

# EUROPEAN JOURNAL OF PHARMACEUTICAL AND MEDICAL RESEARCH

www.ejpmr.com

Research Article
ISSN 2394-3211
EJPMR

# EX-VIVO PERMEABILITY STUDIES OF DIHYDROARTEMISININ-PIPERAQUINE ANTIMALARIAL IN THE PRESENCE OF FOOD COMPONENTS

Sunday O. Awofisayo<sup>1</sup>\*, Ekpedeme Ndem Essien<sup>1</sup> and Ayodeji Akeem Agboke<sup>2</sup>

<sup>1</sup>Department of Clinical Pharmacy and Biopharmacy, Faculty of Pharmacy, University of Uyo, Nigeria.

<sup>2</sup>Department of Pharmaceutics and Pharmaceutical Technology incorporating Pharmaceutical Microbiology, Faculty of Pharmacy, University of Uyo, Nigeria.

\*Corresponding Author: Dr. Sunday O. Awofisayo

Department of Clinical Pharmacy and Biopharmacy, Faculty of Pharmacy, University of Uyo, Nigeria.

Article Received on 25/06/2018

Article Revised on 15/07/2018

Article Accepted on 04/08/2018

#### **ABSTRACT**

The work was aimed at assessing the interactions between dihydroartemisinin-piperaquine (DP) with food components. Food components [*i.e.*, albumin (ALB), starch (STC) and sunflower oil (SFO)] were co-loaded simultaneously with DP in excised intestinal tissues (*i.e.*, duodenum and ileum) from New Zealand white albino male rabbits. Loaded tissues were submerged in 100 mL Tyrode solution (TS) in organ bath. DP equivalent to (100 mg/mL) of PQ was similarly loaded in duodenum and ileum as controls (CRT1 and CRT2), respectively. Sampling (5mL) was performed from organ bath at timed interval post immersion. Analysis was done using Chemstation high pressure chromatographic reverse-phase (HPLC-RP) system with Zorbact XDB C8 column and mobile phase acetonitrile: 10 mM ammonium acetate (70: 30, %v/v). The wavelength of UV detection was at 220 nm and flow rate 0.7 mL/min. PQ permeation kinetics was zero order in the presence of food components. SFO treatment revealed significant reduction in  $C_{max}$  in duodenum (0.2951 versus 0.9800 µg/mL) and ileum (0.6150 versus 1.2170 µg/mL), p<0.05. ALB produced no significant difference in  $P_{eff}$  disappearance from lumen and  $P_{eff}$  appearance in the Tyrode solution for duodenum but higher values for ileum (p<0.05). There was higher rate for appearance (Ka) of PQ in organ bath for STC and SFO treatments in both intestinal segments, p<0.05. Similarly, the extents of permeation at 6 h (AUC<sub>6</sub>) for the treatments were lower in duodenum than the control. ALB, SFO and STC significantly affected the rate and extent of permeation of PQ across the absorptive intestinal membrane.

**KEYWORDS:** Food components, Dihydroartemisinin-piperaquine, Intestinal membrane, Permeability, Antimalarial, Drug-drug interaction.

#### INTRODUCTION

The physiochemical properties of a drug substance, dosage form, physiological function of gastrointestinal tract along with the biochemical and physical properties of the epithetical barrier all influence the complex process of intestinal absorption. The transport of molecules across intestinal membrane can be classified mechanistically into active transport, carrier-mediated or passive transport. Passive transport comprised of simple diffusion, facilitated diffusion and endocytosis or transcytosis. The process of mass transfer across the partition wall is directed towards equilibrating any difference in chemical potential while internal or partial pressure gradient, constitute the driving forces.

