

SERUM CONCENTRATIONS OF SOME ELECTROLYTES (SODIUM, POTASSIUM AND BICARBONATE) IN WISTAR RATS FED WITH CARROT (*DAUCUS CAROTA*) EXTRACT**Onyebuchi Obia*¹, Regina Kalio² and Elile Peace Okpara¹**¹Department of Human Physiology, Faculty of Basic Medical Sciences, College of Medical Sciences, Rivers State University, Nkpolu-Oroworukwo, Port Harcourt, Nigeria.²Department of Human Physiology, Faculty of Basic Medical Sciences, College of Health Sciences, University of Port Harcourt, Nigeria.***Corresponding Author: Onyebuchi Obia**

Department of Human Physiology, Faculty of Basic Medical Sciences, College of Medical Sciences, Rivers State University, Nkpolu-Oroworukwo, Port Harcourt, Nigeria.

Article Received on 03/02/2025

Article Revised on 24/02/2025

Article Published on 16/03/2025

ABSTRACT

Background to the study: Plasma electrolytes regulate many physiological processes necessary for cellular survival. The present study investigated the effects of carrot (*Daucus carota*) extract on serum concentrations of some electrolytes (sodium, potassium and bicarbonates) in wistar rats. **Methodology:** The study involved 28 male wistar rats separated into 4 groups of 7 rats each. Group 1 served as control and was given distilled water, whilst groups 2, 3 and 4 served as test groups and were given aqueous extract of *Daucus carota* at daily doses of 200mg/kg, 400mg/kg and 600mg/kg respectively. The experiment lasted for 28 days and thereafter the animals were sacrificed under anesthesia and blood samples collected for determination of serum potassium, sodium and bicarbonate concentrations using standard laboratory techniques. **Results and Discussion:** The results of our study showed that daily oral administration of 200mg/kg and 400mg/kg carrot extract caused a dose-dependent increase in the serum concentrations of potassium, sodium and bicarbonates, although not significantly. However, administration of 600mg/kg of carrot extract caused a significant increase in sodium concentration. Therefore, increasing the daily dosage of carrot might distort the physiological homeostatic processes with resultant significant increase in plasma sodium concentration. Excessive consumption of dietary sodium is discouraged because of risk of developing cardiovascular and other disorders. **Conclusion:** Conclusively, moderate consumption of carrot did not have any significant effect on the serum concentrations of potassium, sodium and bicarbonates and by implication did not alter the physiologic homeostatic processes. Higher doses of carrot extract significantly increased serum sodium necessitating cautious consumption of large amounts of carrot.

KEYWORDS: Serum electrolytes, Sodium, Potassium, Bicarbonate, *Daucus carota*, Wistar rats.**INTRODUCTION**

Electrolytes in the blood essentially regulate many body functions; water balance, nerve and muscle function, acid-base balance, blood pressure control, maintenance of heart rate and rhythm, bone and teeth function (Jomova et al., 2022). These roles are necessary for the optimal functioning of vital organs of the body. In physiological condition, the concentrations of the different electrolytes in the blood must be maintained within their respective normal ranges. Therefore, adverse deviations from the physiologic ranges could be detrimental to cellular survival. Many of the electrolytes are derived from our diets especially fruits and other plant products (Fernandez & Marette, 2017; Qian, 2018; Kalaycıoğlu & Erım, 2019; Alwis et al., 2020; Sultana et al., 2021; Muhammad et al., 2024). In deficiency disorders or in conditions requiring higher demand for

specific electrolytes, preformed supplements are often administered. Carrots are mainly orange coloured root vegetables (Carlos & Dias, 2014; Bahrami et al., 2018) which are consumed as food and also used in herbal preparations. Some of the health benefits of carrots have been documented in previous studies (Tanaka et al., 2012; Fiedor & Burda, 2014; Bystricka et al., 2015; Obia et al., 2025). Although some researchers have reported significant amounts of electrolytes in various fruits and vegetables, our study investigated the effect of carrot consumption on the serum concentrations of potassium, sodium and bicarbonates.

