



**DETERMINATION OF ANTIOXIDANT PROPERTY OF PLANT EXTRACTS OF  
GERMINATED BROWN RICE (*ORYZA SATIVA L.*) AND COMPARISON WITH NON-  
GERMINATED BROWN RICE**

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**ABSTRACT**

Brown rice (*Oryza sativa L.*), is a most important pigmented rice which is abundant in many bioactive compounds. Due to the presence of bioactive compounds, brown rice is known for different Properties such as antioxidant, anti-inflammatory, anti-diabetic, anti- microbial and anti-malarial properties. Antioxidant property is the capacity to hinder the oxidation of biological compounds, thereby reduce risk of degenerative diseases. Brown rice contain high antioxidant capacity than normal white rice varieties, since it is rich in polyphenols, flavonoids, vitamins, antho-cyanidins, etc. The germination process will enhance the amount of bio active compounds. There are large number of synthetic antioxidants, but taking plant-based food having antioxidant activity is always a good option. In this study, we are evaluating the antioxidant property of GBR through phytochemical analysis, GC-MS analysis, estimation of antioxidant activity by estimation of DPPH scavenging, TPC, TFC, SOD AND H<sub>2</sub>O<sub>2</sub> scavenging.

**KEYWORDS:** Antioxidants, Degenerative disease, GBR, Bioactive compounds.

**INTRODUCTION**

Rice is a monocotyledonous annual grass plant of Poaceae family. The Main species of rice is *Oryza sativa L.*, grown mainly in Asian countries<sup>[1]</sup> Rice (*Oryza sativa L.*) is one of popular staple food consumed by approximately 2.5 billion people.<sup>[2]</sup> According to the color of outer layer, the rice varieties classified in to brown rice, black rice, red rice and purple rice.<sup>[3]</sup> The brown rice is a whole grain and it consist of all parts of a rice kernel such as bran, germ endosperm.<sup>[4]</sup> Hence the brown rice is very nutritious since it contains all vital parts of rice grain so more amount of nutrients. Milling of brown rice yield white rice, which is tastier and have long shelf life. But, it losses the bran and germ portions by the process of milling. So that the nutritional quality of white rice is low compared to white rice.<sup>[3]</sup> The antioxidant property of brown rice is very high compared to white rice because it contains large quantity of bio-active compounds like phenolic compounds, flavonoids, gamma oryzanol, ferulic acid, alpha tocopherol, gamma amino butyric acid (GABA), lignin, etc.<sup>[5]</sup>

**Germinated Brown Rice (GBR)**

The sprouted brown rice is obtained by soaking method. Brown rice soaking in water for 3 hours and leaving wet for 21 hours will produce budding. At the germination time, the nutrient quality, taste, enzyme activity and chemical compositions of rice changes. The quality,

flavour and texture of unpolished brown rice increased through germination also essential nutrients like vitamins, minerals, fiber, ferulic acid, lysin, inositol, GABA, gamma oryzanol, tocotrienol increased<sup>[6]</sup> GBR alleviate health problems like DM, cardio vascular disease, hypertension, Alzheimer's, stress, depression, insomnia, dementia, colon cancer, constipation, etc.<sup>[7]</sup> Optimum amount of gamma amino butyric acid (GABA) obtained through sprouting. GABA is an inhibitory neurotransmitter, which reduce health risks and Presence of high GABA makes GBR popular. GBR contain two times higher content of GABA than normal brown rice and 10 times higher GABA than white rice.<sup>[7]</sup>

**MATERIALS AND METHODS**

Brown rice (1 kg) was collected from domestic market of Malappuram of Kerala state, India (2021 January).

**Preparation of material:** The rice grains were thoroughly washed and soaked in water overnight. Then leave it moist for 20-24 hours at 30° C. The sprouting appeared within 2-3 days, shade dried it and grinded into powder.

**Extraction of sample:** 20 g powder was weighed and suspended in in 200 ml of water, ethanol solvents. Extraction was done by using Soxhlet apparatus for 5 hours.

