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CHROMATOGRAPHIC METHOD FOR DEVELOPMENT AND VALIDATION OF CINNARIZINE USING ICH GUIDELINES

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ABSTRACT

In the present research work, a successful attempt was made for determination of Cinnarizine in Bulk form by High performance liquid chromatography. The method was developed by experimentation, based on literature survey. The simplicity, rapidity, reproducibility and economy of the proposed method completely fulfil the objective of this research work. The HPLC method was developed and validated for estimation of Cinnarizine. The mobile phase was consisting of Methanol: water (87:13) pH3. Detection was done at 255 nm. The method was found to be simple, linear, rapid, accurate, precise, reproducible and robust. The % RSD was found within limit. The result showed that proposed method was suitable for the accurate, precise and rapid determination of in Cinnarizine its bulk form. Before making experimentation with ion-exchange or ion-pair chromatography, ion suppression by pH controls and reverse phase chromatography should be tried for ion forming organic compounds. Ion-pair chromatography should be preferred to Ion Exchange chromatography.

KEYWORD: Cinnarizine, High performance liquid chromatography, Derivative Spectrophotometry, Q absorbance method, Normal Phase Chromatography.

I. INTRODUCTION

Analytical Chemistry is a measurement of science consisting of a set of powerful ideas and methods that are useful in all fields of science and medicine. It seeks ever improved means of measuring the chemical composition of natural and artificial materials. This branch of chemistry, which is both theoretical, and a practical science, is practiced in a large number of laboratories in many diverse ways while analytical method, is a specific application of a technique to solve an analytical problem. Methods of analysis are routinely developed, improved, validated, collaboratively studied and applied. The discipline of analytical chemistry consists of qualitative and quantitative analysis. Modern analytical methods are extremely sensitive and they require only small amount of sample material to provide precise and detailed information. The scope of drug analysis includes the analytical investigation of bulk-drug materials, the intermediates in their synthesis, products of drug research (potential pharmacons), drug formulations, impurities and degradation products of drugs, biological samples containing the drugs and their metabolites. Obtained data of drug analysis can contribute to the maximal efficacy and maximal safety of drug therapy and the maximal economy of the production of drugs. The efficacy, safety and economy of drug therapy are extremely important issues not only from the point of view of public health, but their financial, moreover political, aspects are also immense. As a consequence of this, pharmaceutical and biomedical analysis is among the most important branches of applied analytical chemistry. To fulfill the rapidly increasing demands as regards the number and the quality of analytical measurements, great efforts has to been made in the development of analytical method. Because of these reasons analytical methods are in widespread use. They are widely used in monitoring the use of drugs and medicines, product development and stability studies. Analytical methods often classified are classical / traditional and instrumental method. This classification is largely historical with classical methods and it is replacing by highly sensitive instrumental methods. [1,2]

1.1. Instrumental methods^[1,3]

Instrumental methods are used to investigate analytes scientific instruments. Modern analytical chemistry is dominated by sophisticated instrumentation, the roots of analytical chemistry and some of the principles used in modern instruments are from traditional techniques many of which are still used today. These methods are extremely sensitive and they require only small amount of sample material to provide precise and detailed information. Because of these reasons analytical methods are in widespread use.

1.2. UV- Visible spectrophotometer^[2,4,5]

Absorption spectroscopy is the measurement of the absorption of electromagnetic radiation from definite and narrow wavelength range by molecules, ions and atoms of chemical substance. Techniques most commonly employed in analytical field ultraviolet, visible, infrared and atomic absorption spectroscopy. The ultravioletvisible spectroscopy is one of the most frequently employed techniques in pharmaceutical analysis. It involves the measurements of the amount of ultraviolet (190-380 nm) or visible (380-800 nm) radiation absorbed by a substance in solution. Instruments which measure the ratio or a function of the ratio, of the intensity of two beams of light in the ultraviolet-visible region are called ultraviolet-visible spectrophotometers. Absorption of light in both the ultraviolet and visible regions of the electromagnetic spectrum occurs when the energy of the light matches that required to induce in the molecule an electronic transition and its associated vibrational and rotational transitions. There are four types of transitions observed in UV-visible spectroscopy $\sigma \rightarrow \sigma^*, \pi \rightarrow \pi^*, n \rightarrow \pi^* \text{ and } n \rightarrow \sigma^*.$

1.3. Introduction to Derivative Spectrophotometry^[2]

Derivative spectrophotometry, which consists in the differentiation of a normal spectrum, offers a useful means for improving the resolution of mixtures, because it enhances the detectability of minor spectral features. It involves the conversions of a normal spectrum to its first, second or higher derivative spectrum. In the context of derivative spectrophotometry, the normal absorption spectrum is referred to as the fundamental, zero order, or D0 spectrum whereas for first derivative D1 spectrum, second derivative D2 spectrum and so on. The first derivative spectrum is plot of rate of absorbance with wavelength against wavelength.

