acetylsahcyaldehyde,^[15] 4-Hydroxy-3-[hydroxyisopentyl] acetophenone,^[16] 4-Acroxy-3-[3-hydroxyisopentyl] aetophenone,^[17] 4-Acetoxy-3-[3'-acetoxytisopentyl]acetophenone^[18] [L. A. Loyola et al 1985], Dihydroeuparin [J. I. D. Graw et al 1962], 4-hydroxy-3-[isopenten-2-yl] acetophenone [F. Bohlmann et al 1970], 3-hydroxy-2,2-dimethyl-6-acetylchromane[S. Valverde et al 1972].



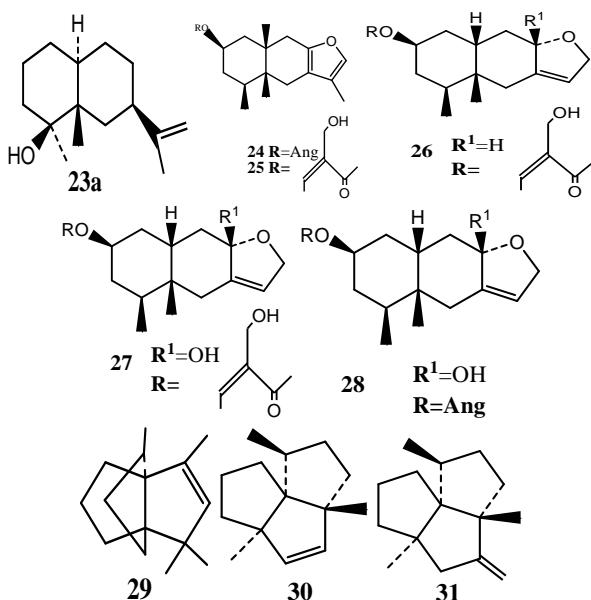
3.5. *Senecio auricula* var. major

6 α -Methoxyeuryopsin,^[19d] 6 β -[2-Methylbutyryloxy]-furanoeremophil-1-one,^[21b] 6 β -Isobutyryloxyenberinone,^[21c] 6 β -[2-Methylbutyryloxy]-senberginone,^[21d] 6 β -Hydroxy-1O α H-furanoeremophil-1-one,^[21e] 6 β -Hydroxy-1O β H-furanoeremophil-1-one,^[21f] 1 β -Hydroxy-1O β H-furanoeremophil-6-one,^[21g] 11 β , 12 β -Epoxy-6 β -[2-methylbutyryloxy]-dihydrobakkenolide A[senauricolid]^[22] furanoeremophilanes,^[19a] [F. Bohlmann, et al 1986]^[19b] [F. Bohlmann et al 1977],^[20a] [J. Harmatha, et al 1969, Z. Samek et al 1969],^[20b] [A. H. Mericli, et al 1989],^[20c] [F. Bohlmann et al 1976] and,^[21a] alkaloids [F. M. Panizo et al 1974].



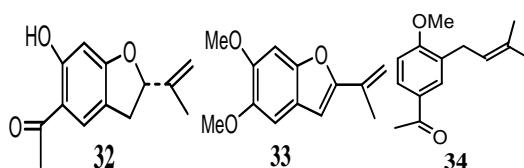
3.6. *Senecio alatus*

3.6. Scheclo aratus
 4-hydroxy-10x-H-eremophil-11[12]en,^[23a] 2 β -Angeloxyloxy-10 β -H furanoeremophilane,^[24] 2 β -[5-hydroxyanggeloxyloxy]-10 β -H-furanoeremophilane,^[25] 2 β -[5-hydroxyanggeloxyloxy]-8 β -H-eremophilanolide,^[26] 2 β -Angeloxyloxy-8 β -hydroxy-10 β -H-eremophilanolide,^[27] 2 β -Angeloxyloxy-8 β -hydroxy-10 β -H-eremophilanolide^[28] [F. Fohmann, et al 1980], eremophilane derivatives,^[29] [L. H. Zalkow et al 1978],^[30] [L. H. Zalkow et al 1977] and^[31] [F. Bohlmann et al 1979].



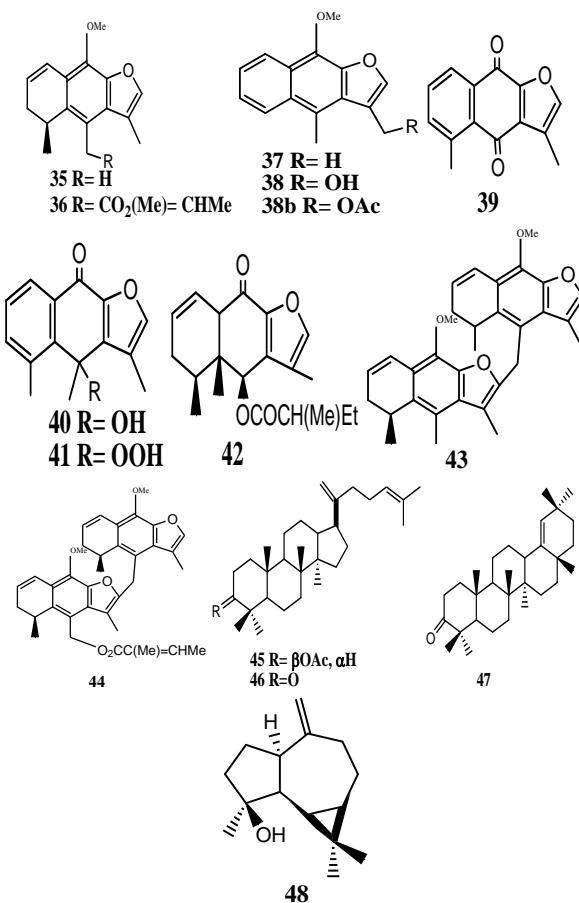
3.7. *Senecio amplexicaulis*

4-hydroxy-10x-H-eremophil-11[12]ene^[23a] afforded α - and β -farnesene [1 and 2], β -squiphellandrene, linalolacetate, p-hydroxyacetophenone derivatives^[32] [B. Kamthory et al 1939],^[33] [T. Murae et al 1968],^[34] [F. Bohlmann et al 1978a].



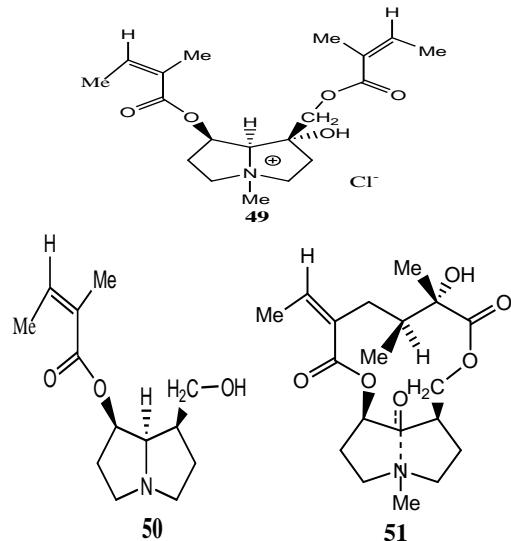
3.8. *Senecio canescens*

3.3 Schleimsäuren
 Cacalohastine^[35] [K. Hayashi et al 1973,F. Bohlmann et al 1978b], lupeol, α -amyrin, sitosterol,stigmasterol, 14-angeloyloxy cacalohastine^[36] [F. Bohlmann et al 1977], dehydrocacalohastine^[37] [K. Hayashi et al 1973,F. Bohlmann et al 1978b], 13-Hydroxy-3,4-dehydrocaelohastine^[38] [S. Abdo, et al 1992], 9,13-acetoxydehydrocacalohastine^[38b] [F. Bohlmann et al 1978b, F. Bohlmann et al 1978c], maturinone^[39] [P. M. Brown et al 1969,H. Kakisawa et al 1969], cacalonol^[40] [T. Takemoto et al 1975,], cacalonol hydroperoxide,^[41] 6 β -[2-methylbutanoyloxy]-9-oxo-1[10]-furanoeremophilene^[42] [F. Bohlmann et al 1984], dimeric sesquiterpenes^[43] [F. Bohlmann et al 1978b], 14-Angezoyloxy-12-[cacalohastin-14-yl]cacalohastine,^[44] dammaradienyl acetate^[45] [D. J. Roy et al 1981, J. D. P.Teresa,et al 1986], dammaradienone^[46] [J. D. P.Teresa, et al 1979, I. Wahlberg et al 1971], germanicione^[47] [A. Gonzales,et al 1981],spathulenol^[48] [T. D. Hubert et al 1985].



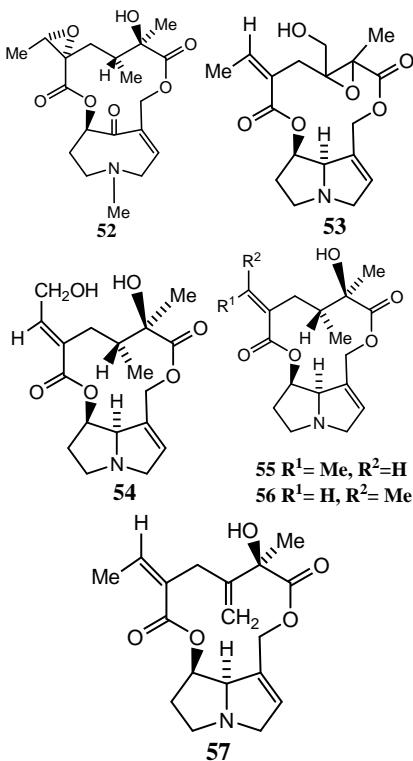
3.9. *Senecio integrifolius* var Fauriri

N-Methyl-O⁷.O⁹-diangeloyl-lα-hydroxyl-platyneciniumchloride,^[49] O⁷-Angeloylturneforcidine,^[50] 1,2-Dihydrosenkirkin,^[51] O⁷-Angeloylheliotridine-N-oxide [E. Roeder et al 1991],neosenkirkine [H. L. Zalkow et al 1988], otonecine alkloides[L. B. Bull, et al 1968,N. H. Amsterdam et al 1962,C.C.J. Culvenor, et al 1967].



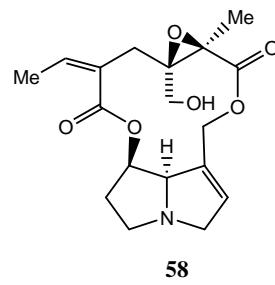
3.10. *Senecio argunensis*

Pyrrolizidine alkaloid-ocoscnine,^[52] erucifoline,^[53] 21-hydroxyintegerrimine^[54] [K. Liu et al 1991], senecionine^[55] [X. T et al 1979],integerrimine^[56] [G. G. Habermehl et al 1988], seneciphylline^[57] [E. Roder et al 1990].



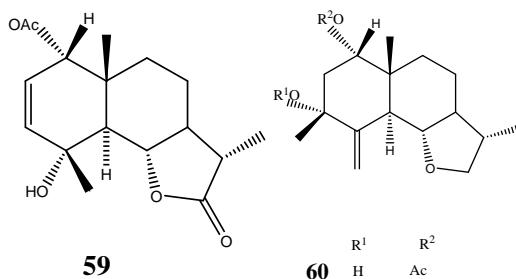
3.11. *Senecio persoonii*

Two toxic pyrrolizidine alkaloids- seneciphylline^[57] [E. Roeder et al 1992], erucifoline^[58] [K. Liu et al 1991],seneciphylline-N-oxide [E. Roeder et al 1993], erucifoline-N-oxide [H. J. Segall et al 1983].



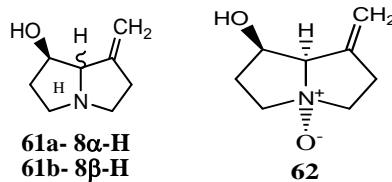
3.12. *Senecio chrysanthemoides*

Two lactones chrysanthemolide-lα-acetoxy-4α-hydroxy-5α,6β,7α,11β-H-eudesm-2-en-12, 6- olide^[59] and 1-acetylerivanin-lα-acetoxy-3α-hydroxy-5α,6β,7α,11β-H-eudesm-4[15]en-12,6-olide.^[60] [N. Mengi et al 1991], pyrrolizidine alkaloid seneciphylline, an acetylenic ester, a triterpene and derivatives of benzoquinone and euparin [K. L. Handa, et al 1957] have been reported.



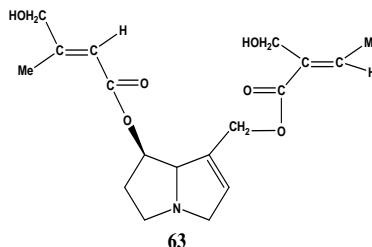
3.13. *Senecio schweinfurthii*

Predominant alkaloid are 7β-hydroxy-l-methylene-8α-pyrrolizidine,^[61a] 7β-hydroxy-l-methylene-8β-pyrrolizidine^[61b] and 7β-hydroxy-l-methylene-8α-pyrrolizidine N-oxide^[62] [M. H. Benn, et al 1995].



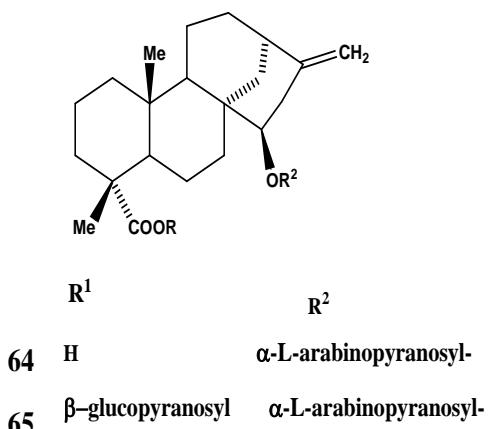
3.14. *Senecio doria*

Pyrrolizidine alkaloid doriasenine^[63] [E. Roder et al 1988].



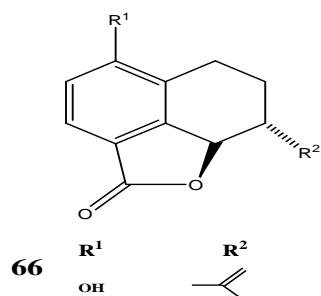
3.15. *Senecio rufus*

Two diterpene glycosides- rufusoside A[ent-15 α -[α -L-arabinopyranosyloxy]-kaur-16-en-19-oic acid],^[64] Rufiisoside B[ent-15 α -[α -L-arabinopyranosyloxy]-kaur-16-en-19-oic acid β -D-glucopyranosyl ester],^[65] a glycoside andglycone[D. L. Cheng,et al 1993].



3.16. *Senecio gilliesiano*

Norsesquiterpene lactone, isocoumarine retronecine, senecionine, platyphyllide, 1-hydroxy platyphyllide^[66] [F. H. Guidugli, et al 1986], pyrrolizidine alkaloids[M. J. Pestchankar et al 1985, Pestchankar, et al 1985, Pestchankar, et al 1986], Furanoeremophilanes sesquiterpene [M. S. Ascheri et al 1980, M. S. A. Salmeron et al 1983].

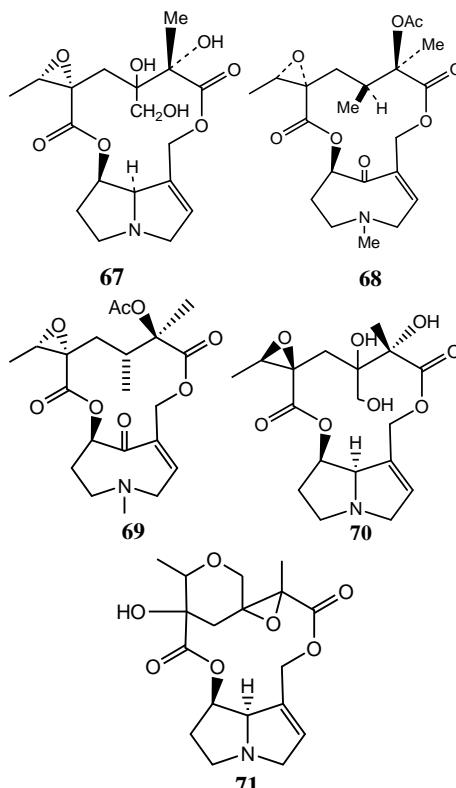


3.17. *Senecio gallicus*

Alkaloids namely ligularzine, senkirine and seneclonine-N-oxide [J. D. Urones et al 1988], flavonoids [R. M. A et al 1981] acetophenones and terpenoids [P. Teresa et al 1988].