### GC-MS Analysis of Bioactive Compounds

Gas Chromatography-Mass Spectrometry (GCMS), is a combination of gas chromatography and mass spectrometry used to identify unknown samples and separate volatile and semi-volatile compounds. Ethanol extract of brown rice used for this technique. Gas chromatography combined with Shimadzu mass spectrometry (GC-MS- QP 2020 NX SHIMADZU). GC consist of a capillary column (DB- 5ms) of fused silica having film thickness of 0.25 micro meter. The carrier gas used was Helium at 1.0 ml/min flow rate. 1 micro litre sample was injected by autoinjector. The temperature of injector and detector was set at 250° C. The primary temperature of column is 40° C raised to 1000 C at 5° C/min for 3 min, again raised temperature 250°C at 10° C/min. The ion source was set at 200° C. Whole process of GC carried out at 39 mins. For the Mass Spectrometric (MS) detection were adjusted to an ionization energy of 70 eV. Temperature of electron impact ion source was set at 260° C and transmission line at 280° C. The mass scan range was m/z 40-300 for volatile compounds, since their molecular weight is low. The ethanolic extract of brown rice was qualitatively detected by a Shimadzu GC-MS solution (version.4) software.

### Identification of Compounds

The mass spectrum of GC interpreted by using Databases of National Institute of Standard and Technology (NIST4) and WILEY9. It was by the comparison of spectrum of a known component with spectrum of unknown component stored in inbuilt library.

### Antioxidant property of germinated brown rice (*Oryza sativa* L.)

#### Estimation of enzymatic antioxidants

##### a) Estimation of SOD

Estimation of SOD was based on inhibition of nitro-blue formation of superoxide ion by plant extracts, measured spectrophotometrically at 560 nm. 3 ml samples were taken, which contain 0.02 ml extract, 0.2 ml EDTA, 0.1 ml NBT, 0.05 of riboflavin and 2.64 ml of phosphate buffer. Control tubes were set up, where DSMO was added instead of plant extract. All tubes were vortexed, initial optical density measured at 560 nm. Tubes were illuminated using fluorescent lamp for 30 minutes. Then absorbance measured at 560 nm. Illumination was indicative of superoxide ion scavenging activity.

##### b) Estimation of hydrogen peroxide scavenging assay

The ability of plant extract to scavenge hydrogen peroxide was estimated by method of *Ruch et.al* (1989). Solution of H<sub>2</sub>O<sub>2</sub> (40mM) in phosphate buffer. Plant extracts at concentration of 10mg/micro litre was added to H<sub>2</sub>O<sub>2</sub> solution (0.6ml), total volume was made up to 3ml. Absorbance of solution was recorded at 230 nm. Blank solution containing phosphate buffer without H<sub>2</sub>O<sub>2</sub> was prepared.

### Estimation of non-enzymatic antioxidants

#### a) Estimation of total flavonoid content

##### ALUMINIUM CHLORIDE METHOD

Aluminium chloride reacts with C-4 keto group and either the C-3 or C-5 hydroxyl groups of flavones and flavanols and forms acid-labile compounds with flavonoids. 0.5 ml sample added into test tube containing 1.25 ml distilled water. Then 0.075 ml of sodium nitrite was added and allowed to stand for 5 minutes. 0.15 The mixture was diluted by 0.75 ml distilled water. The absorbance of solution at 510 nm was measured. The resultant flavonoid content was expressed in milligram catechin.

#### b) Estimation of total phenolic compounds

##### FOLIN-CIOCALTEU METHOD

Phenol reacts with phosphomolybdic acid Folin-cioalteu reagent in alkaline medium, produce bluish coloured complex. Pipetted out different volumes of (0.2-1ml) of standard into test tube and made up to 3 ml with distilled water, added 0.5 ml folin-cioalteu reagent. After 3 minutes, added 2 ml of 20 % sodium carbonate to each tube, mixed well, boiled for exact 1 minute, cooled and read absorbance at 660 nm colorimetrically. Prepared calibration curve to different concentrations.