1.4. MODES OF CHROMATOGRAPHY^[6]

Modes of chromatography are defined essentially according to the nature of the interactions between the solute and the stationary phase, which may arise from hydrogen bonding, Vander walls forces, electrostatic forces or hydrophobic forces are based on the size of the particles (e.g. Size exclusion chromatography)

Different modes of chromatography are as follows -

- Normal Phase Chromatography
- Reverse Phase Chromatography

- Reverse Phase ion pair Chromatography
- Ion Chromatography
- Ion-Exchange Chromatography
- Affinity Chromatography

1.5. Adsorption Chromatography /Normal Phase Chromatography $^{\left[7\right]}$

In normal phase chromatography, the stationary phase is a polar adsorbent and the mobile phase is generally a mixture of non-aqueous solvents.

The silica structure is saturated with silanol groups at the end. These OH groups are statistically disturbed over the whole of the surface. The silanol groups represent the active sites (very polar) in the stationary phase. This forms a weak type of bond with any molecule in the vicinity when any of the following interactions are present.

- Dipole-induced dipole,
- Dipole-dipole,
- Hydrogen bonding,
- π -Complex bonding

1.6. HPLC System^[7,8]

The importance of Chromatography is increasing rapidly in pharmaceutical analysis. The exact differentiation selective identification and quantitative determination of structurally closely related compounds. important field of application of chromatographic methods is the purity testing of final products and intermediates (detection of decomposition products and by-products). As a consequence of the above points, chromatographic methods are occupying an everexpanding position in the latest editions of the pharmacopoeias and other testing standards. The modern form of column chromatography has been called high performance, high pressure, and high-resolution and high-speed liquid chromatography. High-Performance Liquid Chromatography (HPLC) is a special branch of column chromatography in which the mobile phase is forced through the column at high speed. As a result the analysis time is reduced by 1-2 orders of magnitude relative to classical column chromatography and the use of much smaller particles of the adsorbent or support becomes possible increasing the column efficiency substantially.

II.MATERIALS AND INSTRUMENTS

2.1. Procurement of drug sample

Table No. 2.1.: Details of drug sample.

| b of arag sample. | | |
|-------------------|----------|---------------------------------|
| Name of Drug | Quantity | Drug Supplier |
| Cinnarizine | 10 gm. | Chempro Pharma Pvt. Ltd, Mumbai |

2.2. Reagents and chemicals

All the chemicals used are of HPLC and AR grade. Chemicals used are as follows

Table No. 2.2: Reagents and Chemicals.

| Sr. No | REAGENTS | GRADE | MANUFACTURES |
|--------|--------------|-------|--|
| 1 | Methanol | HPLC | Merck specialities private limited, Mumbai |
| 2 | Water | HPLC | Merck specialities private limited, Mumbai |
| 3 | Acetonitrile | HPLC | Merck specialities private limited, Mumbai |

III.RESULTS AND DISCUSSION

3.1. Identification of drug

3.1.1. Organoleptic properties of drug

Table No. 3.1.: Organoleptic properties of drug.

| Sr. No. | Organoleptic Property | Cinnarizine |
|---------|-----------------------|--------------|
| 1 | Colour | White powder |
| 2 | Odor | Odorless |

3.1.2 Melting point of drug

Table No.3.2: Melting point of drug.

| Sr. No. Name of drug | | M.P. (°C) | |
|----------------------|-------------|------------------|--|
| 1 | Cinnarizine | 117-120 ° C | |

3.1.3 Solubility Study

Table No. 3.3.: Solubility Study of Cinnarizine.

