SHORT COMMUNICATION

Enzyme inhibitory and immunomodulatory activities of the depsidone lobaric acid extracted from the lichen *Heterodermia* sp.

Vinitha M. Thadhani^{1*}, Qamar Naaz², M. Iqbal Choudhary², M. Ahmed Mesaik³ and Veranja Karunaratne⁴

Revised: 07 October 2013; Accepted: 11 December 2013

Abstract: This study evaluates the enzyme inhibitory and immunomodulatory activities of the lichen specific depsidone, lobaric acid. Lobaric acid was extracted with methanol from Heterodermia sp. found in Labukelle, Sri Lanka with a yield of 0.67 %. It was subjected to enzyme inhibition assays using acetyl and butyryl-cholinesterase, phosphodiesterase, β -glucuronidase, α -glucosidase and urease. In the β - glucuronidase inhibitory activity it showed an IC₅₀ value of $3.28 \pm 0.05 \mu M$, which was significantly lower than that of the standard, D-saccharic acid 1, 4-lactone (IC₅₀ = $48.4 \pm 1.2 \mu M$). Lobaric acid showed a significant inhibition of phosphodiesterase enzyme with an $IC_{50} = 313.7 \pm 2.2$, when compared with the two standards EDTA (IC₅₀ = 274.0 \pm 0.1 μ M) and cysteine (IC₅₀ = 748.0 \pm 0.1 µM). Lobaric acid showed a moderate actyl and butyryl -cholinesterase inhibitory activity while it showed no activity against the enzymes α -glucosidase and urease.

In the immunomodulatory assay, lobaric acid exhibited a potent oxidative burst inhibitory activity in human polymorphonuclear (PMN) cells. It suppressed both the myloperozidase dependant and myloperozidase independent reactive oxygen species (ROS) production of PMNs. The results indicate the pharmacological potential of lobaric acid as a lead compound for further studies.

Keywords: β-glucuronidase, enzyme inhibitory activity, *Heterodermia* sp., immunomodulatory activity, lobaric acid, phosphodiesterase.

INTRODUCTION

Lichens are a rich source of bioactive secondary metabolites (Mitrovi'c et al., 2011). Previous studies

have reported a number of compounds including the Ambewela amide A, B, sekikaic acid and lecanoric acid from Sri Lankan lichens such as *Parmotrema* sp., *Pyxine consocians* and *Heterodermia leucomelos* with anticancer, antioxidant, α-PLK1 inhibitory and insect herbivoury activities (Karunaratne *et al.*, 2002, 2005, 2008; Kathirgamanathar *et al.*, 2006; Thadhani *et al.*, 2011; Williams *et al.*, 2011). Of the major structural classes of lichen metabolites, depsidones comprise a tricyclic ring system with two benzene rings connected through ether and ester moieties and are relatively rare. The depsidone lobaric acid is one of the most biologically potent secondary metabolites reported from lichens such as *Stereocaulon sasakii*, *Stereocaulon alpinum* Laur and *Sterocaulon azoreum* (Morita *et al.*, 2009).

Various biological activities of lobaric acid have been reported earlier including antitumor, antiproliferative (Burlando et al., 2009), anti-inflammatory, antioxidant (Thadhani et al., 2011), antimicrobial (Thadhani et al., 2012) and selective serine threonine protein kinase (PLK1) inhibitory activity. Lobaric acid inhibits the 5-lipoxygenase pathway by inhibiting the formation of cysteinyl - leukotrines as determined by enzyme immunoassay (Gissurarson et al., 1997). The antimitotic activity of lobaric acid has also been studied and it has been shown to inhibit the polymerization of tubulin (Morita et al., 2009). Through interaction with a dynamic surface of the CBP/p300 GACKIX domain, lobaric acid demonstrated an enormous potential for targeting difficult protein-protein interactions (Majmudar et al., 2012; Stojanovic et al., 2012). Furthermore, it has

¹Department of Chemistry, Faculty of Applied Sciences, University of Sri Jayewardenepura, Gangodawila, Nugegoda.

