



**EFFECTIVE NOVEL PHARMACEUTICAL TECHNOLOGIES FOR
BRAIN DRUG DELIVERY SYSTEMS: AN REVIEW**

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ABSTRACT

The regulated entry of most blood-borne substances into the brain has been recognized for more than a century. The blood-brain barrier shielding function provided by endothelial cells is important in the treatment of neurological diseases because this exclusion of foreign substances also restricts entry of many potentially therapeutic agents into the brain. BBB allows a selective entry of nutrients and minerals across it and limits the entry of foreign substances like drugs as well as neuro pharmaceutical agents. The conventional drug delivery systems which release drug into general circulation fail to deliver drugs effectively to brain and is therefore not very useful in treating certain

diseases that affect CNS including Alzheimer's disease, dementia, Parkinson's disease, mood disorder, AIDS, viral and bacterial meningitis. The various strategies available and under development for enhancing drug delivery to the CNS are reviewed. The present review enlightens about several novel approaches including nanotechnology based approach like nanoparticles, liposomes, antibody mediated delivery approach and application of genomics in brain drug targeting that would give an insight to the researchers, academia and industrialists.

KEYWORDS: BBB, Nanoparticle, Liposomes, Chimeric peptides, Mannitol.

INTRODUCTION

Disorders of the central nervous system are numerous and affect a large part of the world's population. The neurodegenerative diseases, such as Alzheimer's, Parkinson's diseases and multiple sclerosis are characterized by symptoms related to movement, memory, and

dementia due to the gradual loss of neurons. Stroke, ischemia, human immunodeficiency virus infection, epilepsy, and other psychiatric disorders such as anxiety, depression and schizophrenia conditions that markedly affect the morbidity and mortality in modern society. Brain tumors, including gliomas, astrocytomas and glioblastomas, constitute a relevant and unsolved clinical problem and the treatment of brain cancers are major challenges.^[1] A successful therapy for CNS disorders crucial milestones: (1) crossing the BBB into the brain, (2) reaching the specific cells to which it is directed, and (3) performing its ameliorative function there. Further, any therapeutic must have a safety profile that supports chronic dosing. The BBB allows the creation of a unique extracellular fluid environment within the central nervous system (CNS) whose composition can, as a consequence, be precisely controlled.

The extracellular fluid compartments of the CNS comprise the brain and spinal cord parenchymal interstitial fluid (ISF) and the cerebrospinal fluid (CSF), contained within the ventricles of the brain and the cerebral and spinal subarachnoid spaces. The structural BBB is created by the cerebral endothelial cells forming the capillaries of the brain and spinal cord. The fact that dye injected intravenously into animals stains all body tissues except the brain has been known for more than a century; this observation shows that the brain capillaries prevent molecules from entering the intact central nervous tissue. Thus, the concept of the blood-brain barrier (BBB) was established. The BBB is a result of the endothelial cells of normal brain capillaries, which have many unique properties.^[2-7] Endothelial cells of brain capillaries have fewer pinocytotic vesicles and more mitochondria than those of capillaries elsewhere in the body, and the capillaries themselves lack fenestrations and inter endothelial passages. Various transport mechanisms at the BBB have been explained for the transport of these substances (Fig. 1). These transport systems mainly operate in the luminal and abluminal membranes, i.e. from both blood-to-brain and brain-to-blood directions. But the blood-to-brain transport system is of considerable interest in drug delivery for targeting of drug molecules into brain as compared to brain-to-blood transport system.

