

Behavioural Responses of *Cryptolaemus montrouzieri* (Coleoptera: Coccinellidae) to *Paracoccus marginatus* (Papaya Mealybug) Related Cues in Laboratory Conditions

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ABSTRACT

Purpose: Cryptolaemus montrouzieri Mulsant (Coleoptera: Coccinellidae) is a well-known biocontrol agent that is widely used to control soft-bodied insects including papaya mealybug. This study aimed to evaluate the behaviour of C. montrouzieri to chemical simulants associated with honeydew (HD) and crushed papaya mealybugs (CPMBs), under laboratory conditions.

Research Method: The larvae and adult stages of C. montrouzieri were introduced into a Petri dish containing HD or CPMBs treated and untreated Carica papaya (Papaya) leaf discs, and the behaviour was recorded for 20 minutes using video surveillance. Searching time, walking speed, and number of turns on leaf discs were quantified.

Findings: Results revealed that the searching time of C. montrouzieri, on papaya leaf discs was significantly varied with different larval instars and adults ($F_{(4,298)} = 17.21$, p<0.05) as well as leaf-coated substances ($F_{(2,298)} = 78.07$, p<0.05). When considering all larval instars of C. montrouzieri searching time on CPMBs (690.7 ± 25.3 s) was significantly higher compared to HD (524.8 ± 33.5 s) and clean leaf disc (219.6 ± 34.4 s) ($F_{(2,227)} = 60.61$, p<0.05). The searching period of the first larva (L1) was significantly higher compared to adults in all treatments (p<0.05). The walking speed of the fourth larva (L4) and adults of C. montrouzieri on HD and CPMBs leaf discs was significantly lower compared to that of clean leaf discs. The number of turns per unit path length was significantly higher in both HD (1.100 /cm) and CPMBs (1.129 / cm) leaf discs compared to the control (0.501 /cm) concerning larvae and adults.

Originality/value: It leads to the conclusion that HD and CPMBs considerably change the searching behaviour of larvae and adults of C. montrouzieri. Further work should be focused on examining and utilizing these chemical cues for searching to enhance the efficacy of C. montrouzieri in controlling papaya mealybug.

Keywords: Crushed mealybugs, Cryptolaemus, honeydew, Paracoccus marginatus, searching behaviour, walking

INTRODUCTION

Paracoccus marginatus Williams Granara de Willink, better known as the Papaya Mealybug (PMB), is a soft-bodied insect belonging to the family Pseudococcidae (Hemiptera; Sternorrhyncha), poses a substantial threat to agriculture due to its ability to feed on over 200 host plants, including the fruit crop, papaya ^{1*}Postgraduate Institute of Agriculture, University of Peradeniya, Sri Lanka

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(Carica papaya L.) (Finch et al., 2020). PMB as well as other Homopteran insects (i.e., aphids, whiteflies, scale insects, etc.) excrete honeydew which contains sugar, amino acids, and waxes (Leroy et al., 2014). These substances are directly derived from phloem sap and are sometimes synthesized by the insect itself (Buitenhuis et al., 2004). Honeydew acts as a source of carbohydrates for its foragers, (*i.e.*, ants, predators, and parasitoids) (Buitenhuis et al., 2004). Honeydew also acts as an info-chemical for foraging parasitoids and predators (Buitenhuis et al., 2004). The chemicals associated with honeydew are often described as 'kairomones' (Heidari and Copland, 1993). Kairomones are highly volatile compounds by nature and attract foragers or facilitate them for intensive searches in the host locations and subsequently, increase the chance of prey encounter (Heidari and Copland, 1993). Mealybug-infested leaves often get contaminated with wax and possibly with honeydew (Meiracker et al., 1990).

Many studies revealed that honeydew and wax attract adult-stage predators and parasitoids and honeydew and/or wax act as an arresting stimulus (Leroy et al., 2014). An arrestant is a chemical substance that causes insect aggregation by indirect kinetic reactions, either by slowing down or stopping the movement (Leroy et al., 2014). Waage (1978) has categorized the insect arrestment responses into several categories namely: orthokinetic response (dent the walking speed) or boost turning which can be non-directional (klinokinetic response) or unidirectional (klinotactic response). The searching behaviour of predators and parasitoids has been studied by several authors using different methods and developed some mathematical models that can be used to describe the outcome of a predator or parasitoid search (Evans, 1976).

Olfactory cues accelerate the chance of prey encounters and thereby enhance the efficacy of predators or parasitoids (Purandare and Tenhumberg, 2012). Location of prey by immature life stages of predators (*i.e.*, larva), occurs without involving visual cues (Carter and Dixon, 1984). However, coccinellids show increased searching for a location, followed by catching each prey (Carter and Dixon, 1984). Most of the previous studies regarding honeydew and the searching behaviour of foragers have been done for parasitic hymenopterans and few studies have focused on predatory coccinellids (Carter and Dixon, 1984; Purandare and Tenhumberg, 2012; Leroy *et al.*, 2014).