²H.E.J. Research Institute of Chemistry, International Centre for Chemical and Biological Sciences, University of Karachi, Pakistan.

³Dr. Panjwani Centre for Molecular Medicine and Drug Research, University of Karachi, Pakistan.

⁴Department of Chemistry, Faculty of Science, University of Peradeniya, Peradeniya.

^{*} Corresponding author (vinimool@yahoo.com)

194 Vinitha M. Thadhani et al.

been reported that lobaric acid elicit contact allergy in sensitive persons (Thune & Solberg, 1980).

In this study, the pharmacological potential of lobaric acid, particularly its immunomodulatory and inhibitory activities against β - glucuronidase, acetyl - and butyrylcholinesterases, phosphodiesterase, α -glucosidase, and urease enzymes are reported.

METHODS AND MATERIALS

Manually cleaned, air-dried and crushed lichen specimens of a *Heterodermia* sp. collected from Labukelle, Sri Lanka were sequentially extracted with CH₂Cl₂ followed by MeOH. Lobaric acid was isolated with a yield of 0.67 % from the MeOH extracts when fractionated *via* silica gel column using hexane/CH₂Cl₂ to CH₂Cl₂/MeOH as solvents. The dentification of the compound was carried out by using spectral data and its comparison with the reported data (Gonzilez *et al.*, 1992).

Figure 1: Structure of lobaric acid

To determine the inhibitory activity of lobaric acid on acetyl and butyryl-cholinesterases enzymes, electric-eel acetyl-cholinesterase (AChE, EC 3.1.1.7) and horse serum butyryl-cholinesterase (BChE, EC 3.1.1.8) were used. The inhibition was measured using the spectrophotometric method developed by Ellman *et al.* (1961).

 β -Glucuronidase inhibitory activity was determined by measuring the absorbance of p-nitrophenol at 405 nm, formed from the substrate p-nitrophenyl- β -D-glucuronide on addition of β -glucuronidase in the presence of lobaric acid (Khan *et al.*, 2002).

Phosphodiesterases enzyme inhibitory activity was measured by a spectrophotometric method using bis-(p-nitropheny1) phosphate as chromogenic substrate, which was added to a mixture containing a buffer, phosphodiesterase enzyme and the test compound lobaric acid. The release of p-nitrophenol was measured at 410 nm.

Lobaric acid was evaluated for its α -glucosidase inhibitory activity using the method of Oki *et al.* (2000) and against the urease enzyme using the indophenols method (Weatherburn, 1967).

Lobaric acid was screened over a range of concentrations $(3.1 - 50 \mu g/mL)$ for its oxidative burst inhibitory potential. The measurement of chemiluminescence was employed to investigate the different kinds of reactive oxygen species (OH, O-2 and H₂O₂). Luminol-enhanced chemiluminescence assay was performed as described by Helfand et al. (1982). Briefly, whole blood neutrophils and polymorphonuclear leukocytes (PMNs) were suspended in Hank's balance salt solution (HBSS) with calcium and magnesium salts and incubated with lobaric acid for 30 min. To each well, serum opsonized zymosan (Saccharomyces cerevisiae origin) was added, followed by the addition of luminol (3-aminophthalhydrazide), and then HBSS to adjust the final volume to 0.2 mL. HBSS was used as a control. Chemiluminescence peaks were recorded with a luminometer.

RESULTS AND DISCUSSION

Cholinesterase inhibitors are used for the management of Alzheimer's disease. Lobaric acid showed moderate inhibition against acetyl-cholinesterase with an IC value of $26.86 \pm 0.9 \, \mu M$ and butyryl-cholinesterase with an IC value of $36.76 \pm 0.8 \, \mu M$, when compared to the standard gallanthamine (AChE: $0.50 \pm 0.01 \, \mu M$) and BChE: $8.50 \pm 0.01 \, \mu M$). It has been reported that acetylated derivatives of depsidones isolated from Lobaria pulmonaria (L.) Hoffm. (Lobariaceae) possess moderate acetyl-cholinesterase inhibitory activity (Mortia et al., 2009). This is the first report on AChE and BChE inhibitory activity of lobaric acid.