Co-administered agents may influence the mechanism and velocity of passage of drugs with respect to lag time and the time period for equilibrium to be attained. Since drug administration is often linked with meal times, there may be variations in the pharmacokinetic disposition of orally administered drugs basically influenced by the quantity and type of meal ingested. [4]

Dihydroartemisinin-piperaquine (DP) is an antimalarial prescribed for multi-drug resistant *Plasmodium falciparum* malaria.<sup>[5,6]</sup> It has been reported to give variable oral absorption when taken with fatty meals.<sup>[7]</sup> Meal types, composition and volume vary widely in malaria endemic boundaries. Drug-food interactions in recent times have been of major concern because of poor clinical outcomes attendant to significant episodes.<sup>[8]</sup> It has also been noted as a source of inconvenience and non-adherence through disruptions in patients' daily schedule.<sup>[9]</sup>

The mechanisms of interactions and the consequences of different types of meals on the physiological nature of gastrointestinal medium have been reported. In real life malaria conditions, meal types ingested with antimalarial may vary widely. Furthermore, the anorexic condition associated with the episode may prevent a sufferer from ingesting a particular prescribed meal. The array of possible predisposed food usually is not extensively researched into during drug development but the pharmacokinetic Food and Drug Administration

(FDA) experimental prescribed meal. [11] The molecular mechanisms of drug-food interaction have received some concern stemming from the observed grape fruit juice warfarin.<sup>[12]</sup> It therefore interactionwith necessarv to investigate the drug specific biopharmaceutical importance of food-drug interaction on drugs of interest at post-approval and drug marketing stages. Malaria drugs are key candidates for this investigation in the light of the widely reported therapeutic hitch and poor clinical outcomes.

Awofisayo *et al.* reported on the *in vitro* drug-food interaction with DP and concluded that there was no change in the peaks and troughs due to PQ and DHA from Fourier transform infra-red spectra characteristics of dihydroartemisinin (DHA) or PQ in DP. [14] Consequently, this study was aimed at assessing the effect of the basic food components on intestinal permeability of the PQ component in DP following oral administration *post cibos*.

#### **EXPERIMENTAL**

#### **Materials and Chemicals**

PQ and tinidazole reference standard powder were kind donations from Central Research Laboratory, University of Lagos, Nigeria. High pressure liquid chromatographic (HPLC) grade acetonitrile, ammonium acetate and methanol were products of Sigma Aldrich, Germany. Magnesium chloride, calcium chloride, sodium chloride, sodium bicarbonate, potassium chloride were products of Sigma Aldrich, Germany. Glucose (Evans PLC) and P-ALAXIN® (BVS, India) were bought from a registered pharmacy, Lagos Nigeria. Albumin, sunflower oil and soluble starch powder were purchased from a local distributor in Lagos, Nigeria.

# Preparation of Standard Solution/ Working Solutions/ Physiologic solutions

Stock solution of PQ was prepared by dissolving 50 mg accurately weighed reference standard powder in 10 mL volumetric flask. Serial dilutions were made to produce graded concentrations of working solutions in the range 1-100 mg/mL. Tyrode solution (TS) was prepared according to the method of Awofisayo and coworkers. [13]

# Preparation of admixture of drugs with food components

The admixture of DP suspension with the respective food components were prepared from the volume of DP suspension required for intestinal loading equivalent to PQ at 10 mg PQ content/kg body weight of rabbit and the food components at 40 mg/kg body weight.

#### **Handling of Animals**

New Zealand albino male rabbits were employed for the experiment. Animals were fed with standard pellets, free access to water and allowed to acclimatize to the environment for one week, prior to the experiment. The protocol of this study was approved by Faculty of

Pharmacy, University of Uyo Ethics Committee on the Use of Laboratory Animals (UFP012). Good Laboratory Practice Guideline was observed.

# **Loading of Excised Tissue Segments**

The excised tissues were tied at one end with silk suture and loaded with DP (100 mg/mL) at 10 mg of PQ/kg body weight and food components (STC, ALB and SFO) at 40 mg, simultaneously.

