

**TO DEVELOP AND FORMULATION OF FLOATING TABLETS ALLOPURINOL TO
ENHANCE THE BIOAVAILABILITY****Nikhil Pandey*, Praveen Kumar and Dr. Shamim Ahmed**

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ABSTRACT

Drug release and dissolution from the dose form retained in the stomach can occur at the stomach's pH below strictly controlled parameters. One of the most extensively trained drug transport systems with stomach retentive behavior is the floating system. Medication such as furosemide, ciprofloxacin, cyclosporine, and allopurinol should take advantage of gastric maintenance. Medication whose solubility is lower in the higher pH of the small intestine than the stomach (e.g., chlorthalidone and cinnarizine), medication prone to intestinal pH corruption (e.g., captopril), and medication for localized stomach exercise (e.g., misoprostol) can be administered as dose structures with gastric maintenance. The dosage structure of HBS allows for the regulation of anti-infection drugs, catecholamines, opioids, analgesics, anticonvulsants, muscle relaxants, antihypertensive drugs, and vitamins. The following medications are listed as acceptable for use in the definition of coasting measurements buildings: drifting drugs and capsules (acetaminophen, ampicillin, amoxycillin-trihydrate, diltiazem, fluorouracil, isosorbide-mononitrate, paraaminobenzoic corrosive, piretamide, theophylline, and verapamil hydrochloride, etc.); floating microspheres (headache medicine, griseofulvin, p-nitro aniline, ibuprofen, terfenadine, and tranilast).

KEYWORDS: Bioavailability, Allopurinol, Validation, Microspheres.**1.1. INTRODUCTION**

When it comes to ease of use, patient compliance, and formulation flexibility, oral therapy is the most widely used drug delivery technique. To combat the variations in drug plasma levels that are as frequently as possible observable using traditional dosage forms, such as drug release rapid. It led to the development of some kind of innovative drug delivery systems that could change detailed methods and provide a range of therapeutic benefits.^[1]

Such innovative medication delivery systems' primary goals are.

A single dosage of the medication would release the active ingredient over an extended duration.

Delivering the dynamic ingredient straight to the location of activity would minimize or completely eradicate the negative effects of the medication.

The planning of an oral controlled or sustained drug delivery system should be fundamentally altered in order to get a more consistent and increased bioavailability of pharmaceuticals. A few physiological issues, such as the difficulty to contain and limit the DDS inside the desired region of the GIT and the fundamental structure of the gastric emptying process, impede the advancement

process. The factors that affect oral drug delivery include the place of medicine consumption and the length of time that pharmaceuticals transit through the gastrointestinal tract.^[2] A few physiological constraints affect the majority of oral measurement structures, such as gastrointestinal transit due to varying gastric releasing causing non-uniform ingestion profiles, divided medication discharge, and a shorter half-life of the dose structure in the stomach.^[3] This results in insufficient consumption of maintenance-phase drugs, particularly in the upper portion of the small intestine, since the medicine is not fully absorbed once it passes through the osmosis site. A few factors affect the dosage forms in gastric emptying individuals, which leads to the vast intra- and between-subject variations that are seen.^[4]

Given that many drugs are especially concentrated in the upper portion of the gastrointestinal tract, this substantial variability may cause non-uniform retention and unpredictable bioavailability. Therefore, a useful medication delivery system would be one that can regulate and prolong the stomach emptying duration, which can be used to deliver the drugs in higher concentrations to the point of consumption (e.g., the upper portion of the small intestine).^[5]

Numerous synthetic elements have been introduced as a result; some can be ingested anywhere in the gastrointestinal tract (GIT), while others have windows for absorption (such as the upper portion of the small intestine), and some medications have low solubility in intestinal media. A specific delivery mechanism is needed for medications that fall into the second and third categories as well as those that are necessary for local activity in the stomach. Gliding drug delivery systems (FDDS) can be used to meet all of the aforementioned requirements and provide excellent medication delivery to the ingestion window, for nearby action, and for the treatment of gastrointestinal issues such as gastro-esophageal reflux.^[6]

1.2. METHODS

Three phases went into the development of the Allopurinol floating pills. Initially, different polymers were used in varying concentrations to make preliminary batches. These prepared batches' physicochemical characteristics were assessed. Three concentrations (lower, -1; medium, 0; and maximum, 1) were chosen from the two most promising polymers employed in the formulation of the tablets to be evaluated. Using a 32 response surface complete factorial design, optimization batches were created in the second step. Nine batches in all were made. The lag and float times, or the dissolving rate under study, determine how the polymer concentration changes. The most effective formulations in the final step would use optimal polymers to achieve the intended results (lag time of 45 seconds, floating time of 24 hours, and dissolving rate of 90%). This formulation was made, then assessed.^[7]

2.2.1 PRELIMINARY TABLETS PREPARATION

ALLOPURINOLL fueled Because of its many benefits, floating tablets were created utilizing the direct compression approach.^[8]

- Easiest way to manufacture tablets.
- High doses can be accommodated.
- Use of conventional equipment.
- Use of commonly available excipients.
- Fewer steps for processing.

2.2.1.1 DIRECT COMPRESSION TECHNIQUE USED FOR FLOATING TABLET

Allopurinol's gastro-retentive floating tablets were made with a variety of swellable polymers, including K100M, HPMC, K15M, and K4M. Other polymers, such as citric acid or sodium bicarbonate (NaHCO_3), are utilized as gas-generating agents. Certain diluents or binders are utilized, such as talc for glidants and magnesium stearate as lubricant. For consistent tablet punching during preparation, all combined formulations were run through sieve number 44. A single punch direct compression technique was employed, and each tablet weighed 200 mg. The manufactured punch tablets were used to test the parameters of the evaluation.^[9]

3. RESULT AND DISCUSSION

3.1 Identification of Drug

Colour: White crystalline powder

Odour: Odourless

3.2 Spectrophotometric scan of Allopurinol in pH 1.2 HCl buffer

After appropriately diluting the stock solution (A) of allopurinol in pH 1.2 HCl buffer medium, a sample containing 25 µg/ml of the medication was scanned between 200 and 400 nm. 260 nm was determined to be the λ_{max} .