### Estimation of antioxidants by DPPH assay

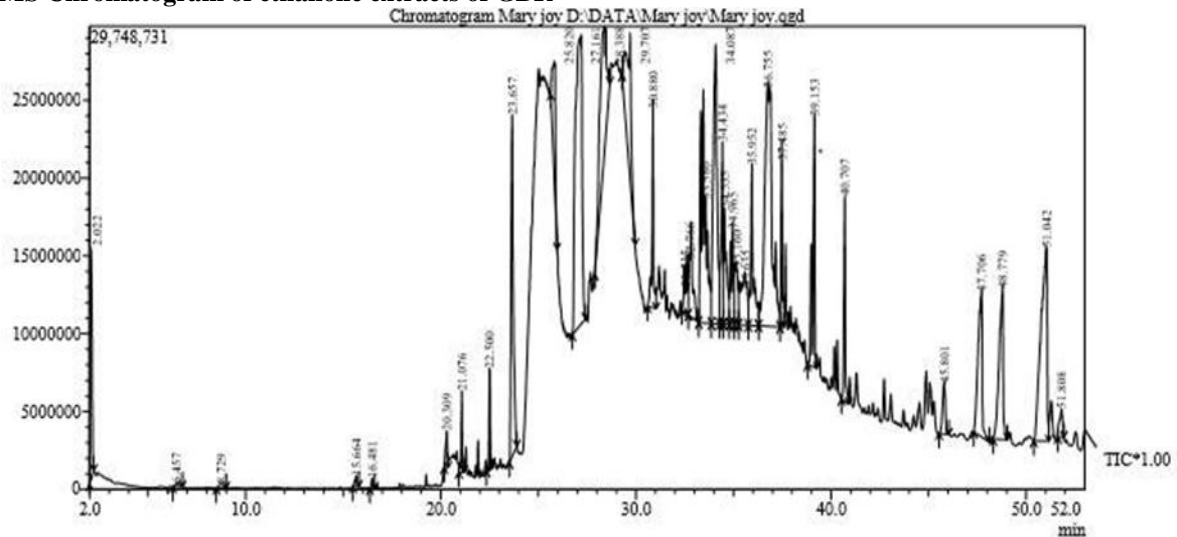
Antioxidants inhibit lipid oxidation, so scavenging of DPPH radical hence determine free radical scavenging ability. it is widely used method. 0.1 mM solutions of DPPH in methanol prepared, 1ml of this solution added into 3 ml extract in methanol (concentrations 50,100,200,400 &800 micro gram/ml). shaken vigorously allowed to stand for 30 minutes at room temperature. Read absorbance at 517 nm. If the absorbance value is lower, the more is the free radical scavenging capacity. The capacity of radical scavenging calculated by following formula

$$\text{DPPH scavenging effect (\% of inhibition)} = \left\{ \frac{(A_0 - A_1)}{A_0} \right\} * 100$$

Where, A<sub>0</sub> is absorbance of control reaction, A<sub>1</sub> is absorbance in presence of all extract sample and reference. All tests performed in three times and results were averaged.