## Sampling and Analysis of Samples

Sampling (5 mL) was taken from the organ bath at 0, 1, 2, 4 and 6 h post immersion of loaded tissues in organ bath. Chromatographic analyses of samples were performed by injecting 1  $\mu$ L of filtered sample into the system. The HPLC system comprised of C8 Zorbact XDB (150 x 4.6 mm, 4.6  $\mu$ m) column and mobile phase was acetonitrile: 10 mM ammonium acetate (70:30 %, v/v). UV detection of analyte at 220 nm and flow rate at 0.7 mL/min.

#### **Statistical Analyses**

Statistical evaluation of data was performed using statistical package for social scientists (SPSS) version 20 (IBM, USA). Paired T-test was employed to analyze for difference between treatments and control set-ups while statistical difference was taken at confidence interval of 5%.

#### **RESULTS**

The representative chromatogram for PQ analysis in HPLC is presented in Figure 1. Peaks due to PQ were shown at elution time of 4.25 min while that of the IS at 2.10 min. Peaks There were no interference with the peaks for the analyte and the IS. The concentrations of PQ versus time profile for the experimental conditions in the duodenum and ileum segments are presented in Figures 2a and 2b, respectively. The AUC measurements in Table 1 revealed information on the rate and extent of permeation under the experimental conditions. AUC<sub>2</sub> values revealed significantly higher permeation wit ALB in duodenum than other treatments (p=0.001, 0.003 and 0.001 for ALB, STC and SFO, respectively). SFO treatment had no significant difference compared with the control but higher than STC and ALB in the ileum (p=0.004 and 0.001, respectively). The control had higher overall (AUC<sub>6</sub>) permeation for the analyte than the food-based treatments in ileum but ALB revealed significantly higher value in duodenum (Table 1).

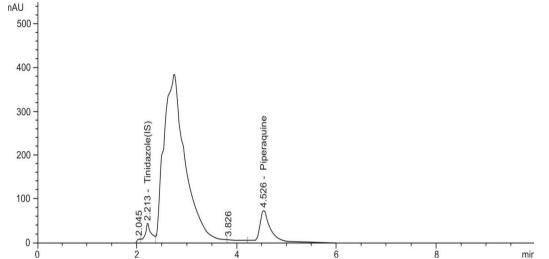


Figure 1: Representative chromatograph of sample from intestinal perfusate containing piperaquine.

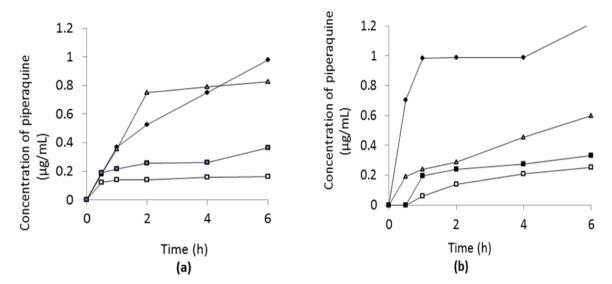


Figure 2: Piperaquine permeation across intestinal epithelia from (a) duodenum and (b) ileum, in the presence of  $\Box$  sunflower oil,  $\triangle$  albumin,  $\blacksquare$  starch and  $\blacklozenge$ DP alone.

Table 1: AUC measurements for the treatments and intestinal regions.

Media condition	AUC <sub>0-2</sub>	(μg.hmL <sup>-1</sup> )	AUC <sub>0-6</sub> (μg.hmL <sup>-1</sup> )		
	Duodenum	Ileum	Duodenum	Ileum	
DP + ALB	0.7457±0.0153	0.4357±0.0154	4.2580±0.4843	2.3657±0.1554	
DP + STC	0.2728±0.0373	0.2798±0.0113	0.7738±0.0828	1.4495±0.0630	
DP +SFO	0.3877±0.0385	1.2325±0.0078	1.6710±0.1442	1.0133±0.0878	
Control	0.6198±0.0083	1.5408±0.4275	3.5228±0.3638	5.6603±0.1073	

NB:  $AUC_{0-2}$  and  $AUC_{0-6}$  represent areas under the curve for PQ permeation from 0-2 h and 0-6 h, respectively.