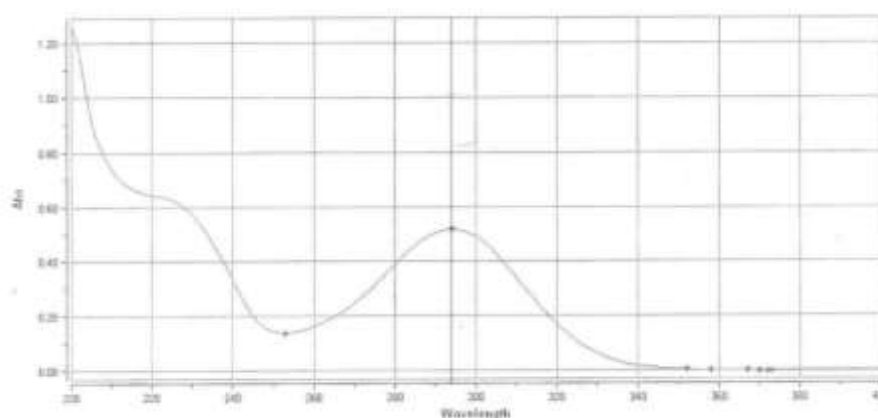


Fig. 3.1 U.V. scan of Allopurinol in pH 1.2 HCl buffer.

3.2.1 Validation of λ_{max}

To determine the λ_{max} value for Allopurinol, a suitably diluted solution of 25 µg/ml of stock A was scanned at a wavelength of 200–400 nm. A pH (1.2) HCl buffer

solution was utilized as a blank, and this was verified and validated by obtaining the drug's overlaid UV spectra at various concentrations ranging from 5–25 µg/ml.

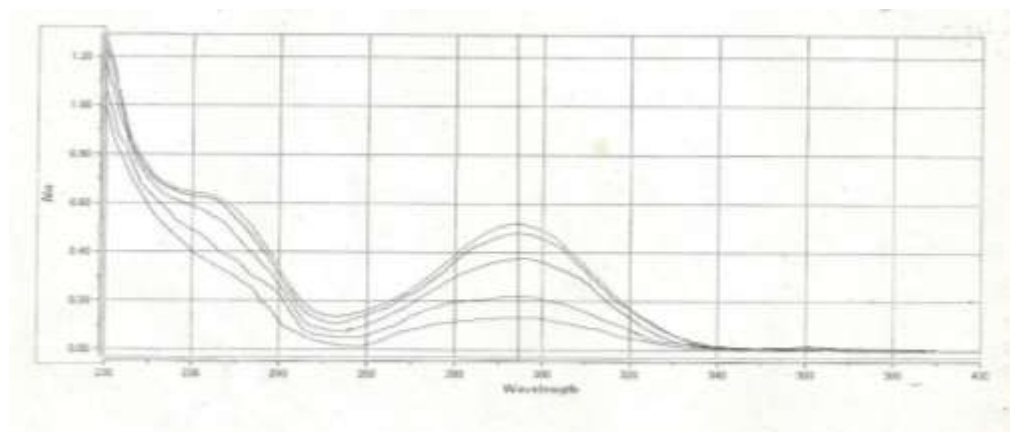


Fig. 3.2: Overlain spectra of Allopurinol.

For as long as anybody can remember, oral medicine has been the most appealing and practical method of medication administration in the modern pharmaceutical and clinical world. The bulk of formulations currently on the market still recommend oral therapy, despite extensive and unreliable research revealing a number of pharmacokinetic and pharmacodynamic challenges standing in the way of a successful public healthcare system. These challenges include the overwhelming need for dose optimization techniques to minimize toxic effects and dosing frequency while also improving the efficacy of the drug in a given formulation.

3.3. EXPERIMENTAL STUDIES

3.3.1. Making 0.1N (pH 1.2) ready HCl buffer: A volumetric flask was filled with 50 milliliters of a 0.2 ml

potassium chloride solution and 85 milliliters of a 0.2 milliliter hydrochloric acid solution. To alter the pH (1.2) as needed, 200 milliliters of distilled water and 0.2 milliliters of hydrochloric acid solution were added to the volume.

3.3.2. Creating an Allopurinol standard curve in a pH 1.2 HCl buffer: A UV spectrophotometer was used to load the different aliquots shown in Table 7 and determine the corresponding absorbances at λ_{max} 260 nm. Plotting concentration versus absorbance on a graph revealed a straight line, indicating that the medication complied with Beer-Lambert's Law at concentrations between 5 and 25 $\mu\text{g/ml}$.

Table 3.1: Concentrations v/s absorbance data of Allopurinol in pH 1.2HCl buffer.

S.No.	Concentration ($\mu\text{g/ml}$)	Absorbance
1.	5	0.113
2.	10	0.218
3.	15	0.328
4.	20	0.441
5.	25	0.553

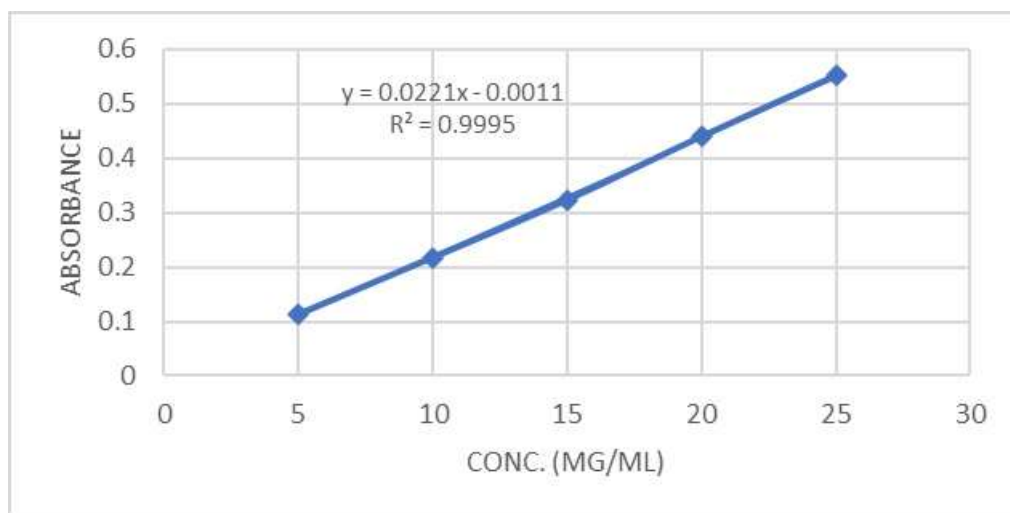


Fig. 3.3: Regression curve of Allopurinol in pH 1.2 HCl buffer.

3.3.3 Preparation of Standard curve of Allopurinol in simulated gastric fluid

By measuring the absorbance of different aliquots in Table 3.2 at 260 nm and drawing the graph between

absorbance v/s concentrations, a standard curve of allopurinol was created. The straight line that resulted indicated that the medication obeyed Beer-Lambert's Law at concentration ranges of 5–25 µg/ml.

Table 3.2: Concentrations v/s absorbance data of Allopurinol in SGF.