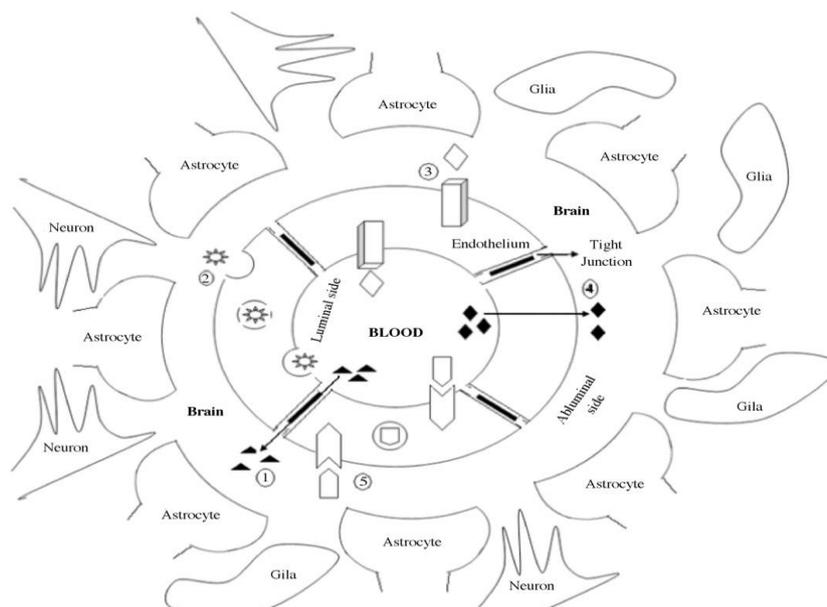


Fig.1

In addition, some regions of the CNS called as circumventricular organs (CVO) are present adjacent to the ventricles of brain where BBB capillary endothelial tight junctions are absent. These brain sites are unique in terms that they are highly vascularised as compared to other brain regions and lacks BBB because the capillary system supplying the CVOs contains fenestrated endothelial cells instead of epithelial tight junction.^[8-10] Examples of such areas include choroid plexus, pineal gland, neurohypophysis, median eminence, organosum vasculosum of lamina terminalis, subfornical organ (SFO), area postrema of the chemoreceptor trigger zone (CTZ) and nucleustractus solitarius (NTS). These sites require intimate contact to closely monitor the composition of the blood and to respond accordingly. Compared to the area of tight BBB capillaries, the relative surface area of the capillaries of CVOs is very less (5000:1) which enables CVOs not to allow a significant diffusion of substances into the CNS.

Physicochemical properties of Brain delivery: The majority of drugs that are used to treat CNS disease have a molecular weight between 150 and 500 Daltons and log octanol /water partition coefficient between 0.5 and 6.0. It is generally assumed that charged molecules cannot readily penetrate the BBB thus, for a drug that is partially ionized at physiological pH 7.4, it is the uncharged fraction that determines the diffusion gradient across the BBB and forms the driving force for any passive diffusive movement of drug. Relatively small chemical modifications to a molecule may enhance the circulatory half-life and increase the area under the curve in plasma. This increase in half-life may stem from a reduced peripheral

distribution volume or resistance to enzymatic hydrolysis in the circulation. In addition, several highly lipid soluble molecules whose CNS penetration would be expected to be significant do not have the expected high BBB penetration. These drugs were initially thought to be excluded from the CNS by virtue of their physical properties, as they are generally all bulky molecules of high molecular weight. However, it is apparent that not all of the lipid-soluble molecules that are excluded from the CNS share these same characteristics.^[11] It is now clear that these drugs are substrates for the ABC group of efflux transporters mentioned earlier, which continually hydrolyze ATP and are able to extrude drugs from the cerebral capillary endothelial cells and the CNS into blood against a concentration gradient. These ABC transporters include Pgp (Pgp /ABCB1), multidrug resistance proteins (MRP/ABCC1–12), and breast cancer resistance protein (BCRP/ABCG2).^[12]

BBB transport system: The BBB rigorously limits transport into the brain. BBB not only functions as a physical barrier, but also a biochemical barrier that expresses certain enzymes like peptidases along with several cytosolic enzymes and efflux p-glycoprotein system that helps effluxing drugs from the endothelial cells back into the blood which helps in its further protecting action towards the brain microenvironment. Cells have different receptors for the uptake of many different types of ligands, including hormones, growth factors, enzymes, and plasma proteins. It occurs at the brain for macromolecular substances, such as transferrin, insulin, leptin, and IGF-I & IGF-II, and is a highly specific type of energy dependent transport.^[13-15]