There was no significant difference in the parameter  $P_{\rm eff}$  appearance in organ bath and  $P_{\rm eff}$  disappearance from intestinal lumen for ALB treatment and its control values in the duodenal set up (Figure 3). However, treatments with STC and SFO revealed significantly higher  $P_{\rm eff}$  appearance in organ bath and  $P_{\rm eff}$  disappearance from intestinal lumen, p<0.05.

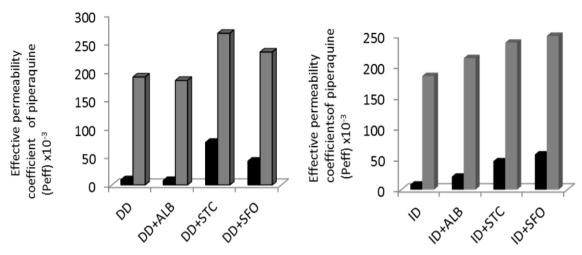


Figure 3: Effective permeability coefficients of piperaquine from ■appearance in organ bath and □disappearance from lumen in (a) duodenum and (b) ileum, in the presence of albumin (DD+ALB), starch (DD+STC) and sunflower oil (DD+SFO).

Similarly, SFO treatment produced significantly higher  $P_{\rm eff}$  appearance and  $P_{\rm eff}$  disappearance compared with other treatments, p<0.05. The rate for PQ appearance (Ka) in organ bath for treatments with STC and SFO were higher compared with the control in duodenum (p=0.019 and 0.012, respectively) while the same parameter had values with no significant difference for PQ disappearance. Similarly in ileum, Ka for PQ appearance values for STC and SFO were significantly higher than the control p=0.021 and 0.004, respectively (Table 2). In the same vein, Ka for PQ disappearance were higher than the control, with p=0.035 and 0.033, respectively.

**Table 2: Kinetics of piperaquine permeation.** 

	Media conditions								
Parameters	Duodenum			Ileum					
	DP alone	DP+ALB	DP+STC	DP+ SFO	DP alone	DP+ALB	DP+STC	DP+SFO	
Ka* (h <sup>-1</sup> )	0.0367±0.0006	0.0317±0.0032	0.3013±0.0076	0.1679±0.0031	0.0327±0.0011	0.0854±0.0021	0.0182±0.0024	0.2297±0.0011	
Ka **(h <sup>-1</sup> )	0.7600±0.0190	0.7360±0.0165	1.0676±0.0105	0.9345±0.0539	0.0327±0.0011	0.0854±0.0021	0.1842±0.0039	0.2297±0.0024	
C <sub>max</sub> ± SEM (μg/mL)	0.9800±0.0025	0.8270±0.0060	0.1640±0.0020	0.2951±0.0030	1.2170±0.0070	0.6000±0.5970	0.3310±0.0040	0.6150±0.0060	
$\mathbb{R}^2$	0.9438	0.7463	0.4438	0.6160	0.8652	0.9270	0.7434	0.8652	

NB: R<sup>2</sup> represents the regression coefficient for PQ permeation across the absorptive membrane; Values presented are the highest R<sup>2</sup> values denoting the kinetics of permeation; Ka\* is the rate of appearance of PQ in Tyrode solution while Ka\*\* is rate of disappearance from intestinal lumen; C<sub>max</sub> is the maximum concentration achieved in the TS.

#### DISCUSSION

In the recent years effect of food on the bioavailability of drugs has received particular importance necessitating food and medication time counseling for optimum therapeutic implications. [15] Since a vast number of medications are absorbed in the intestine, this work has followed other researcher's interests in this regard and considerations are highlighted on the quantitative permeation of drug molecules through the intestinal barrier. [16] This flux or permeability study therefore was designed to assess the effects of food components on the intestinal permeation of PQ from DP antimalarial drug. The major effect of diets on drugs is alteration of absorption by fatty, high protein and fiber-rich diets. The consequent bioavailability variations due to food composition have been correlated with clinical outcomes of drugs.[17] This preliminary study was designed to showcase the salient biopharmaceutical implications of DP antimalarial drug administration's effective absorption in the presence of concomitantly administered agents.

