

EUROPEAN JOURNAL OF PHARMACEUTICAL AND MEDICAL RESEARCH

www.ejpmr.com

Review Article
ISSN 2394-3211
EJPMR

ISO-ENZYMES IN DIAGNOSTICS: A REVIEW

¹Dr. Silky Rai, ²*Dr. Sharique Ahmad, ³Dr. Saeeda Wasim, ⁴Shivani Singh and ⁵Dr. Saba Naziya

¹Junior Resident, Department of Pathology, Era's Lucknow Medical College and Hospital, Era University, Lucknow, Uttar Pradesh, India-226003.

²Professor, Department of Pathology, Era's Lucknow Medical College and Hospital, Era University, Lucknow, Uttar Pradesh, India-226003.

³Nova IVF Fertility, Hazratganj, Lucknow, U.P., India-226001.

⁴Research Scholar, Department of Pathology, Era's Lucknow Medical College and Hospital, Era University, Lucknow, Uttar Pradesh, India-226003.

⁵Junior Resident, Department of Pathology, Era's Lucknow Medical College and Hospital, Era University, Lucknow, Uttar Pradesh, India-226003.

*Corresponding Author: Dr. Sharique Ahmad

Professor, Department of Pathology, Era's Lucknow Medical College and Hospital, Era University, Lucknow, Uttar Pradesh, India-226003.

Article Received on 02/03/2021

Article Revised on 21/03/2021

Article Accepted on 11/04/2021

ABSTRACT

New areas of problems keep surfacing as a result of the complex interactions among the various factors which eventually lead to diseases. Extensive studies of these different factors abnormality under taken so far have been able to provide an insight into the patho-physiological alterations in various diseases. Physiologically, iso-enzymes are structurally different forms of the same enzyme and perform the same enzyme activity. In this review, we tried to provide an overview about the various forms, structure and functions of different iso-enzymes with special focus on their tissue localization.

KEYWORDS: Hepato-biliary diseases, Cancer, Leucocyte, Prostate gland.

INTRODUCTION

New areas of problems keep surfacing as a result of the complex interactions among the various factors which eventually lead to diseases. Extensive studies of the different factors abnormality under taken so far have been able to provide an insight into the pathophysiological alterations in various diseases. [1]

Physiologically, iso-enzymes are structurally different forms of the same enzyme and perform the same enzyme activity. [2,3] The first research according to internet on the iso-enzymes dated as far as back to 1957, where a report mentioned iso-enzymes are the different variants of same enzymes having similar function in the same individual.[4] Later on, curiosity in the role of isoenzymes in medical research has been investigated. An intensive search over the literature through the years reported that iso-enzymes are formed in different ways, attributing to presence of origin from the same locus of the gene and post translational molecular heterogeneity of enzyme protein synthesized. [5-8] Further searches on the identification of iso-enzymes revealed that they are separated by electrophoresis, heat stability, by their inhibitors, by substrate specificity, by co-factor requirements, antibodies specificity, and lastly their tissue localization. [9-15] However, there is a lot more research to be done for coming to a clear conclusion

about the formation of iso-enzymes and their association with different diseases. In this review, we tried to provide an overview about the various forms, structure and functions of different iso-enzymes including adenosine deaminase (EC 3.5.4.4), aldolase (EC 4.1.2.13), alkaline phosphatase (EC 3.1.3.1), betaglucuronidase (EC 3.2.1.31), creatine kinase (EC 2.7.3.2), enolase (EC 4.2.1.11), glucose-6-phosphatase (EC 3.1.3.9), lactate dehydrogenase (EC 1.1.1.27), and prostate specific antigen (EC 3.4.21.77).