In certain diseases such as cancer, inflammatory joint disease, some hepatic diseases and AIDS, the activity of β -glucuronidase increases. An IC₅₀ value of 3.28 \pm 0.05 μ M was observed, which is 12 fold more potent than the standard D-saccharic acid 1,4 - lactone (IC₅₀ = 48.4 \pm 1.25 μ M).

Many β - glucuronidase inhibitors such as 8 - hydroxytricetin-7- glucuronide, isovitexin, trihydroxypipecolic acid and scoparic acid A and C, L - aspartic acid, tectorigenin and benzothiazoles have been isolated from different plants and some are used clinically (Khan *et al.*, 2002). However, there are no reports of β - glucuronidase inhibitory activity of compounds isolated from lichens.

Table 1: Immunomodulatory activities of loba	ic acid
---	---------

Compound	Immunomodulatory activity IC ₅₀ ± SEM (μg)		
	Whole blood + luminol	PMN's + luminol	PMN's + lucigenin
Lobaric acid (1)	37.6 ± 0.9	< 3.1	< 3.1
Ibuprofen	11.8 ± 1.87	2.5 ± 0.6	-
Sodium diethyldithio carbamatetrihydrate	-	1.27 ± 0.23	8.16 ± 1.9

Phosphodiesterase is believed to be involved in a wide variety of processes, such as bone formation, insulin resistance and metastasis of cancer cells. The inhibitors of phosphodiesterase are used in the treatment of some forms of arthritis. Only a few inhibitors have been reported so far, majority of them being of synthetic origin (Ahmad *et al.*, 2003) and none from lichen sources. Lobaric acid showed a significant inhibition of phosphodiesterase enzyme with an $IC_{50} \pm SEM$ (μM) of a 313.7 \pm 2.2 when compared with the two standards, EDTA ($IC_{50} = 274.0 \pm 0.1 \ \mu M$) and cysteine ($IC_{50} = 748.0 \pm 0.1 \ \mu M$).

Lobaric acid did not show α -glucosidase inhibitory activity or any significant inhibition against the urease enzyme.

Immunomodulators are substances capable of interacting with the immune system to up-regulate or down-regulate specific aspects of the host response. Due to the broad application of their action, immunomodulators are becoming very popular in the global natural product based health industry (Yeap et al., 2011). Various disease conditions such as infections, organ transplant rejection, cancer, rheumatoid arthritis, and systemic lupus erythematosus are currently treated with immunomodulating agents (Long et al., 2011). Different immunomodulatory agents have been screened from a variety of plants, including the lichen Thamnolia vermicularis var. subuliformis (Omarsdottir et al., 2007). Lobaric acid was screened over a range of concentrations (3.1–50 μg/mL) for its oxidative burst inhibitory potential. It suppressed both the myeloperoxidase dependant and myeloperoxidase independent ROS production with PMNs at the lowest concentration tested (3.1 ug/mL). when compared with standards ibuprofen and sodium diethyldithiocarbamatetrihydrate (Table 1).

CONCLUSION

Although acetyl and butyryl-cholinesterases inhibitory activities of acetylated and diacytylated depsidones have

been reported (Pejin et al., 2012), this is the first report of these activities of lobaric acid. However, lobaric acid did not show any inhibitory potential against α -glucosidase or urease enzymes. In the β -glucuronidase enzyme inhibitory assay, lobaric acid has shown 12 times higher activity than the available standard D-saccharic acid 1,4lactone further indicating the pharmacological potential of lobaric acid as a lead compound. Even though a few heteroglycans isolated from the lichen *Thamnolia* vermicularis var. subuliformis (Omarsdottir et al., 2007) have shown potential immunomodulatory activity, this is the first report of the promising immunomodulatory activity of a lichen depsidone. The results presented above make lobaric acid with highly diverse functional groups, such as three carbonyls in the form of keto, ester and acid, phenolic OH and an ether group, an excellent candidate for future studies with structural optimization.