| • | , | | | | | |
|---|-----------------|----------|----------------|--|--|--|
| | Sr. No Solvents | | Solubility | | | |
| | 1 | Water | Freely soluble | | | |
| | 2 | Methanol | Freely soluble | | | |
| | 3 | DMSO | Soluble | | | |

3.2. FTIR spectrum of Cinnarizine

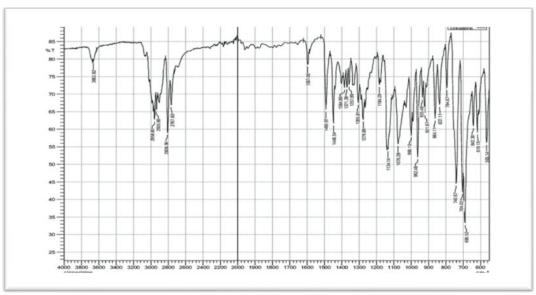


Fig.No.3.1: IR Spectrum of Cinnarizine.

| Sr.No. | Functional group | Standard range (cm-1) | Observed range (cm-1) |
|--------|---|------------------------------|-----------------------|
| 1 | C-H, Aromatic Stretching | 3067-3025 cm ⁻¹ , | 3066 |
| 2 | C-H Alkane Stretching | 2962-2853cm ⁻¹ | 2808 |
| 3 | Characteristic Peaks of Mon substituted benzene | 2000-1667cm ⁻¹ | 1800 |
| 4 | C=C Aromatic | Near 1600cm ⁻¹ | 1595 |
| 5 | Piperazine ring N,N disubstituted | 140-1500cm ⁻¹ | 1492 |

3.3. Development of HPLC method for Cinnarizine High performance liquid chromatographic method was

High performance liquid chromatographic method was developed and validated for determination of Cinnarizine in bulk form. Mobile phase consists of Methanol: Water (87:13) pH3. Chromatogram obtained was shows the maximum wavelength where the drug shows maximum response was 255 nm and is shown in Fig.3.2

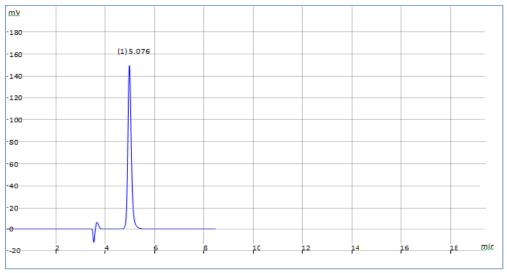


Fig. no.3.2: Typical chromatogram of Cinnarizine.

3.3.1. Linearity

Table No.3.5: Data of calibration curve of Cinnarizine by HPLC method.

| Sr. No. | Conc. (µg/ml) | Area |
|---------|---------------|----------------|
| 1 | 10 | 614839 ±1680 |
| 2 | 20 | 1339880 ±2642 |
| 3 | 30 | 2032564 ±16880 |
| 4 | 40 | 2665713 ±5621 |
| 5 | 50 | 3251263 ±7513 |

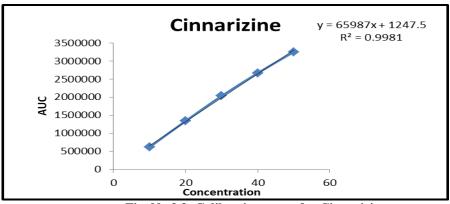


Fig. No.3.3: Calibration curve for Cinnarizine.

3.3.2. Optical characteristics

Table No. 3.6: Optical characteristics for Cinnarizine.

| Sr.No | Parameters | High performance liquid chromatography |
|-------|---|--|
| 1 | λmax (nm) | 255 |
| 2 | Beer's law limit (µg/mL) | 10-50 |
| 3 | Regression equation[y] | y = 65987x + 1247.5 |
| 4 | Slope[m] | 65987 |
| 5 | Intercept [c] | 1247.5 |
| 6 | Correlation coefficient [r ²] | 0.9981 |
| 7 | Limit of detection (LOD) (μg/mL) | 0.084 |
| 8 | Limit of quantitation (LOQ) (μg/mL) | 0.254 |

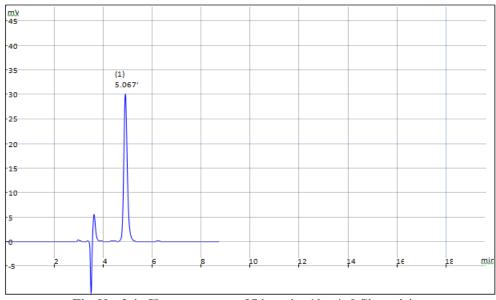


Fig. No. 3.4: Chromatogram of Linearity 10µg/ml Cinnarizine.