RESULTS AND DISCUSSIONS

GC-MS Chromatogram of ethanolic extracts of GBR

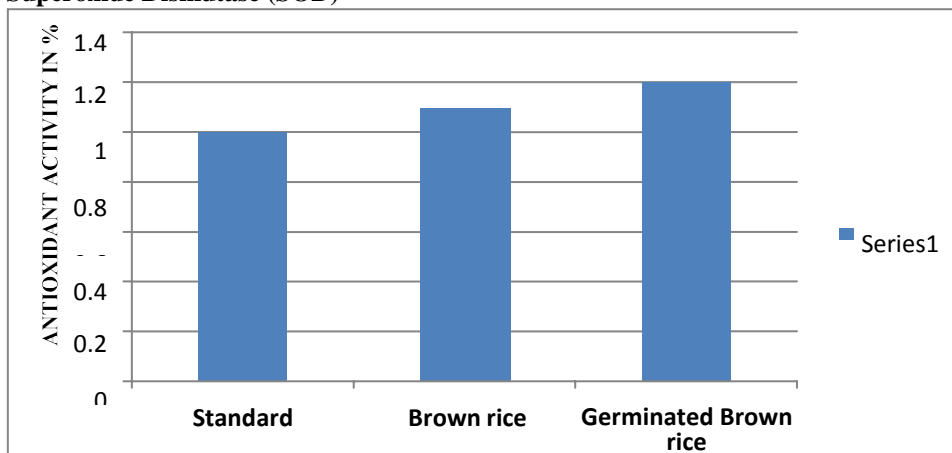


Mass spectrometry of ethanolic extract of GBR

| Peak# | R. Time | I. Time | F. Time | Area      | Height   | A/H Mark | Name   |
|-------|---------|---------|---------|-----------|----------|----------|--|
| 1     | 2.022   | 2.005   | 2.210   | 109029955 | 15231739 | 7.16     | ETHANOL  |
| 2     | 6.457   | 6.260   | 6.770   | 307569312 | 12489670 | 1.82     | MI BENZOFURAN, 2,3-DIHYDRO-  |
| 3     | 8.729   | 8.495   | 9.000   | 486929134 | 13562748 | 2.11     | MI 2-Methoxy-4-vinylphenol   |
| 4     | 15.664  | 15.495  | 15.800  | 527883473 | 1537546  | 3.20     | MI Dodecanoic acid   |
| 5     | 16.481  | 16.360  | 16.615  | 319414721 | 2126453  | 1.58     | MI 1-NONADECENE  |
| 6     | 20.309  | 20.165  | 20.370  | 14434126  | 2366781  | 6.10     | MI Tetradecanoic acid  |
| 7     | 21.076  | 20.925  | 21.180  | 19169803  | 5225611  | 3.67     | MI 1-Nonadecene  |
| 8     | 22.500  | 22.345  | 22.500  | 14796573  | 6569139  | 2.25     | MI 9-Heptadecanone   |
| 9     | 23.657  | 23.520  | 23.895  | 179586423 | 21953719 | 14.30    | HEXADECANOIC ACID, METHYL ESTER  |
| 10    | 25.820  | 25.630  | 25.960  | 79675683  | 7815793  | 10.19    | HEXADECANOIC ACID  |
| 11    | 27.161  | 26.740  | 27.410  | 439531055 | 18491181 | 18.26    | 9-OCTADECENOIC ACID (Z)-, METHYL ESTER   |
| 12    | 28.388  | 27.890  | 28.660  | 217382852 | 7941251  | 27.37    | E,E-3,13-Octadecadien-1-ol   |
| 13    | 29.707  | 29.285  | 29.960  | 142338873 | 8903067  | 15.99    | Z-9-Pentadecenol   |
| 14    | 30.880  | 30.585  | 31.035  | 68129511  | 12654722 | 7.67     | Ethanamine, 2,2'-oxybis[N,N-dimethyl-  |
| 15    | 32.535  | 32.360  | 32.685  | 35900523  | 1628949  | 22.04    | ETHYL DOCOSANOATE  |
| 16    | 32.766  | 32.685  | 33.210  | 66181889  | 3928932  | 16.84    | V Heneicosane  |
| 17    | 33.580  | 33.210  | 33.860  | 232625425 | 7924892  | 29.35    | 9,12-OCTADECADIENOIC ACID (Z,Z)-, 2,3-DIHYDROXYPROPYL ESTER  |
| 18    | 34.087  | 33.860  | 34.285  | 248458971 | 17694836 | 4.81     | V Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester  |
| 19    | 34.434  | 34.285  | 34.485  | 75148130  | 11225974 | 5.05     | V HEXATRIACONTANE  |
| 20    | 34.535  | 34.485  | 34.785  | 70670572  | 7226921  | 9.78     | V 1,2-Benzenedicarboxylic acid, diisooctyl ester   |
| 21    | 34.965  | 34.785  | 35.035  | 56146754  | 6153876  | 9.12     | V 9-Octadecenoic acid (Z)-, tetradecyl ester   |
| 22    | 35.160  | 35.035  | 35.260  | 44715962  | 3900863  | 11.46    | V 1,3-Benzenediol, 5-pentadecyl-   |
| 23    | 35.635  | 35.260  | 35.760  | 76852936  | 2877576  | 26.71    | V Hexadecane   |
| 24    | 35.952  | 35.760  | 36.285  | 91129424  | 9753809  | 9.34     | V Triacontane  |
| 25    | 36.755  | 36.285  | 37.385  | 412729696 | 15435531 | 3.07     | V 9,12-OCTADECADIENOIC ACID (Z,Z)-, 2,3-DIHYDROXYPROPYL ESTER                                      |
| 26    | 37.485  | 37.385  | 37.585  | 56140527  | 11621215 | 6.99     | V HEXATRIACONTANE  |
| 27    | 39.153  | 38.785  | 39.285  | 113656369 | 15884252 | 12.36    | V 2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23-hexamethyl-, (all-E)-                          |
| 28    | 40.707  | 40.535  | 40.910  | 68026677  | 12668315 | 11.36    | V Hexatriacontane  |
| 29    | 45.801  | 45.535  | 46.035  | 32615498  | 3246927  | 10.05    | V 2H-1-BENZOPYRAN-6-OL, 3,4-DIHYDRO-2,7,8-TRIMETHYL-2-(4,8,12,16,20,24,28,32-OCTAMETHYL-3,7,11,15, |
| 30    | 47.706  | 47.310  | 48.110  | 138656629 | 9254872  | 14.98    | V ERGOST-5-EN-3-OL, (3.BETA.,.24R)-  |
| 31    | 48.779  | 48.310  | 49.035  | 137635682 | 9489605  | 14.50    | V Stigmasterol   |
| 32    | 51.042  | 50.385  | 51.160  | 277093698 | 12455156 | 7.80     | V ERGOST-5-EN-3-OL, (3.BETA.)-   |
| 33    | 51.808  | 51.625  | 51.930  | 19779712  | 1829446  | 10.81    | MI Cholest-4-en-3-one  |