MATERIALS AND METHODS

The experiment was carried out at animal house of the department of Human Physiology, faculty of Basic Medical Sciences, University of Port Harcourt in the

year, 2019. Ethical approval was obtained from the university of Port Harcourt Research Ethics Committee with approval number; UPH/CEREMAD/REC/MM67/012. A total of twenty eight male wistar rats weighing 120 to 150g were purchased and acclimatized for a period of two weeks. These animals were separated into 4 groups of 7 rats each, housed in plastic cages and allowed to feed and drink *ad libitum* with Top feed Finisher mash and clean water. Their immediate environment (beddings) was changed daily, the temperature of the environment kept at normal conditions while the external environment was cleaned and disinfected regularly.

Preparation and administration of carrot extract

Mature carrot tubers were bought from Oil Mill market in Obio Akpor Local Government Area of Rivers State, Nigeria. The plant was identified at the department of Plant Science and Biotechnology, University of Port Harcourt; *Daucus carota* L, in the family; Apiceace with assigned herbarium number; UPH/C/132. The tubers were washed with water to remove soil particles. About 2kg of the fresh carrot was cut into tiny pieces and air dried for seven days. The dried carrots were blended using a blender and carefully poured into a maceration jar containing four liters of water. The mixture was

allowed to macerate for 24hours after which a Whatman filter (20-25 μ m, pore size) was used to get a clear filtrate. The filtrate was now poured into an evaporating dish and dried in a water bath at 45°C to obtain a semi-solid aqueous extract of *Daucus carota*. The dosages administered in the study were based on the lethal dose (LD50) of 5000mg/kg which was previously determined (Ayeni et al., 2019). Following acclimatization, the wistar rats were weighed and separated into four groups of seven rats each. Group 1 served as control and was given distilled water, whilst groups 2, 3 and 4 served as test groups and were given aqueous extract of *Daucus carota* at daily doses of 200mg/kg, 400mg/kg and 600mg/kg respectively. The experiment lasted for 28 days. Thereafter the animals were sacrificed under anesthesia and blood samples collected for determination of serum potassium, sodium and bicarbonate concentrations using standard laboratory techniques.

Statistical analysis was done using SPSS software version 22.0. Results were presented in tables. Continuous variables were expressed as mean \pm Standard error of mean (SEM). Statistical difference was determined using analysis of variance (ANOVA) and at $p < 0.05$.

RESULTS AND DISCUSSION

Table 1: Effect of Carrot extract on serum potassium, sodium and bicarbonate concentrations of wistar rats.

Group	Potassium (mmol/l)	Sodium (mmol/l)	Bicarbonate (mmol/l)
Control	5.73 \pm 0.73	128.50 \pm 2.23	25.67 \pm 0.95
200mg/kg	6.45 \pm 0.32	129.17 \pm 3.25	27.17 \pm 0.40
400mg/kg	7.43 \pm 0.45	142.83 \pm 7.93	29.00 \pm 0.86
600mg/kg	8.70 \pm 0.40	163.00 \pm 5.72*	30.67 \pm 0.67

* Significantly different compared to control.

The results of our study showed that oral administration of carrot extract caused a dose-dependent increase in the serum concentrations of potassium and bicarbonates, although not significantly. Also the 200mg/kg and 400mg/kg of carrot extract caused a non-significant increase in sodium concentration. However, administration of 600mg/kg of carrot extract caused a significant increase in serum sodium concentration.

Despite the numerous benefits of electrolytes in the blood, the concentration and type of electrolyte consumed is of utmost importance. Most diets are enriched with sodium either to taste or as a preservative (Sambu et al., 2022; Warner, 2024; Yu & Rhee 2024) to the extent that our daily dietary sodium requirement is often exceeded. Deficiency of plasma sodium resulting from inadequate dietary intake rarely occurs since it is richly available in our daily meals. However, hyponatremia may occur in malnutrition (Baez et al., 2024) or during conditions resulting to excessive loss of sodium (Puckett, 2023). Moderate consumption of carrot as shown in the present study will add just enough sodium in the plasma that is within the physiologic limit. Increasing the daily dosage of carrot up to 600mg/kg/day

distorted the physiological homeostatic processes with resultant significant increase in plasma sodium concentration. Excessive consumption of dietary sodium is discouraged because of risk of developing cardiovascular and other disorders. Therefore, carrot should be consumed moderately to obtain its benefits. Many studies support the moderate consumption of natural products for optimal benefit (Obia et al., 2018; Obia & Eifuobhokhan, 2024; Obia & Emmanuel, 2025).