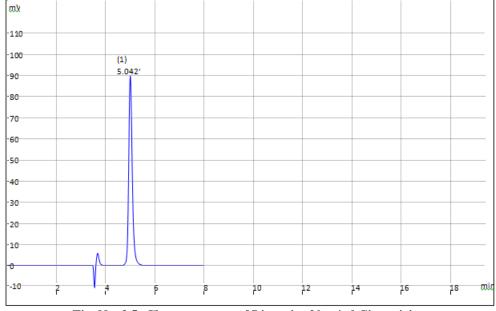


Fig. No. 3.5: Chromatogram of Linearity 20µg/ml Cinnarizine.

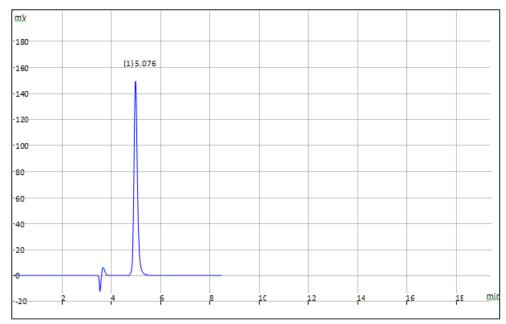


Fig. No. 3.6: Chromatogram of Linearity 30µg/ml Cinnarizine.

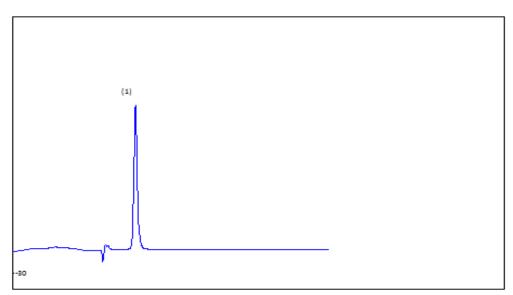


Fig. No. 3.7: Chromatogram of Linearity $40\mu g/ml$ Cinnarizine.

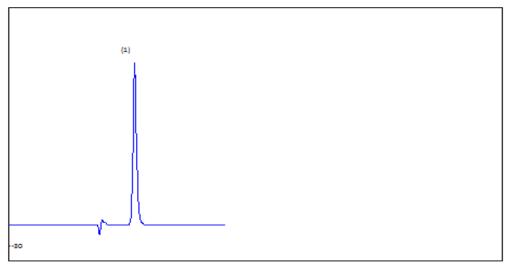


Fig. No. 3.8: Chromatogram of Linearity $50\mu\text{g/ml}$ Cinnarizine.

3.3.3. Accuracy

Table No.3.7: Data for recovery study of Cinnarizine by HPLC method

| Level of addition | Standard added (µg/ml) | conc. (µg/ml) | Total conc. (µg/ml) | Area obtained* | Std Area | Drug recovered (µg/ml) | %Recovery |
|-------------------|------------------------------|------------------|---------------------------|-------------------|----------|------------------------------|------------|
| | 10 | 20 | 30 | 2042365 | | 30.14466 | 100.482199 |
| 50% | 10 | 20 | 30 | 2051326 | 2032564 | 30.276921 | 100.923071 |
| | 10 | 20 | 30 | 2062345 | | 30.439558 | 101.465194 |
| | 20 | 20 | 40 | 2671542 | | 40.087466 | 100.218666 |
| 100% | 20 | 20 | 40 | 2663215 | 2665713 | 39.962517 | 99.9062915 |
| | 20 | 20 | 40 | 2692654 | | 40.40426 | 101.010649 |
| 150% | 30 | 20 | 50 | 3245622 | | 49.913249 | 99.8264982 |
| | 30 | 20 | 50 | 3225659 | 3251263 | 49.606245 | 99.2124907 |
| | 30 | 20 | 50 | 3265595 | | 50.220407 | 100.440813 |

Table No.3.8: Statistical validation of Cinnarizine by HPLC method.