Starvation is an internal factor that alters the hostsearching behaviour of predators (Sengonca *et al.*, 1995). Prolonged starvation leads to intensive prey searching and directional movement towards the host (Grettenberger and Joseph, 2019). Gui and Boiteau (2010) reported that starvation is a trigger of insect dispersal. The starvation level may mediate the searching behaviour of foragers; for instance, predators alter their search path with starvation (Maselou and Fantinou, 2016). In contrast, some authors reported that there is no positive correlation between walking distance or speed, and starvation (Wallin and Ekbom, 1988; Hénaut *et al.*, 2002Pyralidae; Bengtsson *et al.*, 2004 Lamine *et al.*, 2005).

Cryptolaemus montrouzieri Mulsant (Coleoptera: Coccinellidae) is a key biocontrol agent that is widely used to control soft-bodied insects including papaya mealybug (PMB) (Paraiso and Services, 2016). Adult C. montrouzieri utilizes visual and chemical cues to locate its host (Heidari and Copland, 1992; Heidari and Copland, 1993). Further, these cues enable predators to gather information regarding the density and quality of host insects in a specific habitat (Jayanthi et al., 2012). Some researchers argued that the larvae of this coccinellid perceive prey only by physical contact (Heidari and Copland, 1992). It is, however, evident that mealybug secretions such as wax and honeydew act as arresting and oviposition stimulants for C. montrouzieri (Heidari and Copland, 1993; Merlin et al., 1996). In previous research, few studies have focused on the ability of honeydew (kairomones) to mediate host finding in C. montrouzierei. The primary objectives of this study were to examine the behaviour of all predatory stages of C. montrouzieri (L1 to adult) in response to honeydew and chemical substances associated

with whole-body crushed papaya mealybugs. Furthermore, this study compares the results with previous findings to identify any discrepancies and suggest areas for future investigation.

MATERIALS AND METHODS

Insect Cultures

PMB and C. montrouzieri adults were collected by field sampling of infested papaya plants. Colonies of mealybugs and predators were maintained under laboratory conditions at 26 ± 2 °C, 65-85% relative humidity. C. montrouzieri was reared on Pseudococcus viburni (Pseudococcidae) infested fruits of Cucurbitamoschata (pumpkin) according to the method described by Gunawardana and Hemachandra (2020). The ovisacs of PMB required for the experiments were obtained from cultures maintained on potato sprouts (Solanum tuberosum L.) following the method described by Nisha and Kennedy (2017). C. montrouzieri has four larval instars, and those were identified using the morphometric description by Özgökçe et al., (2006). All larval stages and adults used in these tests were uniform in size and belonged to the same generation.

Honeydew Collection and Leaf Disc Preparation

Honeydew was collected by placing Petri dish lids under potato sprouts that were heavily infested with *P. marginatus*, for 2-3 days. One honeydew droplet was carefully taken using a camel hair paintbrush and diluted with distilled water (1:1 v/v) applied on the abaxial surface of the prepared papaya leaf disc and allowed to air dry.

Extraction of Mealybugs

Three mature adult mealybugs were crushed with a pin and releasing body fluid was diluted with

distilled water (1:1 v/v) applied on leaf discs as above and allowed to air dry completely. Distilled water was applied in the control treatment.

Location Behaviour of the Predator, C. montrouzieri

Circular leaf discs of Carica papaya (Papaya) without main veins (30 mm in diameter) were placed upside down and glued in Petri-dishes (9cm, in diameter). The papaya leaf discs were treated separately with PMB honeydew crushed mealybugs and glued in the centre of the Petri dish before the test. The treated leaf discs were then offered to larvae (viz., L1 - L4) and adults of C. montrouzieri, and their activities were recorded by a digital camera (Full HD) for 20 minutes (Fig. 01.). The predator larvae were released into the experimental arena using a soft camel-hair paintbrush. The larvae and adults utilized in the experiments were obtained from rearing units, which were carefully maintained to ensure consistency in the physiological age of the coccinellids and subjected to a shorter starvation period before the test (6h). Each treatment was evaluated for each larval stage and adult with 18-24 replicates.