Although, some refined foods are fortified with potassium, dietary supplementation of this electrolyte is not a common practice in most food cultures. For this reason, potassium-rich foods are to be intentionally consumed to meet the daily potassium requirement. Potassium deficiency has been implicated in many diseases including cardiovascular diseases, osteoporosis and kidney stone (Palmer et al., 2016; Averin et al., 2020). Studies have shown that sodium bicarbonate composition of certain fruits and vegetables are capable of reducing dietary acid by half (Goraya et al., 2012) which is a protective effect against kidney injury. The bicarbonate content of carrot can be beneficial in patients with excessive gastric acidity as it can have possible

antacid potential.

Conclusively, moderate consumption of carrot did not have any significant effect on the serum concentrations of potassium, sodium and bicarbonates and by implication did not alter the physiologic homeostatic processes. Higher doses of carrot extract significantly increased serum sodium necessitating cautious consumption of large amounts of carrot.

REFERENCES

1. Alwis, U. S., Haddad, R., Monaghan, T. F., Abrams, P., Dmochowski, R., Bower, W., Wein, A. J., Roggeman, S., Weiss, J. P., Mourad, S., Delanghe, J., & Everaert, K. Impact of food and drinks on urine production: A systematic review. *International journal of clinical practice*, 2020; 74(9): e13539. <https://doi.org/10.1111/ijcp.13539>
2. Averin, E. E., Nikitin, A. E., Pozdnyak, A. O., Fedorova, E. L., Zhuk, V. S., Davydov, S. I., Fridman, I. L., Kompaniets, O. G., Kirpichnikova, N. V., Dudarenkova, M. R., Ginzburg, M. L., El Sharif, M. A., Martemyanova, E. G., & Sozykin, A. V. *Kardiologiia*, 2020; 60(2): 155–164. <https://doi.org/10.18087/cardio.2020.2.n972>
3. Ayeni, A., Abubakar, A., Aliyu, N., Uhomoibhi, L., & Garba, I. 'Acute and sub-acute toxicity of the crude extracts of the aerial parts of *Daucus carota* L. in laboratory rats'. *Journal of Medicinal Plants for Economic Development*, 2019; 3(1): a69.
4. Baez, G., Chirio, M., Pisula, P., Seminario, E., Carasa, N., Philippi, R., Aroca-Martinez, G., & Musso, C. G. Hyponatremia and malnutrition: a comprehensive review. *Irish journal of medical science*, 2024; 193(2): 1043–1046. <https://doi.org/10.1007/s11845-023-03490-8>
5. Bahrami, R., Ghobadi, A., Behnoud, N., & Akhtari, E. Medicinal properties of *Daucus carota* in traditional Persian medicine and modern phytotherapy. *Journal of Biochemical Technology. Special issue*, 2018; 2: 107–114.
6. Bystrická, J., Kavalcová, P., Musilová, J., Vollmannová, A., Tóth, T., Lenková, M. Carrot (*Daucus carota* L. ssp. *sativus* (Hoffm.) Arcang.) as source of antioxidants. *Acta Agric. Slov.* 2015; 105: 303–311.
7. Carlos, J., Dias, S. Nutritional and Health Benefits of Carrots and Their Seed Extracts. *Food and Nutrition Sciences*, 2014; 5: 2147–2156.
8. Fernandez, M. A., & Murette, A. Potential Health Benefits of Combining Yogurt and Fruits Based on Their Probiotic and Prebiotic Properties. *Advances in nutrition* (Bethesda, Md.), 2017; 8(1): 155S–164S. <https://doi.org/10.3945/an.115.011114>