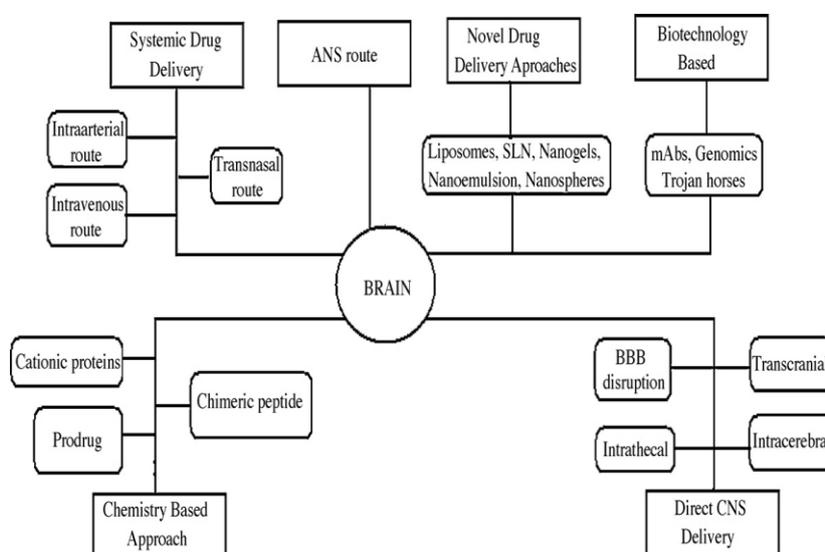


Fig.2

STRATEGIES FOR ENHANCED CNS DELIVERY

1. Direct systemic delivery

Several strategies have been studied shown in Fig. 2 for effective clinical outcome for different CNS conditions.

a. Intravenous delivery

It delivers drugs directly into general circulation by avoiding its first-pass metabolism and has potential to transport drugs to the brain. However, there is little accumulation of the drug in the brain because of the BBB and rapid clearance from the ECF (extracellular fluid). In addition, the brain availability of drug through IV route is largely affected by the half life of the drug in the plasma, rapid metabolism, level of non-specific binding to plasma proteins and the permeability of the compound across the BBB and into peripheral tissues.^[16] The outcome of the route was found to be quite effective in delivery of drugs to brain when administered using a suitable carrier system like polymeric depots, liposomes or lipid carriers.

b. Olfactory route

Delivering drugs for systemic action as the drug directly reaches to blood by crossing the nasal mucosa. The highly permeable nasal epithelium allows rapid drug absorption to the brain due to high total blood flow, porous endothelial membrane, large surface area and avoidance of first-pass metabolism. The olfactory region of respiratory region^[17], is the foremost site from where drug can be absorbed directly into the brain by different mechanisms including transcellular, paracellular, olfactory and trigeminal neural pathways. The olfactory region of nasal mucosa contains olfactory cells which extend up into cranial cavity. When the drug formulation on nasal installation, comes in contact with the mucosa they are rapidly transported directly into the brain, skipping the BBB achieving very rapid cerebrospinal fluid levels.

c. Intra-arterial delivery

The basic mechanism behind the bioavailability of drug in brain may be due to movement of drug in capillaries, then to choroid plexus epithelium and finally reaching CSF or by falling into arterial blood and then going to CSF through white matter and perivascular pathway.^[18]

2. Direct CNS delivery

a. Intracerebral delivery

Intracerebral delivery involves delivery of drug directly into parenchymal space of the brain. The major problem with bolus injection is slower movement of compounds within the brain due to the limited diffusion coefficient.^[19] The reason is due to the closely packed arrangement of cells in both gray as well as white matter microenvironment and due to the concentration dependent diffusion phenomena in brain. Intracerebral implants are devices for controlled release of drugs at the target site in the brain. Polymer depots have been used for the delivery of drug into cerebral environment in the tumor cavity of the brain having drug being present inside the polymer matrix as a core material. It offers sustained release of drugs by the biodegradation of polymer.^[30-31]

b. Trans cranial drug delivery

This route is best suited for meningioma treatment and metastatic cells of CSF^[20] as it distributes drugs mainly into ventricles and subarachnoid area of brain. The major disadvantages are the chance of causing subependymal astroglial reaction due to high drug exposure at the ependymal surface of brain.

c. Intrathecal delivery

Intrathecal route involves delivery of neurotherapeutic agents to brain by direct administration of drugs through intrathecal route into cisterna magna of brain. Intrathecal route is best suited for drug delivery for treatment of spinal diseases and disseminated meningeal diseases but not for large parenchymal diseases like parenchymal tumors such as glioblastoma.^[21]

d. BBB disruption

Drug substances are directly delivered to CNS by the use of certain chemical substance or by the application of energy like ultrasonic waves or electromagnetic radiations externally which helps in the opening of tight junctions and provides successful entry of drugs into the brain. In the first method various chemical substances are used which have hyperosmolar nature, i.e., have higher hypertonicity or osmotic pressure.^[22] Hypertonic solution causes opening of tight junctions due to higher osmotic pressure, which leads to shrinking of endothelial cells, by which disarrangement of extracellular proteins occurs and finally entry of drug takes place paracellularly. Various osmotic substances have already been tested and amongst these

mannitol has been found to be effective and safe . It is administered by intracarotid arterial infusion along with drug substances with the help of cannula.