Mass transport of molecules in a solution or molecular transport across biomembrane barriers is measured in fluxes, in this experiment, net movement of PQ molecules are considered in the presence of concentration gradient as the major diffusion factor. Permeability data were therefore obtained and used to predict drug bioavailability and its consequent clinical outcomes in-vivo. Permeability coefficient, related to flux, is the transport flux of molecules through biomembranes per unit driving force per unit membrane thickness. [18]

Previous studies on transepithelial transport of drugs by Gershkovich et al and other researchers have concluded that the absorption of PQ from DP was enhanced when taken with fatty meal. [19,21] This study however believes that malaria drugs may be taken with varying meal types with differing proportions of food components. Therefore, the study design was to assess the different components of meals as to project at the probable outcome on bioavailability on different food component with varying proportions. The drug components of DP (i.e., DHA and PQ) being lipophilic in nature are expected to dissolve effectively in lipid medium and by inference the outcome in any mix of food components with which the drug may possibly be taken. becomes very important as real life circumstances in malaria presents with anorexia in patients and differing preferences for food types.

SFO treatment in the two regions of the intestine studied agreed with the findings of the earlier researchers. The physicochemical basis for solubility includes higher degree of association of solute molecules with triglyceride-rich lipoproteins (TRL) leading to enhanced solubilizing power. [22] Since meals taken with drugs, will be composed of different food components in varying proportions, we expect an inter- and intra-individual

variability with a specified dosage of medication at defined physiological and physicochemical conditions. It therefore will be necessary to determine the effects of the different meal types, so as to predict the nature of interaction of a mix of food components, obtainable in different geographical settings. Therefore, ALB, STC and SFO were adopted as basic treatments representing the classes of food (*i.e.*, protein, carbohydrate and fat/oil).

This present study also reflects on the hypotheses of coexisting passive transcellular diffusion and carrier mediated mechanisms of PQ molecules in the presence of food components. [20] ALB did not cause any significant difference in PO diffusion in duodenum in the study. ALB is a protein with high molecular weight. It does not transit biomembranes because of its inherent physicochemical limitations. However, a significant reduction in PQ permeation was observed due to ALB in the ileum. The ileum is a distal segment of the intestine with absorptive functions similar to the duodenum. Previous workers have reported on the effect of proteinrich food on drug absorption from intestinal regions. [23] Protein-rich foods cause reduced gastric emptying and will increase the absorption of acidic drugs in the stomach. The reduced gastric emptying may however lead to reduced rate and bioavailability of drugs that are susceptible to acid hydrolysis and enzymatic degradation.

STC is a polymer of glucose units that possess significant gumming or adhesive capacities with molecules in solution with which it is in contact, hence this treatment caused a significant reduction in PO permeation across duodenal epithelium. Previous work by Awofisavo et al. revealed similar reduction in drug absorption of artemether-lumefantrine, a similar antimalarial of the fixed-dose combination regimen, due to co-administration with STC. [24] Similarly coadministration of STC resulted in significant reduction in PQ permeation across ileal epithelium and the same argument is proffered for the mechanism. SFO treatment caused significant increase in PQ permeation across the duodenal and intestinal epithelium, respectively considering its output with the other food components. in agreement with the manufacturer's recommendation that the drug be taken with fatty meal.

Where the effect of co-loading with DP in both regions of the intestine considered caused reduction sequentially, it is believed that food components will pose an interaction likely to cause treatment failure, in the additive effects. The rate of drug permeation observed with food components in this work showed wide variations among the treatments. Similarly, the AUC values at 2 and 6 h revealed that food components caused significant reduction in the overall amount of drug that transited the intestinal membranes. Antimalarial drugs like other anti-infectives are meant for sufficient systemic exposure for therapeutic success.