9. Fiedor, J., & Burda, K. Potential role of carotenoids as antioxidants in human health and disease. *Nutrients*, 2014; 6(2): 466–488. <https://doi.org/10.3390/nu6020466>
10. Goraya, N., Simoni, J., Jo, C., & Wesson, D. E. Dietary acid reduction with fruits and vegetables or bicarbonate attenuates kidney injury in patients with a moderately reduced glomerular filtration rate due to hypertensive nephropathy. *Kidney international*, 2012; 81(1): 86–93. <https://doi.org/10.1038/ki.2011.313>
11. Jomova, K., Makova, M., Alomar, S. Y., Alwasel, S. H., Nepovimova, E., Kuca, K., Rhodes, C. J., & Valko, M. Essential metals in health and disease. *Chemico-biological interactions*, 2022; 367: 110173. <https://doi.org/10.1016/j.cbi.2022.110173>
12. Kalaycıoğlu, Z., & Erım, F. B. Nitrate and Nitrites in Foods: Worldwide Regional Distribution in View of Their Risks and Benefits. *Journal of agricultural and food chemistry*, 2019; 67(26): 7205–7222. <https://doi.org/10.1021/acs.jafc.9b01194>
13. Muhammad, A. I., Ibrahim, R. G., Adamu, S. B., Ubali, S. ., Mustapha, K., & Ibrahim, F. G. Effect of Dietary Turmeric-Ginger Combination on Serum Electrolytes Of Broiler Chicken. *Nigerian Journal of Animal Production*, 2024; 505–508.
14. Obia, O., Odum, J. E., & Chuemere, A. N. Nephroprotective and antihyperlipidemic activity of honey in alloxan induced diabetic wistar rats. *International Journal of Biochemistry Research and Review*, 2018; 22(1): 1–7.
15. Obia, O., & Eifuobhokhan, J. Effect of *Justicia carnea* leaf extract on plasma and fecal lipid profile in high-fat diet fed wistar rats. *International Journal of Health and Pharmaceutical Research*, 2024; 9(4): 64–70.
16. Obia, O., & Emmanuel, F. D. Effect of Oral Administration of Common Pepper Types on the Liver Enzymes of Wistar Rats Fed with High-fat Diet. *East African Scholars Journal of Medical Sciences*, 2025; 8(3): 92–95.
17. Obia, O., Kalio, R. O., Tee, P. G. P., & Onyeso, G. Plasma Lipid Lowering Potential of Carrot (*Daucus carota*) Extract in Male Wistar Rats. *Asian Journal of Research in Medical and Pharmaceutical Sciences*, 2025; 14(1): 18–23.
18. Palmer, B. F., & Clegg, D. J. Achieving the Benefits of a High-Potassium, Paleolithic Diet, Without the Toxicity. *Mayo Clinic proceedings*, 2016; 91(4): 496–508.
19. Puckett L. Renal and electrolyte complications in eating disorders: a comprehensive review. *Journal of eating disorders*, 2023; 11(1): 26. <https://doi.org/10.1186/s40337-023-00751-w>
20. Qian Q. Dietary Influence on Body Fluid Acid-Base and Volume Balance: The Deleterious "Norm" Furthers and Cloaks Subclinical Pathophysiology. *Nutrients*, 2018; 10(6): 778. <https://doi.org/10.3390/nu10060778>
21. Sambu, S., Hemaram, U., Murugan, R., & Alsofi, A. A. Toxicological and Teratogenic Effect of Various Food Additives: An Updated Review. *BioMed research international*, 2022; 2022: 6829409. <https://doi.org/10.1155/2022/6829409> (Retraction published Biomed Res Int., Jan. 9, 2024; 2024: 9792751. doi: 10.1155/2024/9792751.)