Adenosine Deaminase (ADA)

ADA is an enzyme that converts adenosine to inosine, and deoxyadenosine to deoxyinosine. [16] It is best known in the medical science for its role in cell-mediated immunity and helps in insulin activity. [17] Research reported low levels of ADA are observed in individuals with malnutrition and patients with human immunodeficiency virus. [18] Higher values of ADA are reported in diseases including brucellosis, leprosy, infective mononucleosis, viral hepatitis, and liver cirrhosis. [16-19] Till now literature has demonstrated two isoforms of ADA present in human individuals, ADA1 and ADA2. ADA1 is found mainly in lymphocytes and macrophages. [20] The two isoforms are interconverted to each other in the lung. ADA2 was also found in macrophage and co-exists with ADA1. [21] Reports have

been found that ADA2 iso-enzyme activity is of considerable prognostic value in AIDS and adult T-cell leukemia (ATL) cases. [18-22]

Aldolase

Aldolase is an enzyme that helps convert glucose (sugar) into energy and exists in three isoforms, Aldolase A, Aldolase B and Aldolase C. [23] These are found throughout the body. Aldolase A converts fructose 1,6 Bisphosphate to glyceraldehydes 3 phosphate and dihydroxyacetone phosphate, present in liver kidney and intestine. [24] Reports suggested that Aldolase-A deficiency is attributed to myopathy and hemolytic anemia. [23-25] Aldolase-B helps in fructolysis that is by converting fructose-1-phospahte to glyceraldehydes and dihydroxyacetone phosphate present preferentially in liver. [25] Deficiency of this iso-enzyme leads to a condition called as hereditary fructose intolerance. Aldolase-C helps in aldose cleavage and present abundantly in brain and nervous tissue. [26] In a report by Asaka et al^[25], patients affected with muscle abnormalities and cancer found increased levels of aldolase A, but not increased in the patients affected with liver diseases. On the contrary in the same report, patients affected with liver diseases found increased levels of aldolase B, but not increased in the patients affected with muscle abnormalities and cancer patients. Further, the report observed a decrease in serum aldolase B levels in cancer patients. In another report by Ojika et al [26], all the three iso-enzymes of aldolase were estimated in tumor tissues and sera of patients with lung cancer. In patients affected with lung cancer, the report observed aldolase A and aldolase C tissue levels were increased but not aldolase B tissue levels. However, no increase in the sera of these iso-enzymes were observed in the sera of patients with lung cancer.

Alkaline Phosphatase (ALP)

Alkaline phosphatase (ALP) is a glycoprotein that removes the phosphate group from various proteins and nucleotides at basic pH values. [27] ALP is divided into six iso-enzymes depending upon the site of tissue expression that are Alpha-1 ALP, Alpha-2 heat labile ALP, Alpha-2 heat stable ALP (Regan), pre beta ALP, ALP, the leucocyte ALP. [5] From accumulated research it is believed that ALP isoenzymes originated from the duplications of primordial tissue non-specific ALP (TN-ALP) gene into six different forms. [28] Reports have also been demonstrated that alteration in the handling of a specific gene (TN-ALP gene) product during and after its formation gives rise to specific isoforms of ALP enzymes. [28-30] These modifications include amount of carbohydrate content attached to different iso-enzymes, thus differing isoenzymes between each other. [28] Further research has also revealed that only humans and our ancestral origin have placental ALP.[30]

Individuals affected with hepato-biliary diseases including cholestasis or hepatic carcinoma have very

high levels of ALP. [31-33] Increase in the same isoform is noticed in the patients affected with bile duct obstruction.^[31] Slight increase in the ALP is observed in the hepatic tissue diseases, however in patients affected with hepatitis, and also in patients where inflammatory edema produces an obstructive phase, increase in ALP is seen.^[32] In patients affected with gall stones or in patients where bile duct is obstructed by the cancer of head of pancreas or in patients suffering with intra-hepatic obstruction due to drugs observe very high levels of ALP. [32,33] In patients affected with bone diseases or cancers of bone or in hyperparathyroidism observe very high levels of ALP. [34,35] Placental origin ALP can be easily identified if an iso-enzyme of ALP is inhibited by phenylalanine and such iso-enzyme increase is seen normal pregnancy. [36] An iso-enzyme of ALP similar to placental ALP is characteristically seen in circulation about 15% cases of carcinoma of pulmonary tissue, hepatic tissue and digestive tissue and named as Regan iso-enzyme or carcinoplacental iso-enzyme. [37]

Utmost care has to be inducted into interpretation of ALP concentration in the laboratory as there is an intestinal form of ALP where the same can alter the actual concentration of ALP during fasting or anorexia or in postprandial. [30, 32-37]

Beta-Glucuronidase (β-Glu)