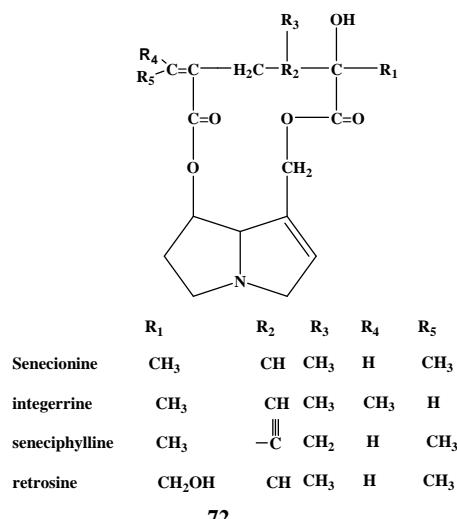
3.18. *Senecio adonisfolius*

Polyphenols [S. Ferry et al 1977], alkaloids [S. Ferry et al 1976], 12,13,19-trihydroxy-15,20-epoxy-15,20-dihydro [12S,15R,20R] senecionan-11,16-dione,^[67] 4,8-secosenecionane[J. D. Urones et al 1988], florosenine,^[69] 12,13,19-trihydroxy-15,20-epoxy-15,20-dihydro-[12S,1SR,20R]-senecionan-11,16-dione^[70] [E. Roeder, et al 1990b], adonifoline^[71] [L. Witte et al 1992].



3.19. *Senecio Longilobus*

Senecionine,^[72] integerrimine, seneciphylline [C. A. Ray, et al 1987], retrorsine [R. Adams et al 1949], Riddelliine [H. J. Segall et al 1978].



3.20. *Senecio glabellus*

Senecionine^[72] [R. Adams et al 1953], integerrimine^[72] [C. A. Ray, et al 1987].

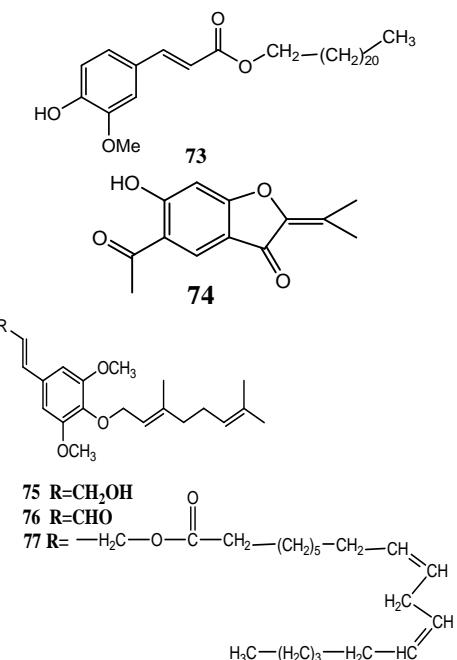
3.21. *Senecio platyphylloides*

Lactone [F. Bohlmann et al 1977].

3.22. *Senecio lumbifolius*

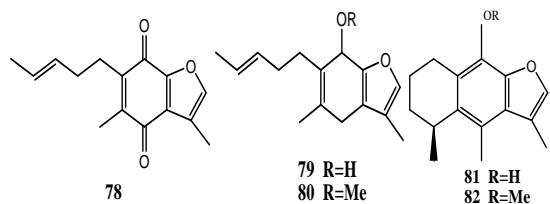
Ferulic acid, n-docosyl ester,^[73] 2-isopropylidene-3-oxo-5-acetal-6-hydroxy-benzodihydrofuran,^[74] 4-O-geranyl-

sinapyl alcohol,^[75] 4-O-geranyl-sinapyl aldehyde,^[76] linoleic acid 4-O-geranyl sinapyl ester^[77] [Y. Zhao et al 1994].



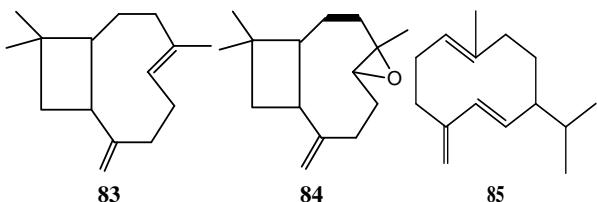
3.23. *Senecio virgaureus*

Benzofuranosesquiterpenes: 2-[3-pentenyl]-3,7-dimethylbenzofuran-1,4-dione,^[78] 1-hydroxy-2-[3-pentenyl]-3,7-dimethylbenzofuran,^[79] 1-methoxy-2-[3-pentenyl]-3,7-dimethylbenzofuran,^[80] furanoeremophilanes^[81,82] [Z. J. Jia et al 1991].



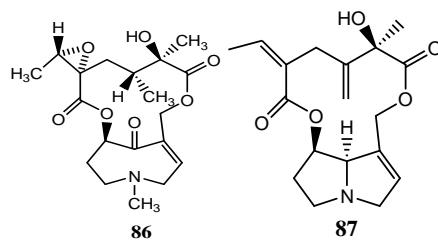
3.24. *Senecio bonariensis*

Pyrrolizidine alkaloids [J. A. Paiva, et al 2004], sesquiterpenoides-β-caryophyllene,^[83] β-caryophyllene oxide,^[84] germacrene D^[85] [C. M. Silva et al 2010].



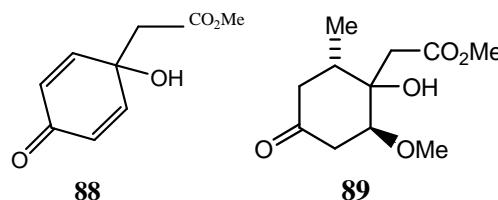
3.25. *Senecio lorentzii*

Otosenine^[86] [M. Noorwala, et al 2000], seneciphylline^[87] [R. J. Molyneux, et al 1982].



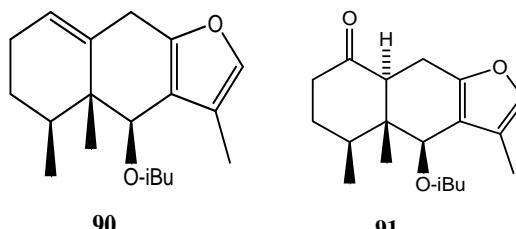
3.26. *Senecio minutes*

Friedelin [J. D. P. Teresa, et al 1980], 3-epifriedelinol [J. D. P. Teresa, et al 1979], fatty acids: lauric; myristic; palmitic; and Stearic, faradiol, maniladiol [P. Torres, et al 1992], arnidiol [J. S. Pyrek, et al 1973], jacaranone,^[88] methyl-1-hydroxy-2,6-dimethoxy-4-oxocyclohexanacetate^[89] [P. Torresb, et al 2000].



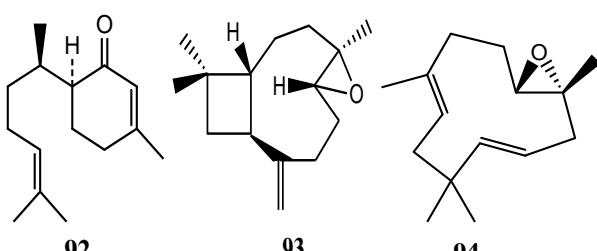
3.27. *Senecio boissieri*

Isohumulene [F. Bohlmann, et al 1974], lupeol [G. Ourisson, et al 1962, E. Wenkert, et al 1978], luponone [A. Chaterjee, et al 1966, M. Reina, et al 2002], 3-epifriedelinol [J. D. P. Teresa, et al 1979], 6β-isobutyryloxy-1[10]-furanoeremophilene,^[90] 6β-isobutyryloxy-1-oxo-10α-furanoeremophilene^[91] [P. Torres, et al 1998], triterpenoids [P. Torresb, et al 2000].



3.28. *Senecio palmensis*

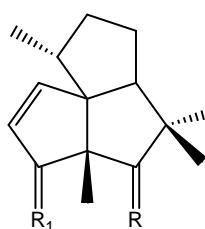
Sesquiterpenes, [6S]-2,10-Bisaboladien-1-one,^[92] 6,7-Epoxy-3[15]-caryophyllene,^[93] 6,7-Epoxy-2,9-humuladiene,^[94] 5α-angeloyloxyisilphin-3-one,^[95] 5α-senecioyloxyisilphin-3-one,^[96] 5α-acetoxyisilfinen-3-one,^[97] 5α-tigloyloxyisilphin-3-one,^[98] 3β-hydroxy-5α-angeloyloxyisilphinene,^[99] silphin-3,5-dione,^[100] 5α-hydroxysilphin-3-one,^[101] 5β-hydroxysilphin-3-one,^[102] 5β-acetoxyisilphin-3-one,^[103] 5β-isobutyryloxyisilphin-3-one,^[104] 5α-isobutyryloxyisilphin-3-one [M. Reina, et al 2002].



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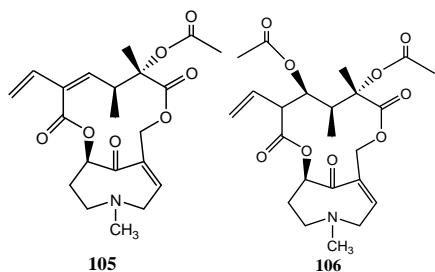
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3.29. *Senecio kaempferi*

Two hepatotoxic otonecine-type pyrrolizidine alkaloids- Clivorine^[105] and ligularine,^[106] [G. Lin, et al 2000].



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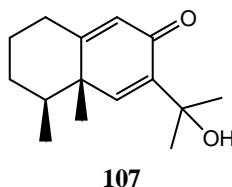
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3.30. *Senecio brasiliensis* populations

N-oxide pyrrolizidine-alkaloids [G. Schmeda, et al 1987].

3.31. *Senecio desfontaznez*

Eremophilanes, furoeremophilanes and pyrrolizidines, 11-hydroxyeremophil 6[7], 9[10]-dien-8-one^[107] [A. A. Ahmed et al 1991].



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3.32. *Senecio pandurifolius*

Essential oils: α -cuprenene, borneol, β -eudesmol, 1-undecene, [E]-caryophyllene, nonadecane, hexadecane, α -zingiberene, borneol, 1-undecene, E- γ -bisabolene bicyclogermacrene dehydroaromadendrene, γ -curcumene, undecane, α -zingiberene, [E,E]- α -farnesene, [E]-caryophyllene, 6-methoxy-2-[1-buten-3-yl]-naphthalene, β -eudesmol, α -Longipinene, silphiperfol-6-ene, β -elemene, β -curcumene, docosane, tricosane, pentacosane,

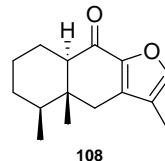
benzene acetaldehyde and decanal [N. Kahriman et al 2011].

3.33. *Senecio tenuifolius*

3-methyl-6,7-dihydrobenzofuran-4[5H]-one, methyl ester hydroquinone and 1,2- benzenedicarbocyclic acid [M. Manubolu, et al 2013].

3.34. *Senecio flaginoides* var. *flaginoides*

10H-9-oxofuranoeremophilane^[108] [L. Arancibia et al 2013].



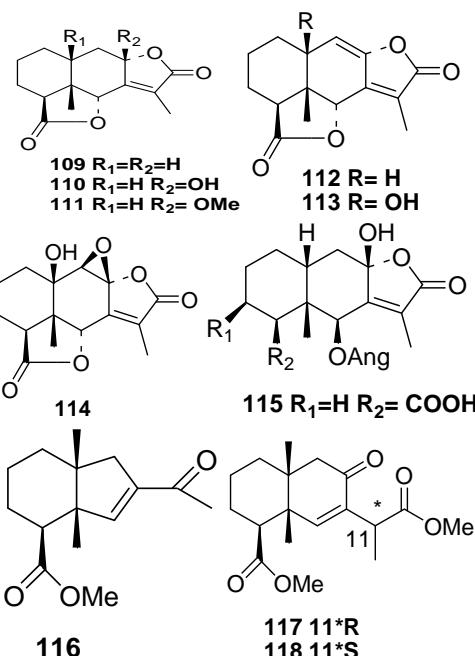
108

3.35. *Senecio delphinifolius*

Flavonoids, terpenes, alkaloids and saponins [Zellagui, et al 2004, Z. Amaral, et al 2012].

3.36 *Senecio przewalskii*

8β -Methoxyermophil-7[11]-en-6 α ,15,8 α ,12-diolide,^[111] eremophil-8[9]7,[11]-dien-6 α 15;8,12-diolide,^[112] 10 β -hydroxyeremophil-8[9]7[11]-dien-6 α ,15;8,12-diolide,^[113] 10 β -hydroxy-8 β ,9 β -epoxyermophil-7[11]-en-6 α ,15;8 α ,12-diolide,^[114] 8 β -hydroxy-6 β -angeloyloxyertnzophil -7-[11]-en-8 α ,12-olide-15-oic acid,^[115] 2-acetyl-3 α , β -methyl-3 α ,4,5,6,7,7a-hexahydroinden-4 β -carboxylic methyl ester,^[116] 11[RS]-8-oxoeremophil-6[7]-en-dimethyl-12,15-dioate,^[117,118] 2-acetyl-5,6-dimethoxybenzofuran,2-propenyl-5-acetyl-7-hydroxy-2,3-dihydroxybenzofuran,5-acetyl-7-methoxybenzofuran, and 1,3-dimethoxy-4,6,1-l-trimethylnaphthofuran [Y. Zhao et al 1995, Z. J. Jia et al 1994].



3.37. *Senecio faurie*

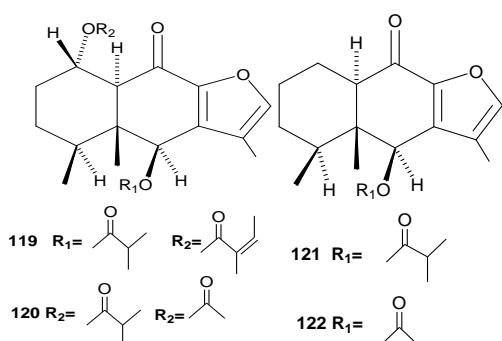
Eremophilanolides,^[109,110] [Y. Moriyama et al 1976].

3.38. *Senecio graveolens*

Essential oils, hydrocarbons [α -pinene, α -phellandrene, α -terpinene, p-cymene, sabinene, γ -terpinene, 1-methyl-4-isopropenylbenzene and terpinolene] also alcohols [terpinen-4-ol, α -eudesmol and β -eudesmol], ketones [acetone and piperitenone] and an aldehyde [isovaleraldehyde] [C. Perez, et al 1999].

3.39. *Senecio patagonicus*

α -angeloxy-6 β -isobutyroxy-9-oxo-10 α H-furanoeremophilane^[119] α -acetoxy-6 β -isobutyroxy-9-oxo-10 α H-furanoeremophilane,^[120] 6 β -isobutyroxy-9-oxo-10 α H furanoeremophilane^[121] 6 β -acetoxy-9-oxo-10 α H furanoeremophilane^[122] [L. Villarroel et al 1987,L. Villarroel et al 1991].

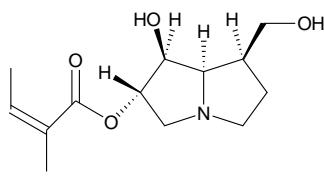


3.40. *Senecio chilensis*

α -angeloxy-6 β -isobutyroxy-9-oxo-10 α H-furanoeremophilane,^[119] α -acetoxy-6 β -isobutyroxy-9-oxo-10 α H-furanoeremophilane,^[120] 6 β -isobutyroxy-9-oxo-10 α H-furanoeremophilane^[121] [L. Villarroel et al 1991].