22. Sultana, R., Alashi, A. M., Islam, K., Saifullah, M., Haque, C. E., & Aluko, R. E. Chemical composition and in vitro antioxidant properties of water-soluble extracts obtained from Bangladesh vegetables. *Journal of food biochemistry*, 2021; 45(3): e13357.
23. Tanaka, T., Shnimizu, M., & Moriwaki, H. Cancer chemoprevention by carotenoids. *Molecules (Basel, Switzerland)*, 2012; 17(3): 3202–3242. <https://doi.org/10.3390/molecules17033202>
24. Warner, J. O. Artificial food additives: hazardous to long-term health?. *Archives of disease in childhood*, 2024; 109(11): 882–885. <https://doi.org/10.1136/archdischild-2023-326565>
25. Yu, H., & Rhee, M. S. Potential of phytic acid in synergy with sodium chloride as a natural-borne preservative to inactivate *Escherichia coli* O157:H7 and inhibit natural microflora in fresh noodles at room temperature. *Current research in food science*, 2024; 9: 100868. <https://doi.org/10.1016/j.crfs.2024.100868>
26. Do, C., Vasquez, P. C., & Soleimani, M. Metabolic Alkalosis Pathogenesis, Diagnosis, and Treatment: Core Curriculum 2022. *American journal of kidney diseases : the official journal of the National Kidney Foundation*, 2022; 80(4): 536–551. <https://doi.org/10.1053/j.ajkd.2021.12.016>
27. Que, F., Hou, X. L., Wang, G. L., Xu, Z. S., Tan, G. F., Li, T., Wang, Y. H., Khadr, A., & Xiong, A. S. Advances in research on the carrot, an important root vegetable in the Apiaceae family. *Horticulture research*, 2019; 6: 69. <https://doi.org/10.1038/s41438-019-0150-6>
28. Purkiewicz, A., Ciborska, J., Tanska, M., Narwojsz, A., Starowicz, M., Przybyłowicz, K. E., & Sawicki, T. The Impact of the Method Extraction and Different Carrot Variety on the Carotenoid Profile, Total Phenolic Content and Antioxidant Properties of Juices. *Plants*. 2020; 9: 1759.
29. Augpole, I., Rackejeva, T., Kruma, Z., Dimins, F. Shredded carrots quality providing by treatment with Hydrogen peroxide. 9th Baltic Conference on “Food for Consumer Well-Being” *FOODBALT*, 2014; 2014: 150-154.
30. Alasalvar, C., Grigor, J. M., Zhang, D., Quantick, P. C., & Shahidi, F. Comparison of volatiles, phenolics, sugars, antioxidant vitamins, and sensory quality of different colored carrot varieties. *Journal of agricultural and food chemistry*, 2001; 49(3): 1410–1416.
31. Molldrem, K. L., Li, J., Simon, P. W., & Tanumihardjo, S. A. Lutein and beta-carotene from lutein-containing yellow carrots are bioavailable in humans. *The American journal of clinical nutrition*, 2004; 80(1): 131–136. <https://doi.org/10.1093/ajcn/80.1.131>
32. Stange, C., & Rodriguez-Concepcion, M. Carotenoids in Carrot. In *Pigments in Fruits and Vegetables*; Springer Science and Business Media LLC: Berlin/Heidelberg, Germany, 2015; 217–228.
