



SHORT REPORT

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Brain-derived neurotrophic factor receptor TrkB exists as a preformed dimer in living cells

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Abstract

Background: Neurotrophins (NTs) and their receptors play crucial roles in the development, functions and maintenance of nervous systems. It is widely believed that NT-induced dimerization of the receptors initiates the transmembrane signaling. However, it is still controversial whether the receptor molecule has a monomeric or dimeric structure on the cell surface before its ligand binding.

Findings: Using chemical cross-linking, bimolecular fluorescence complementation (BiFC) and luciferase fragment complementation (LFC) assays, in this study, we show the brain-derived neurotrophic factor (BDNF) receptor TrkB exists as a homodimer before ligand binding. We have also found by using BiFC and LFC that the dimer forms in the endoplasmic reticulum (ER), and that the receptor lacking its intracellular domain cannot form the dimeric structure.

Conclusions: Most, if not all, of the TrkB receptor has a preformed, yet inactive, homodimeric structure before BDNF binding. The intracellular domain of TrkB plays a crucial role in the spontaneous dimerization of the newly synthesized receptors, which occurs in ER. These findings provide new insight into an understanding of a molecular mechanism underlying transmembrane signaling mediated by NT receptors.

Keywords: BDNF, Chemical crosslinking, Protein fragment complementation assay, Neurotrophin receptor, Pre-formed homodimer

Findings

The neurotrophin (NT) receptor family consists of the tropomyosin-related kinase receptors (Trk) A, B and C, and p75^{TNR}, a member of the tumor necrosis factor (TNF) receptor superfamily. TrkA preferentially interacts with the nerve growth factor (NGF), TrkB with BDNF and NT-4/5, and TrkC with NT-3. NT-3 can also interact with TrkA and TrkB with relatively low affinity, and all the NTs can bind p75^{TNR} with low affinity [1,2]. The Trk receptor kinases play crucial roles in the development and maintenance of the central and peripheral nervous system, and consist of an extracellular ligand-binding domain, a single transmembrane domain and an intracellular tyrosine kinase domain. Upon activation, the receptor kinases initiate downstream signaling cascades mediated by Ras/Raf/MAP kinase, PI3K/Akt and PLC- γ [2-4]. TrkB and its ligand BDNF are highly expressed in

biologically unfavourable neuroblastomas, and TrkB expression is associated with drug resistance and expression of angiogenic factors [5].

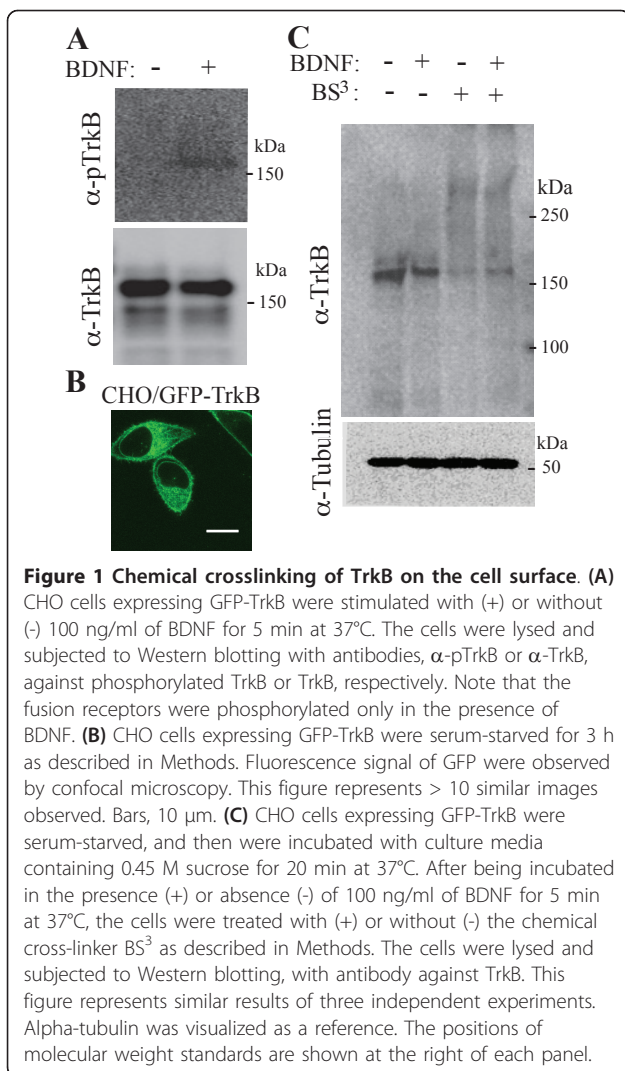
It is widely believed that the Trk receptor kinases are activated through NT-induced receptor dimerization. Because the NT exists in solution as a stable homodimer [6,7], it is thought that the NT dimer acts as a bridge to induce the dimerization of two Trk monomers. However, we have recently found that TrkA exists as a preformed, yet inactive, dimer in living cells [8]. In the present study, therefore, we also examined if TrkB exists as a preformed dimer in living cells before BDNF binding.