3.41. *Senecio nemorensis*

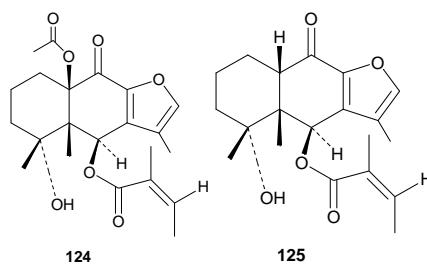
6 α -Angeloylplatynecine^[123] [V. Christov et al 2005], 7-angeloylplatynecine, neosarracine [L. Witte et al 1993], 9-angeloylplatynecine, sarracine [V. Christov et al 1997], mono- and open-chain diester platynecine and retronecine [T. Hartmann et al 1997].



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3.42. *Senecio fistulosus*

Furanoeremophiles- 4 α -hydroxy-6 β -angeloxy- 10 β -acetoxy-9-oxofuranoeremophilane^[124] 4 α -hydroxy-6 β -angeloxy-9-oxofuranoeremophilane^[125] [L. Villarroel et al 1985].



3.43. *Senecio vira-vira*

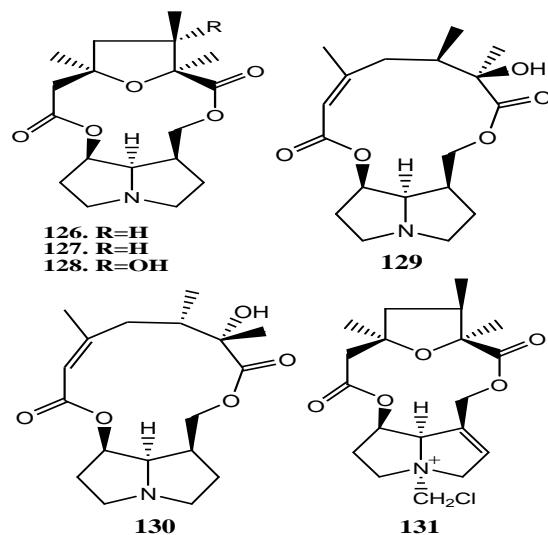
Quercetin 3-0- α -chamnopycanoside, quercetin 3-0- β -rutinoside [T. J. Mabry, et al 1970, K. R. Markham, et al 1978], isorhamnetin 3-0- β -robinobioside [C. A. Buxhi et al 1982], anacrotine [C. K. Atal, et al 1966], neoplatyphylline [C. C. J. Culvenor, et al 1968], uspallatine [M. J. Pestchanker, et al 1985], sitosterol, campesterol, stigmasterol, α - and β -arnyrrins, stigmasta-3,5dien-7-one, and stigmasta4,6-dien-3-one [E. Jares et al 1986].

3.45. *Senecio brasiliensis*

Pyrrrolizidine alkaloids- Integerrimine and retrorsine [G. Schmeda, et al 1987].

3.46. *Senecio bracteatus*

Pyrrolizidine alkaloid-iodanthine,^[130] retroisoseneine,^[126] chloromethylretroisoseneine chloride^[131] [A. L. Perez-Castorena et al 1999], bulgarsenine^[129] [P. Castorena, et al 1998], mulgediifoliine^[127] [R. D. Vivar, et al 1995], [12S]-12-hydroxyretroisoseneine^[128] [P. Castorena, et al 1998].



3.47. *Senecio iordanthus*

Pyrrolizidine alkaloid-iodanthine,^[130] retroisoseneine^[126] [A. L. Perez-Castorena et al 1999], bulgarsenine^[129] [P. Castorena, et al 1998].mulgediifoliine^[127] [R. D. Vivar, et al 1995,].

3.48. *Senecio trapezuntinus*

[E]- β -farnesene [O. Uçuncu, et al 2008].

3.49. *Senecio vernalis*

Spathulenol, 1,8-cineole, m-cymene, isobicyclogermacrene and α -phellandrene, [D. Nori-Shargh, et al 2008]. β -pinene and α -pinene, Δ -3-carene, germacrene D,Z- β -ocimene, and α -humulene [A. Usta, et al 2009].

3.50. *Senecio platyphyllus* var. *platyphyllus*

E-caryophyllene, germacrene D and E- β -farnesene [A. Usta, et al 2009].

3.51. *Senecio glaucus* subsp. *Coronopifloios*

Myrcene and dehydrofukinone [H. L. D. Pooter, et al 2006].

3.52. *Senecio leucostachys*

Sabinene, α -phellandrene, germacrene D and β -caryophyllene [R. R. Vera et al 1994 ,N. Mengi, et al 1995, A. M. El-Shazly et al 1999, V. T. Balzaretti, et al 2000, M. Mirza et al 2008, A. G. Belaunde et al 2007, G. F. Zuniga et al 1996].

3.53. *Senecio squalidus*

P-cymene and α -phellandrene [J. C. Chalchat, et al 2004].

3.54. *Senecio aegyptius* var. *discoideus*

1,10-epoxyfuranoeremophilane [E.Burgueno-Tapia et al 2004].

3.55. *Senecio graveolens*

α -Terpinene, p-cymene, terpinen-4-ol, and α -phellandrene [C. Perez, et al 1999], p-hydroxyacetophenone [L. A. Loyola et al 1985].

3.56. *Senecio farfarifolius*

α -Pinene and 1, 8-cineole [K. H. C. Baser et al 2004].

3.57. *Senecio nutans*

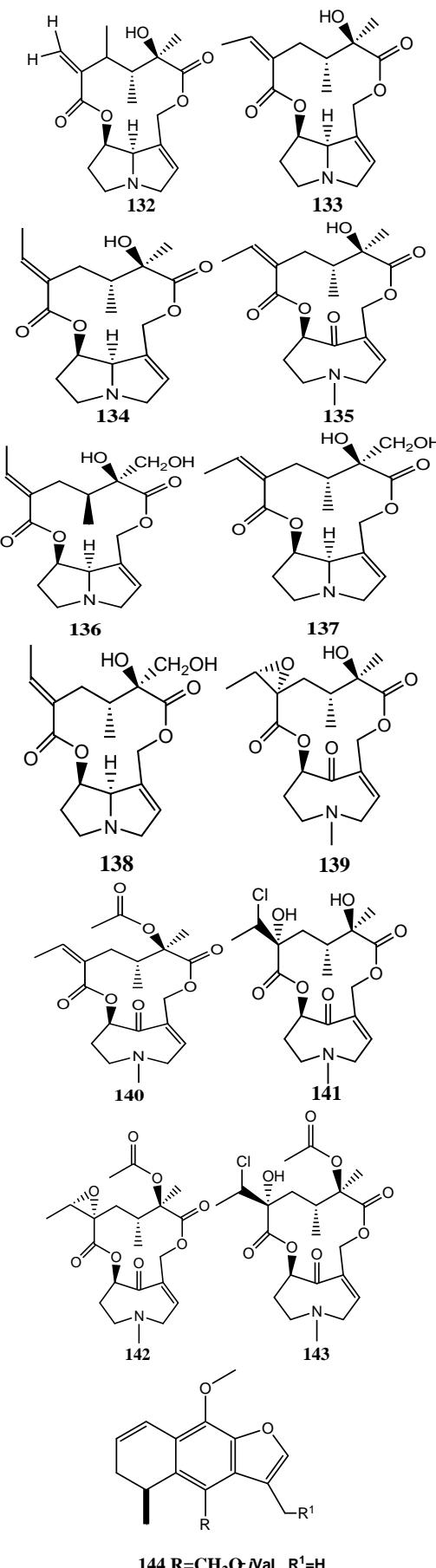
Oxygenated monoterpene hydrocarbons predominate [V. D. Feo, et al 2003].

3.58. *Senecio longipenicillatus*

α -pinene, α -humulene and germacrene D [M. Rondon et al 2006].

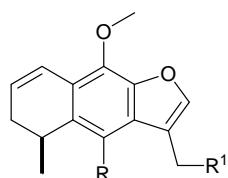
3.59. *Senecio madagascariensis*

Senecivernine,^[132] senecionine,^[133] integerrimine,^[134] senkirkine,^[135] mucronatinine,^[136] retrorsine,^[137] usaramine,^[138] otosenine,^[139] acetylsenkirkine^[140] desacetylidoronine^[141] florosenine,^[142] doronine^[143] [D. R. Gardner,et al 2006], [14-isovaleryloxy-1,2-dehydrocatalol methyl ether,^[144] [E. B. Tapia et al 2007, E. Burgueno- Tapia, et al 2001].



3.60. *Senecio barba-johannis*

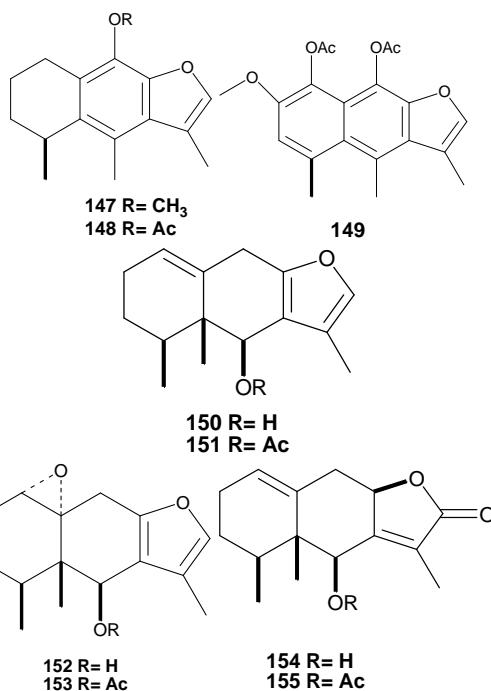
13-hydroxy-14-oxocacalohastine,^[145] 13-acethoxy-14-oxocacalohastine,^[146] [E. Burgueno- Tapia,et al 2004,E. Burgueno- Tapia,et al 2006].



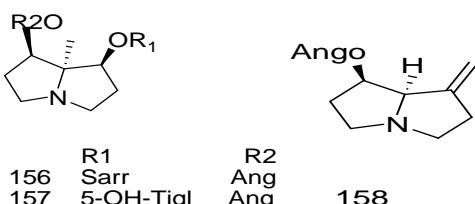
145 R= CH=O R¹= OH
146 R= CH=O R¹= OAc

3.61. *Senecio toluccanus*

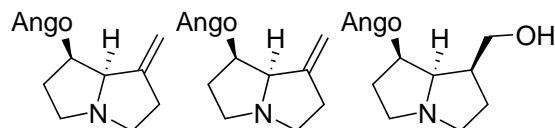
Cacalol methyl ether,^[147] cacalol acetate,^[148] 1-acethoxy-2-methoxy-1,2,3,4-tetrahydrocacalol acetate,^[149] 6-hydroxyeuryopsin,^[150] 6-acetyloxyeuryopsin,^[151] 1[10]-epoxy-6-hydroxyeuryopsin,^[152] 6-acethoxy-1[10]-epoxyeuryopsin,^[153] toluccanolide A,^[154] toluccanolide A acetate,^[155] [E. Burgueno- Tapia,et al 2004,E. Burgueno- Tapia,et al 2006]

**3.62. *Senecio doratophyllum***

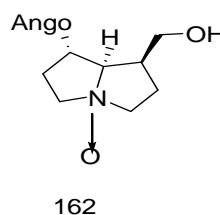
Sarracine,^[156] Neosarracine,^[157] 7b-[Angeloxyloxy]-1-methylidene-8a-pyrrolizidine^[158] [A.-L. Pe rez-Castorena, et al 1999].

**3.63. *Senecio.chrysocoma***

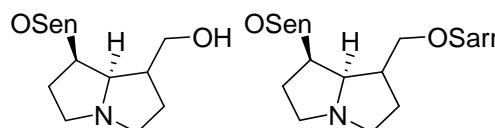
7b-[Angeloxyloxy]-1-methylidene-8a-pyrrolizidine^[159] [A.-L. Pe rez-Castorena,et al 1999], 7a-[Angeloxyloxy]-1-methylidene-8a-pyrrolizidine^[160] [J. R. Liddell,et al 1993], 7-[Angeloxyloxy]platynecine^[161][M. R. Grue et al 1993],7a-[Angeloxyloxy]-1-methylidene-8a-pyrrolizidine N-oxide^[162][J. R. Liddell, et al 1993].



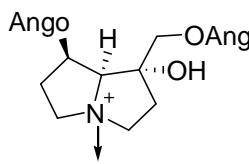
159 160 161

**3.64. *Senecio.cacaliaste***

7-[Senecioyoxy]retronecine,^[163] 9-[Sarracinoxyloxy]-7-[senecioyoxy]-retronecine,^[164] 7,9-Bis[angeloyloxy]-1a-hydroxy-Nmethylplatynecinium Chloride^[165][E. Roeder, et al 1984].



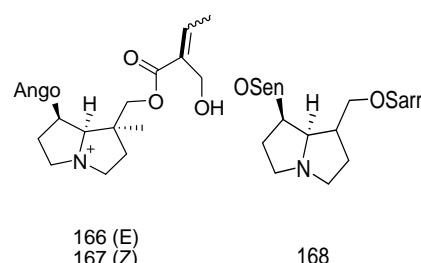
163 164



165

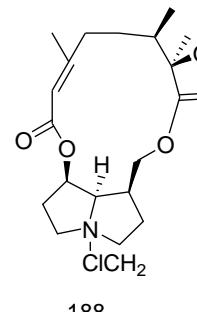
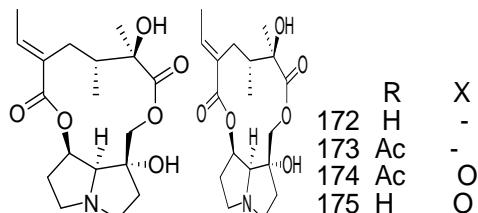
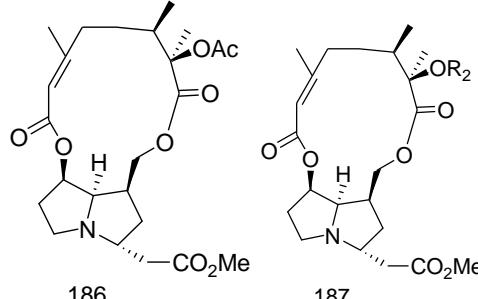
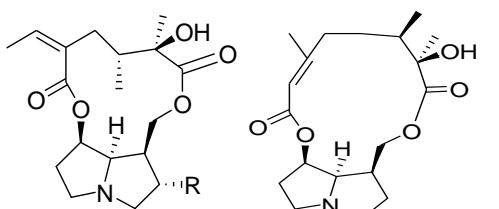
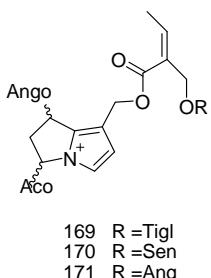
3.65. *Senecio.kaschkarovii*

Triangularine,^[166] Neotriangularine,^[167] 9-[Sarracinoxyloxy]-7-[senecioyoxy]-retronecine^[168][D.-L. Cheng, et al 1992].

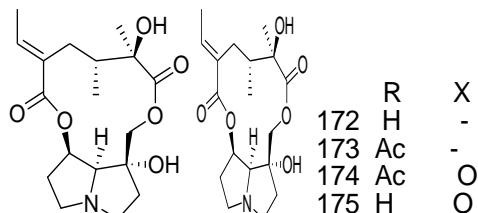


3.66. *Senecio.mikanoides*

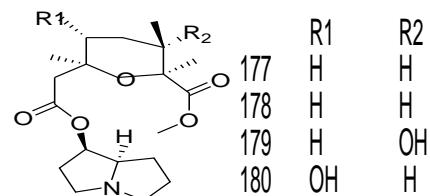
Senampelin E,^[169] Senampelin F,^[170] Senampelin G^[171] [F. Bohlmann et al 1979].