33. Hager, T.J., Howard, L.R. Processing Effects of Carrot Phytonutrients. *Horticultural Science*, 2006; 41: 74-79.
34. Sharma, K. D., Karki, S., Thakur, N. S., & Attri, S. Chemical composition, functional properties and processing of carrot-a review. *Journal of food science and technology*, 2012; 49(1): 22–32. <https://doi.org/10.1007/s13197-011-0310-7>
35. Leja, M., Kamińska, I., Kramer, M., Maksylewicz-Kaul, A., Kammerer, D., Carle, R., & Baranski, R. The content of phenolic compounds and radical scavenging activity varies with carrot origin and root color. *Plant foods for human nutrition (Dordrecht, Netherlands)*, 2013; 68(2): 163–170.
36. Ismail, J., Shebaby, W. N., Daher, J., Boulos, J. C., Taleb, R., Daher, C. F., & Mroueh, M. The Wild Carrot (*Daucus carota*): A Phytochemical and Pharmacological Review. *Plants (Basel, Switzerland)*, 2023; 13(1): 93. <https://doi.org/10.3390/plants13010093>
37. Myojin, C., Enami, N., Nagata, A., Yamaguchi, T., Takamura, H., & Matoba, T. Changes in the radical-scavenging activity of bitter melon (*Momordica charantia* L.) during freezing and frozen storage with or without blanching. *Journal of food science*, 2008; 73(7): C546–C550. <https://doi.org/10.1111/j.1750-3841.2008.00886.x>
38. Phan, M. A. T., Paterson, J., Bucknall, M., & Arcot, J. Interactions between phytochemicals from fruits and vegetables: Effects on bioactivities and bioavailability. *Critical reviews in food science and nutrition*, 2018; 58(8): 1310–1329.
39. Zhu, F., Du, B., & Xu, B. Anti-inflammatory effects of phytochemicals from fruits, vegetables, and food legumes: A review. *Critical reviews in food science and nutrition*, 2018; 58(8): 1260–1270. <https://doi.org/10.1080/10408398.2016.1251390>
40. Lecerf, J. M., & de Lorgeril, M. Dietary cholesterol: from physiology to cardiovascular risk. *The British journal of nutrition*, 2011; 106(1): 6–14.
41. Xu, Z., McClure, S. T., & Appel, L. J. Dietary Cholesterol Intake and Sources among U.S Adults: Results from National Health and Nutrition Examination Surveys (NHANES), 2001–2014. *Nutrients*, 2018; 10(6): 771.
42. Wilde, D. W., Massey, K. D., Walker, G. K., Vollmer, A., & Grekin, R. J. High-fat diet elevates blood pressure and cerebrovascular muscle Ca(2+) current. *Hypertension (Dallas, Tex.: 1979)*, 2000; 35(3): 832–837.
43. Ingelsson, E., Schaefer, E. J., Contois, J. H., McNamara, J. R., Sullivan, L., Keyes, M. J., Pencina, M. J., Schoonmaker, C., Wilson, P. W., D'Agostino, R. B., & Vasan, R. S. Clinical utility of different lipid measures for prediction of coronary heart disease in men and women. *JAMA*, 2007; 298(7): 776–785. <https://doi.org/10.1001/jama.298.7.776>
44. Duan, Y., Zeng, L., Zheng, C., Song, B., Li, F., Kong, X., & Xu, K. Inflammatory Links Between High Fat Diets and Diseases. *Frontiers in*