In all the experiments, Petri dishes containing leaf discs were placed above a fluorescent light and under a high-resolution video camera. Tests were carried out between 8:00 and 16:00 hours, to avoid any diurnal variation. The treatment order was randomized on each tested day. At the start of each experiment, one individual was placed inside the lid of the Petri dish. The individuals who were not coming into contact with the edge of a leaf disc within the video recording time were discarded. Termination of one visit was decided when the predators walked off the treated leaf disc for more than 3 minutes when it flew off from the arena or when it reached the sides or the top of the Petri dish. The arena was cleaned after each run with clean water successively and allowed to air dry after each replicate and all leaf discs were used only once. The movements of the predatory stage inside the Petri dish on

papaya leaf discs were traced onto a transparent sheet and photographed. Total path length and net travel distance in tracing photographs were measured using '*ImageJ*' software. Then, the traced search paths were used to calculate the walking speed (mm/s). Turning (>90°) per unit distance travelled (1 cm) was calculated for all the treatments. (Eq-01).

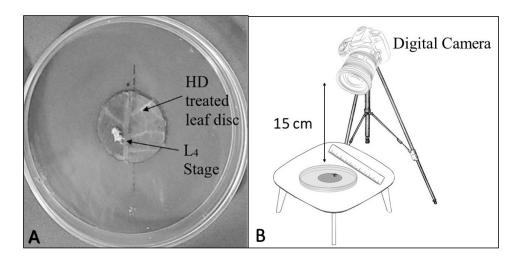
Turns per cm= No. of turns (>90 degree)/Path Length Eq01

Data Analysis

The data relating to different behavioural parameters were analysed in one-way and twoway analysis of variance (ANOVA) and means were separated using Tukey's HSD multiple range test at p < 0.05. All statistical tests were conducted using Minitab software (version 19, Minitab Inc., State College, PA, USA).

RESULTS AND DISCUSSION

The searching time of *C. montrouzieri*, on papaya (*Carica papaya* L.) leaf discs was significantly varied with different larval instars and adults (F_(4,298) = 17.21, *p*<0.05) as well as leaf-coated substances (F_(2,298) = 78.07, *p*<0.05). The interaction between leaf coat substances and actively feeding stages; larval instars and adults, was also found significant (F_(8,298) = 2.74, *p* = 0.006).



- Figure 01: L4 larvae exposed to honeydew treated leaf disc (A) and video recording set-up with fixed camera (B).
- Table 01:The searching period of different stages of C. montrouzieri on honeydew, crushed papaya
mealybug, and clean leaf discs.

Lanual instan/life stage	Searching time on leaf disc (s)			
Larval instar/ life stage —	HD	CPMBs	Clean (control)	
Ι	$674.3\pm71.0^{\mathtt{aA}}$	$747.8\pm51.4^{\mathrm{aA}}$	$270.1\pm71.0^{\text{bA}}$	
Π	$536.6\pm48.1^{\rm aAB}$	$613.5\pm42.3^{\mathtt{aA}}$	$195.0\pm71.0^{\mathrm{bA}}$	
III	$494.3\pm63.3^{\rm bAB}$	$714.8\pm46.2^{\mathtt{aA}}$	$222.1\pm60.8^{\text{cA}}$	
IV	$393.8\pm57.1^{\rm bAB}$	$686.9\pm43.7^{\mathrm{aA}}$	$191.1\pm58.9^{\rm cA}$	
Adult	$374.7\pm44.5^{\mathtt{aB}}$	$278.8\pm38.2^{\mathrm{aB}}$	$45.7\pm57.1^{\rm bB}$	

When the larval instars were on honeydewcoated leaf disc, searching time was not significantly different among larval instars and the average searching time was 524.8 ± 33.5 s. Moreover, the searching time of larval instars on CPMBs leaf coats was also not significantly different; the average searching time of larval instars on CPMBs was 690.7 ± 25.3 s. The average searching time on clean leaf discs was 219.6 ± 34.4 s. The searching time of adults was significantly different to all larval instars when on both in CPMBs ($F_{(4, 140)} = 21.88, p < 0.05$) and clean leaf coat treatments ($F_{(4, 70)} = 8.00, p < 0.05$) (Table 01).

When the first larval instar (L1) was considered across the treatments: honeydew, CPMBs, and clean leaf discs, the searching time was significantly higher both in honeydew and CPMBs leaf coats compared to clean leaf disc $(F_{(2, 40)} = 10.78, p < 0.05)$. Similar observation was observed with second (L2) ($F_{(2, 68)} = 12.55$, p < 0.05), third (L3) (F_(2, 51) = 18.26, p < 0.05) and fourth (L4) larval instars ($F_{(2,59)} = 26.33, p < 0.05$). The searching times of L3 and L4 instars on CPMBs coated leaf disc were significantly higher compared to both HD and clean leaf discs (Table 01). Further, the searching period of L1 was significantly higher compared to adults in all treatments (p < 0.05). This behavioural difference may enable first instar larvae to locate prey immediately which is essential for their survival (Leroy *et al.*, 2014). Additionally, when considering all larval instars of *C. montrouzieri* searching time on CPMBs (690.7 ± 25.3 s) was significantly higher compared to HD (524.8 ± 33.5 s) and clean leaf disc (219.6 ± 34.4 s) (F_(2, 227) = 60.61, *p*<0.05) (Fig. 02.).