**3.67. *Senecio.hadiensis***

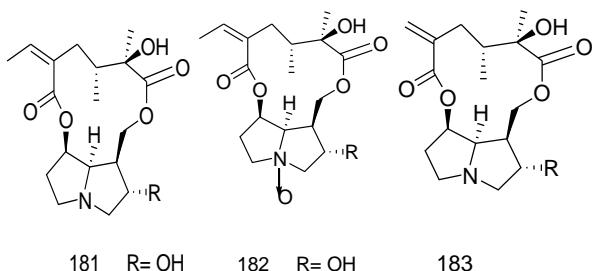
Hadiensine,^[172] 12-O-Acetylhadiensine^[173] 12-O-Acetylhadiensine N-oxide,^[174] Hadiensine N-oxide,^[175] Neorosmarinine^[176] [O. Were, et al 1993].

**3.68. *Senecio.roseus***

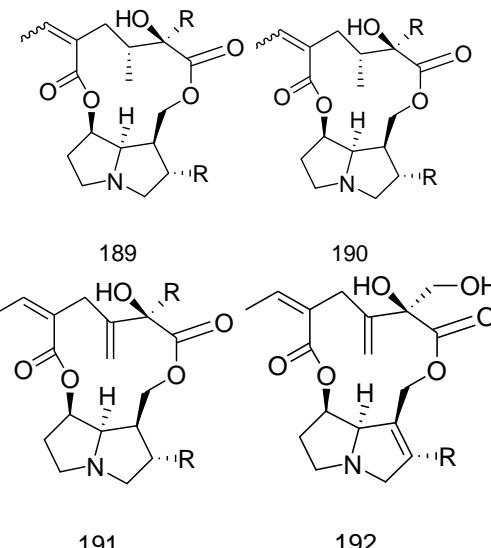
Retroisosenine,^[177] Mulgediifoliine,^[178] [12S]-Hydroxyretroisosenine,^[179] [13R]-Hydroxyretroisosenine^[180] [A.-L. Pe rez-Castorena et al 1997].

**3.69. *Senecio.syringifolius***

Rosmarinine^[181] [H. L. DeWaal, et al 1940], Rosmarinine N-oxide^[182] Neoangularine^[183] [O. Were et al 1993].



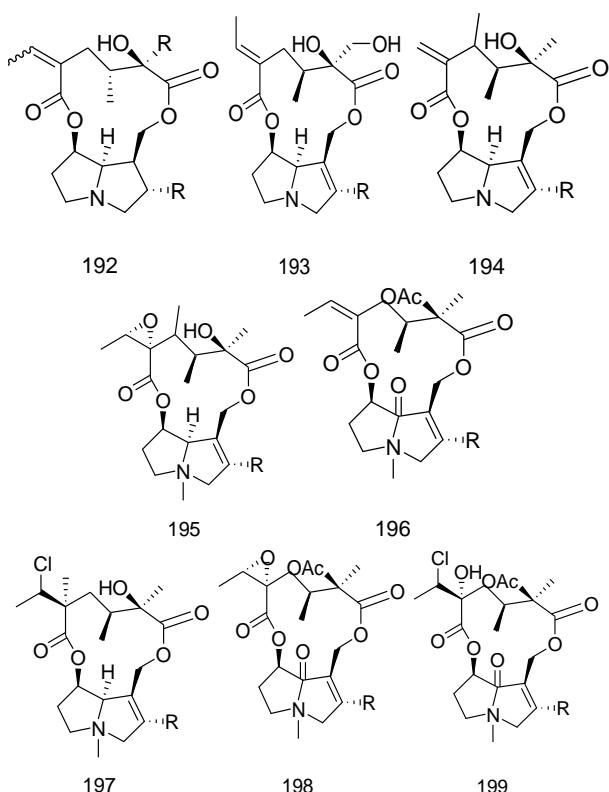
3.71. *Senecio.oxiphyllus*
Retrorsine,^[189] Usaramine,^[190] Seneciphylline,^[191] Riddelline^[192] [H. C. Krebs, et al 1996].

**3.70. *Senecio.callosus***

Rosmarinine,^[184] Bulgarsenine,^[185] Callosine. 11-O-Acetylbulgarsenine^[186] [A. RomoDeVivaretal 1996]. [11OAcetylbulgarsenineNoxide. N [Chloromethyl] bulgarsenine,^[187] N[Chloromethyl]bulgarsenine^[188] [A.-L. Pe rez-Castorena, et al 1998].

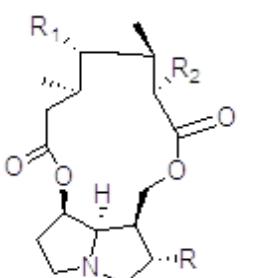
3.72. *Senecio.Madagascariensis*

Usaramine,^[192] Mucronatinine,^[193] Senecivernine,^[194] Otosene,^[195] Acetylsenkirkine,^[196] Desacetylodonine,^[197] Florosenine,^[198] Doronine^[199] [D. R. Gardner, et al 2006].



3.73. *Senecio. Bracteatus*

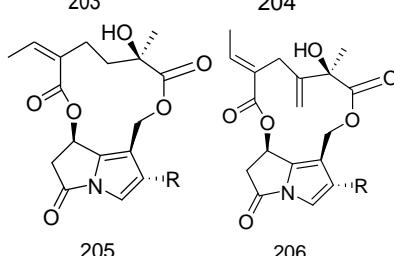
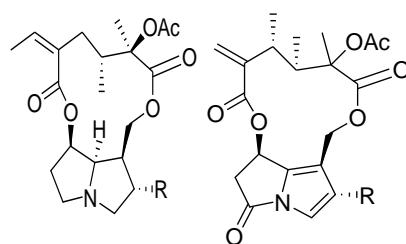
Retroisosenine^[200] [A.-L. Pérez-Castorena, et al 1999],
Mulgediifoline,^[201] [12S]-Hydroxyretroisosenine^[202] [A. R. D. Vivar et al 1995].



	R1	R2
200	H	H
201	H	H
202	H	OH

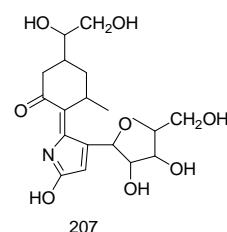
3.74. *Senecio. pterophorus*

Acetylseneciphylline^[203] [J. R. Liddell, et al 1993],
Pterophorin^[204] [F. Bohlmann, et al 1977],
Seneciphylline,^[205] Spartiodine^[206] [J. R. Liddell, et al 1993].



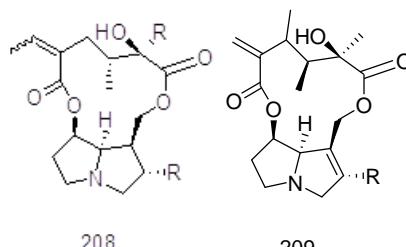
3.75. *Senecio.cannabifolius*

Cannabiloid B^[207] [B. Wu, W et al 2006].



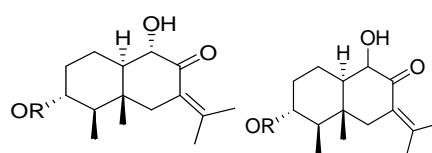
3.76. *Senecio.gilliesiano*

Retrorsine,^[208] Senecionine^[209] [F. H. Guidugli et al 1986].



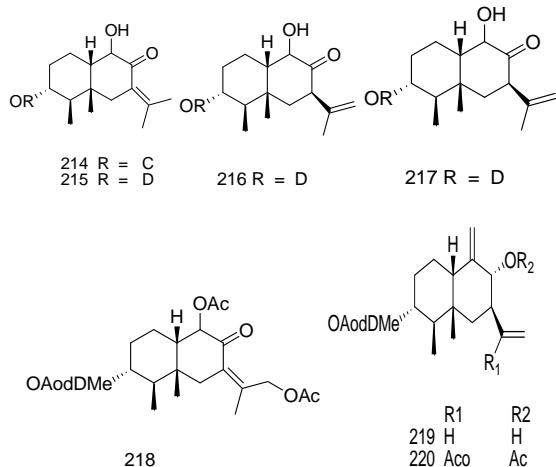
3.77. *Senecio lividus*

3a-[4'-[Angelyloyloxy]angelyloyloxy]-9a-hydroxy-10aHeremophil-7[11]-en-8-one,^[210] 3a-[5'[Angelyloyloxy]angelyloyloxy]-9a-hydroxy-10aHeremophil-7[11]-en-8-one,^[211] 3a[4'[Angelyloyloxy]angelyloyloxy]9hydroxyeremophila7[11],9dien8one,^[212] 3a[5'[Angelyloyloxy]angelyloyloxy]-9-hydroxyeremophila-7[11],9-dien-8-one^[213] [J. M. Cardoso et al 1987].



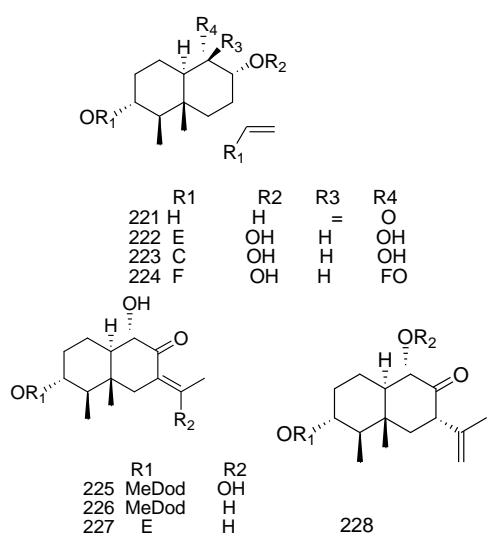
3.78. *Senecio. Speciosus*

4'-[Angeloxyloxy]isosenspeciosone,^[214]
 5'[Angeloxyloxy]isosenspeciosone,^[215]
 5'[Angeloxyloxy]8a,Odihydrosenspeciosone,^[216]
 5'[Angeloxyloxy]senspeciosone,^[217]
 9b,13Diacetoxy8oxoisosenspeciosoltrieneester,^[218]
 8aHydroxy9oxosenspeciosol triene ester,^[219] 8a,13-Diacetoxy-9-oxosenspeciosol triene ester^[220][F. Bohlmann, et al 1978].



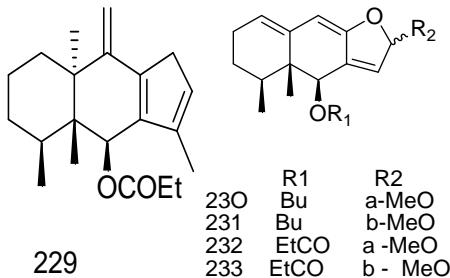
3.79. *Senecio.erubescens* var *crepidifolius*

3a-[{[2Z]-4-[[2Z]-Hex-2-enoyl]oxy}hex-2-enoyl]oxy]-7a,10aH-eremophil-11-ene-8a,9a-diol,^[221] 3a-{[2Z]-4-[Angeloxyloxy]-hex-2-enoyl]oxy}-7a,10aH-eremophil-11-ene-8a,9a-diol,^[222] 3a,9a-{[2Z]-Dihex-2-enoyl]oxy}-7a,10aH-eremophil- 11-en-8a-ol,^[223] 9,13-Dihydroxy-3a-{[2E,4E,6E]-5-methylidodeca,-^[224] 2,4,6-trienoyl]oxy}-10aH-eremophil-7[11]-en-8-one,^[225] 9a-Hydroxy-3a-{[2E,4E,6E]-5-methylidodeca-2,4,6-trienoyl]oxy}-10aH-eremophil-7[11]-ene-8-one,^[226] 3a-[{[2Z]-4-{{[2Z]-Hex-2-enoyl]oxy}hex-2-enoyloxy}-9a-hydroxy-10aH-eremophil-7[11]-en-8-one],^[227] 3a-[{[2Z]-4-{{[2Z]-Hex-2-enoyl]oxy}hex-2-enoyloxy}-9a-hydroxy-7a,10aH-eremophil-11-en-8-one]^[228][F. Bohlmann, et al 1985].



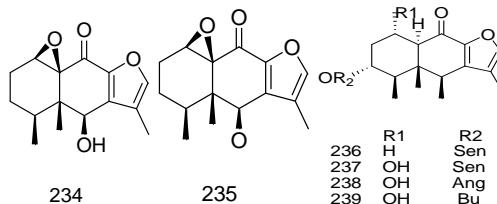
3.80. *Senecio.pachyphylllos*

6b-[Propionyloxy]-10aH-furanoeremophilan-9-one.^[229]
 8,12-Epoxy-6b-[isobutyryloxy]-12a-methoxyeremophilane-1[10],7[11],8-triene,^[230] 8,12-Epoxy-6b-[isobutyryloxy]-12b-methoxyeremophilane-1[10],7[11],8-triene,^[231] 8,12-Epoxy-12amethoxy6b[propionyloxy]eremophilane-1[10],7[11],8-triene,^[232] 8,12-Epoxy-12b-methoxy-6b-[propionyloxy]eremophilane-1[10],7[11],8-triene^[233][M. Ahmed, et al 1991].



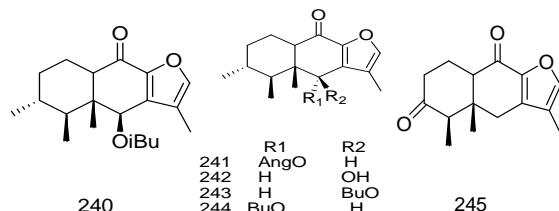
3.81. *Seneciosmithii*

1b,10b-Epoxy-6b-hydroxyfuranoeremophilane-6,9-dione,^[234]
 1b,10b-Epoxyfuranoeremophilane-6,9-dione,^[235]
 Euryopsonol senecioate,^[236] 1a-Hydroxyeuryopsonol senecioate,^[237] 1a-Hydroxyeuryopsonol angelate,^[238] 1a-Hydroxyeuryopsonol isobutyrate^[239][F. Bohlmann et al 1981].



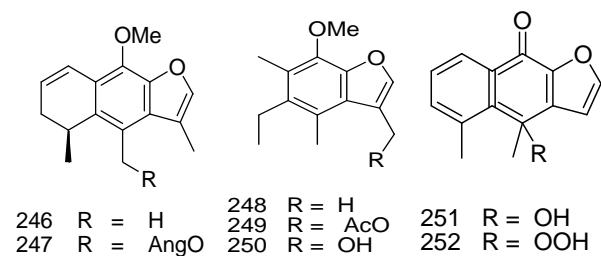
3.82. *Senecio pseudoorientalis*

6β-[Isobutyryloxy]furanoeremophil-1[10]-en-9-one,^[240]
 6α[Angeloxyloxy]furanoeremophil-9-en-1-one,^[241]
 6βHydroxyfuranoeremophil9en1one,^[242]
 6β[Isobutyryloxy]furanoeremophil9-en-1-one,^[243]
 6α[Isobutyryloxy] furanoeremophil-9-en-1-one,^[244]
 Furanoeremophil-1-en-3-one^[245][F. Bohlmann, et al 1980].



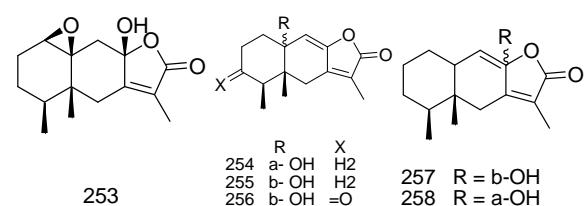
3.83. *Senecio.canescens*

Cacalohastine,^[246] 14-[Angeloxyloxy]cacalohastine,^[247]
 3,4-Dehydrocacalohastine,^[248] 13-Acetoxy-3,4-dehydrocacalohastine,^[249] 13-Acetoxy-3,4-Dehydro-13-hydroxycacalohastine,^[250] 3,4-Dehydro-13-Cacalonol,^[251]
 Peroxycacalonol^[252][S. Abdo, et al 1992].