| Level of addition | Level of addition % Mean recovery* | | % RSD |
|-------------------|--------------------------------------|--------|----------|
| 50% | 101 | 0.4924 | 0.487699 |
| 100% | 100.4 | 0.5693 | 0.567125 |
| 150% | 99.83 | 0.6142 | 0.615228 |

^{*}Average of three determination

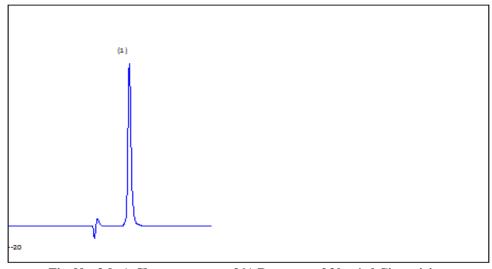


Fig. No. 3.9: A Chromatogram of % Recovery of $30\mu\text{g/ml}$ Cinnarizine.

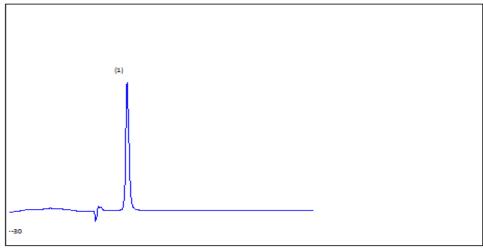


Fig. No. 3.10: A Chromatogram of % Recovery of $40\mu g/ml$ Cinnarizine.

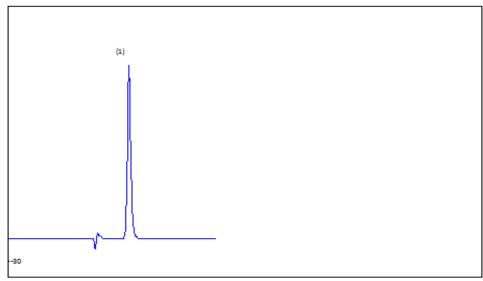


Fig. No. 3.11: A Chromatogram of % Recovery of $50\mu g/ml$ Cinnarizine.

3.3.4. Precision

Table No.3.9: Data for intraday precision of Cinnarizine by HPLC method.

| Sr. No. | Conc. (µg/mL) | Area | Mean* | SD | %RSD |
|---------|---------------|---------|------------|----------|----------|
| 1 | 10 | 614959 | | | |
| 2 | 10 | 615785 | 614432 | 1679.693 | 0.273373 |
| 3 | 10 | 612552 | | | |
| 4 | 30 | 2015685 | | | |
| 5 | 30 | 2032564 | 2023969.33 | 16887.56 | 0.834378 |
| 6 | 30 | 2023659 | | | |
| 7 | 50 | 3256235 | | | |
| 8 | 50 | 3251263 | 3257551 | 7038.878 | 0.216079 |
| 9 | 50 | 3265155 | | | |

Fig. No. 3.12: A Chromatogram of Intraday Precision of 10µg/ml Cinnarizine.

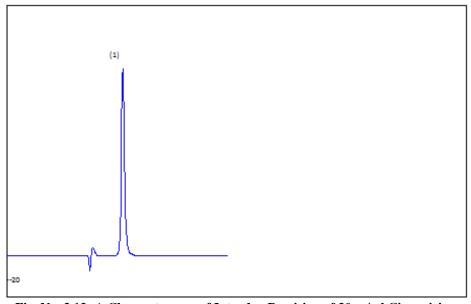


Fig. No. 3.13: A Chromatogram of Intraday Precision of 30µg/ml Cinnarizine.

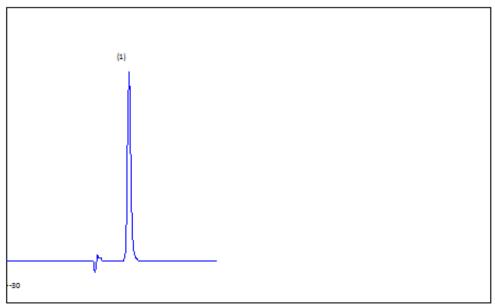


Fig. No. 3.14: A Chromatogram of Intraday Precision of 50µg/ml Cinnarizine.