AUC values for SFO treatment revealed higher values than STC and ALB but lower value than the control. This is as a result of the solubilisation of the poorly watersoluble PQ by a fatty milieu. This study therefore supports DP drug manufacturer's recommendation for DP's optimum absorption. Furthermore, as all the treatments presented lower AUC values than the respective control at 2 and 6 h post-loading, it could therefore be suggested that DP be taken before meals for improved absorption. The foregoing recommendation ensues as it appears from this study that food reduced components significantly the intestinal permeability of PO following oral absorption as compared with the drug alone. The lower values from SFO treatment compared with the control were due to the sequestrating effect of lipid medium on lipophilic compounds when in excess of the solubilisation properties. [26] This study therefore recommends that DP is taken on empty stomach for predictable absorption.

#### **CONCLUSION**

This study revealed a reduction in intestinal permeability of PQ due to the presence of food components (*i.e.*, ALB, STC and SFO) in the absorptive lumen of the duodenal and ileal intestinal region. This is suggestive of better bioavailability of PQ when DP was administered before food. The study also confirmed the significant increase in PQ permeation due to SFO than other food components.

## ACKNOWLEDGEMENTS

The authors express their thanks to PD Ojobor for the technical assistance and to the Management of Central Research Laboratory, University of Lagos for their equipments.

## **Conflicts of Interest**

Authors have no financial and non-financial conflicts to declare.

## **Funding**

This work was funded by TetFund 2015-2016 Merged intervention Award given to Sunday O. Awofisayo.

# REFERENCES

- Reddy LHR, Murthy RSR. Lymphatic transport of orally administered drugs. Indian J Exp Biol, 2002; 40: 1097-1109.
- 2. Tuma PL, Hubbard AL. Transcytosis: crossing cellular barriers. Physiol Rev, 2003; 83: 871-932.
- 3. Zakeri-Milani P, Valizadeh H. Mass transfer phenomena and biological membranes. In: Mass transfer in multiphase systems and its application. Prof Mohamed El-Amin (ed.). ISBN 978-953-307-215-9. http://www.iintechupan.com/books/mass-transfer-in-multilayersystem-and-its-applications/mass-transfer-phenomena-and-biological-membranes.
- 4. Liu W, Pan H, Zhang C, Zhao L, Zhao R, Zhu Y, Weisan P. Developments of methods for measuring