Mannitol has been used for several years for treating brain tumors where barrier is opened for 30 min when 25% solution of drug is allowed for 30 s . Similarly, enhanced methotrexate delivery to the CNS has been attained by intra-arterial administration combined with osmotic disruption of the BBB by mannitol compared with simple intra-arterial or intravenous administration several other substances have been found that can alter the BBB permeability including bradykinin, alkylglycerols.

Secondly, BBB disruption can be achieved by the application of ultrasound^[23] and electromagnetic radiation. The important advantage of this approach is its specificity for targeting to a specific area of brain. There are three basic mechanisms proposed for describing the path of entry of drug molecules into brain using this method. Firstly the temperature of ultrasonic waves induces thermal lesions leading to alteration of permeability and generalized opening of BBB. The normal dose of ultrasonic energy applied is very low, approximately around 200–300 kHz. Secondly the cavitation effect produced by the injected fluid leads to formation of small air filled cavity in barrier's luminal membrane making the entry of drugs easier to brain and the third possible mechanism describes the formation of micro-bubbles by ultrasonography contrast agent. This method is now used mostly as diagnostic tool as a contrast agent for imaging the brain microenvironment as well as tumors.^[26-29]

Permeation enhancers such as surfactants, sodium dodecyl sulphate, Dimethyl sulfoxide, polysorbate, PEG-Hydroxy stearate.^[32]

3. Prodrugs and chemical delivery systems

- a. Inert alternatives of original API to get a product of large bulky structure which has neither biological toxicity nor activity.^[24] These prodrugs give active moiety at the site of action, in brain due to the enzyme specific nature of them. Whenever these moieties reach the brain through blood, the enzymes present on the surface of BBB help in metabolism of prodrugs to give active agents, which then cross the BBB and attains concentration in brain. These are several approaches such as

- Prodrug “Lock-in” mechanism for drug targeting^[33]
- Cyclodextrin complexes for drug delivery
- Biotin-avidin conjugated system for drug delivery
- Antibody directed enzyme prodrug therapy (ADEPT)

b. Chimeric peptides

The word “chimeric” obtained from the Greek word chimera means an animal having body of lion and head of human. The conjugated vector maybe endogenous peptides, monoclonal antibodies (mAbs), modified proteins, peptidomimetic antibodies. These chimeric peptides are formed by covalent binding of a BBB non-permeable neuropeptide with the vector. For example, insulin and transferrin are the two circulating peptides which undergo transcytosis by their insulin and transferrin receptor present at BBB.^[25]

c. Cationic proteins

It is the best suited technique for delivering proteins and peptides with a basic isoelectric point to the brain. Hence this method provides an additional advantage for delivering them by making them charged into cationic form, which can easily enter brain by electrostatic interaction with anionic functional groups present on brain surface. After cationization they easily enter by using the transcellular adsorptive-mediated endocytosis pathways.^[26]

4. Novel approaches for brain targeting

a. Liposomes

Related strategy for achieving transit of neuropharmaceuticals across the BBB uses the attachment of immunoreactive moieties to liposomes. The formation of immunoliposomes designed to target specific tissues has been known for two decades.^[35-37] They possess advantages of carrying hydrophilic, lipophilic as well as amphoteric drug molecules either entrapped inside it or on its micellar surface. The brain distribution of long circulating liposomes can be modulated by conjugation of appropriate targeting vectors.

b. Nanoparticles

Nanosystems employed for the development of nano drug delivery systems in the treatment of CNS disorders include polymeric nanoparticles, nanospheres, nanosuspensions, nanoemulsions, nanogels, nano-micelles and nano-liposomes, carbon nanotubes, nanofibers and nanorobots, solid lipid nanoparticles (SLN), nanostructured lipid carriers (NLC) and lipid drug conjugates (LDC).