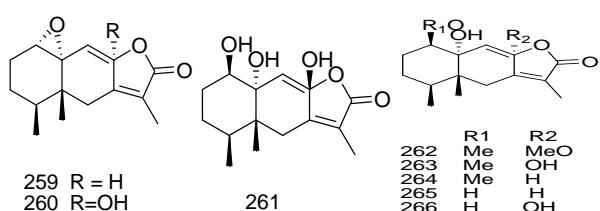
3.84. *Senecio.tsoongianus*

Tsoongianolide F,^[253] Tsoongianolide A,^[254]
 Tsoongianolide B,^[255] Tsoongianolide E,^[256]
 Tsoongianolide C,^[257] Tsoongianolide D^[258] [Y. Zhao, et al 2004].



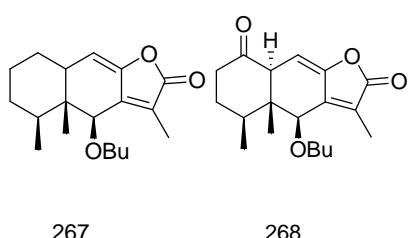
3.85. *Senecio.mairetianus*

Mairetolide A,^[259] Mairetolide B,^[260] Mairetolide H,^[261]
 Mairetolide C,^[262] Mairetolide D,^[263] Mairetolide E,^[264]
 Mairetolide F,^[265] Mairetolide G^[266] [A.-L. Pe'rez-Castorena, et al 2006].



3.86. *Senecio.boissieri*

6β-[Isobutyryloxy] furanoeremophil-1[10]-ene,^[267] 6β-[Isobutyryloxy]-10aH-furanoeremophilan-1-one^[268] [P. Torres et al 2000].

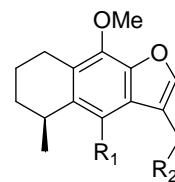


267

268

3.87. *Senecio.picardae*

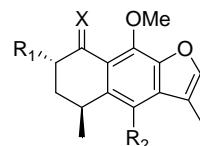
13-Acetoxycacalol methyl ether,^[269] 14-Oxocacalol methyl ether,^[270] 13-Acetoxy-14-oxocacalol methyl ether,^[271] 13-Acetoxy-14-[angeloyloxy]cacalol methyl ether^[272] [F. Bohlmann, et al 1990].



269	R1 Me	R2 AcO
270	CHO	H
271	CHO	AcO
272	AngOCH2	AcO

3.88. *Senecio.fuertesii*

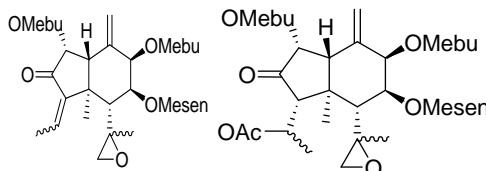
1-Oxocacalol methyl ether,^[273] 1,14-Dioxocacalol methyl ether,^[274] 2-Acetoxycacalol methyl ether^[275] [F. Bohlmann, et al 1990].



273	R1 H	R2 Me	X O
274	H	CHO	O
275	AcO	Me	H2

3.89. *Senecio.implexus*

[3Z]-Implexin,^[276] [3E]-Implexin,^[277] 14-Acetoxy-3,14-dihydroimplexin^[278] [F. Bohlmann, et al 1981].



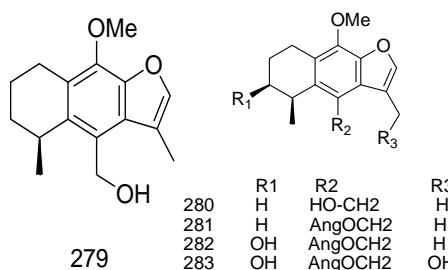
276 3Z

277 3E

278

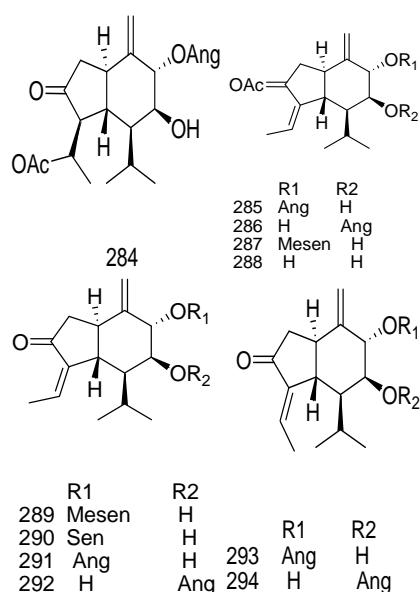
3.90. *Senecio.othonnae*

14-Hydroxycacalol methyl ether,^[279] 1,2-Dehydro-14-hydroxycacalol methyl ether,^[280] 14-[Angeloyloxy]-1,2-dehydrocacalol methyl ether,^[281] 14-[Angeloyloxy]-1,2-dehydro-3b-hydroxycacalolmethyl ether,^[282] 14-[Angeloyloxy]-1,2-dehydro-3b,13-dihydroxycacalolmethyl ether^[283] [F. Bohlmann, et al 1977].



3.91. *Senecio.yugua*

14-Acetoxy-7 α -[angeloyloxy]-6 β -hydroxyoplop-8[10]-en-2-one,^[284] [3Z]-2 β -Acetoxy-7 α -[angeloyloxy]-6 β -hydroxyoplop-3[14]-8[10]-diene,^[285] [3Z]-2 β -Acetoxy-6 β -[angeloyloxy]-7 α hydroxyoplop-3[14],8[10]-diene,^[286] [3Z]2 β Acetoxy6bhydroxy7a[[4methylsenecioyl]oxy]oplop-3[14],8[10]-diene,^[287] [3Z]-2 β -Acetoxy-6 β ,7 α dihydroxyoplop-3[14],8[10]-diene,^[288] [3Z]-6 β -Hydroxy-7 α -[[4-methylsenecioyl]oxy]oplop-3[14],8[10]-dien-2-one,^[289] [3Z]-6 β -Hydroxy-7 α -[senecioyloxy]oplop-3[14],8[10]-dien-2-one,^[290] [3Z]-7 α -[Angeloyloxy]-6 β -hydroxyoplop-3[14],8[10]-dien-2-one,^[291] [3Z]-6 β -[Angeloyloxy]-7 α -hydroxyoplop-3[14],8[10]-dien-2-one,^[292] [3E]-7 α -[Angeloyloxy]-6 β -hydroxyoplop-3[14],8[10]-dien-2-one,^[293] [3E]-6 β -[Angeloyloxy]-7 α -hydroxyoplop-3[14],8[10]-dien-2-one^[294] [M. A. Aal,et al 1988].



4. Biological Activities.

Various biological activities reported from the plants of the genus *Senecio*.

4.1. Antibacterial and Antifungal Activities. In the folk medicine, *Senecio* species were used for the treatment of wounds and as antiemetic, anti-inflammatory, and vasodilatory preparations. More recently, antibacterial and antifungal activities of extracts and compounds isolated from *Senecio* species were reported. The MeOH extract of *S. vulgaris* showed antimicrobial activity against the Gram-positive bacteria *Bacillus subtilis* [minimal-inhibitory concentration [MIC] 0.5 mg/ml] and *Staphylococcus aureus* [MIC 0.125 mg/ml]. The MeOH extracts from both *S. inaequidens* and *S. vulgaris* showed low activity against dermatophytes. The hexane extract of *S. vulgaris* showed significant activity against *Trichophyton tonsurans* [MIC 0.031 mg/ml] [M. R. Loizzo, et al 2004]. Although their activities were less potent than those of chloramphenicol, Cannabiloid B and CannabisideB S.Cannabiside C Cannabilactone A isolated from *S. cannabifolius*,

showed antibacterial activities against the Gram-positive bacteria *S. aureus* and *B. subtilis*, but not against Gram-negative bacteria [B. Wu, W. et al 2006]. Assessed by the agar-well diffusion method, the isolated essential oil of *S. graveolens* showed antimicrobial activity against *Micrococcus luteus* ATCC 9341, oxacillin-sensitive and oxacillin-resistant *S. aureus*, as well as antifungal effects against clinically isolated *Candida albicans*. The MIC values for *M. luteus*, oxacillin-sensitive *S. aureus*, and *C. albicans* were 8.73, 10.91, and 2.13_10_2 mg/ml, respectively. The antimicrobial activity related to known antibiotics was calculated. These results were compatible with a potential concentration-dependent selectivity of the antifungal effect of *S. graveolens* essential oil. Moreover, the minimal bactericidal concentration [MBC] was above 87.3 mg/ml. Thus, the MBC/MIC ratio was clearly higher than 1 [above 8], indicating a bacteriostatic effect of the essential oil [C. Perez, et al 1999]. The aerial parts of *S. aegyptius* var. *discoideus* afforded six new eremophilane derivatives, 8a-Methoxy-1-oxo-10aH-eremophil-7[11]-ene-12,8-lactam and 1b-Hydroxy-8aH-eremophil-7[11],9-dien-12,8-olide 1b,8a-Dihydroxyeremophil-7[11],9-dien-12,8-olide 1b-Hydroxy-8a-methoxyeremophil-7[11],9-dien-12,8-olide 1b,10b-Epoxy-8a-hydroxyeremophil-7[11]-en-12,8-olide 250 1b,10b-Epoxy-8a-methoxyeremophil-7[11]-en-12,8-Olide The antibacterial activities of these compounds were tested against two microorganisms, a Gram-positive [*Bacillus cereus*] and a Gram-negative bacterium [*Serratia sp.*], at concentrations of 200 and 400 mg/ml. The growth of both microorganisms was inhibited by compounds 1b-Hydroxy-8aH-eremophil-7[11], 9-dien-12,8-olide 1b,8a-Dihydroxyeremophil-7[11],9-dien-12,8-olide 1b-Hydroxy-8a-methoxyeremophil-7[11], 9-dien-12,8-olide 1b,10b-Epoxy-8a-hydroxyeremophil-7[11]-en-12,8-olide 250 1b,10b-Epoxy-8a-methoxyeremophil-7[11]-en-12,8-Olide inhibited the growth of *B. cereus*, but had no effect on the growth of *Serratia sp.* [E. H. H. et al 2005].

4.2. Antitubercular Activity. *S. chionophilus* is a small, woody shrub growing in the Mountains in South America, above 1500 m. Infusions of its aerial parts and roots are traditionally used in Chile for treating heavy colds and runny noses. All compounds isolated from this species were evaluated for their antitubercular potential against *Mycobacterium tuberculosis*, in a microplate Alamar Blue assay. Compounds 1a-Hydroxy-6b-[[2-methylbutyryl] oxy]-10aH-furanoeremophil-9-one, 6b-[Angeloyloxy]-1a-hydroxy-10aH-furanoeremophil-9-one and 4-Hydroxyacetophenone exhibited mild antitubercular activity, with MIC values of 119, 114 and 121 mg/ml, respectively. This is the first evidence that sesquiterpenoids of the furanoeremophilane type may be considered as potential antitubercular leads. In addition, the comparison of the antitubercular activities of betulinic, oleanolic, andursolic acids, with MICs of 32, 64, and 32 mg/ml, respectively, with those of inactive analogues indicated that the presence of a COO group in a polycyclic triterpeneskeleton appears to be

necessary for the observed activity against *M. tuberculosis* [J.-Q. Gu, et al 2004].

4.3. Anti-Inflammatory Effects. To investigate the anti-inflammatory effect of the total flavonoids of *S. scandens*, various inflammatory models, including the swelling of fear induced by xylene in mice, the permeability increase of blood capillary by acetic acid in mice, and the cotton-pellet granuloma in mice, were used. In addition, air-sac models of synovitis in mice were set up and the amount of white blood cells [WBCs] and the content of prostaglandin E2 [PGE2] were measured in the inflammatory exudate. The total flavonoids had significant inhibitory action on auris swell induced by xylene in mice and the penetration of capillary vessels in mice. The flavonoids also significantly decreased the proliferation of granuloma caused by implantation of cotton pellets in mice. The amount of WBCs and the content of PGE2 in the experimental groups were obviously lower than those in the control groups. The total flavonoids showed significant anti-inflammatory activity, which is relevant to the inhibition of the production and release of the inflammatory factor PGE2 [W. P. Zhang, et al 2008].

4.4. Antiulcer Activity. The leaves and inflorescences of *S. brasiliensis* are utilized in the traditional medicine for the treatment of inflammatory processes as a blood regulator. Moreover, this plant is also used in the folk medicine to relieve stomach pain. The crude alkaloid extract of *S. brasiliensis* inflorescences, containing a mixture of the PasIntegerrimine, Senecionine, Retrorsine, Usaramine, and Seneciphylline, was evaluated for preventive antiulcer effects on standard rodent models of induced gastric and duodenal ulcers. In the HCl/EtOH-, indomethacin/bethanechol-, and hypothermic restraint-induced gastric ulcers, the lesion was significantly inhibited [$p<0.001$] by the PAs [po]. In the pylorus-ligation, PAs [id] significantly increased the gastric juice content and the pH values and decreased the acid output. In the cysteamine-induced duodenal ulcers, PAs [po] showed significant inhibition [$p<0.001$] of the duodenal lesions when compared to the respective control. The levels of the somatostatin hormone in the blood samples of animals pretreated with the PA mixture [12.5mg/kg] and the free mucus and prostaglandin synthesis after the administration of the PA extract [po] also increased [$p<0.001$]. The results suggested that the PA extract from *S. brasiliensis* inflorescences presents a significant antiulcer effect in the selected ulcer models. One of the mechanisms involved in the action of the PA extract is cytoprotection. Additional studies are in progress to determine other possible mechanisms involved in the antiulcer effect of PAs [W. Toma, et al 2004].