These findings strongly suggest that HD and CPMBs coatings contain some compounds which may be utilised as chemo stimuli by larval and adult stages of C. montrouzieri. Further, searching more intensively in areas that contain HD or CPMBs, increases the chance of locating prey (klinokinatic response). Previous studies have shown that adult C. montrouzieri was significantly attracted to mealybug sex pheromones and showed a high searching period when mealybugs were in the vicinity (Heidari and Copland, 1992). Honeydew and other substances associated with the host mealybugs facilitate sustaining the predator in the contaminated areas. Heidari and Copland (1993) observed that the adult C. montrouzieri spent 98 % of their observed time (viz., 300 s) on the honeydewtreated leaf, whereas it spent 45 % on clean leaves. Purandare and Tenhumberg (2012) found that Hippodamia convergens Guérin-Méneville (Coleoptera: Coccinellidae) larvae spend a longer period in honeydew-coated patches than in clean patches after contact.

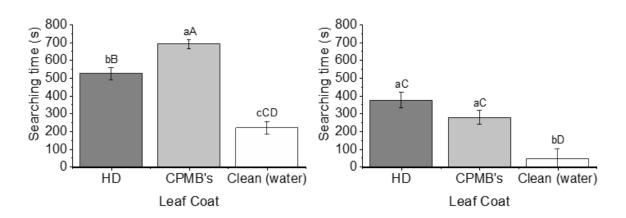


Figure 02: Searching period of *C. montrouzieri* larval stages (left) and adults (right) on papaya leaf discs. (All bars indicate SE. Means followed by the same lowercase letter within the same graph are not significantly different (p>0.05). Means followed by the same uppercase letter between two graphs are not significantly different (p>0.05) by Tukey's HSD test)

Resting time (on leaf disc) was significantly varied with predatory life stage ($F_{(4, 298)} = 9.19$, p<0.05), but not significantly varied with leafcoated material. Young instars of C. montrouzieri (L1, L2, and L3) showed a significantly higher resting time in all treatments whereas L4 and adults showed a significantly lower resting period (Fig. 03.). Similar observations for adults were reported by (Heidari and Copland, 1993), where the resting time was not significantly different between honeydew treated and clean leaves. In contrast, the same authors reported that the resting period of L4 significantly differed between honeydew-treated leaves and clean leaves. These differences probably resulted due to differences in host mealybug, leaf material, and maturity of L4 larvae.

speed was Walking significantly varied concerning different larval instars and adults (F (4, 736) = 647.3, p < 0.05) and leaf-coated substances (F $_{(2, 736)}$ = 121.9, p<0.05) in all treatment combinations. The interaction between leaf-coated substances and the predatory life stage also significantly affected C. montrouzieri walking speed on papaya leaf discs (F $_{(8, 736)}$ = 75.7, p<0.05). Adults of C. montrouzieri showed the highest walking speed in all treatments while the first larval instar (L1) showed the lowest walking speed in all treatments. Moreover, there was no significant difference in walking speed between L1 and L2 as well as between L3 and L4 larval instars in all treatments (Table 02).

The walking speed of L4 and adults of *C.* montrouzieri on HD and CPMBs leaf discs was significantly lower compared to that of control/ clean leaf discs. Further, larval instars and adults of *C. montrouzieri* walk significantly faster outside the leaf disc in HD and CPMBs coated papaya leaf discs (p<0.05), but significant difference was not observed in and outside the leaf disc for L2, L3, and L4 in control/ water coated leaf discs. Moreover, adults of *C. montrouzieri* were walking faster outside the papaya leaf disc (12.35 \pm 0.14 mm/s) compared to larval stages. Adults showed the highest walking speed on clean leaf discs (9.26 \pm 0.21 mm/s) and it was significantly reduced to 3.98 \pm 0.12 mm/s on HD-coated leaf