Beta-Glucuronidase enzyme (β-Glu) is a lysosomal hydrolase involved in the stepwise catabolism of glucuronic acid-containing glycosaminoglycans. [38] The first report on this isozyme was reported in the year 1965. [39] Despite the role of β -Glu has been studied for several years, the physiological role is still remains obscure. Nevertheless, three isoforms have been identified depending on the pH of its surrounding medium. Fondo et al^[40], found three peaks of activity at pH 4.5; pH 5.2; pH 6.1-6.3, respectively in normal premenopausal female mammary gland and lung tissues the report further demonstrated that the enzyme with a pH optimum of 6.1-6.3 disappears in homogenates of human cancer tissue (lung and mammary gland), and increase in its activity is seen at pH optimum 4. Kakizoe et al., in his study, observed six types of iso-enzymes in the rat liver, numbered Type I to Type VI. [43] Some reports have also shown the altered levels of β-Glu enzyme in the patients suffering with cutaneous cancers, prostate carcinoma and hepatomas as well. [41-43] Further research is wanting to understand the exact pattern of this enzyme.

Creatine Kinase (CK)

Creatine kinase (CK) is an enzyme that conjugates phosphate group to creatine in the presence of adenosine triphosphate (ATP), the hydrolysis of same reaction brings the muscle contraction. [44-46] This enzyme exists in three different forms and each isoform is specific to heart, brain, skeletal muscle, and other tissues. [44,45] The CK enzyme is a dimer composed of subunits (Fig. 1) derived from either muscle (M) or brain (B). Three iso-

enzymes have been identified: striated muscle consists of subunit M and designated as MM and it is predominantly present in the blood, heart tissue consists of subunit M & B and designated as MB, and brain consists of subunit B and designated as BB. [44,47] The CK concentrations in

serum are directly proportional to the muscle mass and thus males to have increased levels when compared to females.^[45] Individuals affected with muscle dystrophies and also people affected with acute kidney failure, altered levels in serum CK levels are seen.^[44]

Figure 1: Characteristic features of creatine kinase iso-enzymes.

Iso-enzyme	Subunit make up of iso-enzyme	Tissue from which iso-enzyme has originated
CK-MM	MM	Striated muscle
CK-BB	BB	Brain
CK-MB	MB	Heart Tissue

Enolase (ENO)

Enolase is a glycolytic enzyme, which catalyzes the conversion of 2-phosphoglycerate to phosphoenolase pyruvate. It was identified first by the researchers Lohmann & Meyerhof in the year 1934. Studies have reported three subunits (Fig. 2) in the ENO enzymes which are α , β , and γ . These three subunits make up five iso-enzymes belonging to ENO family. ENO1 is also called as $\alpha\alpha$ or non-neuronal enolase (NNE) present

in hepatic tissue, nervous tissue, and fat tissue. ENO2 is also called as $\gamma\gamma$ or neuron-specific enolase (NSE) present in nervous tissues abundantly. ENO3 is also called as $\beta\beta$ or muscle-specific enolase (MSE) present in muscle tissue in very high levels. Literature showed the increase of ENO concentrations in terms of tumor severity, existence of cancerous growths in the neuronal tissues, and in the individuals suffering with myocardial infarctions. $^{[52-54]}$

Figure 2: Characteristic features of enolase iso-enzymes.

Iso-enzyme	Subunit make up of iso-enzyme	Tissue from which iso-enzyme has originated
ENO1	Αα	Liver, Kidney, Spleen, Adipose tissue
ENO2	Вβ	Muscle
ENO3	Γγ	Nervous tissue

Glucose-6-phosphatase (GluP)

Glucose-6-phosphatase (GluP), is an enzyme that helps in gluconeogenesis process where it is useful to produce glucose during the fasting state. [55-57] It removes phosphate group from the glucose-6-phosphate, thus producing glucose and present only in liver tissue. [57] Absence or deficiency of this enzyme leads to Glycogen storage diseases (Type 1, Type 1A, and Type 1b). [56] Until now three iso-enzymes have been identified in Glucose-6-phosphatase-α, phosphatase-β, and Glucose-6-phosphatase-2. [58] It is reported that two iso-enzymes Glucose-6-phosphatase-α and Glucose-6-phosphatase-β have found to shown the similar effect of gluconeogenesis. [59] On the other hand, isoform Glucose-6-phosphatase-2 have found to show no effect on gluconeogenesis but has significant role in the pancreatic insulin secretion. [60] Studies have shown altered levels of this enzyme in the patients affected with glioblastoma and colorectal carcinoma. [61,62]