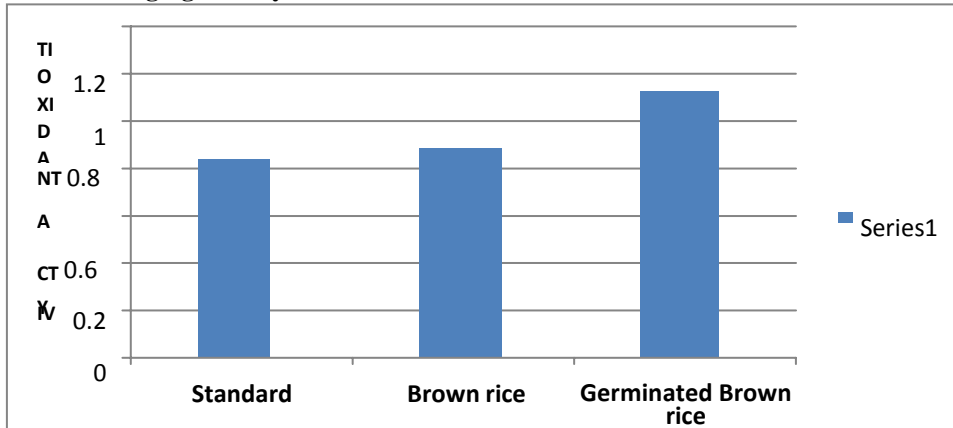
Antioxidant activity analysis

Estimation of Superoxide Dismutase (SOD)