- the intestinal absorption of nanoparticle-bound drugs. Int J Mol Sci, 2016; 17(7): 1171-9.
- Ashley EA, McGready R, Hutagalung R, Phaiphun L, Slight T, Proux S, Thwai KL et al, Once-daily regimen of dihydroartemisinin-piperaquine for the treatment of uncomplicated multidrug-resistant falciparum malaria. Clin Infect Dis, 2005; 41(4): 425-432.
- 6. Poespoprodjo JR, Fobia W, Kanagalem E, Lampah DA, Sugiarto P, Tjitra E, Anstey NM, Price RN. Dihydroartemisinin-piperaquine treatment of multidrug resistant *falciparum* and *vivax* malaria in pregnancy. PLoS ONE, 2014; 9(1): e84976. Doi .10:1371/journal.pore0084976.ecollection2014.
- 7. Price RN, Hasugian AR, Ratcliff A, Siswantoro H, Purba HLE, Kenangalem E, Lindegardh N, Rentinen P, Laihad F, Ebsworth EP, Anstey WM, Tjitra E. Clinical and pharmacological determinants of the therapeutic response to dihydroartemisinin-piperaquine for drug resistant malaria. Antimicrob Agents Chemother, 2007; 51(11): 4090-97.
- 8. Harris KA, Garey KW, Rodvold KA. Drug-food interactions In: Piscitelli SC, Rodvold KA (eds). Drug interactions in infectious diseases. Infectious Diseases. Humana Press. Pg 383-430.
- 9. Marcum ZA, Gellad WF. Medication Adherence to multi-drug regimens. Clin Geriatr Med, 2012; 28(2): 287-300.
- 10. Farre R, Tack J. Food and symptom generation in functional gastrointestinal disorders: Physiological Aspects. Am J Gastroenterol, 2013; 108: 698-706.
- 11. Awofisayo SO, Okhamafe AO. The effect of coadministered drugs and food on the absorption of artemether-lumefantrine tablet. J Adv Med Pharm Sci. 2015; 2(4): 126-134.
- 12. Sullivan DM, Ford MA, Boyden TW. Grapefruit juice and the response to warfarin. Am J Health Syst Pharm, 1998; 55(15): 1581-1583.
- 13. Awofisayo SO, Umoh E, Igwe CN, Ojobor PD. Effect of metronidazole on piperaquine permeability from dihydroartemisinin-piperaquine antimalarial product across intestinal membranes. Saudi J Med Pharm Sci, 2017; 3A(8): 862-867.
- 14. Awofisayo SO, Arhewoh MI, Okhamafe AO. *In vitro* interaction of food component with AL tablet. Int J Curr Res Rev, 2018; 10(1): 22-26.
- 15. Yasuki T, Kondo H, Sako K. The effect of food on the oral bioavailability of drugs: a review of current development and pharmaceutical technologies for pharmacokinetic control. Ther Deliv, 2012; 3(1): 81-90.
- 16. Tscheik C, Blasig IE, Wimkler L. Trends in drug delivery through tissue barriers containing tight junctions. Tissue Barriers, 2013; 1(2): e24565. Doi: 10.4161/tisb.24565.
- 17. Chow S. Bioavaiabiity and bioequivalence in Drug Development. Wiley Interdiscip Rev Comput Stat, 2014; 6(4): 304-312.
- 18. Gershkovich P, Hoffman A. Effect of a high-fat meal on absorption and disposition of lipophilic

- compounds: the importance of degree of association with triglyceride-rich lipoprotein. Eur J Pharm Sci, 2007; 32(1): 24-32.
- Ghaffarian R, Muro S. Models and Methods to Evaluate Transport of Drug Delivery Systems across Cellular Barriers. J Vis Exp, 2013; 80: 50638. Doi: 10. 3791/50638(2013).
- Reuter SE, Evans AM, Shakib S, Lungershausan Y, Francis B, Valentini B, Bacchieri A, Ubben D, Pace S. Effect of food on the pharmacokinetics of piperaquine and dihydroartemisinin. Clin Drug Investig, 2015; 35(9): 559-67.
- 21. Sugero K, Kansy M, Artursson P, Avdeef A, Bendels S et al. Co-existence of passive and carrier-mediated processes in drug transport. Nature Reviews, 2010: 597-614.
- 22. Frankel EH. Basic Concepts. In: Handbook of food-drug interactions, McCabe RJ, Frankel EH, Wolfe JJ (Eds), CRL Press, Boca Raton, 2003.
- 23. Tamargo J, Le-Heuzey J, Mabo P. Narrow therapeutics index drugs: a clinical pharmacological consideration to flecainamide. Eur J Clin Pharmacol, 2015; 71(5): 549-567.
- 24. Awofisayo SO, Arhewoh MI, Okhamafe AO. Studies on in vitro interaction of artemether-lumefantrine with food components. Int J Curr Res Rev, 2018; 10(1): 22-26.
- 25. Wurtz N pascaul A, Marin-Jauffre A, Bouchiba H, Benoit N, Desbordes M, Martelloni M, de Santi VP, Richa G, Taudon N, Pradines B, Briolant S. Early treatment failure drug treatment of Plasmodium falciparum malaria with atovaquone-proguanil in the Republic of Ivory Coast. Malar J, 2012; 11: 146. Doi: 10.1186/1475-2875-11-146.
- Obitte NC, Rohan LC, Adeyeye CM, Parniak MA, Esimone CO. The utility of self-emulsifying oil formulation to improve the poor solubility of the anti HIV drug CSIC. AIDS Res Ther, 2013; 10: 14. htpp://doi.org/10.1186/1742-6405-10-14