$Lactate\ Dehydrogenase\ (LDH)$

Lactate dehydrogenase (LDH) enzyme convert lactate to pyruvate and back, with the help of co-factor NAD⁺ to NADH and present in almost all living cells. [63-67]

LDH enzyme structure is made of four subunits and all of them have similar molecular weight (32kD) with a slight variation in amino acid sequence in each chain. [65-67] Each subunit is composed of either H or M polypeptide chain. [63-64] Different combinations of these two subunits or polypeptide chains give rise to five isoforms of LDH enzyme (figure 3). Therefore, the five

combinations are H4, H3M, H2M2, M3H and M4 varieties and all these isoforms are seen in all individuals. [63,67] All these iso-enzymes can be isolated with the help of electrophoresis technique maintaining the pH at 8.6. [64] H polypeptide chain denotes heart and M polypeptide chain stands for muscle (Fig. 3)^[65], so M4 form is seen in skeletal muscles while H4 form is seen in heart. [67] In normal conditions, iso-enzyme LDH-2, which is composed of combination of polypeptide chains of H3 and M1 is more in blood than iso-enzyme LDH-1 which is having a composition of 4 polypeptide chains of H. However, this normal pattern is altered and called flipped pattern when LDH-1 is higher than the LDH-2 iso-enzyme in patients affected with myocardial infarction. [63-68] Lately a report by van Wilpe et al [69] has reported the importance of LDH enzyme as a marker of diminished anti-tumor activity and another report by Bittar et al^[68] in suggested the importance of LDH in cancer therapy. The reports^[68,69] demonstrated that the increased LDH levels in the tumor is due to the hypoxia and the levels of LDH in the cancer therapeutically treated patients can be taken as the progression of the disease.

Figure 3:	Characteristic	features of	LDH	iso-enz	ymes.
-----------	----------------	-------------	-----	---------	-------

Iso-enzyme	Subunit make up of iso-enzyme	Tissue from which iso-enzyme has originated
LDH-1	H4	Heart muscle
LDH-2	H3M1	Red blood cell
LDH-3	H2M2	Brain
LDH-4	H1M3	Liver
LDH-5	M4	Skeletal muscle

Prostate Specific Antigen (PSA)

PSA is an enzyme normally produced by the glandular tissue of the prostate and secreted into the seminal fluid. [70-75] PSA was first isolated by Chu in 1980. [70,71] It is a glycoprotein with a molecular weight of 32kD. [71] The main function of the PSA is to liquefy seminal coagulum.^[70] Two iso-forms of PSA have been observed, one is complexes with alpha-1-antitrypsin and the other one is free PSA which is not bound.^[71] Physiologically, bound PSA is present in the blood predominantly than free PSA. [72,73] Although the isoforms of PSA are not diagnostically available, but research through the years studies have been published. The studies reported three iso-enzyme forms of PSA. [74,75] Lately, research through the years has reported three PSA isoforms, 10%-30% of PSA is unbounded as free PSA (earlier mentioned) in the serum and composed of various isoforms. [74-77] First isoenzyme is named as ProPSA, which comprises 33% of free PSA and increases with cancer, while the second is termed as BPSA, which is regarded as a nicked form of PSA, is secreted from the transitional zone and makes up 28% of free PSA. The third isoform is termed inactive or intact PSA (iPSA), which decreases with cancer. [74-77]

CONCLUSION

The purpose of this review was to view the latest improvements in the field of iso-enzymes from the past decades and see how the patterns of the iso-enzymes are changing in patho-physiology of different diseases. It is clear from the research reviewed that some enzymes like ALP, CK, and LDH have clear specific patterns in particular in some diseases. In the future studies, pattern of such iso-enzymes should be put into thought process in other diseases rather than in already known diseases. Further in some cases, despite the roles of adenosine deaminase, enolase and B-Glu have been studied for several years, the physiological role is still remains obscure in-terms of their respective iso-enzymes pattern in various diseases. In such cases, it is important to conduct more studies across the globe on the results and derive the facts to a conclusion.