discs and 3.68 ± 0.13 mm/s on CPMBs coated leaf disc. These findings are in accordance with Banks (1957) who reported that Propylea quatuordecimpunctata (Coleoptera: (L.) Coccinellidae) larvae moved slowly on honeydew-coated bean leaves compared to clean bean leaves (orthokinesis) and increased turnings (klinokinesis). The walking speed of early instars (viz., L1, L2 and, L3) was not significantly varied concerning different leaf-coated substances (p>0.05), which was not observed in L4 and adults. Wratten (1973)Adalia bipunctata (L. also reported that larger instars of Adalia bipunctata (L.) (Coleoptera: Coccinellidae) move more rapidly than small ones. It was noted that the walking speed of the fourth instar larvae of C. montrouzieri is somewhat low (2.41 mm/s) in the honeydew-treated area compared to the value reported by Heidari and Copland (1993) (i.e., 4.81 mm/s). This is probably due to the maturity level of larvae (middle age fourth instar larvae). Heidari and Copland (1993) used fourth young instar larvae – that were just after moult – which move faster. Moreover, several internal and external factors determine insect movements, such as temperature, relative humidity, airflow, host density, level of starvation, the intensity of light, prey secretions, and host-bearing plants (Heidari and Copland, 1992). Heidari and Copland (1993) argued that the availability of honeydew is caused by intense searching by both adults and larvae of C. montrouzieri with an increased number of turns and reduced walking speed.

The number of turns (>90°) for 1 cm of the walking path is given in Table 03. The number of turns per 1 cm of the walking path was significantly varied with leaf-coated substances ($F_{(2, 457)} = 58.5$, *p*<0.05), but not with predatory life stages. The number of turns per unit path length was significantly higher in both HD (1.100 /cm) and CPMBs (1.129 /cm) coated papaya leaf discs compared to clean/control leaf discs (0.501 /cm) concerning all *C. montrouzieri* stages (klinokinesis).

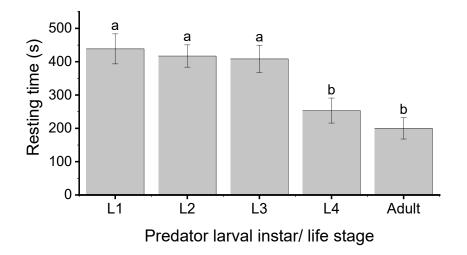


Figure 03: Resting time of *C. montrouzieri* on papaya leaf discs. (All bars indicate SE. mean values capped by the same letter are not significantly different (Tukey's HSD test; p>0.05)).

Table 02:	Walking speed (mm/s) of C. montrouzieri inside and outside the leaf disc in different
	treatments.

Treatment	Larval instar/ life stage	(a) Walking speed Inside the treated zone (mm/s)	(b) Walking speed outside the treated zone (mm/s)
HD	Ι	$0.55\pm0.23^{\rm fg}$	$1.67 \pm 0.67^{*}$
	Π	$0.91\pm0.11^{\rm fg}$	$1.85 \pm 0.35^{*}$
	III	$2.31\pm0.19^{\rm de}$	$3.39\pm0.32^*$
	IV	$2.41\pm0.18^{\rm de}$	$3.58\pm0.42^{\ast}$
	Adult	$3.98\pm0.08^{\rm b}$	$11.67\pm0.18^*$
CPMBs	Ι	$0.56\pm0.13^{\rm g}$	$1.63 \pm 0.42^{*}$
	Π	$1.11\pm0.09^{\rm fg}$	$1.97\pm0.31^{\ast}$
	III	$2.24\pm0.21^{\rm de}$	$3.08\pm0.25^*$
	IV	$2.39\pm0.15^{\rm de}$	$3.24\pm0.42^{\ast}$
	Adult	$3.68\pm0.09^{\rm bc}$	$12.35 \pm 0.19^{*}$
Control (Water)	Ι	$0.69\pm0.20^{\rm fg}$	$1.63\pm0.67^*$
	Π	$1.57\pm0.20^{\rm ef}$	$1.82\pm0.49^{\rm ns}$
	III	$2.91\pm0.23^{\rm cd}$	$3.03\pm0.47^{\text{ns}}$
	IV	$3.65\pm0.20^{\rm bc}$	$3.32\pm0.43^{\rm ns}$
	Adult	$9.26\pm0.14^{\rm a}$	$12.12 \pm 0.21^{*}$

Notes- HD, and CPMBs indicate honeydew and crushed papaya mealybug-coated leaf discs, respectively. Different letters in column (a) have significant differences.^{ns} and * in column (b) indicate no significant and significant differences within the same raw at p<0.05 by Tukey's HSD test.