Table No.3.10: Data for interday precision of Cinnarizine by HPLC method.

| | or meet any process | | -y | | | |
|---------|---------------------|---------|------------|------------|------------|--|
| Sr. No. | Conc. (µg/mL) | Area | Mean* | SD | %RSD | |
| 1 | 10 | 613258 | | | | |
| 2 | 10 | 610256 | 613279.667 | 3034.55801 | 0.49480819 | |
| 3 | 10 | 616325 | | | | |
| 4 | 30 | 2023526 | | | | |
| 5 | 30 | 2032556 | 2037444.67 | 16901.8362 | 0.8295605 | |
| 6 | 30 | 2056252 | | | | |
| 7 | 50 | 3245622 | | | | |
| 8 | 50 | 3212552 | 3238386.33 | 23083.3062 | 0.71280273 | |
| 9 | 50 | 3256985 | | | | |

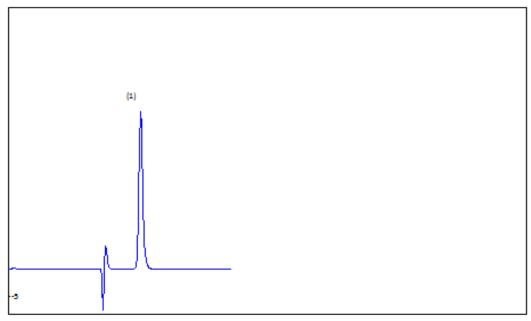


Fig. No. 3.15: A Chromatogram of Interday Precision of $10\mu\text{g/ml}$ Cinnarizine.

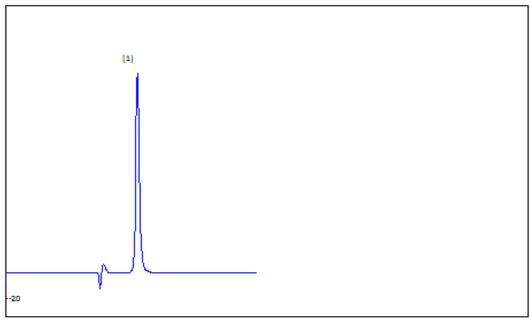


Fig. No. 3.16: A Chromatogram of Interday Precision of 30µg/ml Cinnarizine.

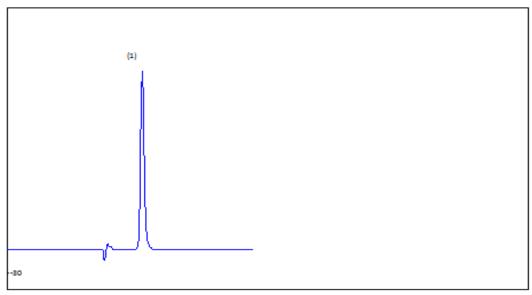


Fig. No. 3.17: A Chromatogram of Interday Precision of 50µg/ml Cinnarizine.

3.4. Limit of detection and limit of Quantitation

Table No. 3.11: Results of LOD and LOQ values of Cinnarizine.

| Drugs | LOD (µg/ml) | LOQ (µg/ml) |
|-------------|-------------|-------------|
| Cinnarizine | 0.084 | 0.254 |

3.4.1. Robustness

Robustness was studied by different deliberate variations in the chromatographic conditions. Results are shown in Table no. 49.

| Sr.No | Parameter | Condition | Area | Mean* | SD | %RSD |
|-------|---------------------------|-----------|---------|---------|---------|------|
| 1 | Change in Flow | 0.9 | 1333913 | 1362556 | 4329.28 | 0.31 |
| 2 | rate (ml/min) | 1 | 1339880 | | | |
| 3 | | 1.1 | 1337757 | | | |
| 1 | Change in Wavelength (nm) | 253 | 1340385 | | | |
| 2 | | 255 | 1339880 | 1363715 | 2175.33 | 0.15 |
| 3 | | 257 | 1341622 | | | |

^{*}Average of three determination

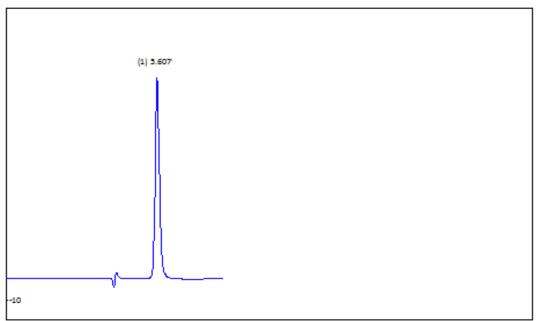


Fig. No. 3.18: Chromatogram of 20µg/ml Cinnarizine at flow rate 0.9 ml/min.