4.5. Antifeedant Effects. Species belonging to the family Asteraceae are an important source of terpenes and alkaloids with biological activity. The tricyclopentanoids sesquiterpene 11b-acetoxy-5a-[angeloyloxy] silphinene-3-one [487], isolated from *S.*

palmensis [A. Gonzalez-Coloma, et al 1995], as well as related C[5]/C[11]-substituted derivatives were found to be very efficient antifeedants against several divergent insect species [A. Gonzalez-Coloma, et al 1995]. To demonstrate the importance of this class of molecules as model insect antifeedants and their potential as new GABA modulators, [M. Reina, et al 2002] carried out additional chemical work on *S. palmensis*, resulting in the isolation of the known sesquiterpenes [6S]-Bisabola-3,7[11]-dien-2-one, 5a-[Angeloyloxy]silphinene-3-one, 5a-Acetoxy silphinene-3-one, and 6,7-Epoxyhumula-2,9-diene. The new silphinenes 5a-[Senecioyloxy] silphinene-3-one, 5a-[Tigloyloxy] silphinene-3-one, and 5a-[Angeloyloxy]-3b-hydroxysilphinene]Caryophyllanes. A series of semisynthetic analogues, such as silphinene-3,5-dione and 5-hydroxysilphinene-3-one, was also generated, to carry out a preliminary structure – activity study on the antifeedant action of these molecules against several divergent insect species, including the lepidopteran *Spodoptera littoralis*, the chrysomelid *Leptinotarsa decemlineata* [Colorado potato beetle, CPB], and five aphid species with diverse host adaptations. In summary, C[5]-substituted silphinenes, such as [6S]-Bisabola-3,7[11]-dien-2-one, 5a-[Angeloyloxy]silphinene-3-one, 5a-Acetoxy silphinene-3-one, and 6,7-Epoxyhumula-2,9-diene. The new silphinenes 5a-[Senecioyloxy]silphinene-3-one, 5a-[Tigloyloxy]silphinene-3-one, and 5a-[Angeloyloxy]-3b-hydroxysilphinene]Caryophyllanes, are more efficient CPB and aphid antifeedants than their biogenetic precursors 6,7-Epoxy caryophyll-3[15]-ene Humulanes 6,7-Epoxyhumula-2,9-diene. A comparative study of their activities with that of C[5]/C[11]-substituted analogues showed that esterification at C[5] and acetylation at C[11] are important structural requirements for the antifeedant activity of these molecules [M. Reina, et al 2002]. Compounds 1a-Acetoxy-8b-methoxyeremophil-7[11]-en-12,8-olide and 1a-[Angeloyloxy]-6b-hydroxy-8b-methoxyeremophil-7[11]-en-12,8-olide, isolated from *S. miser*, were found to be effective antifeedants against *Myzus persicae* and *L. decemlineata*, respectively [M. Reina, et al 2001]. Similarly, several sesquiterpene lactones with a g-butenolactone group have been reported as having moderate to high potency as *L. decemlineata* antifeedants. The PA 37 was found to be an effective antifeedant against *L. decemlineata* [not adapted to PAs] [M. Reina, et al 2001]. Previous data have shown that the CPB is sensitive to the macrocyclic diesters senecionine, the open diester PA echimidine, and the saturated monoester PA 3'-acetyl trachelanthamine, but with too few structural features in common to draw any conclusions on structure – activity relationships. Also the PA N-oxide Integerrimine N-oxide was found to be a strong CPB antifeedant [M. Reina, et al 2001]. Previous reports have shown that some PA N-oxides, including senecionine N-oxide, are active antifeedants against some aphid species and *S. littoralis*, but they were found to be less potent than their tertiary bases. The data presented by [M. Reina, et al 2001] suggest that the CPB putative taste receptors can interact with different

PAs [and N-oxides] of different structural classes with high molecular selectivity. Little is known, however, about the molecular mechanisms that modulate PA insect taste reception. Several PAs, including N-oxides and Senecionine, have demonstrated significant binding activity to muscarinic and serotonin receptors, indicating that these compounds can affect several molecular targets besides long-term toxicity through DNA alkylation by PA metabolites generated in the liver. Therefore, the interference of PAs with neuronal signal transduction could mediate insect taste regulation as proposed for chrysomelid beetles. The PAs 37 and Senecionine were not toxic to *S. littoralis*. Tertiary PAs are deleterious to organisms with a microsomal cytochrome P450 system, but *S. littoralis* larvae can clearly tolerate PAs. These larvae prevent PA poisoning by rapid and efficient excretion of the absorbed tertiary alkaloid. In contrast, compound Integerrimine was moderately toxic to *L. decemlineata*, but Senecionine was not. A previous experiment has shown that ten other PAs were not toxic to this insect. Oreina beetles [Chrysomelidae] are able to take up plant alkaloid N-oxides and eliminate tertiary PAs. Similarly, *L. decemlineata* beetles could eliminate tertiary PAs efficiently enough to avoid poisoning, with some exceptions [such as Senecionine] [M. Reina et al 2001].

4.6. Cytotoxicity towards the Human Hepatoma Cell Line Huh-7.

A number of traditional remedies used in South Africa contain PAs, some of which are hepatotoxic [V. Steenkamp, et al 2001]. Investigated the effect on human Huh-7 cells of *S. latifolius*, a plant that is a component of some traditional remedies and known to contain toxic PAs. The cells were also treated with solutions of the standard pyrrolizidine retrorsine. The changes in the gross morphology of the cells were studied using light microscopy after haematoxylin and eosin staining. The cytoskeleton was investigated using fluorescence-labelled anti- β -tubulin antibody and the nuclear organisation was studied using fluorescence-labelled antinuclear antibodies. The plant extracts induced dose-dependent gross morphological changes. At high doses, necrosis was observed and at lower doses, destruction of the cytoskeleton, nuclear fragmentation, and apoptosis were detected. Doses of less than the equivalent of 330 ng/ml retrorsine led to multinucleated cells with failure in spindle formation and clumping of nuclear chromatin. This latter finding suggests that chronic low-dose treatment with such traditional remedies could induce teratogenic and/or carcinogenic effects [V. Steenkamp, et al 2001].

4.7. Antimitotic Effects. the antimitotic activity of PAs extracted from *S. brasiliensis*, stored for more than 23 years under variable conditions of temperature and humidity and exposed to light. Both the crude alkaloid [containing Integerrimine, Retrorsine, and impurities] and pure integerrimine conserved the ability to induce acute toxicity in mice, leading to the death of the animals in less than 24 h. The alkaloids also conserved the potential

to induce significant increases in micronucleus frequencies in polychromatic erythrocytes of mouse bone marrow, compared to the negative control. The administration of alkaloids to lymphocyte cultures blocked with cytochalasin-B showed no significant increase in the micronucleus frequency in binucleated cells, probably due to the lack of a metabolic activation mechanism. However, an antimitotic effect was observed [R. Santos-Mello, et al 2002].

4.8. Angiotensin-Converting Enzyme Inhibitory Activity.

Extracts of *S. samnitum* and the derived methyl ester of chlorogenic acid have been shown to inhibit the angiotensin-converting enzyme [ACE] by using an in vitro bioassay based on the enzymatic cleavage of the chromophore-fluorophore labeled substrate dansyltriglycine into dansylglycine, which was quantitatively measured by HPLC. The most effective fraction of the *S. samnitum* extract, obtained in AcOEt, inhibited the activity of ACE to 52.56_0.23% [SD] at 300 mg/ml. The major constituent of this fraction, Chlorogenic acid methyl ester, showed a significant ACE inhibition of 56.78_0.25% at a concentration of 82.5 mg/ml. The ACE inhibitory property of different extracts of *S. samnitum* may imply that in vivo, these extracts may have a hypotensive effect [R. Tundis et al 2005].

4.9. Insecticidal, Neurotoxic, and Glutathione-Depleting Activities.

The active insecticidal component of *S. palmatus* was identified as jacaranone, a neurotoxicant and glutathione-reactive quinol previously known to have insect antifeedant activity. Further, it was observed that mono- and bisglutathione [GSH] adducts were formed on incubation of Jacaranone with GSH and rat liver GSH S-transferase. The toxic action of Jacaranone in mice [intraperitoneal LD50/4150– 200 mg/kg] was associated with both neurological signs and GSH depletion in the liver 90 min after the treatment. The neurotoxic effect [tail raising, tremors, lachrymation, and ataxia] led to death in 30–90 min [H. Xu et al 2003].

4.10. Phytotoxic activity

The cacalolides, eremophilolides of *Senecio madagascariensis*, *S. barba-johannis*, *S. toluccanus* demonstrated potential phytotoxic effects on *L. sativa* [E. Burgueno-Tapia et al 2004].

4.11. Anti-proliferative activity

Effect of *S. palmatus* extract on cell proliferation and differentiation was studied performed to investigate the effect of intermedeol on proliferation and differentiation of human leukemia-derived HL-60 cells as well as the underlying mechanisms for these effects. Intermedeol exhibited a potent antiproliferative activity against HL-60 cells. In addition, this compound was found to be a potent inducer for HL-60 cell differentiation as assessed by nitroblue tetrazolium reduction test, esterase activity assay, phagocytic activity assay, morphology change, and expression of CD14 and CD66b surface antigens.

These results suggest that intermedeol induces differentiation of human leukemia cells to granulocytes and monocytes/macrophage lineage. Moreover, the expression level of c-myc was down-regulated during intermedeol-dependent HL-60 cell differentiation, whereas p21 [CIP1] was up-regulated. Taken together, our results suggest that intermedeol may have potential as a therapeutic agent in human leukemia [S. H. Jeong, et al 2002].

4.12. Antiviral activity

S. tsoongianus is a widely distributed Chinese medicinal herb traditionally used for the treatment of hepatitis B, dermatosis and inflammation. The sesquiterpene lactone isolated from it showed suppressive activity on the expression of hepatitis B virus surface antigen [HBsAg] and hepatitis B virus e antigen [HBeAg] in the HepG 2.2.15 cell lines. The sesquiterpenes also decreased the number of infectious virions released without having effect on intracellular hepatitis B virus DNA. These results suggest that enantiomeric sesquiterpene lactones may possess the potential to work synergistically with other antiviral compounds for the treatment of HBV infection [L. H. Zalkow, et al 1985].

4.13. Antioxidant activity

The antioxidant properties of the methanol, n-hexane and ethyl acetate extracts from *Senecio stabianus* Lacaita were studied. The antioxidant activities were carried out using two different in vitro assays, namely 2,2-diphenyl-1-picrylhydrazyl [DPPH] test and 2, 2-azinobis-[3-ethylbenzthiazoline-6-sulphonate] [ABTS] test. The ethyl acetate extract showed the highest activity with inhibitory concentration 50% [IC_{50}] values of 35.5 and 32.7 mg/mL on DPPH test and ABTS test, respectively [E. Burgueno-Tapia, et al 2006]. Methanolic and ethyl acetate extract of *Senecio gibbosus* demonstrated good radical scavenging activity as evaluated by the 2,2-diphenyl-1-picrylhydrazyl [DPPH] test with inhibitory concentration 50% [IC_{50}] values of 0.02 and 0.01 mg/mL, respectively.

The ethyl acetate extract of *S. inaequidens* showed better antioxidant activity than extract of *S. vulgaris* at the concentration of 0.312 mg/mL. The ethyl acetate extract of *S. inaequidens* inhibited 61.60% of the DPPH free radical whereas the extract of *S. vulgaris* showed 44.57% inhibition [F. Conforti, et al 2006a, R. Tundis, et al 2007].

Total phenolic content and total flavonoid content in alcoholic extract of aerial parts of *Senecio tenuifolus burm* was determined in addition to antioxidant activity. The extract with highest amount of phenolic compounds exhibited the maximum antioxidant activity [S. Aparna, et al 2013].

The antioxidant activity of extract of *Senecio splendens* was determined using ABTS method and the chemical structures present were identified as 5-O-caffeoylequinic acid [5-CQA], 3,4-di-O-

caffeoylequinic acid [3,4-DCQA], 3,5-di-O-caffeoylequinic acid [3,5-DCQA] and 4,5-di-O-caffeoylequinic acid [4,5-DCQA]. 3, 5-DCQA being the most abundant compound among the four isomers. The evaluation of radical scavenging activity of each isomer revealed that 3,4-DCQA possessed best antioxidant activity while 3,5-DCQA accounted for the highest radical scavenging capacity owing to it being present in the highest amount [Y. F. Shang, et al 2010].

4.14. Anti-gout Activity

Leaf extract of *Senecio splendens* is efficacious against collagen-induced arthritis [CIA] in mice. Also, the levels of rheumatoid factor, anti-type II collagen antibody, tumor necrosis factor-alpha, interleukin-1, and interleukin-6 in serum were reduced by its administration. The study suggested that leaf extract of might be effective for the treatment of inflammatory arthritis like human rheumatoid arthritis [E. M. Choi, et al 2008].

4.15. Anti-hepatotoxic activity

Hepatotoxicity produced by carbon tetrachloride, D-galactosamine, alpha-naphthylisothiocyanate and DL-ethionine was inhibited in rats by pretreatment with methanolic extract of *Senecio splendens* [Compositae]. Ethyl acetate extract fractionated from the methanolic extract showed a strong inhibitory effect. 3, 4-dicaffeoylquinc acid [DCQA], isolated from the methanolic extract was examined for its anti-hepatotoxicity. Pretreatment with DCQA [5 and 10 mg/kg, p.o.] significantly reduced serum aminotransferases [alanine and aspartate], sorbitol dehydrogenase, gamma-glutamyltransferase, alkaline phosphatase, and lactate dehydrogenase activities during carbon tetrachloride or galactosamine induced hepatotoxicity, suggesting that DCQA is responsible for the antihepatotoxic activity of *Ligularia fischeri*. DCQA also partially restored bile flow and reduced total bilirubin and cholic acid concentrations in rats. Treatment with DCQA inhibited the increase in triglyceride, cholesterol, and total lipids in DL-ethionine-induced fatty liver. The study justifies the traditional use of this plant in the treatment of jaundice and hepatic failure [E. M. Choi, et al 2008].

CONCLUSION

The genus *Senecio* [Asteraceaca], widely distributed throughout the world, is known to be a source of PAs, eremophilolides and furanoeremophilanes [M. Reina, et al 2001]. PAs are widespread in this genus. They are responsible for the hepatotoxic and carcinogenic effects of this genus [D.-L. Cheng, et al 1992]. These sesquiterpenoids with eremophilane, cacialol, bisabolane, eudesmane, oplopnone, and germacrane skeletons are among the most studied sesquiterpenoids isolated from *Senecio* species. Other types of sesquiterpenoids as phomalaIRDanes, caryophyllanes, humulanes, presilphiperfolenes, aromadendranes, himachalanes, africananes, pentalenanes, bakkanes, and

valerenes, along with diterpenoids, triterpenoids, steroids, and flavonoids, are also important components. The present review compiles the majority of the components isolated from *Senecio* species and summarizes the biological activities of this genus.