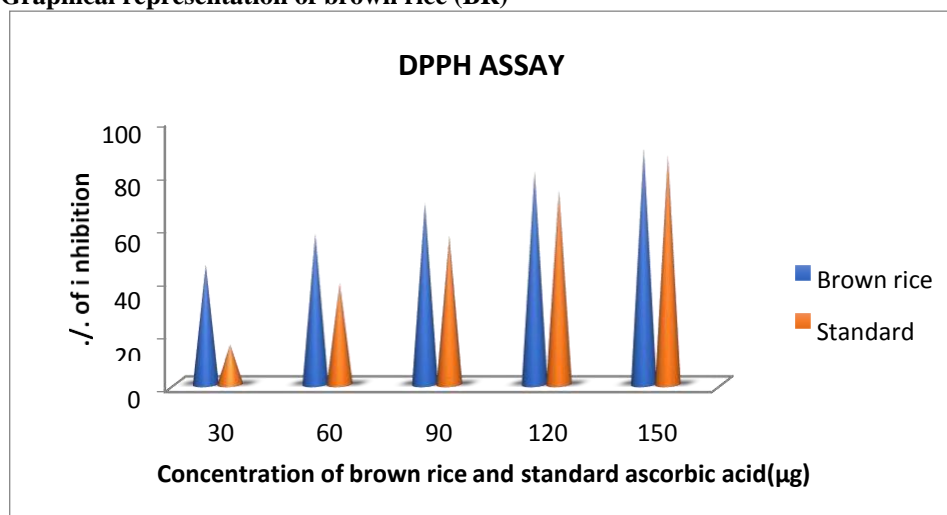
The result shows the GBR contain higher activity of SOD, compared with normal brown rice.

**Estimation of H<sub>2</sub>O<sub>2</sub> scavenging activity**

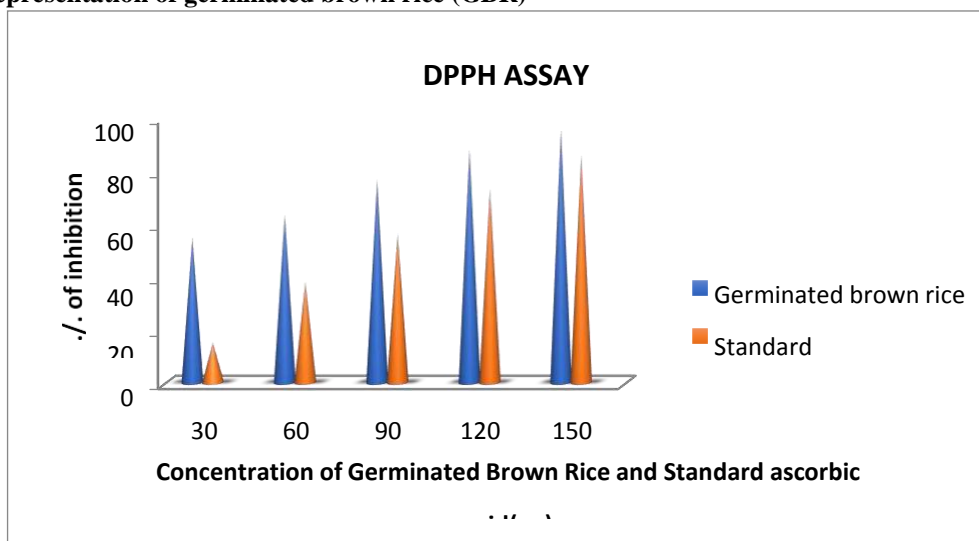


From the graph, the H<sub>2</sub>O<sub>2</sub> scavenging capacity of GBR was higher when compared with non-germinated brown rice.