S.No.	Concentration (µg/ml)	Absorbance
1.	5	0.131
2.	10	0.265
3.	15	0.417
4.	20	0.548
5.	25	0.685

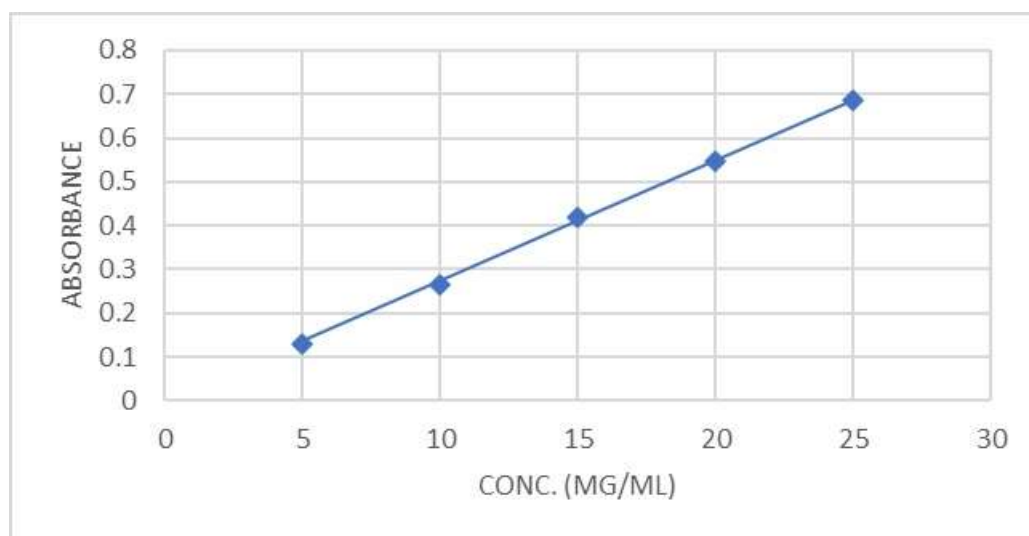


Fig. 3.4: Standard Curve of Allopurinol in SGF.

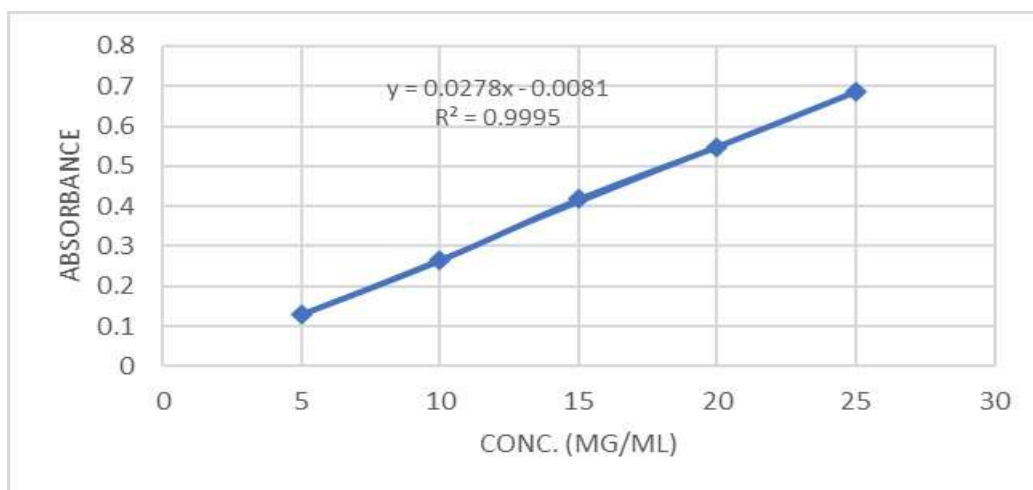


Fig. 3.5: Regression curve of Allopurinol in SGF.

3.3.4 Preparation of simulated gastric fluid (SGF)

Table 3.3: Composition of Simulated gastric fluid.

S.No.	Ingredients	Quantity (for 1000ml)
1.11	Pepsin	3.2 gm
2.12	Sodium chloride	2.0 gm
3.3	HCl	7.0 ml

A precise weight of 2.0 grams of sodium chloride and 3.2 grams of pure pepsin were thoroughly dissolved in 7.0 milliliters of hydrochloric acid. Distilled water was added to the mixture to bring the volume up to 1000 milliliters, and the pH was adjusted to 1.2.

3.3.5 Preparation of standard stock solution

(A) In 100 ml of pH 1.2 HCl buffer and stock solution (B) in simulated gastric fluid: 10 mg of allopurinol was dissolved in each of the two solutions separately to create stock solutions (100µg/ml). For additional research, stock solutions A and B were diluted.

3.3.6 Preparation of standard curve by using pH 1.2 HCl buffer

Serial dilutions (5–25µg/ml) of stock solution A were made, and the absorbance was measured at a preset λ_{max} . Plotting the absorbance v/s concentration (µg/ml) graph yielded the standard curve.

3.3.7 Preparation of standard curve by using simulated gastric fluid

Serial dilutions (5–25µg/ml) of the stock solution B were made, and the absorbances were measured at a predetermined λ_{max} . To create the standard curve, plot absorbance v/s concentration (µg/ml).

3.4 DRUG EXCIPIENTS COMPATABILITY STUDIES

3.4.1 Study using FTIR Technique

FTIR (Shimadzu IR Affinity I) spectrophotometer was used to conduct the drug polymer interaction investigation. The drug and potassium bromide (KBr) mixture was crushed into a fine powder using a mortar and pestle in a 1:9 ratio. The mixture was then compressed to create pellets at a pressure of 75 kg/cm², and it was scanned at a resolution of 2 cm⁻¹. The distinctive summits were noted.



Fig. 3.6 FTIR Spectra of Allopurinol (pure drug).

Table 3.4: Represents interpretation of IR spectra of Allopurinoll.

S.NO.	PEAK	WAVE NUMBER (cm ⁻¹)
1.	C-H	3100-2990
2.	C=O	1300-1100
3.	N-H stretching	3300-3030
4.	N-H bending	1620-1560

3.5 FORMULATION DESIGN

The formulation was classified into 09 batches (F₁ to F₉) prepared with different ratios of polymers as depicted in the table below.

Table 3.5: Formulation design for tablets (200 mg) prepared by Direct compression.

INGREDIENTS	FORMULATION CODE (quantity in mg)								
	F ₁	F ₂	F ₃	F ₄	F ₅	F ₆	F ₇	F ₈	F ₉
Allopurinol	20	20	20	20	20	20	20	20	20
HPMC K4M	80	--	--	40	--	40	20	--	20
HPMC K15M	--	80	--	40	40	--	60	20	--
HPMC K100M	--	--	80	--	40	40	--	60	60
MCC	45	45	45	45	45	45	45	45	45
NaHCO ₃	40	40	40	40	40	40	40	40	40
Citric Acid	10	10	10	10	10	10	10	10	10
Magnesium Stearate	02	02	02	02	02	02	02	02	02
Talc	03	03	03	03	03	03	03	03	03

3.5.1 Pre-compression evaluation of formulation

3.5.1.1 Bulk density

During a graduation, a certain amount of sample was meticulously injected. Three times, at two-second intervals, the cylinder was born onto a hard picket surface from a height of one inch. After that, the majority density was computed by dividing the sample load in grams by the final volume in cm³.