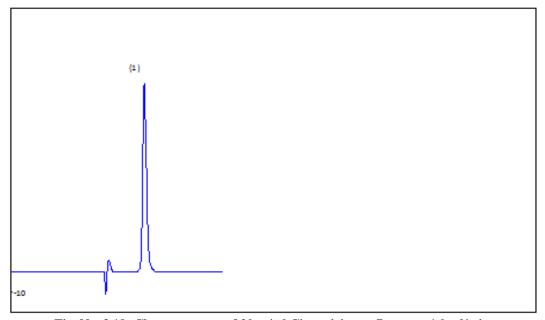


Fig. No. 3.19: Chromatogram of $20\mu g/ml$ Cinnarizine at flow rate 1.0 ml/min.

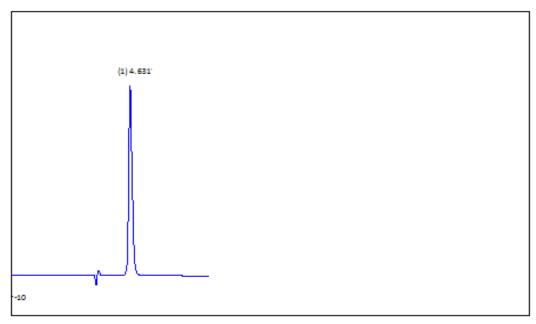


Fig. No. 3.20: Chromatogram of 20µg/ml Cinnarizine at flow rate 1.1 ml/min.

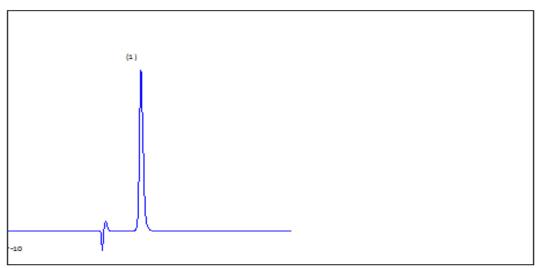


Fig. No. 3.21: Chromatogram of 20µg/ml Cinnarizine at Wavelength 253nm.

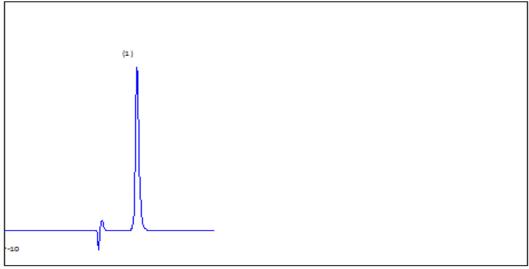


Fig. No. 3.22: Chromatogram of 20µg/ml Cinnarizine at Wavelength 255nm.

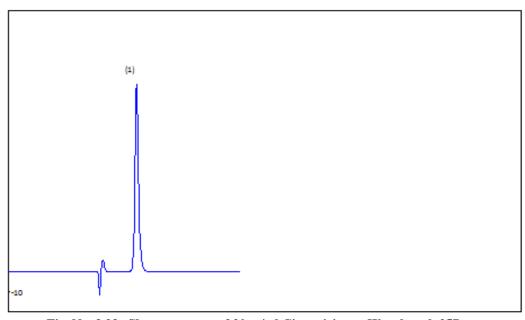


Fig. No. 3.23: Chromatogram of $20\mu\text{g/ml}$ Cinnarizine at Wavelength 257nm.

3.4.2. Ruggedness

Table No.3.13: Data for ruggedness study of Cinnarizine by HPLC method.

| Sr. No | Analyst | Conc. (µg/ml) | Area | Mean area* | SD | % RSD |
|--------|------------|------------------|---------|---------------|------------|------------|
| | | | 2032565 | | | |
| 1 | Analyst-I | 30 | 2023659 | 2031067.33 | 6784.62927 | 0.33404256 |
| | | | 2036978 | | | |
| | | | 2032659 | | | |
| 2 | Analyst-II | 30 | 2045265 | 2036694 | 7426.92945 | 0.36465613 |
| | | | 2032158 | | | |