5. Biotechnology based approach

a. Monoclonal antibodies for targeting

b. Application of genomics in brain drug delivery

Possible methods for delivery of gene therapeutics specifically to the human brain include invasive craniotomy, cationic-liposome–DNA complexes, or viral vectors; but each technique has some limitations.^[34] Gene therapy would ideally use a non-viral formulation, intravenously administered, with widespread gene expression throughout the brain, and specific localization to the brain with no expression in peripheral tissues. In theory, gene expression in the brain could be altered with the targeting techniques discussed above, in combination with gene promoters specific to particular cells.

c. Molecular Trojan Horses (monocytes) for brain targeting

6. Drug delivery through autonomic nervous system

The autonomic nervous system has now come into existence as an ideal route for delivering drugs to brain.

CONCLUSION

Further physicochemical modification of these more potent drugs may provide a small but vital increment in their CNS permeability, which results in a very significant increase in the therapeutic index. It is armed with this improved knowledge that drug development can move into a more informed and rational phase aimed at optimizing drug design for delivery to the CNS. An effectual advance for achieving capable drug delivery would be to judiciously develop delivery strategies based on the understanding of their interactions with the biological environment, changes in cell receptors that occur with progression of disease, mechanism and site of drug action, multiple drug administration, and pathobiology of the disease under consideration. The search for new methods of achieving BBB transit of specific brain-targeted compounds is just beginning, but the remarkable progress of the past decade suggests that the ability to deliver BBB-impermeable agents across the brain endothelium is an attainable goal.

REFERENCES

1. Mayeux, R. Epidemiology of neurodegeneration. *Annu. Rev. Neurosci*, 2003; 26: 81-104.
2. Cornford EM, Hyman S. Blood-brain barrier permeability to small and large molecules. *Adv Drug Del Rev*, 1999; 36: 145–63.

3. Pardridge WM. Drug and gene targeting to the brain with Trojan horses. *Nature Rev* 2000; 1: 131–39.
4. Stewart PA, Hayakawa K, Farrell CL. Quantitation of blood-brain barrier ultrastructure. *Microsc Res Tech* 1994; 27: 516–27.
5. Minn A, Ghersi-Egea JF, Perrin R, Leninger B, Siest G. Drug metabolizing enzymes in the brain and cerebral microvessels. *Brain Res Rev*, 1991; 16: 65–82.
6. Abbott NJ, Romero IA. Transporting therapeutics across the blood-brain barrier. *Mol Med Today*, 1996; 2: 106–13.
7. Pardridge WM, Golden PL, Kang YS, Bickel U. Brain microvascular and astrocyte localization of P-glycoprotein. *J Neurochem*, 1997; 68: 1278–85.
8. Rapoport, S. I. (2000). Osmotic opening of the blood-brain barrier: principles mechanism and therapeutic applications. *Cell Mol Neurobiol*, 20, 217–230.
9. Rennels, M. L., Gregory, T. F., & Fugimoto, K. (1983).
10. Innervation of capillaries by local neurons in the cat hypothalamus: a light microscopic study with horseradish peroxidase. *J Cereb Blood Flow and Metab*, 3; 535–542.
11. Richard, J. P., Melikov, K., Vives, E., Ramos, C., Verbeure, B., Gait, M. J., Chemomordik, L. V., & Lebleu, B. (2003). Cell penetrating peptides: a re-evaluation of the mechanism of cellular uptake. *J Biol Chem*, 2003; 278: 585–590.
12. Rousselle, C., Clair, P., Lefauconnier, J. -M., Kaczorek, Michel, Scherrmann, Jean-Michel, & Temsamani, Jamal (2000). New advances in the transport of doxorubicin through the blood-brain barrier by a peptide vector-mediated strategy. *Mol Pharmacol*, 2000; 57: 679–686.
13. Morita, K., Sasaki, H., Furuse, M., & Tsukita, S. (1999). Endothelial claudin: claudin 5 TMVCF, constitutes tight junction strands in endothelial cells. *J Cell Biol*, 1999; 147: 185–194.
14. Neuwelt, E. A. (2004). Mechanisms of disease: the blood-brain barrier. *Neurosurgery*, 2004; 54: 131–142.
15. Granholm AC, Backman C, Bloom F, et al. NGF and anti-transferrin receptor antibody conjugate: short and long term effects on survival of cholinergic neurons in intraocular septal transplants. *J Pharmacol Exp Ther*, 1994; 268: 448–59.
16. Wu D, Boado R, Pardridge WM. Pharmacokinetics of blood-brain barrier transport of [3H]-biotinylated phosphorothioate oligodeoxynucleotide conjugated to a vector-mediated drug delivery system. *J Pharmacol Exp Ther*, 1996; 276: 206–11.