REFERENCES

- B. Nordenstam, V.H. Heywood, J. B. Harborne and B.L. Turner, Academic. In *The Biology and Chemistry of the Compositae*, eds: Press, London, England, 1977; 799-830.
- E. Burgueno-Tapia, L. R. Hernandez, A. Y. Resendiz-Villalobos, P. Joseph-Nathan, Magn. RESON Chem, 2004; 42: 887.
- G. B. Hammond, I. D. Fernandez, L. F. Villegas and A.J. Vaisberg, J. Ethnopharmacol, 1998; 61: 17-30.
- E. Uzun, G. Sariyar, A. Adsersen, B. Karakoc, G. Otuk, E. Oktayoglu and S. Pirildar, J. Ethnopharmacol, 2004; 95: 287-296.
- F. Bohlmann, J. Jakupovic, U. Warning, M. Grenz, T. V. Chau-Thi, R. M. King and H. Robinson, Bulletin des Societes Chimiques Beiges, 1986; 95: 707.
- W. H. Heywood, C. J. Humphries, *the Biology and Chemistry of the Compositae*. Academic Press, London, 1977.
- D. J. Robins, Fortschritte der Chemie Organischer Naturstoffe, 1982; 41: 115.
- F. Bohlmann, C. Zdero, J. Jakupovic, L. N. Misra, S. Benarjee, P. Singh, R. N. Baruah, M. A. Metwally, G. Schmeda-Hirschmann, L. P. D. Vincent, R. M. King and H. Robinson, Phytochemistry, 1985; 24: 1249.
- S. Dupre, M. Grenz, J. Jakupovic, F. Bohlmann, H.M. Niemeyer, Phytochemistry, 1991; 30: 1211-1220.
- C. Dong-Liang, C. Xiao-Ping, C. Jie-Kai, E. Roeder, Phytochemistry, 1992; 32: 151-153.
- P. Torres, J. Ayala, C. Grande, M. J. Macias, M. Grande, Phytochemistry, 1998; 47(1): 57-61.
- Y. Zhao, L. Wang, C. Yu-Fang and H. Man-Li, Chemistry & Biodiversity, 2011; 8(1): 13-72.
- B. E. Juarez, M. E. Mendiondo and P. Seeligmann, Biochemical Systematics and Ecology, 1995; 23(3): 335-6.
- P. Torres, C. Grande, J. Anaya, M. Grande, Phytochemistry, 1997; 52: 1507-1513.
- E. M. Suleimenov, R. A. Jose, S. B. Rakhamadieva, W. Borggraeve, W. L. Dehaen, L. Susaga, M. Parveza, S. Mathengeb and M. H. Benna, Phytochemistry, 2000; 54: 933-935.
- H. Zhong-Mei, H. Ying, S. Jia-Ming and Feng-Yan, Yingyong Huaxue, 2010; 27(12): 1486-1488.
- T. Dao-peng, C. Gui-xin and W. Zheng-tao, Biochemical Systematic and Ecology, 2010; 38(1): 122-124.
- A. B. Pomilio and E. A. Jarres, Int J Pharmacogn 1997; 35: 207-211.
- E. A. Jares, M. C. Tettamanzi and A. B. Pomilio, Phytochemistry, 1990; 29: 340-341.
- M. C. Tettamanzi, E. A. Jares and A. B. Pomilio, Fitoterapia, 1992; 63: 551-552.
- J. A. Paiva, L. E. S. Barata and J. R. Trigo, Biochem Syst Ecol, 2004; 32: 1219-1222.
- C. M. Silva, A. A. Bolzan and B. M. Heinzmann, Quim Nova, 2006; 29: 1047-1053.
- The Encyclopedia of Traditional Chinese Medicine, Peoples Hygeian Press, Shanghai, 1977; 1323.
- A. White, Hierbas de Ecuador, Plantas Medicinales. ZIKR Publication, Imprenta Mariscal, Quito [Ecuador], 1976.
- The Encyclopedia of Traditional Chinese Medicine Science and Technology Press, Shanghai, 1977; 2955.
- J. Bautista Peres, G. Stubing and R. Figuerola, Guia de las plantas medicinales de la comunitat valenciana. Las Provincias, Valencia, 1991.
- S. Abdo, M. D. Bernardi, G. Marinoni, G. Mellerio, S. Samaniego, G. Vidari and P. V. Finzi, Phytochemistry, 1992; 31: 3937.
- L. A. Loyola, S. Pedreros, G. Morales, Phytochemistry, 1985; 24: 1600-1602.
- A. A. Bolzan, C. M. Silva, L. N. Francescato, A. L. Murari, G. N. S. Silva, C. G. Heldwein, B. Heinzmann, Planta Med, 2007; 62: 427-430.
- C. K. Parikh, Textbook of medical jurisprudence and toxicology. India: CBS Publisher, 1999.
- D. S. Bhakuni and S. Gupta, Planta Medica, 1982; 46: 251.
- P. Torresa, J. Ayala, C. Grande, J. Anaya and M. Grande, Phytochemistry, 1999; 52: 1507-1513.
- G. R. Ckera, D. Mannsa, E. P. Schenkelb, R. Hartmann and B. M. Heinzmann, Phytochemistry, 1999; 52: 1587-1591.
- J. I. D. Graw and W. A. Bonner, J. Org. Chem, 1962; 27: 3917.
- F. Bohlmann, and M. Grenz, Chem. Ber, 1970; 103: 90.
- S. Valverde, T. G. D. Quezada and B. Rodriguez, Phytochemistry, 1972; 11: 446.
- F. Bohlmann, C. Zdero, J. Jakupovic, M. Grenz, V. Castro, R. M. King, H. Robinson and P. D. V. Leszek, Phytochemistry, 1986; 25: 1151-1159.
- F. Bohlmann, K. H. Knoll, C. Zdero, P. K. Mahanta, M. Grenz, A. Suwita, D. Ehlers, N. Le Van, W. R. Abraham, A. A. Natu, Phytochemistry, 1977; 16: 965.
- J. Harmatha, Z. Samek, L. Novotny, V. Herout, F. Soma, Collection of Czechoslovak Chemical Communications, 1969; 34: 1739.
- Z. Samek, J. Harmatha, L. Novotny and F. Sorm, Collection of Czechoslovak Chemical Communications, 1969; 34: 2792.
- A. H. Mericli, F. Mericli, J. Jakupovic, F. Bohlmann, X. A. Dominguez and H. S. Vega, Phytochemistry, 1989; 28: 1149.
- F. Bohlmann and C. Zdero, Chemische Berichte, 1976; 109: 819.
- F. M. Panizo and B. Rodriguez, Anales de Quimica, 1974; 70: 1043.

44. F. Fohlmann, and J. Ziesche, Phytochemistry, 1980; 19: 2681-2684.
45. L. H. Zalkow, R. N. Harris and D. Van Derveer, Chem. Comm, 1978; 420.
46. L. H. Zalkow, R. N. Harris, D. Van Derveer, and J. A. Bertrand, Chem Comm, 1977; 456.
47. F. Bohlmann, N. Le Van, T. Cuong Pham, A. Schuster, V. Zabel and W. H. Waston, Phytochemistry, 1979; 18: 1831.
48. B. Kamthory and A. Roobertson, J. Chem. Ber., 1939; 103: 90.
49. T. Murae, Y. Tanahashashi and T. Takahashi, Tetrahedron, 1968; 24: 2177.
50. F. Bohlmann and B. Grenz, Chem. Comm, 1978; 420.
51. K. Hayashi, H. Nakamura and H. Mitsuhashi, Phytochemistry, 1973; 12: 2931.
52. F. Bohlmann and C. C. Zdero, Chem. Ber., 1978; 111: 3140.
53. F. Bohlmann, and C. Zdero, Phytochemistry, 1978; 17: 565.
54. P. M. Brown and R. H. Thomson, J. Chem. Soc., 1969; 1184.
55. H. Kakisawa and Y. Inouye, Tetrahedron Letters, 1969; 1929.
56. T. Takemoto, G. Kusano, K. Aota, M. Kaneshina and N. A. ElEmary, 1974 Yakugaku Zasshi, 94, 1593. C. A., 1975; 82: 135676.
57. F. Bohlmann, V. Castro, C. Zdero, R. M. King and H. Robinson, Rev. Latinoam. Quim, 1984; 14: 101.
58. D. J. Roy and Mukhopadhyay, Indian J. Gem., 1981; 20: 628.
59. J. D. P. Teresa, I. S. Bellido, M. S. Gonzales and S. Vincente, Phytochemistry, 1986; 25: 185.
60. J. D. P. Teresa, A. S. Feliciano, A. F. Barrero and M. Medarde, a Quim, 1979; 75: 422.
61. I. Wahlberg, M. B. Hjelte, K. Karlsson and C. R. Enzell, Acta Chem. Stand, 1971, 25, 3285.
62. A. Gonzales, B. M. Fraga, P. Gonzales, M. G. Hernandez and A. G. Ravelo, Phytochemistry, 1981; 20: 1919.
63. T. D. Hubert and D. F. Weimer, Phytochemistry, 1985; 24: 1197.
64. E. Roeder and K. Ltut, Phytochemistry, 1991; 30: 1734-1737.
65. H. L. Zalkow, C. F. Asibal, J. A. Glinski, S. J. Bonetti, L. T. Gelbaum, D. Van Derveer and G. Powis, J. Nat. Prod, 1988; 51: 690.
66. L. B. Bull, C. C. J. Culvenor, A. T. Dick, The Pyrolizidine Alkaloids, North Holland Amsterdam, 1968; 82-86.
67. N. H. Amsterdam and L. A. Wunderlich, Chem. Ind., 1962; 2089.
68. C. C. J. Culvenor, G. M. Donovan and L. W. Smith, Aust. J. Chem, 1967; 20: 801.
69. K. Lru and E. Roder, Phytochemistry, 1991; 30: 1303-1305.
70. X. T. Liang and E. Roder, Planta Med, 1979; 51: 362.
71. G. G. Habermehl, W. Martz, C. H. Tokarnia, J. Doeberleiner and M. C. Mendez, Toxicol, 1988; 26: 275.
72. E. Roder, H. Wiedenfeld and R. Kersten, Sci. Pharm, 1990; 58: 1.
73. E. Roeder and T. Bourauel, Nat. Toxins, 1992; 1: 35.
74. K. Liu and E. Roder, Phytochemistry, 1991; 30: 1303.
75. E. Roeder, T. Bourauel and R. Kersten, Phytochemistry, 1993; 32: 1051-1053.
76. H. J. Segall and J. L. Dallas, Phytochemistry, 1983; 22: 1271.
77. N. Mengi, S. C. Taneja, V. P. Mahajan and C. S. Mathela, Phytochemistry, 1991; 30: 2329- 2330.
78. K. L. Handa, I. C. Chopra, and S. N. Sobti, J. Sci. Ind. Res., 1957; 16: 22.
79. M. H. Benn, S. Mathenge, R. M. Munavu, S. O. Were, Phytochemistry, 1995; 40: 1327-1329.
80. E. Roder, H. Wiedenfeld, A. Pfizer, Phytochemistry, 1988; 27: 4000-4001.
81. D. L. Cheng, X. I. Cao, J. K. Cheng and E. Roeder, Phytochemistry, 1993; 32: 151-153.
82. F. H. Guidugli, M. H. Pestchankar, M. S. A. Desselmeron and O. S. Giordano, Phytochemistry, 1986; 25: 1923-1926.
83. M. J. Pestchankar, M. S. Ascheri and O. S. Giordano, O.S., Planta Med., 1985; 2: 165.
84. Pestchankar, M.J., Ascheri, M.S., Giordano, O.S., Planta Med, 1985; 2: 1622.
85. Pestchankar, M.J., Ascheri, M.S., Giordano, O.S., J. Nat. Prod. [in press], 1986.
86. M. S. Ascheri and O. S. Giordano, An. Asoc. Quim.Argentina, 1980; 68: 105.
87. M. S. A. Salmeron, J. Kavka and O. S. Giordano, Planta Med., 1983; 47: 221.
88. J. D. Urones, P. B. Barcala, I. S. Marcos, R. S. Moro, M. L. Esteban and A. F. Rodriguez, Phytochemstry, 1988; 27: 1507-1510.
89. R. M. A. Mansour and N. A. M. Salem, Phytochemstry, 1981; 20: 1180.
90. P. Teresa, J. D. Urones, J. G. Basabe, P. S. Marcos, I. Fernandez, R. S. Moro and M. J. Cuadrado, Phytochemistry [In press], 1988.
91. S. Ferry, Planta Med Phytother, 1977: 11- 25.
92. S. Ferry and I. L. Brazier, Ann. Pharma, 1976; 34: 133.
93. E. Roeder, Phytochemistry, 1990; 29: 11.
94. L. Witte, L. Ernst, V. Wray and T. Hartmann, Phytochemistry, 1992; 31: 1027 -1028.
95. C. A. Ray, J. H. Williams and C. J. Reagor, Phytochemistry, 1987; 26: 2431-2433.
96. R. Adams and T. R. Govindachari, J. Am. Chem. Soc., 1949; 71: 1180.
97. H. J. Segall and R. J. Molyneux, Res. Comm. Gem.Path. Pharmacol, 1978; 3: 545.
98. R. Adams and D. B. L. Van, J. Ant. Chem. Soc, 1953; 75: 4631.
99. F. Bohlmann and C. Zdero, Phytochemistry, 1977; 16: 135.

- 100.Y. Zhao, Z. J. Jia and L. Yang, Phytochemistry, 1994; 37: 1149-1152.
- 101.Z. J. Jia and H. M. Chen, Phytochemistry, 1991; 30: 3132-3134.
- 102.C. M. Silva, A. A. Bolzan, C. A. Mallmann, P. Pozzatti, S. H. Alves and B. M. Heinzmann, Brazilian Journal of Pharmacognosy, 2010; 20: 87-92.
- 103.M. Noorwala, F. V. Mohammada, V. U. Ahmad, U. B. Senerb, F. Ergunb and D. Deliorman, Fitoterapia 2000; 71: 618-620.
- 104.R. J. Molyneux, J. N. Roitman, M. Benson and R. E. Lundin, Phytochemistry, 1982; 21: 439.
- 105.J. D. P. Teresa, I. S. Bellido, V. J. R. Salado, F. Moliner and M. R. Alberdi, Riv Ital EPOS, 1980; 62: 236.
- 106.P. Torres, R. Chinchilla and M. Grande, Studia Chemica, 1992, XVII: 53.
- 107.J. S. Pyrek and E. Baranowska, Tetrahedron Lett, 1973; 809.
- 108.P. Torresb, C. Grandea, J. Anayaa and M. Grande, Fitoterapia, 2000; 71: 91-93.
- 109.F. Bohlmann, C. Zdero and M. Grenz, Chem Ber, 1974; 107: 3928.
- 110.G. Ourisson and J. M. Lehn, Bull Soc Chim Fr, 1962; 1137.
- 111.E. Wenkert, G. V. Baddeley, I. R. Burfitt and L. N. Moreno, Org Magnetic Resonance, 1978; 11: 337.
- 112.A. Chaterjee, R. Mukherjee, S. K. Srimany and S. J. Bhattacharjee, Ind Chem. Soc, 1966; 63.
- 113.M. Reina, M. Nold, O. Santana, J. C. Orihuela and A. G. Coloma, J. Nat. Prod., 2002; 65: 448-453.
- 114.G. Lin, P. Rose, K. B. Chatson, E. M. Hawes, X. G. Zhao and Z. T. Wang, Journal of Natural Products, 2000; 63: 857-860.
- 115.G. Schmeda, Hirschmann, E. A. Ferro, L. Franco, L. Recalde, and C. Theoduloz, Journal of Natural Products, 1987; 50: 770-772.
- 116.A. A. Ahmed, Journal of Nature product, 1991; 54: 271.
- 117.N. Kahriman, G. Tosun, S. Terzioglu, S. A. Karaoglu and N. Yayli, Rec. Nat. Prod., 2011; 5: 82-91.
- 118.M. Manubolu, L. Goodla, S. Ravilla, V. R. Obulum, Asian Pac J Trop Biomed, 2013; 3: 191-195.
- 119.L. Arancibia, C. Naspic, G. Pucci and M. Arce, Bol. Latinoam Caribe Plant Med Aromát, 2013; 12: 18-23.
- 120.Zellagui, S. Rhouati, C. Joel, T. Gabor, A. A. Ahmed and W. P. Paul, Rev. Latinoamer. Quim, 2004; 32: 376-381.
- 121.Z. Amarl, T. Soukaina, G. Noureddine and R. Salah, Der Pharma Chemica, 2012; 4: 2080.
- 122.Y. Zhao, Z. J. Jia and H. Peng, Journal of Nature product, 1995; 58: 1358-1364.
- 123.Z. J. Jia and Y. Zhao, J. Nat. Prod., 1994; 57: 146.
- 124.Y. Moriyama and T. Takahashi, Bull. Chem. Soc. Japan, 1976; 49: 3196.
- 125.C. Perez, A. M. Agnese and J. L. Cabrera, J. Ethnopharmacol, 1999; 66: 91-96.
- 126.L. Villarroel and R. Torres, Rev. Latinoam Quim, 1987; 18: 73.
- 127.L. Villarroel and R. Torres, Journal of Natural Product, 1991; 54: 588-590.
- 128.V. Christov, N. Kostova, and L. Evstatieva, Natural Product Research, 2005; 19: 389-392.
- 129.L. Witte, P. Rubiolo, C. Bicchi and T. Hartmann, Phytochemistry, 1993; 32: 187.
- 130.V. Christov, M. Simeonov, and L. Evstatieva Compt. Rend. Acad. Bulg. Sci., 1997, 50, 47.
- 131.T. Hartmann, L. Witte, A. Ehmke, C. Theuring, M. R. Rahier and J. Pasteels, Phytochemistry, 1997; 45: 489.
- 132.L. Villarroel and R. Torres, Journal of Natural Products, 1985; 48: 841-842.
- 133.T. J. Mabry, K. R. Markham and M. B. Thomas, "The Systematic Identification of Flavonoids," Springer, New York, 1970.
- 134.K. R. Markham, B. Ternai, R. Stanley, H. Geiger, T. J. Mabry, Tetrahedron, 1978; 34: 1389.
- 135.C. A. Buxhi, and A. B. Pomilio, Jou. Nat. Prod., 1982; 45: 557.
- 136.C. K. Atal, K. K. Kapur, C. C. J. Culvenor and L. W. Smith, Tetrahedron Letter 1966, 537.
- 137.C. C. J. Culvenor, N. I. Koretskaya, L. W. Smith, L. M. Utkin, Awt. J. Cbem, 1968; 21: 1671.
- 138.M. J. Pestchanker, M. S. Ascheri and O. S. Giordano, Phytochemistry, 1985; 24: 1622.
- 139.E. Jares and A. B. Pomilio, Journal of Natural Products, 1986; 50(5): 14.
- 140.A. L. Perez-Castorena, A. Arciniegas, R. P. H. Gutierrez, R. A. Toscano, J. L. Villasenor, and A. R. D. Vivar, J. Nat. Prod., 1999; 62: 1039-1043.
- 141.P. Castorena, A. L. Arciniegas, A. Perez, A. R. Villasenor, J. L. R. de Vivar, J. Nat. Prod., 1998; 61: 1288-1291.
- 142.R. D. Vivar, A. Perez, A. L. Arciniegas, A. Vidales, P. Gavino and R. Villasenor, Tetrahedron, 1995; 51: 12521-12528.
- 143.O. Uçuncu, N. Yayli, A. Yasar, S. Terzioglu and N. Yayli, Nat. Prod. Com., 2008; 3: 925-928.
- 144.D. Nori-Shargh, S. Raftari and F. Deyhimi, Flav. Fragr. J., 2008; 23: 357-359.
- 145.A. Usta, O. Uçuncu, T. B. Cansu, S. Terzioglu and N. Yayli, Asian J. Chem., 2009; 21: 6369-6374.
- 146.H. L. D. Pooter, L. F. D. Buyck, N. M. Schamp, E. Aboutabi, A. D. Bruyn and S. Z. Husain, Flav. Fragr. J., 2006; 1: 159-163.
- 147.R. R. Vera, S. J. Laurent and D. J. Fraisse, J. Essent. Oil Res, 1994; 6: 21-25.
- 148.N. Mengi, S. N. Garg, S. K. Agarwal and C. S. Mathela, J. Essent. Oil Res., 1995; 7: 511-514.
- 149.A. M. El-Shazly, Zagazig J. Pharm. Sci., 1999; 8: 1-8.
- 150.V. T. Balzaretti, A. Arancibia, A. M. E. Marchiaro and M. S. Feijoo, Molecules, 2000; 5: 459-461.
- 151.M. Mirza and N. Z. Baher, J. Essent. Oil Bear. Plant, 2008; 11: 179-183.
- 152.A. G. Belaunde, J. G. Sandoval, L. De Martino, F. Senatore and V. De Feo, J. Essent. Oil Bearing Plants 2007; 10: 332-338.