- immunology*, 2018; 9: 2649.
45. Carson, J. A. S., Lichtenstein, A. H., Anderson, C. A. M., Appel, L. J., Kris-Etherton, P. M., Meyer, K. A., Petersen, K., Polonsky, T., Van Horn, L., & American Heart Association Nutrition Committee of the Council on Lifestyle and Cardiometabolic Health; Council on Arteriosclerosis, Thrombosis and Vascular Biology; Council on Cardiovascular and Stroke Nursing; Council on Clinical Cardiology; Council on Peripheral Vascular Disease; and Stroke Council. Dietary Cholesterol and Cardiovascular Risk: A Science Advisory From the American Heart Association. *Circulation*, 2020; 141(3): e39–e53.
 46. Wali, J. A., Jarzebska, N., Raubenheimer, D., Simpson, S. J., Rodionov, R. N., & O'Sullivan, J. F. Cardio-Metabolic Effects of High-Fat Diets and Their Underlying Mechanisms-A Narrative Review. *Nutrients*, 2020; 12(5): 1505.
 47. Guo, Z., Ali, Q., Abaidullah, M., Gao, Z., Diao, X., Liu, B., Wang, Z., Zhu, X., Cui, Y., Li, D., & Shi, Y. High fat diet-induced hyperlipidemia and tissue steatosis in rabbits through modulating ileal microbiota. *Applied microbiology and biotechnology*, 2022; 106(21): 7187–7207.
 48. Stellaard F. From Dietary Cholesterol to Blood Cholesterol, Physiological Lipid Fluxes, and Cholesterol Homeostasis. *Nutrients*, 2022; 14(8): 1643.
 49. Zhao, B., Gan, L., Graubard, B. I., Männistö, S., Albanes, D., & Huang, J. Associations of Dietary Cholesterol, Serum Cholesterol, and Egg Consumption With Overall and Cause-Specific Mortality: Systematic Review and Updated Meta-Analysis. *Circulation*, 2022; 145(20): 1506–1520.
 50. Kirkpatrick, C. F., Sikand, G., Petersen, K. S., Anderson, C. A. M., Aspry, K. E., Bolick, J. P., Kris-Etherton, P. M., & Maki, K. C. Nutrition interventions for adults with dyslipidemia: A Clinical Perspective from the National Lipid Association. *Journal of clinical lipidology*, 2023; 17(4): 428–451.
 51. Anne Moorhead, S., Welch, R. W., Barbara, M., Livingstone, E., McCourt, M., Burns, A. A., & Dunne, A. The effects of the fibre content and physical structure of carrots on satiety and subsequent intakes when eaten as part of a mixed meal. *The British journal of nutrition*, 2006; 96(3): 587–595.
 52. Nicolle, C., Cardinault, N., Aprikian, O., Busserolles, J., Grolier, P., Rock, E., Demigné, C., Mazur, A., Scalbert, A., Amouroux, P., & Rémésy, C. Effect of carrot intake on cholesterol metabolism and on antioxidant status in cholesterol-fed rat. *European journal of nutrition*, 2003; 42(5): 254–261.
 53. Feingold, K. R. (2024). The Effect of Diet on Cardiovascular Disease and Lipid and Lipoprotein Levels. In K. R. Feingold (Eds.) *et. al., Endotext*. MDText.com, Inc.
 54. Obia, O., & Eifuobhokhan, J. Effect of *Justicia carnea* leaf extract on plasma and fecal lipid profile in high-fat diet fed wistar rats. *International Journal of Health and Pharmaceutical Research*. 2024; 9(4): 64-70.
 55. Eifuobhokhan, J., Obia, O., & Charles, C. *Justicia carnea* leaf extract improves intestinal transit in high-fat diet induced delayed gut motility in wistar rats. *European Journal of Pharmaceutical and Medical Research*. 2024; 11(12): 50-54.
 56. Cheng, H. M., Koutsidis, G., Lodge, J. K., Ashor, A., Siervo, M., & Lara, J. Tomato and lycopene supplementation and cardiovascular risk factors: A systematic review and meta-analysis. *Atherosclerosis*, 2017; 257: 100–108. <https://doi.org/10.1016/j.atherosclerosis.2017.01.009>
 57. Obia, O., & Asuquo, E.A. “Endogenous Antioxidant Responses to Dietary Honey Supplementation in Alloxan induced Diabetic Wistar Rats”. *IOSR Journal of Nursing and Health Science (IOSRJNHS)*, 2018; 7(6): 37-40.
 58. Obia, O., E. Odum, J., & N. Chuemere, A. Nephroprotective and Antihyperlipidemic Activity of Honey in Alloxan Induced Diabetic Wistar Rats. *International Journal of Biochemistry Research & Review*, 2018; 22(1): 1–7. <https://doi.org/10.9734/IJBCRR/2018/41585>
 59. Aune D. Plant Foods, Antioxidant Biomarkers, and the Risk of Cardiovascular Disease, Cancer, and Mortality: A Review of the Evidence. *Advances in nutrition (Bethesda, Md.)*, 2019; 10(4): S404–S421. <https://doi.org/10.1093/advances/nmz042>
 60. Chinko, B. C., Pughikumo, D. T., Obia, O., Udeh, W. C., & Hart, V. O. Honey Attenuates Phenylhydrazine-Induced Hematotoxicity and Oxidative Stress in Male Wistar Rats. *International Blood Research & Reviews*, 2023; 14(3): 10–18.
 61. Marston, N. A., Giugliano, R. P., Im, K., Silverman, M. G., O'Donoghue, M. L., Wiviott, S. D., Ference, B. A., & Sabatine, M. S. Association Between Triglyceride Lowering and Reduction of Cardiovascular Risk Across Multiple Lipid-Lowering Therapeutic Classes: A Systematic Review and Meta-Regression Analysis of Randomized Controlled Trials. *Circulation*, 2019; 140(16): 1308–1317.
 62. Sirtori, C. R., & Fumagalli, R. LDL-cholesterol lowering or HDL-cholesterol raising for cardiovascular prevention. A lesson from cholesterol turnover studies and others. *Atherosclerosis*, 2006; 186(1): 1–11.
 63. Kjeldsen, E. W., Nordestgaard, L. T., & Frikke-Schmidt, R. HDL Cholesterol and Non-Cardiovascular Disease: A Narrative Review. *International journal of molecular sciences*, 2021; 22(9): 4547.