^{*}Average of three determination

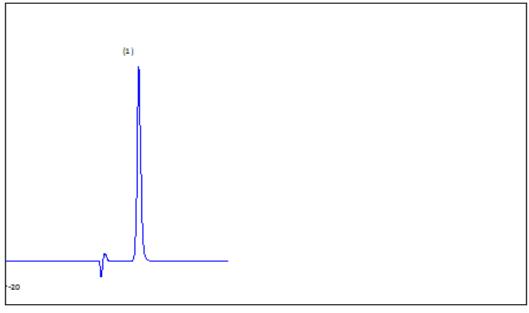


Fig. No. 3.24: Chromatogram of $30\mu g/ml$ Cinnarizine by Analyst –I.

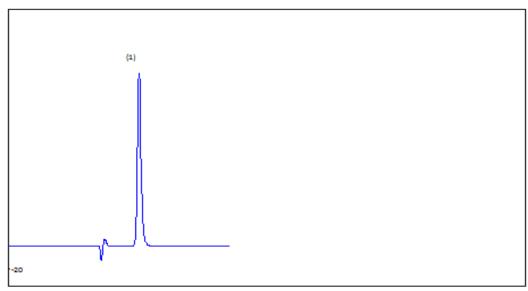


Fig. No. 3.25: Chromatogram of 30µg/ml Cinnarizine by Analyst –II.

3.4.3. Specificity

Table No.3.14: Data for specificity study of Cinnarizine by HPLC method.

| total in Butta for specificity study of chimarizane by the Be method. | | | | | | |
|---|--------------------|---------------------------|---------|-------------|-------------|------------|
| Drug conc. (µg/ml) | Excipients (µg/ml) | Total conc. (µg/ml) | Area | Mean* | SD | %RSD |
| 10 | 20 | 30 | 672869 | | | |
| 10 | 20 | 30 | 673645 | 672057 2222 | 519.7541085 | 0.07722286 |
| 10 | 20 | 30 | 672658 | 673057.3333 | 319.7341083 | 0.07722286 |
| 20 | 20 | 40 | 1370589 | | | |
| 20 | 20 | 40 | 1375624 | 1371722.333 | 3476.428963 | 0.25343533 |
| 20 | 20 | 40 | 1368954 | 13/1/22.333 | 3470.428903 | 0.23343333 |
| 30 | 20 | 50 | 1921659 | | | |
| 30 | 20 | 50 | 1939855 | 1930366.667 | 9123.085242 | 0.47260893 |
| 30 | 20 | 50 | 1929586 | 1930300.007 | 9123.063242 | 0.47200893 |

3.4.4. System Suitability

System suitability parameters were measured to verify the system, method and column performance.

Standard solution of Cinnarizine was injected into the system for five times and system suitability parameters were checked.

Table No.3.15: Data for System suitability study of Cinnarizine by HPLC Method.

| 110101201201 | TOT SJETCHE SCHOOL STATE | staaj of Chimarizmic s | THE ELC MICHIGAN | | |
|-----------------------|--------------------------|------------------------|--------------------|---------------------|--|
| Sr. No. conc. (μg/ml) | | Retention Time (min) | Theoretical plates | Asymmetry Factor | |
| 1 | 30 | 5.05 | 8487 | 1.25 | |
| 2 | 30 | 5.02 | 8552 | 1.24 | |
| 3 | 30 | 5.02 | 8462 | 1.25 | |
| 4 | 30 | 5.01 | 8359 | 1.23 | |
| 5 | 30 | 5.03 | 8252 | 1.24 | |
| 6 | 30 | 5.1 | 8539 | 1.25 | |
| Mean | | 5.038 | 8441.83 | 1.243 | |
| SD | | 0.033115958 | 115.691688 | 0.00816497 | |
| %RSD | | 0.657280011 | 1.37045691 | 0.65669966 | |

IV.CONCLUSION

The HPLC method was developed and validated for estimation of Cinnarizine. The mobile phase was consisting of Methanol: water (87:13) pH3. Detection was done at 255 nm. The method was found to be

simple, linear, rapid, accurate, precise, reproducible and robust. The % RSD was found within limit. The result showed that proposed method was suitable for the

accurate, precise and rapid determination of in Cinnarizine its bulk form.

V. ACKNOWLEDGEMENT

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