REFERENCES

- Ahmad V.U., Abbasi M.A., Hussain H., Akhtan M.N., Farooq U., Fatima N. & Choudhary M.I. (2003). Phenolic glycosides from *Symplocosracemosa*: natural inhibitors of phosphodiesterase I. *Phytochemistry* 63: 217 – 220.
- Burlando B., Ranzato E., Volante A., Appendino G., Pollastro F. & Verotta L. (2009). Antiproliferative effects on tumour cells and promotion of keratinocyte wound healing by different lichen compounds. *Planta Medica* 75: 607 – 613.
 - DOI: http://dx.doi.org/10.1055/s-0029-1185329
- Ellman G.L., Courtney K.D., Andres V. & Featherstone R.M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology* 7: 88 – 95.
- Gissurarson S.R., Sigurdsson S.B., Wagner H. & Ingolfsdottir K. (1997). Effect of lobaric acid on cysteinylleukotriene formation and contractile activity of guinea pig Taenia coli. The Journal of Pharmacology and Experimental Therapeutics 280(2): 770 – 773.
- Gonzilez G.A., Rodriguez-Perez E.M., Conseulelo E., Padron H. & Barrera J.B. (1992). Chemical constituents of the lichen Sterocaulon azoreum. Zeitschrift fur Naturforschung C – A Journal of Biosciences 47c: 503 – 507.

196 Vinitha M. Thadhani et al.

 Helfand S.L., Werkmeister J. & Roder J.C. (1982). Chemiluminescence response of human natural killer cells: the relationship between target cell binding, chemiluminessence and cytolysis. *Journal of Experimental Medicine* 156: 492 – 505.

- DOI: http://dx.doi.org/10.1084/jem.156.2.492
- Karunaratne V., Bombuwela K., Kathirgamanathar S., Kumar V., Karunaratne D.N., Ranawana K.B., Wijesundara D.S.A., Weerasooriya A. & De Silva E.D. (2002). An association between the butterfly, *Talicadanyseus* and the lichen as evidenced from chemical studies. *Current Science* 83(6): 741 – 745.
- Karunaratne V., Bombuwela K., Kathirgamanathar S. & Thadhani V.M. (2005). Lichens: a chemically important biota. *Journal of the National Science Foundation of* Sri Lanka 33(3): 169 – 186.
- Karunaratne V., Kathirgamanathar S., Wijesekera A., Wijesundara D.S.A. & Wolseley P. (2008). Insights into the unique butterfly-lichen association between *Talicada* nyseus nyseus and *Leproloma sipmanianum*. *Journal of* Plant Interactions 3: 25 – 30.
 DOI: http://dx.doi.org/10.1080/17429140701740061
- Kathirgamanathar S., Ratnasooriya W.D., Baekstrom P., Andersen R.J. & Karunaratne V. (2006). Chemistry and bioactivity of Physciaceae lichens: *Pyxine consocians* and *Heterodermia leucomelos*. *Pharmaceutical Biology* 44: 217 – 220.
 - DOI: http://dx.doi.org/10.1080/13880200600686624
- Khan K.M., Shujaat S., Rahat S., Hayat S., Atta-ur-Rahaman & Choudhary M.I. (2002). β-N-Cyanoethyl acyl hydrazide derivatives: a new class of β-glucuronidase inhibitors. *Chemistry and Pharmaceutical Bulletin* 50: 1443 1446. DOI: http://dx.doi.org/10.1248/cpb.50.1443
- Long M.D., Kappelman M.D. & Pipkin C.A. (2011).
 Nonmelanoma skin cancer in inflammatory bowel disease: a review. *Inflammatory Bowel Disease* 17(6): 1423 – 1427.
- Majmudar C.Y. et al. (15 authors) (2012). Sekikaic acid and lobaric acid target a dynamic interface of the coactivator CBP/p300. Angewandte Chemie International Edition 51(45): 11258 – 11262.
- Mitrović T., Stamenković S., Cvetković V., Nikolić M., Tošić S. & Stojičić D. (2011). Lichens as source of versatile bioactive compounds. *Biologica Nyssana* 2(1): 1 – 6.
- Morita H., Tsuchiya T., Kishibe K., Noya S., Shiro M. & Hirasawa Y. (2009). Antimitotic activity of lobaric acid and a new benzofuran, sakisacaulon A from Stereocaulon sasakii. Bioorganic and Medicinal Chemistry Letters 19 (13): 3679 – 3681.