Conflict of Interest

The authors have no conflict of interest among them.

REFERENCES

- 1. de Wildt SN. Profound changes in drug metabolism enzymes and possible effects on drug therapy in neonates and children. Expert opinion on drug metabolism & toxicology, 2011; 7(8): 935-948.
- 2. Shannon LM. Plant isoenzymes. Annual review of plant physiology, 1968; 19(1): 187-210.

- Kottel RH, Hanford WC. Differential release of membrane-bound alkaline phosphatase isoenzymes from tumor cells by bromelain. Journal of Biochemical and Biophysical Methods, 1980; 2(6): 325-330.
- 4. Markert CL, Møller F. Multiple forms of enzymes: tissue, ontogenetic, and species specific patterns. Proceedings of the National Academy of Sciences of the United States of America, 1959; 45(5): 753.
- Moss DW. Isoenzymes. Springer Science & Business Media, 2012.
- Rioux JA, Lanotte G, Serres E, et al. Taxonomy of Leishmania. Use of isoenzymes. Suggestions for a new classification. Annales de parasitologie humaine et compare, 1990; 65(3): 111-125.
- 7. O'Hare MC, Doonan S. Purification and structural comparisons of the cytosolic and mitochondrial isoenzymes of fumarase from pig liver. Biochimica et Biophysica Acta (BBA)-Protein Structure and Molecular Enzymology, 1985; 827(2): 127-134.
- 8. Kokkinidis M, Glykos NM, Fadouloglou VE. Catalytic activity regulation through post-translational modification: the expanding universe of protein diversity. Advances in Protein Chemistry and Structural Biology, 2020; 122: 97-125.
- Norbeck J, Påhlman AK, Akhtar N, et al. Purification and characterization of two isoenzymes of dl-glycerol-3-phosphatase from Saccharomyces cerevisiae identification of the corresponding GPP1 and GPP2 genes and evidence for osmotic regulation of Gpp2p expression by the osmosensing mitogenactivated protein kinase signal transduction pathway. Journal of Biological Chemistry, 1996; 271(23): 13875-13881.
- Cadeau BJ, Malkin A. A relative heat stability test for the identification of serum alkaline phosphatase isoenzymes. Clinica Chimica Acta., 1973; 45(3): 235-242.
- 11. Celebioglu HU, Erden Y, Hamurcu F, et al. Cytotoxic effects, carbonic anhydrase isoenzymes, α-glycosidase and acetylcholinesterase inhibitory properties, and molecular docking studies of heteroatom-containing sulfonyl hydrazone derivatives. Journal of Biomolecular Structure and Dynamics, 2020; 22: 1-2.
- of 12. Golpar G. Evaluation serum level organophosphates (Diazinon and Malathion) and their metabolits and its relation with genetic polymorphism of isoenzymes (CYP2C8, CYP19A1, CYP1B1, CYP1A1) and GSTP1 in breast cancer patients referring to Mazandaran University of Medical Sciences clinics:. a case control