17. Pardridge WM, Boado R, Kang YS. Vector mediated delivery of a peptide nucleic acid through the blood-brain barrier in vivo. *Proc Natl Acad Sci USA*, 1995; 92: 5592–96.
18. Deguchi Y, Kurihara A, Pardridge WM. Retention of biologic activity of human epidermal growth factor following conjugation to a blood-brain barrier drug delivery vector via an extended poly (ethylene glycol) linker. *Bioconj Chem*, 1999; 10: 32–37.
19. Manninger, S.P., Muldoon, L.L., Nesbit, G., Murillo, T., Jacobs, P.M., Neuwelt, E.A., 2005. An exploratory study of ferumoxtran-10 nanoparticles as a blood–brain barrier imaging agent targeting phagocytic cells in CNS inflammatory lesions. *AJNR Am. J. Neuroradiol.* 2005; 26; 2290–2300.
20. Ommaya, A.K., 1984. Implantable devices for chronic access and drug delivery to the central nervous system. *Cancer Drug Deliv.* 1984; 1:169–179.
21. Pan, J., Feng, S.S., 2008. Targeted delivery of paclitaxel using folate- decorated poly(lactide)-vitamin E TPGS nanoparticles. *Biomaterials*, 2008; 29 (17): 2663–2672
22. Pandit, J.K., Singh, S., Muthu, M.S., 2006. Controlled release formulations in neurology practice. *Ann. Indian Acad. Neurol.* 2006; 9 (4): 207–216
23. Pardridge, W. M. (2002). Drug and gene targeting to the brain with molecular Trojan horses. *Nat Rev Drug Discov*, 2002; 1: 131– 139.
24. Vive's, E., Brodin, P., & Lebleu, B. (1997). A truncated HIV-1 Tat protein basic domain rapidly translocates through the plasma membrane and accumulates in the cell nucleus. *J Biol Chem*, 1997; 272: 16010–16017.
25. Warrington, J. S., Greenblat, D. J., & von Moltke, L. L. (2004). The effect of age on P-glycoprotein expression and function in the Fischer-344 rat. *J Pharm Exp Ther*, 2004; 309: 730–736.
26. Zhao, M., & Weissleder, R. (2004). Intracellular cargo delivery using Tat peptide and derivatives. *Med Res Rev*, 2004; 24: 1 – 12.
27. Zlokovic, B. V. (2004). Clearing amyloid through the blood-brain barrier. *J Neurochem*, 2004; 89: 807– 811.
28. Lee HJ, Engelhardt B, Lesley J, Bickel U, Pardridge WM. Targeting rat anti-mouse transferring receptor monoclonal antibodies through blood-brain barrier in mouse. *J Pharmacol Exp Ther*, 2000; 292: 1048–52.
29. Shi N, Boado RJ, Pardridge WM. Receptor-mediated gene targeting to tissues in vivo following intravenous administration of pegylated immunoliposomes. *Pharm Res*, 2001; 18: 1091–95.

30. Schronder, U., Sommerfeld, P., Ulrich, S., Sabel, B.A., 1998. Nanoparticle technology for delivery of drugs across the blood–brain barrier. *Int. J. Pharm. Sci.* 1998; 87: 1305–1307.
31. Wu, H., Hu, K., Jiang, X., 2008. From nose to brain: understanding transport capacity and transport rate of drugs. *Expert Opin. Drug Deliv.* 2008; 5 (10): 1159–1168.
32. Keep RF. Editorial comment. *Stroke*, 2001; 32: 1383–84.
33. Schwarze, S. R., Ho, A., Vocero-Akbani, A., and Doway, S.(1999). In vivo protein transduction: delivery of a biologically active protein into the mouse. *Science*, 1999; 285: 1569–1572.
34. Pardridge WM. Brain drug targeting. New York: Cambridge University Press, 2001: 353.
35. Huang A, Kennel SJ, Huang L. Immunoliposome labeling: a sensitive and specific method for cell surface labeling. *J Immunol Methods* 1981; 46: 141–51.
36. Huang A, Kennel SJ, Huang L. Interactions of immunoliposomes with target cells. *J Biol Chem*, 1983; 258: 14034–40.
37. RJ, Huang L. Interactions of antigen-sensitized immunoliposomes with immobilized antibody: a homogeneous solid phase immunoliposome assay. *J Immunol*, 1985; 134: 4035–40.