Bulk Density = mass of powder / volume of powder

3.5.1.2 Tapped density

By mechanically recording the measuring cylinder containing an amount of material, the Tapped Density was determined. The cylinder produced a mechanical sound and ran for 100 hard and quick faucets before the bed volume dropped to an absolute minimum. We calculated the broached density as follows.

Tapped density = mass of powder / volume of powder after tapping

3.5.1.3 Carr's index (% compressibility):

It demonstrated the advantage of being able to portray proportion and flow using a cloth. The following formula was used to compute it without a doubt.

Carr's Index = (Tapped density – Bulk density)/Tapped density X 100

Table 3.6: Relation between % compressibility and Flow ability.

S.No.	%compressibility	Flow ability
1.	5-15	Excellent
2.	12-16	Good
3.	18-21	Fair to Passable
4.	23-35	Poor
6.	33 -38	Very Poor
7.	>40	Very-Very Poor

3.5.1.4 Hausner's ratio: Hausner's ratio= Tapped density / Bulk density

3.5.1.5 Angle of Repose

The greatest angle that may exist between the surface of a powder pile and a horizontal plane is known as the angle of repose. The funnel technique was used to calculate each formulation's granules' angle of repose.

$$\tan\theta = h/r$$

Hence, $\theta = \tan^{-1} h/r$

Where,

θ - Angle of repose.

h- height and r- radius

Table 3.7: Relation between Angle of repose and Flow-ability.

S. No.	Angle of repose (degree)	Flow-ability
1.	<25	Excellent
2.	25-30	Good
3.	30-40	Passable
4.	>40	Very Poor

3.6 DRUG-POLYMER INTERACTION STUDIES

3.6.1 FTIR analysis

Utilizing FTIR analysis, the drug's compatibility with the polymers (HPMC K4M, HPMC K15M, and HPMC K100M) was investigated. Using the KBr pellet method, the IR absorption spectra of allopurinol were produced. The distinctive peaks of several functional groups contained in the medicine were shown in the table, and the peaks were compared with the reference.



Fig. 3.7: FTIR Spectra of Allopurinol (pure drug).

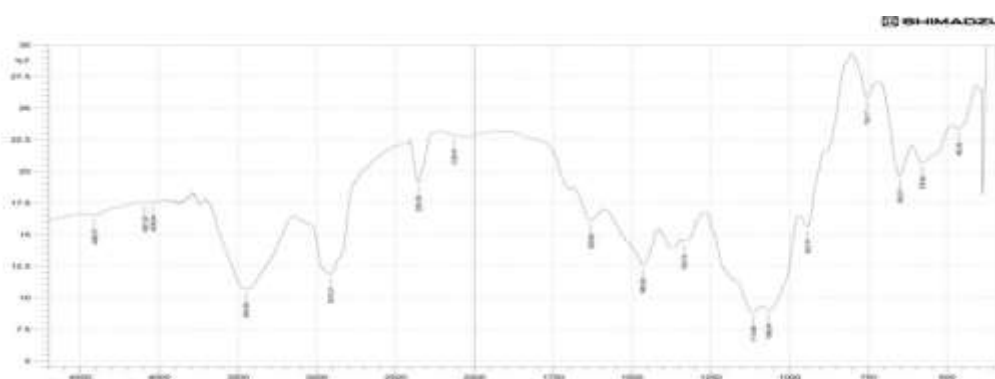


Fig. 3.8 FT-IR spectra of Allopurinol with HPMC K4M.

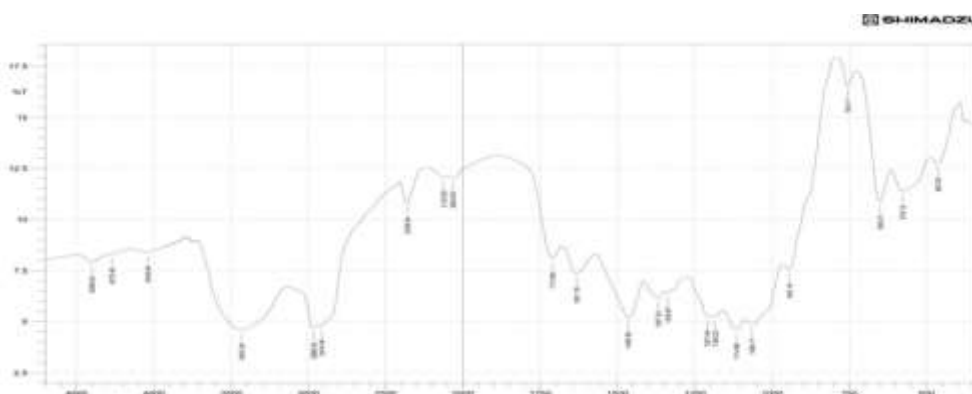


Fig. 3.9: FT-IR spectra of Allopurinol with HPMC K15M.

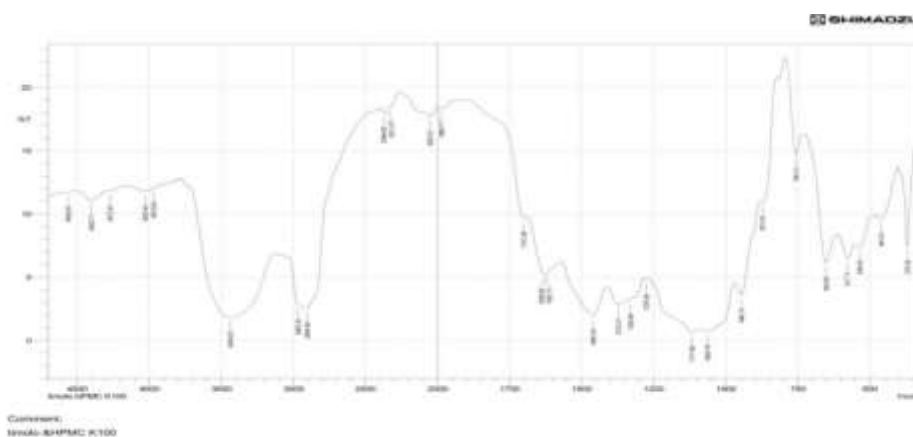


Fig. 3.10. FT-IR spectra of Allopurinol with HPMC K100M.