www.ejpmr.com Vol 8, Issue 5, 2021. ISO 9001:2015 Certified Journal 281

- study (Doctoral dissertation. Mazandaran: Mazandaran university of medical sciences, 2020.
- 13. Bunik V, Kaehne T, Degtyarev D, et al. Novel isoenzyme of 2-oxoglutarate dehydrogenase is identified in brain, but not in heart. The FEBS journal, 2008; 275(20): 4990-5006.
- 14. Mimori T, Grimaldi Jr G, Kreutzer RD, et al. Identification, using isoenzyme electrophoresis and monoclonal antibodies, of Leishmania isolated from humans and wild animals of Ecuador. The American journal of tropical medicine and hygiene, 1989; 40(2): 154-158.
- 15. Rahim SM, Delaunoy JP, Laurent P. Identification and immunocytochemical localization of two different carbonic anhydrase isoenzymes in teleostean fish erythrocytes and gill epithelia. Histochemistry, 1988; 89(5): 451-459.
- 16. Van der Weyden MB, Kelley WN. Human adenosine deaminase. Distribution and properties. Journal of Biological Chemistry, 1976; 251(18): 5448-5456.
- 17. Giblett E, Anderson J, Cohen F, et al. Adenosine-deaminase deficiency in two patients with severely impaired cellular immunity. The Lancet., 1972; 300(7786): 1067-1069.
- 18. Flinn AM, Gennery AR. Adenosine deaminase deficiency: a review. Orphanet journal of rare diseases, 2018; 13(1): 1-7.
- 19. Antonyan AA, Karapetyan LG, Sharoyan SG, et al. Isoforms of adenosine deaminase1 in synovial fluids at different arthritis. Հայшиտшնի կենսաբանական հանդես Biological Journal of Armenia Биологический журнал Армении, 2017; 69(1): 92-96.
- 20. Ombrello AK, Qin J, Hoffmann PM, et al. Treatment strategies for deficiency of adenosine deaminase 2. The New England journal of medicine, 2019; 380(16): 1582.
- 21. Moens L, Hershfield M, Arts K, et al. Human adenosine deaminase 2 deficiency: A multi-faceted inborn error of immunity. Immunological Reviews, 2019; 287(1): 62-72.
- 22. Tardif V, Muir R, Cubas R, et al. Adenosine deaminase-1 delineates human follicular helper T cell function and is altered with HIV. Nature communications, 2019; 10(1): 1-5.
- 23. Chang YC, Yang YC, Tien CP, et al. Roles of aldolase family genes in human cancers and diseases. Trends in Endocrinology & Metabolism, 2018; 29(8): 549-559.
- 24. Kusakabe T, Motoki K, Hori K. Mode of interactions of human aldolase isozymes with cytoskeletons. Archives of biochemistry and biophysics, 1997; 344(1): 184-193.
- 25. Asaka M, Kimura T, Meguro T, et al. Alteration of aldolase isozymes in serum and tissues of patients with cancer and other diseases. Journal of clinical laboratory analysis, 1994; 8(3): 144-148.
- 26. Ojika T, Imaizumi M, Abe T, et al. Immunochemical and immunohistochemical studies

- on three aldolase isozymes in human lung cancer. Cancer, 1991; 67(8): 2153-2158.
- McComb RB, Bowers Jr GN, Posen S. Alkaline phosphatase. Springer Science & Business Media, 2013.
- 28. Komoda T, Koyama I, Nagata A, et al. Ontogenic and phylogenic studies of intestinal, hepatic, and placental alkaline phosphatases: evidence that intestinal alkaline phosphatase is a late evolutionary development. Gastroenterology, 1986; 91(2): 277-286.
- 29. Crofton PM. Biochemistry of alkaline phosphatase isoenzymes. CRC Critical Reviews in Clinical Laboratory Sciences, 1982; 16(3): 161-194.
- 30. Goltzman D, Miao D. Alkaline phosphatase.
- 31. Wiwanitkit V. High serum alkaline phosphatase levels, a study in 181 Thai adult hospitalized patients. BMC family practice, 2001; 2(1): 2.
- 32. Yu MC, Chan KM, Lee CF, et al. Alkaline phosphatase: does it have a role in predicting hepatocellular carcinoma recurrence?. Journal of Gastrointestinal Surgery, 2011; 15(8): 1440-1449.
- 33. Singh AK, Pandey A, Tewari M, et al. Advanced stage of breast cancer hoist alkaline phosphatase activity: risk factor for females in India. 3 Biotech, 2013; 3(6): 517-520.
- 34. Withold WO, Schulte U, Reinauer H. Method for determination of bone alkaline phosphatase activity: analytical performance and clinical usefulness in patients with metabolic and malignant bone diseases. Clinical chemistry, 1996; 42(2): 210-217.
- 35. Garnero P, Delmas PD. Assessment of the serum levels of bone alkaline phosphatase with a new immunoradiometric assay in patients with metabolic bone disease. The Journal of clinical endocrinology & metabolism, 1993; 77(4): 1046-1053.
- 36. Byers DA, Fernley HN, Walker PG. Studies on Alkaline Phosphatase: Inhibition of Human-Placental Phosphoryl Phosphatase by 1-Phenylalanine. European Journal of Biochemistry, 1972; 29(2): 197-204.
- 37. Bukowczan J, Pattman S, Jenkinson F, et al. Regan isoenzyme of alkaline phosphatase as a tumour marker for renal cell carcinoma. Annals of clinical biochemistry, 2014; 51(5): 611-614.
- 38. Jefferson RA, Burgess SM, Hirsh D. beta-Glucuronidase from Escherichia coli as a genefusion marker. Proceedings of the National Academy of Sciences, 1986; 83(22): 8447-8451.
- 39. SADAHIRO R, TAKANASHI S, KAWADA M. Studies on the Isozyme of β-Glucuronidase. The Journal of Biochemistry, 1965; 58(1): 104-106.
- 40. Fondo Jr EY. Beta-glucuronidase. The significance of its isozymes. Journal of the National Medical Association, 1968; 60(4): 312.
- 41. Pearson JP, Pretlow TP, Bradley Jr EL, et al. Beta-glucuronidase activity in prostatic carcinoma and benign prostatic hyperplasia. Cancer, 1989; 64(4): 911-915.