Treatment	Larval instar/ life stage	Number of turns (>90°) for 1 cm of the walking path
HD	Ι	$1.24\pm0.10^{\rm a}$
	II	$1.06\pm0.05^{\rm ab}$
	III	$0.98\pm0.07^{\rm ab}$
	IV	$1.13\pm0.12^{\rm ab}$
	Adult	$1.08\pm0.11^{\rm ab}$
CPMBs	Ι	$1.16\pm0.07^{\rm a}$
	II	$1.12\pm0.05^{\rm a}$
	III	$1.13\pm0.05^{\rm a}$
	IV	$1.16\pm0.05^{\mathrm{a}}$
	Adult	$1.05\pm0.06^{\rm ab}$
Control (Water)	Ι	$0.55\pm0.09^\circ$
	II	$0.36\pm0.16^{\circ}$
	III	$0.44\pm0.19^{ ext{bc}}$
	IV	$0.46\pm0.11^\circ$
	Adult	$0.67\pm0.12^{\mathrm{bc}}$

Table 03:Number of turns (>90°) for 1 cm of walking path of C. montrouzieri under leaf coatings of
HD and CPMBs.

Notes- HD, and CPMBs indicate honeydew and crushed papaya mealybug coated leaf discs, respectively. Different letters indicate no significant and significant differences within the column at p < 0.05 by Tukey's HSD test.

However, since 1980, host searching behaviour of coccinellid was considered random and do not perceive their prey until contact, which has been challenged by several authors (Carter and Dixon, 1984; Sengonca, et al., 1995; Purandare and Tenhumberg, 2012; Urbina et al., 2018; Chunli et al., 2019). The length of time a coccinellid spends searching for food is not the same for all individuals of the same developmental stage (instar). Instead, it may vary and be influenced by the hunger level of the individual coccinellid. In other words, a hungry ladybird beetle may search for food more intensively and for a longer period than a well-fed one. (Bond, 1980; Carter and Dixon, 1982). Further, Sengonca et al., (1995) reported that the maximum olfactory response was observed in the first to fourth instar larvae and adults of C. montrouzieri after a period of starvation lasting 4, 8, 12, 12, and 24 hours, respectively. Previous studies have shown that predatory coccinellid larvae use honeydew as a kairomone while searching for host aphids. Stubbs (1980) found that adults of a seven-spotted ladybird, Coccinella septempunctata (Coleoptera: Coccinellidae) can identify host aphids at a short distance by visual cues whereas their larvae move towards crushed prey at a short distance which was identified as an olfactory detection. Stubbs (1980) and Carter and Dixon, (1984) showed C. septempunctata larvae stay a longer period on plants grown on honeydew-contaminated soil compared to the control and found that the arrestment response of this coccinellid larva was elicited by the chemicals associated with the haemolymph of crushed prey. Carter and Dixon (1984) found C. septempunctata larvae increase their searching time in ears of wheat that carry honeydew than the control. Further, Pettersson et al. (2008) showed that chemical stimuli are important in the foraging behaviour of C. septempunctata L.

Merlin *et al.*, (1996) found that the wax secretions and honeydew of host mealybugs were used as an attractant and oviposition stimulants by *C. montrouzieri*. An olfactory study done by Sengonca *et al.* (1995) pointed out that the first instar larvae and adults of *C. montrouzieri* were significantly attracted to the kairomones of *Planococcus citrl* (Risso). They also revealed that all the stages of *C. montrouzieri* were attracted to kairomones produced by mealybug *P. citri* significantly (p<0.05) indicating *C. montrouzieri* uses the kairomones of mealybug as cues in reaching the prey. all feeding stages of *C. montrouzieri* spend more time searching in leaf discs coated with honeydew and CPMBs compared to clean leaf discs (klinokinatic response). Hence, honeydew, and whole-body crushed papaya mealybug serve as behavioural stimulants of *C. montrouzieri*, under laboratory conditions. Further work on the examination or synthesis of these compounds may enhance the efficient use of *C. montrouzieri* for population suppression of mealybugs.

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CONCLUSIONS

The study revealed that the searching time of *C*. *montrouzieri*, on papaya leaf discs significantly varies with different feeding stages of the predator, and also with leaf-coated substances. Further,

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REFERENCES

- Banks, C. J. (1957). The behaviour of individual coccinellid larvae on plants. *The British Journal Of Animal Behaviour*. 1, 12–24. DOI: https://doi.org/10.1016/S0950-5601(57)80039-2.
- Bengtsson, G., Nilsson, E., Rydén, T. and Wiktorsson, M. (2004). Irregular walks and loops combine in small-scale movement of a soil insect: implications for dispersal biology. *Journal of Theoretical Biology*. 231(2), 299–306. DOI: https://doi.org/10.1016/j.jtbi.2004.06.025.
- Bond, A. B. (1980). Optimal foraging in a uniform habitat: the search mechanism of the green lacewing. *Animal Behaviour*. 28, 10–19. DOI: https://doi.org/10.1016/S0003-3472(80)80003-0.
- Buitenhuis, R., Mcneil, J. N., Boivin, G. and Brodeur, J. (2004). The role of honeydew in host searching for aphid hyperparasitoids. *Journal of Chemical Ecology*. 30 (2), 273–285. DOI: https://doi.org/10.1023/b:joec.0000017977.39957.97
- Carter, M. C. and Dixon, A. F. G. (1984). Honeydew: an arrestant stimulus for coccinellids. *Ecological Entomology*. 9(4), 383–387. DOI: https://doi.org/10.1111/j.1365-2311.1984. tb00834.x.
- Carter, M. C. and Dixon, A. F. G. (1982). Habitat quality and the foraging behaviour of coccinellid larvae. *Journal of Animal Ecology*. 51(3),865–878. DOI: https://doi.org/10.2307/4011