The ability of the pure medication to maintain its distinctive peak in various combinations demonstrated

that all of the polymers used in the formulations were compatible with it.

3.7 PRE-COMPRESSION EVALUATION

The pre-compression characteristics, namely angle of repose, bulk density, tapped density, Carr's index, and

Hausner's ratio, were assessed for the created formulation mix (F1-F9).

Table 3.8. Flow Properties of Formulation blends (F₁-F₉).

FORMULATION CODE	Bulk Density(gm/ml)	Tapped Density(gm/ml)	Carr's Index	Hausner's Ratio	Angle of repose (°)
F ₁	0.50	0.63	20.63	1.26	30.12
F ₂	0.51	0.61	16.40	1.19	25.26
F ₃	0.53	0.65	18.46	1.22	28.10
F ₄	0.50	0.59	15.25	1.18	23.54
F ₅	0.49	0.60	18.33	1.22	28.62
F ₆	0.52	0.62	16.12	1.19	25.00
F ₇	0.53	0.64	17.18	1.21	27.70
F ₈	0.50	0.61	18.03	1.22	28.95
F ₉	0.52	0.63	17.46	1.21	26.85

The following results were deduced: angle of repose (23.54-30.12), bulk density (0.49-0.53 gm/ml), tapped density (0.59 - 0.65 gm/ml), Carr's index (15.25 – 20.63%), and Hausner's ratio (1.18 - 1.26).

F1–F9 were as tabulated below. Nine batches of floating tablets prepared by the direct compression technique were evaluated for post compression parameters like weight variation, diameter, thickness, hardness, friability, floating lag time, total floating time, and swelling index.

3.8 EVALUATION OF DIRECT COMPRESSED TABLETS

After-compression assessments for batches F1–F9: In addition, the results of various analysis parameters for

Table 3.9: Evaluation data of batch F₁-F₉.

Evaluation Parameters	Formulation Code								
	F ₁	F ₂	F ₃	F ₄	F ₅	F ₆	F ₇	F ₈	F ₉
Wt variation (mg)	200±7.6	200±4.6	200±5.6	200±6.7	200±7.3	200±7.6	200±6.6	200±7.6	200±7.7
Hardness	5.50	5.75	5.80	6.30	6.20	6.50	5.75	5.80	6.30
Friability (%)	0.63	0.57	0.61	0.64	0.54	0.63	0.53	0.60	0.61
Thickness (mm)	2.75	2.83	2.80	2.85	2.95	2.78	2.88	2.81	2.88
Diameter (mm)	8.05	8.04	8.03	8.04	8.05	8.05	8.04	8.03	8.04
Floating lag time	36	40	68	45	50	48	52	57	55
Total floating	6	9	>12	10	12	11	>12	>12	>12
Swelling Index	55	57	65	68	70	67	60	75	70
Drug content (%)	98.22	97.43	98.71	97.43	98.49	98.23	97.52	98.60	98.45

The following results were inferred: diameter (8.03–8.05), floating lag time (36–68 sec), total floating duration (8–12 hrs), hardness (5.5–6.5 kg/cm²), friability (0.53–0.64%), weight variation (192.5–207.3 mg), drug content (97.43–98.71), thickness (2.75–2.95 mm), and swelling index (55–75%).

3.15 In-vitro comparative study of finalized formulation F₃, F₅ and F₈ in pH 1.2 HCl buffer

Three batches (F₃, F₅ and F₈) were selected which produced significant results for different evaluation parameters.

Table 3.18 Release kinetic data of formulation F₃, F₅ and F₈ in pH 1.2 HCl buffer.

Time (min)	% drug release		
	F ₃	F ₅	F ₈
30	8.18181	8.38181	10.2272
60	14.3223	12.2772	16.3693
90	22.5125	16.3756	22.5147
120	30.7068	20.4754	30.7090
180	34.8147	32.4312	42.9988
240	43.0191	43.0034	51.2045
300	53.2605	51.2090	61.4602
360	62.4437	59.4193	71.7215
420	73.5683	67.6340	79.9431
480	80.3478	75.8534	86.1238
540	87.0456	84.0772	90.2625
600	92.3227	92.3056	94.40341
660	98.7147	98.4931	96.5011
720	----	-----	98.601

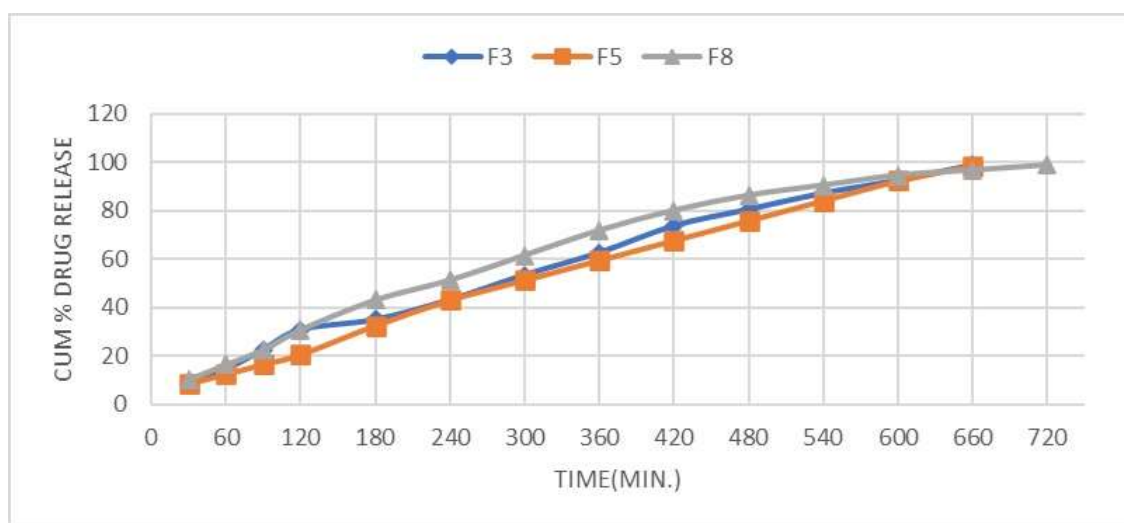


Fig. 3.11: Comparative Zero order release of formulation F₃, F₅ and F₈ in pH 1.2 HCl buffer.