- 42. Vaquero C, Masson C, Guigon M, et al. Beta-glucuronidase in human cutaneous tumours. European Journal of Cancer (1965), 1975; 11(10): 739-746.
- 43. KAKIZOE T, KAWACHI T, SUGIMURA T. β-Glucuronidase isozyme patterns of experimental hepatomas of rats. GANN Japanese Journal of Cancer Research, 1976; 67(2): 289-294.
- 44. Wu AH. Creatine kinase isoforms in ischemic heart disease. Clinical chemistry, 1989; 35(1): 7-13.
- 45. Clarkson PM, Apple FS, Byrnes WC, et al. Creatine kinase isoforms following isometric exercise. Muscle & nerve., 1987; 10(1): 41-44.
- 46. Wallimann T, Hemmer W. Creatine kinase in non-muscle tissues and cells. Molecular and cellular biochemistry, 1994; 133(1): 193-220.
- 47. Wallimann T. Bioenergetics: dissecting the role of creatine kinase. Current biology, 1994; 4(1): 42-46.
- 48. Gerlt JA, Babbitt PC, Rayment I. Divergent evolution in the enolase superfamily: the interplay of mechanism and specificity. Archives of biochemistry and biophysics, 2005; 433(1): 59-70.
- 49. Lohman K, Meyerhof O. Enzymatic transformation of phosphoglyceric acid into pyruvic and phosphoric acid. Biochem. Z., 1934; 273: 60-72.
- 50. Peshavaria M, Day IN. Molecular structure of the human muscle-specific enolase gene (ENO3). Biochemical journal, 1991; 275(2): 427-433.
- 51. Pancholi V. Multifunctional α-enolase: its role in diseases. Cellular and Molecular Life Sciences CMLS., 2001; 58(7): 902-20.
- 52. Royds JA, Timperley WR, Taylor CB. Levels of enolase and other enzymes in the cerebrospinal fluid as indices of pathological change. Journal of Neurology, Neurosurgery & Psychiatry, 1981; 44(12): 1129-1135.
- 53. Roine RO, Somer H, Kaste M, et al. Neurological outcome after out-of-hospital cardiac arrest: prediction by cerebrospinal fluid enzyme analysis. Archives of Neurology, 1989; 46(7): 753-756.
- 54. Hay EL, Royds JA, Davies-Jones GA, et al. Cerebrospinal fluid enolase in stroke. Journal of Neurology, Neurosurgery & Psychiatry, 1984; 47(7): 724-729.
- 55. Baginski ES, Foà PP, Zak B. Glucose-6-phosphatase. InMethods of enzymatic analysis. Academic Press, 1997.
- 56. Van Schaftingen E, Gerin I. The glucose-6-phosphatase system. Biochemical Journal, 2002; 362(3): 513-532.
- 57. Swanson MA. [83] Glucose-6-phosphatase from liver.
- 58. Hutton JC, O'Brien RM. Glucose-6-phosphatase catalytic subunit gene family. Journal of Biological Chemistry, 2009; 284(43): 29241-29245.
- 59. Ghosh A, Shieh JJ, Pan CJ, et al. Histidine 167 is the phosphate acceptor in glucose-6-phosphatase-β forming a phosphohistidine enzyme intermediate during catalysis. Journal of Biological Chemistry, 2004; 279(13): 12479-12483.