- Chun-li, XIU, Hong-sheng, PAN., Wei, Z., Yi-zhong, Y. and Yan-hui, L. (2019). Volatiles from Sophora japonica, flowers attract Harmonia axyridis adults (Coleoptera: Coccinellidae). Journal of Integrative Agriculture. 18(4), 873–883. DOI: https://doi.org/10.1016/S2095-3119(18)61927-6.
- Evans, H. F. (1976). The searching behaviour of *Anthocoris confuses* (Reuter) in relation to prey density and plant surface topography. *Ecological Entomology*. 1, 163–169. DOI: https://doi.org/10.1111/j.1365-2311.1976.tb01219.x
- Finch, E. A., Beale, T., Chellappan, M., Goergen, G., Gadratagi, B.G., Khan, M.A.M., Rehman, A., Rwomushana, I., Sarma, A.k., Wyckhuys, K.A. and Kriticos, D.J. (2020). The potential global distribution of the papaya mealybug, *Paracoccus marginatus*, a polyphagous pest. *Sci.* 77(3), 1361-1370. DOI: https://doi.org/10.1002/ps.6151.
- Gui, L. and Boiteau, G. (2010). Effect of food deprivation on the ambulatory movement of the Colorado potato beetle, *Leptinotarsa decemlineata*. *Entomologia Experimentalis et Applicata*. 134, 138–145. DOI: https://doi.org/10.1111/j.1570-7458.2009.00946.x
- Gunawardana, D. U. M. and Hemachandra, K. S. (2020). Mass rearing of mealybug predator, *Cryptolaemus montrouzieri* Mulsant (Coleoptera: Coccinellidae) on two mealybug species, *Planococcus minor* and *Pseudococcus viburni*. *Tropical Agricultural Research*. 31(1), 31–41. DOI: https://doi.org/10.4038/tar.v31i1.8342.
- Grettenberger, I. M. and Joseph, S. V. (2019). Influence of starvation on walking behavior of Bagrada hilaris (Hemiptera: Pentatomidae). PLoS ONE. 14(4), 1–16. DOI: https://doi.org/10.17605/OSF.IO/FQXWB.Funding.
 - Heidari, M., and Copland, M. J. W. (1993). Honeydew: a food resource or arrestant for the mealybug predator *Cryptolaemus montrouzieri*. *Entomophaga*. 38(1), 63–68. DOI: *https://doi.org/*10.1007/BF02373140
- Heidari, M. and Copland, M. J. W. (1992). Host finding by *Cryptolaemus montrouzieri* (Coleoptera : Coccinellidae) a predator of mealybusgs (Hom., Pseudococcidae). *Entomophaga*. 37(4), 621–625. DOI: https://doi.org/10.1007/BF02372333.
- Hénaut, Y., Alauzet, C., and Lambin, M. (2002). Effects of starvation on the search path characteristics of Orius majusculus (Reuter) (Het., Anthocoridae). *Journal of Applied Entomology*. 126(9), 501–503. DOI: https://doi.org/10.1046/j.1439-0418.2002.00694.x
- Jayanthi, P. D. K., Sangeetha, P., and Verghese, A. (2012). Ovipositional responses of *Cryptolaemus montrouzieri* Mulsant (Coleoptera : Coccinellidae) to the presence of prey, *Maconellicoccus hirsutus* (Green). *Journal of Biological Control*. 26(3), 240–244. DOI: https://doi.org/10.18311/jbc/2012/3495
- Lamine, K., Lambin, M., and Alauzet, C. (2005). Effect of starvation on the searching path of the predatory bug *Deraeocoris lutescens*. *BioControl*. 50(5), 717–727. DOI: https://doi.org/10.1007/ s10526-005-2899-9