Preformed TrkB homodimers detected by chemical crosslinking

We first established a Chinese hamster ovary (CHO) cell line stably expressing green fluorescent protein (GFP)-fused TrkB at the receptor's N-terminus. The GFP-TrkB fusion protein was inactive in the absence of BDNF, and could be activated by BDNF added to the culture media,

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as shown by Western blotting using antibody against phosphorylated TrkB (Figure 1A). The cells were serum-starved for 2 h at 37°C, followed by an-hour incubation on ice, in order to increase the cell-surface expression of the receptor chimera (Figure 1B). Furthermore, sucrose, 0.45 M at a final concentration, was added to the culture media in order to inhibit the clathrin-dependent endocytosis of GFP-TrkB upon BDNF binding [9,10]. Then, the receptor molecules on the cell surface were chemically cross-linked with BS³, a membrane-impermeable cross-linker, in the presence or absence of BDNF. After the cross-linking, the cell lysates were subjected to Western blotting, and immunostained with anti-TrkB antibody. In the absence of the cross-linker, only the lower molecular mass band, ~170 kDa, was detected, while in the presence of the cross-linker higher molecular mass band, ~340 kDa, was also recognized by anti-TrkB antibody in addition to the lower band. Based on the apparent molecular weights, we assigned the

lower and higher bands as the monomer and dimer of TrkB, respectively. The band intensity of the TrkB dimer cross-linked was similar in the absence and presence of BDNF (Figure 1C). These results suggest either that a fraction of TrkB exists as a preformed dimer, or that only a fraction of the receptor dimers was expressed on the cell surface and was chemically cross-linked. Furthermore, the results also indicate that the dimer fraction was not increased after being incubated with BDNF.

Preformed TrkB homodimers detected by BiFC

To directly observe spontaneous dimerization and subcellular localization of TrkB in living cells, we employed BiFC [11,12]. The N-terminal and C-terminal fragments of the Venus fluorescent protein [13], a derivative of yellow fluorescent protein, were fused to the C-terminus of TrkB, separately, to make TrkB-VN and TrkB-VC, respectively. When these constructs were expressed together in the human embryonic kidney HEK 293 cells, Venus fluorescence was clearly observed, whereas no fluorescence was observed in HEK 293 cells expressing either TrkB-VN or TrkB-VC, or in cells co-expressing TrkB-VN and the epidermal growth factor receptor (EGFR)-VC as a negative control (Figure 2A). To exclude again the possibility that the TrkB dimers were activated due to the BiFC fusion, we examined the receptor phosphorylation in the absence or presence of BDNF. In the absence of BDNF, only very low level of basic autophosphorylation of the fusion receptors was observed, and BDNF treatment could enhance the phosphorylation of the fused TrkB receptor molecules (Figure 2B). Together with the results shown in Figure 1A, these results confirm that the BiFC fusion receptors behave like the wild-type receptor, and suggest again that TrkB exists as an inactive dimer, which can be activated by BDNF binding.