- 60. Shieh JJ, Pan CJ, Mansfield BC, et al. In islet-specific glucose-6-phosphatase-related protein, the beta cell antigenic sequence that is targeted in diabetes is not responsible for the loss of phosphohydrolase activity. Diabetologia, 2005; 48(9): 1851-1859.
- 61. Abbadi S, Rodarte JJ, Abutaleb A, et al. Glucose-6–phosphatase Is a Key Metabolic Regulator of Glioblastoma Invasion. Molecular Cancer Research, 2014; 12(11): 1547-1559.
- 62. Burt BM, Humm JL, Kooby DA, et al. Using positron emission tomography with [18F] FDG to predict tumor behavior in experimental colorectal cancer. Neoplasia, 2001; 3(3): 189-195.
- 63. Farhana A, Lappin SL. Biochemistry, Lactate Dehydrogenase (LDH). StatPearls Publishing, 2020.
- 64. Hoffman NE, Bent AF, Hanson AD. Induction of lactate dehydrogenase isozymes by oxygen deficit in barley root tissue. Plant physiology, 1986; 82(3): 658-663.
- 65. Read JA, Winter VJ, Eszes CM, et al. Structural basis for altered activity of M-and H-isozyme forms of human lactate dehydrogenase. Proteins: Structure, function, and bioinformatics, 2001; 43(2): 175-185.
- 66. Markert CL, Faulhaber I. Lactate dehydrogenase isozyme patterns of fish. Journal of Experimental Zoology, 1965; 159(3): 319-332.
- 67. Wróblewski F, Gregory KF. Lactic dehydrogenase isozymes and their distribution in normal tissues and plasma and in disease states. Annals of the New York Academy of Sciences, 1961; 94(3): 912-932.
- Bittar PG, Charnay Y, Pellerin L, et al. Selective distribution of lactate dehydrogenase isoenzymes in neurons and astrocytes of human brain. Journal of Cerebral Blood Flow & Metabolism, 1996; 16(6): 1079-1089.
- 69. Van Wilpe S, Koornstra R, Den Brok M, et al. Lactate dehydrogenase: a marker of diminished antitumor immunity. OncoImmunology, 2020; 9(1): 1731942.
- 70. Rittenhouse HG, Finlay JA, Mikolajczyk SD, et al. Human kallikrein 2 (hK2) and prostate-specific antigen (PSA): two closely related, but distinct, kallikreins in the prostate. Critical reviews in clinical laboratory sciences, 1998; 35(4): 275-368.
- 71. Polascik TJ, Oesterling JE, Partin AW. Prostate specific antigen: a decade of discovery-what we have learned and where we are going. The Journal of urology, 1999; 162(2): 293-306.
- 72. Freedland SJ, Platz EA, Presti Jr JC, et al. Obesity, serum prostate specific antigen and prostate size: implications for prostate cancer detection. The Journal of urology, 2006; 175(2): 500-504.
- 73. Hudson ML, Bahnson RR, Catalona WJ. Clinical use of prostate specific antigen in patients with prostate cancer. The Journal of urology, 1989; 142(4): 1011-1017.
- 74. Özen H, Sözen S. PSA isoforms in prostate cancer detection. european urology supplements, 2006; 5(6): 495-499.

- 75. Hori S, Blanchet JS, McLoughlin J. From prostate-specific antigen (PSA) to precursor PSA (proPSA) isoforms: a review of the emerging role of proPSAs in the detection and management of early prostate cancer. BJU international, 2013; 112(6): 717-728.
- 76. Jansen FH, van Schaik RH, Kurstjens J, et al. Prostate-specific antigen (PSA) isoform p2PSA in combination with total PSA and free PSA improves diagnostic accuracy in prostate cancer detection. European urology, 2010; 57(6): 921-927.
- 77. Guazzoni G, Nava L, Lazzeri M, et al. Prostate-specific antigen (PSA) isoform p2PSA significantly improves the prediction of prostate cancer at initial extended prostate biopsies in patients with total PSA between 2.0 and 10 ng/ml: results of a prospective study in a clinical setting. European urology, 2011; 60(2): 214-222.

www.ejpmr.com Vol 8, Issue 5, 2021. ISO 9001:2015 Certified Journal 284