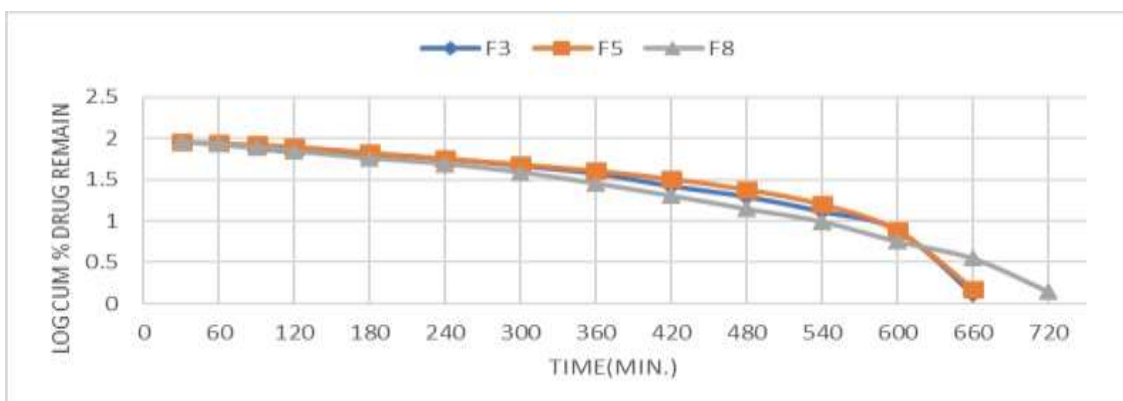


Fig. 3.12 Comparative first order release of formulation F₃, F₅ and F₈ in pH 1.2 HCl buffer.

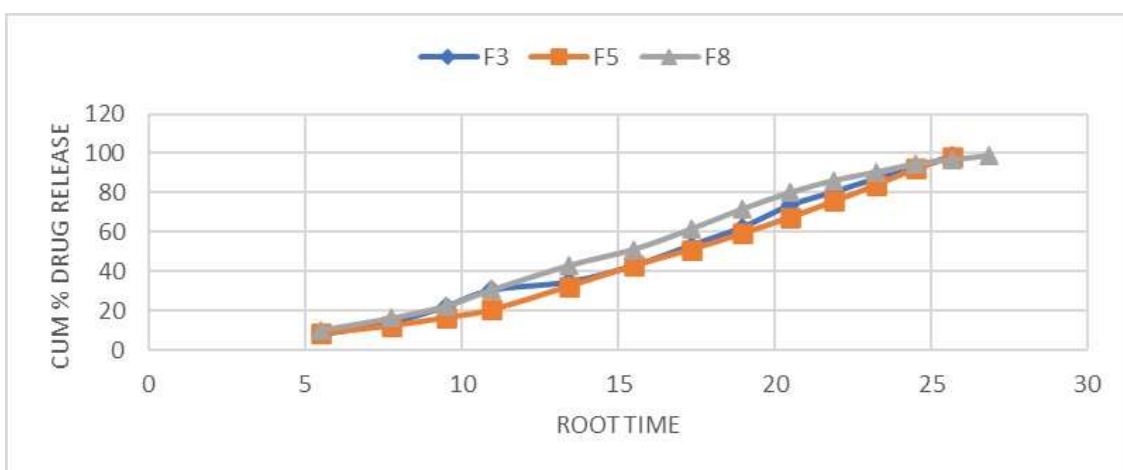


Fig. 3.13 Comparative Higuchi model release of formulation F₃, F₅ and F₈ in pH 1.2 HCl buffer.

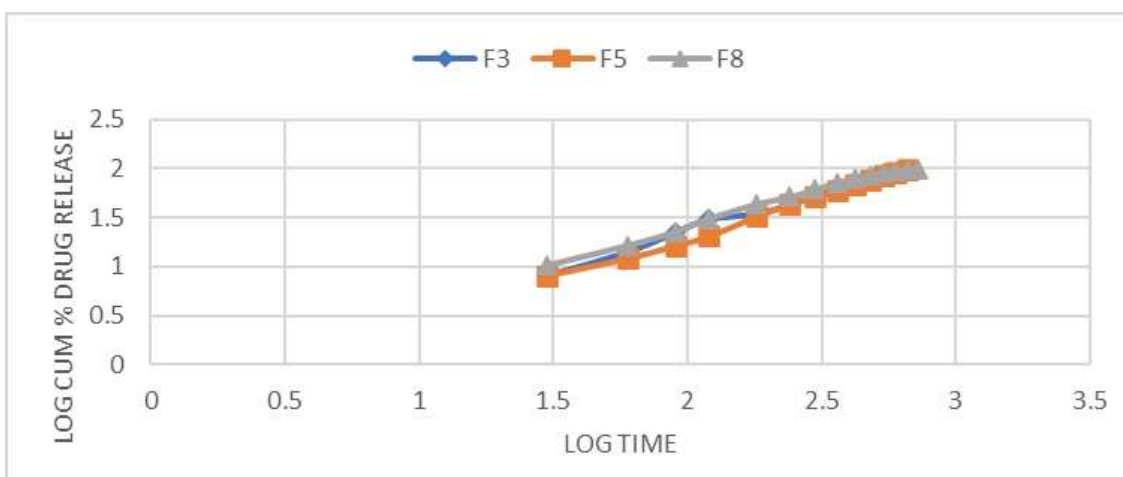


Fig. 3.14 Comparative Korsmeyer-Peppas model release of formulation F₃, F₅ and F₈ in pH 1.2 HCl buffer.

3.16. *In-vitro* comparative study of finalized formulation F₃, F₅ and F₈ in simulated gastric fluid.

Table 3.10. Release kinetic data of formulation F₃, F₅ and F₈ in SGF.

Time (min)	% drug release		
	F ₃	F ₅	F ₈
30	9.5317	7.6215	11.6432
60	14.3227	11.6428	18.4321
90	22.5125	16.3757	24.4254
120	30.7068	21.5243	33.4213
180	34.8147	32.4312	44.3256
240	43.0159	43.0034	54.3212
300	53.2670	51.2090	64.2375
360	62.4737	61.4321	72.8532
420	71.6432	67.6340	80.4522
480	80.3478	76.4256	86.12386
540	87.0456	84.0772	91.5342
600	92.3227	92.30568	95.2363
660	98.7147	98.4931	97.4247
720	-----	-----	99.5832