- Oki Y., Okubo M., Tanaka S., Nakanish K. & Murase T. (2000). Diabetes mellitus secondary to glycogen storage disease type III. *Diabetic Medicine* 17: 810 812.
 DOI: http://dx.doi.org/10.1046/j.1464-5491.2000.00378.x
- 17. Omarsdottir S., Freysdottir J. & Olafsdottir E.S. (2007). Immunomodulating polysaccharides from the lichen *Thamnolia vermicularis* var. *subuliformis. Phytomedicine* **14**(2-3): 179 184.
 - DOI: http://dx.doi.org/10.1016/j.phymed.2006.11.012
- Pejin B., Tommonaro G., Iodice C., Tesevic V. & Vajs V. (2012). Acetylcholinesterase inhibition activity of acetylated depsidones from *Lobaria pulmonaria*. *Natural Products Research* 26(17): 1634 – 1637.
 - DOI: http://dx.doi.org/10.1080/14786419.2011.585989
- Stojanovic G., Stojanovic I. & Smelcerovic A. (2012).
 Lichen depsidones as potential novel pharmacologically active compounds. *Mini-Reviews in Organic Chemistry* 9(2): 178 184.
 - DOI: http://dx.doi.org/10.2174/157019312800604689
- Thadhani V.M., Choudhary M.I., Sajjad A., Iman O., Siddique H. & Karunaratne V. (2011). Antioxidant activity of some lichen metabolites. *Natural Products Research* 25(19): 1827 – 1837.
 - DOI: http://dx.doi.org/10.1080/14786419.2010.529546
- Thadhani V.M., Choudhary M.I., Khan S. & Karunaratne V. (2012). Antimicrobial and toxicological activities of some depsides and depsidones. *Journal of the National Science Foundation Sri Lanka* 40(1): 43 48.
 - DOI: http://dx.doi.org/10.4038/jnsfsr.v40i1.4167
- 22. Thune P.O. & Solberg Y.J. (1980). Photosensitivity and allergy to aromatic lichen acids, compositae oleoresins and other plant substances. *Contact Dermatitis* **6**(1):64 71.
- Weatherburn M.W. (1967). Phenol-hypochlorite reaction for determination of ammonia. *Analytical Chemistry* 39: 971 – 974.
 - DOI: http://dx.doi.org/10.1021/ac60252a045
- Williams D.E., Lauren F.L., Whitney J., Singh T.M., McDonald L., Kathirgamanathar S., Karunaratne V. & Andersen R.J. (2011). Depsidones isolated from the Sri Lankan lichen *Parmotrema* sp. exhibit selective PLK1 inhibitory activity. *Pharmaceutical Biology* 49(3): 296 – 301.
 - DOI: http://dx.doi.org/10.3109/13880209.2010.517540
- Yeap S.K, Rahman M.B.A., Alitheen N.B., Young Ho W., Omar A.R., Beh B.K. & Huynh Ky. (2011). Evaluation of immunomodulatory effect: selection of the correct targets for immuno stimulation study. *American Journal of Immunology* 7(2): 17 – 23.
 - DOI: http://dx.doi.org/10.3844/ajisp.2011.17.23