- 153.G. F. Zuniga, I. F. Valderrama, G. B. Hammond, Rev. Latinoam.Quim, 1996; 25: 14-16.
- 154.J. C. Chalchat, Z. A. Maksimovic, S. D. Petrovic andM. S. Gorunovic, J. Essent. Oil Res., 2004; 16: 227-228.
- 155.K. H. C. Baser andB. Demirci, J. Essent. Oil Res., 2004; 16: 558-559.
- 156.V. D. Feo, E. U. Soria, R. U. Soria and F. Senatore, Flav. Fragr. J., 2003; 18: 234-236.
- 157.M. Rondon, M. Araqe, A. Morales, M. Gualtieri, J. Rojas, K. Veres andI. Mathe, Nat. Prod. Com, 2006; 1: 113-115.
- 158.D. R. Gardner, M. R. Thorne, R. J. Molyneux, J. A. Pfister andA. A. Seawright, Biochemical Systematics and Ecology, 2006; 34: 736-744.
- 159.E. B. Tapia, A. G. Coloma, D. M. Benito andP. J. Nathan, Z. Naturforsch, 2007; 62: 362-366.
- 160.E. Burgueno- Tapia,M. A. Bucio, A. Rivera and P. Joseph- Nathan, J. Nat. Prod, 2001; 64: 518-521.
- 161.E. Burgueno- Tapia,M. A. Bucio,A. Rivera, E. Joseph-Burgueno-Tapia, L. R. Hernandez,A. Y. Resendiz-Villalobos,P. Joseph-Nathan,Magn. Reson. Chem, 2004; 42: 887-892.
- 162.E. Burgueno-Tapia,B. Hernandez-Carlos,P. Joseph- Nathan,J. Mol. Struct., 2006; 825: 115-123.
- 163.A.-L.Pe'rez-Castorena, A. Arciniegas, J. L. Villasenor, A. Romo de Vivar, Biochem. Syst. Ecol, 1999; 27: 835.
- 164.A.-L.Pe'rez-Castorena, A. Arciniegas, J. L. Villasenor, A. Romo de Vivar, Biochem. Syst. Ecol, 1999; 27; 835.
- 165.J. R. Liddell, C. G. Logie, Phytochemistry, 1993; 34: 1198.
- 166.M. R. Grue, J. R. Liddell, Phytochemistry, 1993; 33: 1517.
- 167.J. R. Liddell, C. G. Logie, Phytochemistry, 1993; 34; 1198.
- 168.E. Roeder, H. Wiedenfeld, R. Britz-Kirstgen, Phytochemistry, 1984; 23: 1761.
- 169.D.-L.Cheng, J.-K. Niu, E. Roeder, Phytochemistry, 1992; 31: 3671.
- 170.F. Bohlmann, C. Zdero, D. Berger, A. Suwita, P. Mahanta, C. Jeffrey, Phytochemistry, 1979; 18: 79.
- 171.O. Were, M. Benn, R. M. Munavu, Phytochemistry, 1993; 32: 1595.
- 172.A.-L. Pe'rez-Castorena, A. Arciniegas,A. Castro, J. L. Villasenor, R. A. Toscano,A. Romo de Vivar,J. Nat. Prod, 1997; 60: 1322.
- 173.H. L. DeWaal, Onderstepoort J. Vet.Sci. Anim. Ind, 1940; 15: 241.
- 174.O. Were, M. Benn, R. M. Munavu, Phytochemistry, 1993; 32: 1595.
- 175.O. Were, M. Benn, R. M. Munavu, Phytochemistry, 1993; 32: 1595.
- 176.A. Romo De Vivar, A.-L. Perez, P. Vidales, D. A. Nieto, J. L. Villasenor, Biochem. Syst. Ecol, 1996; 24: 175.
- 177.A.-L.Pe'rez-Castorena, A. Arciniegas, R. Pe'rez Alonso, J. L. Villasenor, A. Romo de Vivar, J. Nat. Prod, 1998; 61: 1288.
- 178.H. C. Krebs, T. Carl, G. G. Habermehl, Phytochemistry, 1996; 43: 1227.
- 179.D. R. Gardner, M. S. Thorne, R. J. Molyneux, J. A. Pfister, A. A. Seawright, Biochem. Syst. Ecol, 2006; 34: 736.
- 180.A.-L. Pe'rez-Castorena, A. Arciniegas, R. Perez, H. Gutierrez, R. A. Toscano, J. L. Villasenor, A.Romo de Vivar, J. Nat. Prod, 1999; 62: 1039.
- 181.A. R. D. Vivar, A. L. Perez, A. Arciniegas, P. Vidales, R. Gavino, J. L. Villasenor, Tetrahedron, 1995; 51: 12521.
- 182.J. R. Liddell, C. G. Logie, Phytochemistry, 1993; 34: 1629.
- 183.F. Bohlmann, C. Zdero, M. Grenz, Chem. Ber, 1977; 110: 474.
- 184.J. R. Liddell, C. G. Logie, Phytochemistry, 1993; 34: 1629.
- 185.B. Wu, W. H. Lin, H. Y. Gao, L. Zheng, L. J. Wu, C. S. Kim, Pharm. Biol, 2006; 44: 440.
- 186.F. H. Guidugli, M. J. Pestchanker, M. S. A. De Salmeron, O. S. Giordano, Phytochemistry, 1986; 25: 1923.
- 187.J. M. Cardoso, J. Jakupovic, F. Bohlmann, Phytochemistry, 1987; 26: 2321.
- 188.F. Bohlmann, A. Suwita, C. Zdero, Phytochemistry, 1978; 17: 1763.
- 189.F. Bohlmann, C. Zdero, J. Jakupovic, L. N. Misra, S. Banerjee, P. Singh, R. N. Baruah, M. A. Metwally, G. Schmeda-Hirschmann, L. P. D. Vincent, R. M. King, H. Robinson, Phytochemistry, 1985; 24: 1249.
- 190.M. Ahmed, H.-M. Niemeyer, Phytochemistry, 1991; 30: 2078.
- 191.F. Bohlmann, C. Zdero, R. M. King, H. Robinson, Phytochemistry, 1981; 20: 2389.
- 192.F. Bohlmann, J. Ziesche, Phytochemistry, 1980; 19: 1851.
- 193.S. Abdo, M. de Bernardi, G. Marinoni, G. Mellerio, S. Samaniego, G. Vidarit, P. Vita Finzit, Phytochemistry, 1992; 31: 3937.
- 194.Y. Zhao, H. Jiang, M. MacLeod, S. Parsons, D.W. H. Rankin, P. Wang, C. H. K. Cheng, H. Shi, X.Hao, F. Gue'ritte, Chem. Biodiversity, 2004; 1: 1546.
- 195.A.-L.Pe'rez-Castorena, A. Arciniegas, S. L. Guzman, J. L. Villasenor, A. Romo de Vivar, J. Nat.Prod, 2006; 69: 1471.
- 196.P. Torres, C. Grande, J. Anaya, M. Grande, Fitoterapia, 2000; 71: 91.
- 197.F. Bohlmann, S. Dupre, B. Nordenstam, Phytochemistry, 1990; 29: 3163.
- 198.F. Bohlmann, S. Dupre, B. Nordenstam, Phytochemistry, 1990; 29: 3163.
- 199.F. Bohlmann, M. Ahmed, J. Jakupovic, C. Jeffrey, Phytochemistry, 1981; 20: 251.
- 200.F. Bohlmann, K.-H. Knoll, C. Zdero, P. K. Mahanta, M. Grenz, A. Suwita, D. Ehlers, N. L. Van,W.-R. Abraham, A. A. Natu, Phytochemistry, 1977; 16: 965.
- 201.M. A. Aal, F. Bohlmann, T. Sarg, M. El-Domiatiy, B. Nordenstam, Phytochemistry, 1988; 27; 2599.

- 202.M. R. Loizzo, G. A. Statti, R. Tundis, F. Conforti, M. Bonesi, G. Autelitano, P. J. Houghton, A. Miljkovic-Brake, F. Menichini, *Phytother. Res.*, 2004; 18: 777.
- 203.B. Wu, W. H. Lin, H. Y. Gao, L. Zheng, L. J. Wu, C. S. Kim, *Pharm. Biol.*, 2006; 44: 440.
- 204.C. Perez, A. M. Agnese, J. L. Cabrera, *J. Ethnopharmacol.*, 1999; 66: 91.
- 205.E. H. H. M. Abou, A. A. Ahmed, *J. Nat. Prod.*, 2005; 68: 439.
- 206.J.-Q. Gu, Y. Wang, S. G. Franzblau, G. Montenegro, B. N. Timmermann, *J. Nat. Prod.*, 2004; 67: 1483.
- 207.W. P. Zhang, H. Q. Chen, W. S. Zhang, G. L. Cao, Z. Huang, *Li Shizhen Med. Materia Medica Res.*, 2008; 19: 605.
- 208.W. Toma, J. R. Trigo, A. C. Bensuaski de Paula, A. R. M. Souza Brito, *J. Ethnopharmacol.*, 2004; 95: 345.
- 209.A. Gonzalez-Coloma, M. Reina, R. Cabrera, P. Castan˜era, C. Gutierrez, *J. Chem. Ecol.*, 1995; 21: 1255.
- 210.A. Gonzalez-Coloma, M. Reina, R. Cabrera, P. Castan˜era, C. Gutierrez, *J. Chem. Ecol.*, 1995; 21: 1255.
- 211.M. Reina, M. Nold, O. Santana, J. C. Orihuela, A. González-Coloma, *J. Nat. Prod.*, 2002; 65: 448.
- 212.M. Reina, M. Nold, O. Santana, J. C. Orihuela, A. González-Coloma, *J. Nat. Prod.*, 2002; 65: 448.
- 213.M. Reina, A. González-Coloma, C. Gutie˜rrez, R. Cabrera, M. L. Rodr_guez, V. Fajardo, L. Villarroel, *J. Nat. Prod.*, 2001; 64: 6.
- 214.M. Reina, A. González-Coloma, C. Gutie˜rrez, R. Cabrera, M. L. Rodr_guez, V. Fajardo, L. Villarroel, *J. Nat. Prod.*, 2001; 64: 6.
- 215.M. Reina, A. González-Coloma, C. Gutie˜rrez, R. Cabrera, M. L. Rodr_guez, V. Fajardo, L. Villarroel, *J. Nat. Prod.*, 2001; 64: 6.
- 216.M. Reina, A. González-Coloma, C. Gutie˜rrez, R. Cabrera, M. L. Rodr_guez, V. Fajardo, L. Villarroel, *J. Nat. Prod.*, 2001; 64: 6.
- 217.M. Reina, A. González-Coloma, C. Gutie˜rrez, R. Cabrera, M. L. Rodr_guez, V. Fajardo, L. Villarroel, *J. Nat. Prod.*, 2001; 64: 6.
- 218.V. Steenkamp, M. J. Stewart, S. van der Merwe, M. Zuckerman, N. J. Crowther, *J. Ethnopharmacol.*, 2001; 78: 51.
- 219.V. Steenkamp, M. J. Stewart, S. van der Merwe, M. Zuckerman, N. J. Crowther, *J. Ethnopharmacol.*, 2001; 78: 51.
- 220.R. Santos-Mello, L. I. Deimling, C. M. Lauer Ju˜nior, A. Almeida, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2002; 516: 23.
- 221.R. Tundis, M. R. Loizzo, G. A. Statti, B. Deguin, R. Amissah, P. J. Houghton, F. Menichini, *Pharm. Biol.*, 2005; 43: 605.
- 222.H. Xu, N. Zhang, J. E. Casida, *J. Agric. Food Chem.*, 2003; 51: 2544.
- 223.E. Burgueno-Tapia, M. A. Bucio, A. Rivera, E. Joseph-Burgueno-Tapia, L. R. Hernandez, A. Y. Resendiz-Villalobos, P. Joseph-Nathan, Magn. Reson. Chem., 2004; 42: 887-892.
- 224.S. H. Jeong, S. J. Koo, J. H. Choi, H. J. Park, J. Ha, J. H. Park, K. T. Lee, *Planta Med.*, 2002; 68: 881-5.
- 225.L. H. Zalkow, J. A. J. Glinski, L. T. Gelbaum, T. J. Fleischmann, L. S. McGowan and M. M. Gorden, *J. Med. Chem.*, 1985; 28: 687.
- 226.E. Burgueno-Tapia, B. Hernandez-Carlos, P. Joseph-Nathan, *J. Mol. Struct.*, 2006; 825: 115-123.
- 227.F. Conforti, M. R. Loizzo, G. A. Statti, P. J. Houghton and F. Menichini, *International Journal of Food Sciences and Nutrition*, 2006; 57: 1-8.
- 228.R. Tundis, M. R. Loizzo, G. A. Statti, P. J. Houghton, A. Miljkovic-Brake, F. Menichini, *Natural Product Research*, 2007; 21: 396-400.
- 229.S. Aparna, K. N. V. Rao, A. Begami and D. Banji, *International Journal of Pharmacy and Biological Sciences*, 2013; 3: 17-23.
- 230.Y. F. Shang, S. M. Kim, D. G. Song, C. H. Pan, W. J. Lee and B. H. Um, *J. Food Sci.*, 2010; 75: C530-5.
- 231.E. M. Choi and Y. H. Kim, *Food Chem. Toxicol.*, 2008; 46: 375-379.
- 232.E. M. Choi and Y. H. Kim, *Food Chem. Toxicol.*, 2008; 46: 375-379.
- 233.M. Reina, A. González-Coloma, C. Gutie˜rrez, R. Cabrera, M. L. Rodr_guez, V. Fajardo, L. Villarroel, *J. Nat. Prod.*, 2001; 64: 6.
- 234.D.-L. Cheng, J.-K. Niu, E. Roeder, *Phytochemistry*, 1992; 31: 3671.