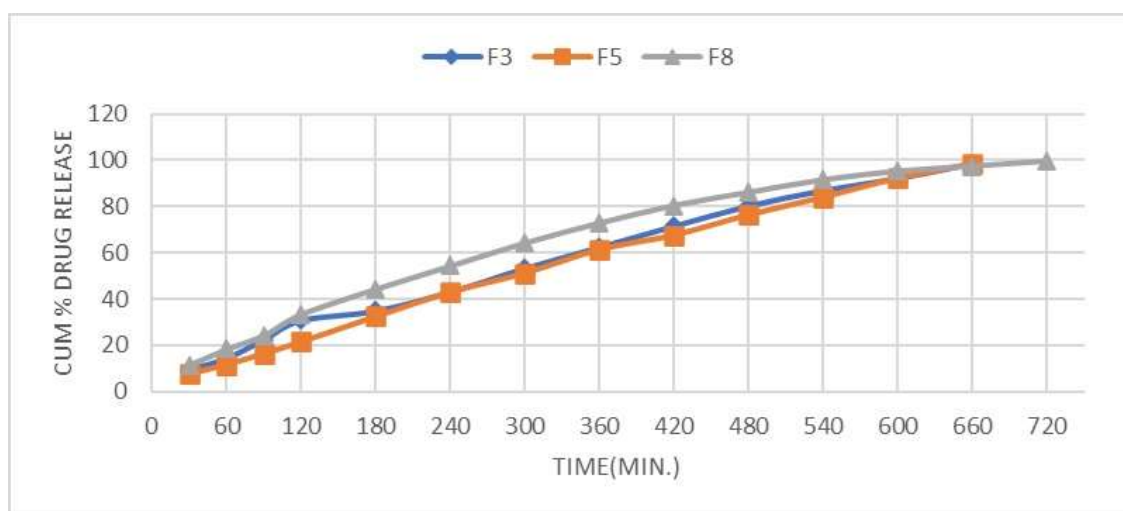


Fig. 3.15 Comparative Zero order release of formulation F₃, F₅ and F₈ in SGF.

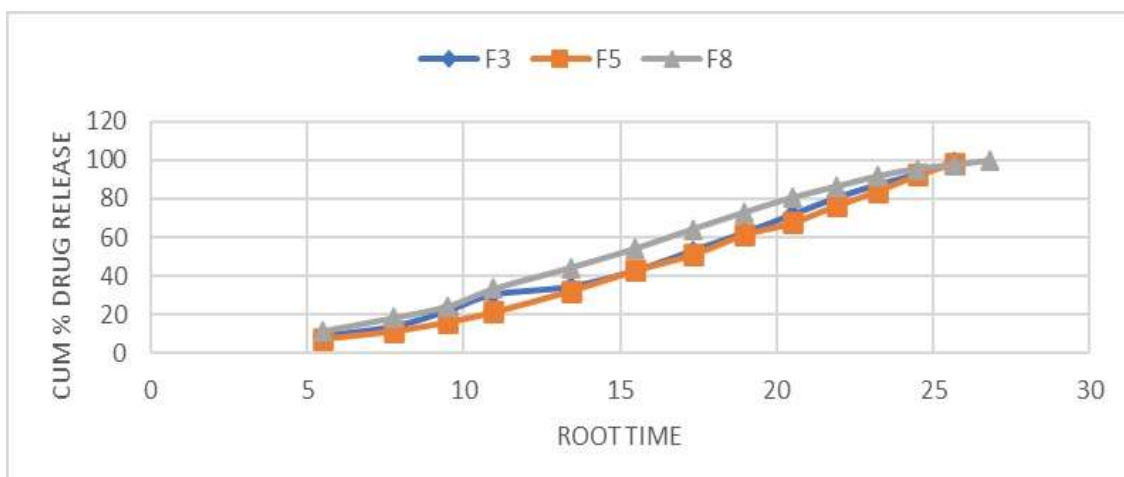


Fig. 3.16 Comparative Higuchi model release of formulation F₃, F₅ and F₈ in SGF.

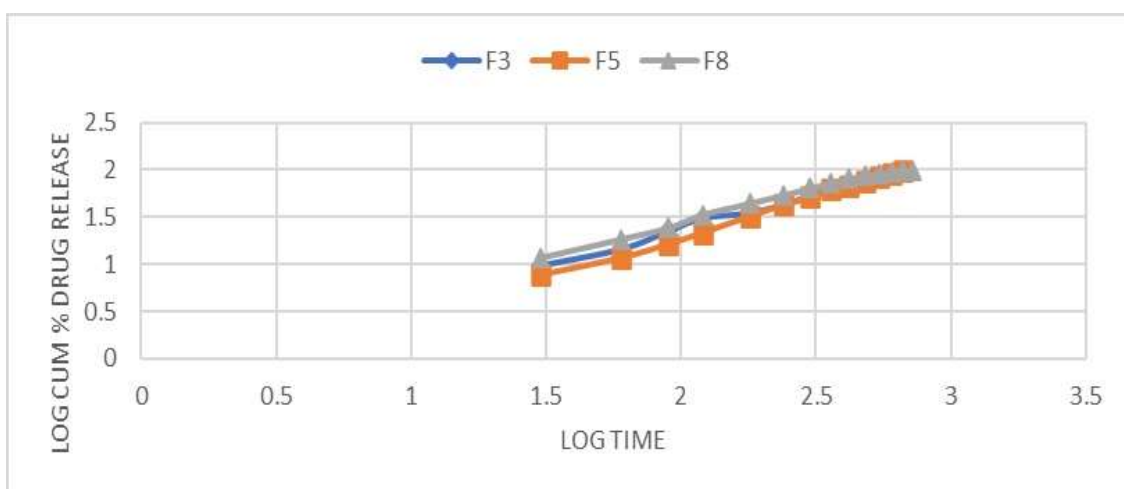


Fig. 3.17 Comparative Korsmeyer-Peppas model release of formulation F₃, F₅ and F₈ in SGF.

3.17 Stability data of Allopurinol floating tablet

Formulation F₈, which was chosen as the most optimized formulation based on the release characteristics, underwent expedited stability investigations. The optimal formulation (F₈) was subjected to a stability analysis in

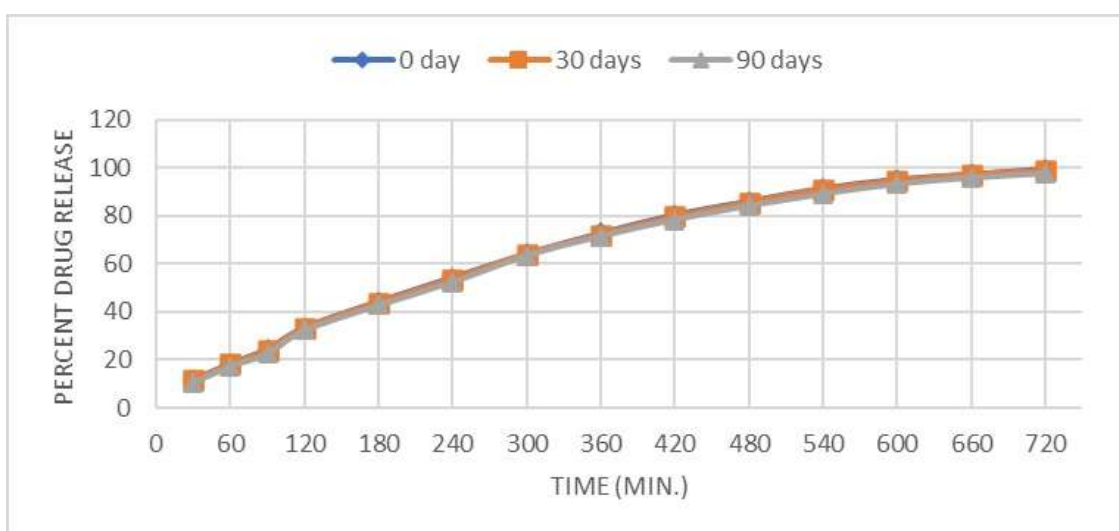
accordance with ICH criteria under accelerated settings (40 ± 2 °C, $75 \pm 5\%$ RH). The results indicated that neither a physical alteration nor a notable decrease in drug contents had occurred in the formulation (F₈).