- Leroy, P. D., Almohamad, R., Attia, S. and Capella, Q. (2014). Aphid honeydew : an arrestant and a contact kairomone for *Episyrphus balteatus* (Diptera : Syrphidae) larvae and adults. *European Journal of Entomology*. 111(2), 237–242. DOI: https://doi.org/10.14411/eje.2014.028
- Maselou, D. and Fantinou, A. (2016). Effect of hunger level on prey consumption and functional response of the predator *Macrolophus pygmaeus*. *Bulletin of Insectology*. 68(2), 211–218.
- Meiracker, R. A. F. Van Den, Hammond, W. N. O. and Alphen, M. Van. (1990). The role of kairomones in prey finding by *Diomus* sp . and *Exochomus* sp ., two coccinellid predators of the cassava mealybug , *Phenacoccus manihoti*. *Entomologia Experimentalis et Applicata*. 56, 209–217. DOI: https://doi.org/10.1111/j.1570-7458.1990.tb01399.x
- Merlin, J., Lemaitre, O. and Grégoire, J. C. (1996). Chemical cues produced by conspecific larvae deter oviposition by the coccidophagous ladybird beetle, *Cryptolaemus montrouzieri*. *Entomologia Experimentalis et Applicata*. 79(2), 147–151. DOI: https://doi.org/10.1111/j.1570-7458.1996. tb00820.x
- Nisha, R. and Kennedy, J. S. (2017). Life cycle of the parasitoid *Acerophagus papayae* Noyes and Schauff on papaya mealybug *Paracoccus marginatus* Williams and Granara de Willink vis-a-vis local adaptation with co- evolutionary "Arms Race". *Journal of Entomology and Zoology*. 5(3), 1711–1719.
- Özgökçe, M. S., Atlihan, R. and Karaca, I. (2006). The life table of *Cryptolaemus montrouzieri* Mulsant (Coleoptera: Coccinellidae) after different storage periods. *Journal of Food, Agriculture and Environment*. 4(1), 282–287.
- Paraiso, O. and Services, C. (2016). Cryptolaemus montrouzieri (Mulsant) (Coccinellidae: Scymninae): a review of biology, ecology, and use in biological control with particular reference to the potential impact on non-target organisms. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources. 8(5): 1–20. DOI: https://doi.org/10.1079/PAVSNNR20138005.
- Pettersson, J., Ninkovic, V., Glinwood, R., Al Abassi, S., Birkett, M., Pickett, J. and Wadhams, L. (2008). Chemical stimuli supporting foraging behaviour of *Coccinella septempunctata* L. (Coleoptera: Coccinellidae): volatiles and allelobiosis. *Applied Entomology and Zoology*. 43(3), 315–321. DOI: https://doi.org/10.1303/aez.2008.315
- Purandare, S. R. and Tenhumberg, B. (2012). Influence of aphid honeydew on the foraging behaviour of *Hippodamia convergens* larvae. *Ecological Entomology*. 37, 184–192. DOI: https://doi.org/10.1111/j.1365-2311.2012.01351.x
- Sengonca, C., Kotikal, Y. K. and Schade, M. (1995). Olfactory reactions of *Crptolaemus montrouzieri* Mulsant (Col., Coccinellidae) and *Chrysoperla carnea* (Stephens) (Neur., Chrysopidae) in relation to period of starvation. *Anzeiger fur Schadlingskunde, Pflanzenschutz, Umweltschutz.* 68(1), 9–12.
- Stubbs, M. (1980). Another look at prey detection by coccinellids. *Ecological Entomology*. 5(2), 179–182. DOI: https://doi.org/10.1111/j.1365-2311.1980.tb01139.x

- Urbina, A., Verdugo, J. A., López, E., Bergmann, J., Zaviezo, T. and Flores, M. F. (2018). Searching behavior of *Cryptolaemus montrouzieri* (coleoptera: Coccinellidae) in response to mealybug sex pheromones. *Journal of Economic Entomology*. 111(4), 1996–1999. DOI: https://doi. org/10.1093/jee/toy168
- Waage, J. K. (1978). Arrestment responses of the parasitoid, Nemeritis canescens, to a contact chemical produced by its host, *Plodia interpunctella*. *Physiological Entomology*. 3, 135–146. DOI: https://doi.org/10.1111/j.1365-3032.1978.tb00143.x
- Wallin, H. and Ekbom, B. S. (1988). Movements of carabid beetles (Coleoptera: Carabidae) inhabiting cereal fields: a field tracing study. *Oecologia*. 77(1), 39–43. DOI: https://doi.org/10.1007/ BF00380922
- Wratten, S. D. (1973). The effectiveness of the coccinellid beetle, Adalia bipunctata (L.), as a predator of the lime aphid, Eucallipterus tiliae L. The Journal of Animal Ecology. 42(3), 785. DOI: https://doi.org/10.2307/3139