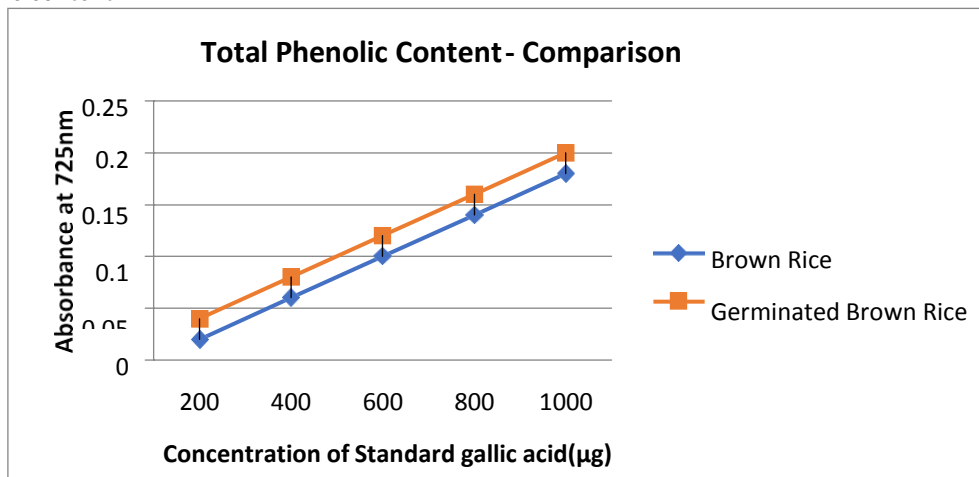
**DPPH assay: Graphical representation of brown rice (BR)**



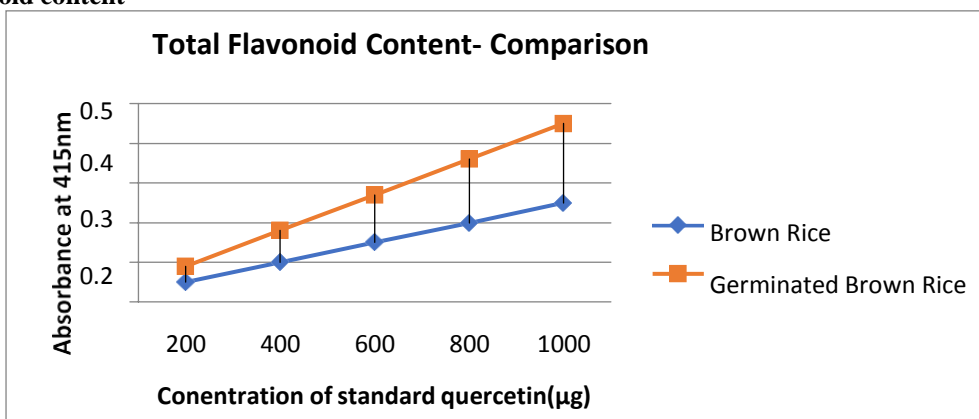
**Graphical representation of germinated brown rice (GBR)**



The results show that the percentage of inhibition rate is more in GBR when compared to non-germinated brown rice.

**Total phenolic content**

From this result shows that the total phenolic content is more in germinated brown rice than non-germinated brown rice.

**Total flavonoid content**

Results shows that the total flavonoid content is higher in germinated brown rice than non-germinated brown rice.

**CONCLUSION**

In this study, the *Antioxidant property of germinated brown rice* was determined and also *Comparison of GBR with non-germinated brown rice* were conducted. The germinated brown rice contains high levels of bioactive compounds like phenolics, flavonoid, flavanols, anthocyanidins, vitamins and more amino acid content than non-germinated brown rice, this makes the GBR more potent in antioxidant activity.

The bioactive compounds of GBR were evaluated by GC-MS analysis thus obtained chromatogram of ethanolic extract of GBR. The antioxidant property of GBR was assessed by using estimation of enzymatic (SOD, H<sub>2</sub>O<sub>2</sub> scavenging activity) and non-enzymatic (total phenolic content, total flavonoid content, DPPH scavenging assay) activities. The free radical scavenging capacity of GBR was determined by DPPH scavenging and H<sub>2</sub>O<sub>2</sub> radical scavenging assays.

In both assays GBR shows very high scavenging activity than standard and non-germinated brown rice. Thus, it can be concluded that germinated brown rice (*Oryza*

*sativa* L.) have potent antioxidant property and this is higher than non-germinated brown rice.

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