Table 3.11 Stability study of formulation F₈ at $40^\circ \pm 2^\circ\text{C}$, $75\% \pm 5\%$ RH.

Stability study	Formulation F ₈		
	0 Days	30 Days	90 Days
Physical appearance	No change	No change	No change
Average weight (mg)	201	201	199
Hardness (Kg/cm ²)	5.5	52.4	5.3
Assay (% drug remaining)	99.54%	99.23%	98.82%
Swelling index (%)	75	73	72
% drug release	99.58	98.58	97.36

3.18 In-vitro dissolution profile of formulation F₈ on stability studies.Table 3.12 Kinetic release profile of F₈ on stability studies at different time intervals in SGF.

Time (min)	% drug release		
	0 Day	30 Days	90 Days
30	11.6432	11.6174	10.5732
60	18.4321	18.1572	17.3625
90	24.4256	23.9535	22.6432
120	33.4212	33.0421	32.2546
180	44.3256	43.8742	42.7453
240	54.3211	53.5435	52.34818
300	64.2378	64.0326	63.3617
360	72.8532	72.0562	71.4215
420	80.4525	79.5371	78.2683
480	86.1238	85.3218	84.0312
540	91.5347	90.8464	88.9631
600	95.2362	94.3628	93.0532
660	97.4247	97.0345	95.6326
720	99.5832	98.5835	97.3643

Fig. 3.18 Drug release profile of formulation F₈.

In traditional dose forms such tablets, capsules, suspensions, etc., a significant portion of the active medication is either not absorbed at all or absorbed very slowly from the gastrointestinal system, requiring more frequent administration and having a relatively lower bioavailability of the drug. The drug's compatibility with carefully chosen polymers was confirmed using FTIR Spectrophotometric experiments using FTIR Affinity-1. Following a comparison of the distinctive drug peaks achieved with HPMC K4M, HPMC K15M, and HPMC K100M, formulations from each of the nine batches were generated and documented.

Thin layer chromatography (TLC) was used, and combinations and R_f values were computed in comparison to the pure drug and polymer. The densitometry TLC tests and FTIR spectrum analysis verified that the medication was compatible with the polymer(s) in use. The compatibility of the medication with the polymer was ascertained using FTIR and densitometry TLC experiments. The distinctive peaks of the pure drug were compared to those produced with tablets from other batches, which stayed almost the same. The R_f values of the pure drug's and different polymers' thin layer chromatographs were almost equal. Using the drug and HPMC K4M, HPMC K15M, and HPMC K100M ratios of 1:4, 1:2:2, and 1:1:3, respectively, nine batches (F1–F9) of floating tablets were manufactured by the direct compression technique.

Every batch's floating tablets underwent a number of assessment tests, including ones measuring the tablets' hardness, friability, weight fluctuation, thickness, diameter, drug content, swelling index, floating lag time, and overall floating time. The powder mix was used to estimate other physicochemical (pre-compression) characteristics, such as bulk density (0.49–0.53), tapped density (0.59–0.65), Hausner's ratio (1.18 - 1.26), angle of repose (23.54–30.12), and Carr's index (15.25–20.63 %).

The following post-compression evaluation parameters were applied to these formulations: diameter (8.03–8.05), floating lag time (36–68sec), total floating time (8–12 hrs), hardness (5.5–6.5 kg/cm²), friability (0.53–0.64%), weight variation (192.5–207.3 mg), drug content uniformity (97.43–98.71), thickness (2.75–2.95 mm), and swelling index (55–75%). The medication with the best formulation (F8) demonstrated a sustained release over a 12-hour period of 99.58% in simulated stomach juice, although its in-vitro release in pH 1.2 HCl buffer was reported to be 98.60%. Stability tests on the best formulation, F8, revealed that the formulations were stable and held onto their medicinal qualities for three months at 40°C±20°C and 75%±5% relative humidity.

According to the ICH recommendations, a stability study was conducted using the improved formulation (F8) under officially defined circumstances. The results indicated that the formulation was stable and met the

dosage compliance requirement. The aforementioned statistics all met the formulation's distinctive characteristics for gastroretentive floating tablets in a satisfactory manner. The current worker tends to provide future researchers motivation to create such cutting-edge medication delivery systems that can replace traditional dosage forms with notable pharmacokinetic and pharmacodynamic qualities.

CONCLUSION

For as long as humans have been in the pharmaceutical and therapeutic fields today, oral medicine has been the most appealing and practical way to apply drugs. Even though extensive research has shown several pharmacokinetic and pharmacodynamic challenges standing in the way of a successful public healthcare system, the majority of formulations currently in use still suggest oral therapy. These challenges include the overwhelming need for dose-optimization techniques to minimize toxic effects and dosing frequency while also improving the efficacy of the drug in a given formulation.

The gastrointestinal system does not absorb a significant portion of the active medication included in typical dose forms such tablets, capsules, liquids, etc.; as a result, the medicine must be administered more often and has a reduced bioavailability. "Formulation and evaluation of gastroretentive floating tablets of allopurinol" was the idea of the current employee. This would lengthen the time that food is in the stomach, increase absorption, and result in sustained pharmacological activity. Eventually, this would also boost the drug's bioavailability.

A typical xanthine oxidative inhibitor used to treat gout is allopurinol. The densitometry TLC tests and FTIR spectrum analysis verified that the medication was compatible with the polymer(s) in use. Using the drug and HPMC K4M, HPMC K15M, and HPMC K100M ratios of 1:4, 1:2:2, and 1:1:3, respectively, nine batches (F1–F9) of floating tablets were made using the direct compression technique. Every batch's floating tablets underwent a number of assessment tests, including ones measuring hardness, Stability tests on the best formulation, F8, revealed that the formulations were stable and maintained their medicinal qualities for three months at 40°C±20°C and 75%±5% relative humidity. Allopurinol floating tablets were made using three different polymers: HPMC K4M, HPMC K15M, and HPMC K100M.

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