The Venus fluorescence was observed not only on the cell surface but also inside the cells (Figure 2A, B). To examine where the Venus reconstitution occurs during the receptor synthesis, we treated the cells co-expressing TrkB-VN and TrkB-VC with Brefeldin A (BFA), a lactone antibiotic that disassembles the Golgi apparatus and blocks anterograde transport of membrane proteins from ER to Golgi [14]. In the absence of BFA, Venus fluorescence was clearly observed both on the cell surface and inside the cells (Figure 2C, upper panels), while in the presence of BFA, Venus fluorescence was observed only inside the cells (Figure 2C, lower panels). The fluorescence inside the cells was mainly co-localized with calnexin, an ER marker [15]. These results indicate that TrkB spontaneously dimerizes in ER before the newly synthesized receptor reaches the cell surface. In contrast, no Venus fluorescence was observed in HEK 293 cells co-expressing TrkBΔICD-VN and TrkBΔICD-VC, both

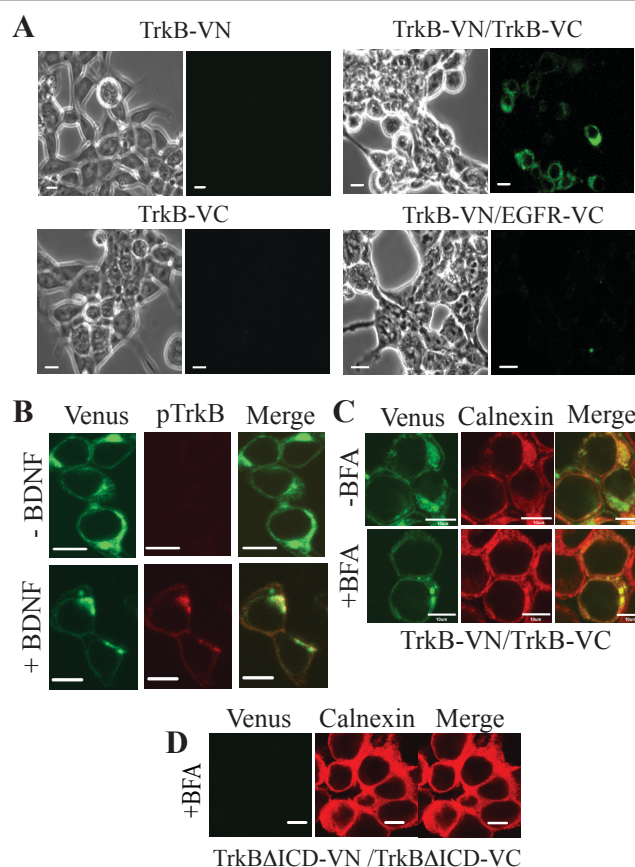


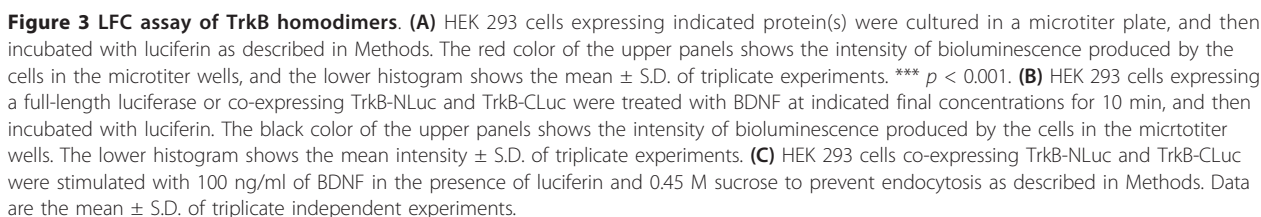
Figure 2 BiFC assay of TrkB homodimers. (A) HEK 293 cells were transfected with TrkB-VC alone, TrkB-VN alone, TrkB-VN and TrkB-VC, or TrkB-VN and EGFR-VC. Twenty-four hours after the transfection, Venus fluorescence signals were observed by confocal microscopy. This figure represents > 20 similar images observed. (B) HEK 293 cells co-expressing TrkB-VN and TrkB-VC were treated with (+) or without (-) 100 ng/ml of BDNF. The cells were fixed, and immunostained with anti-pTrkB primary antibody and then with fluorescently labeled secondary antibody. Fluorescence signals of Venus (green) and pTrkB (red) were observed by confocal microscopy. Note that the green and red fluorescence images do not completely overlap since TrkB-VN and TrkB-VC can form non-fluorescent homodimers. This figure represents > 20 similar images derived from four independent experiments. Bars, 10 μm. (C, D) HEK 293 cells co-expressing TrkB-VN and TrkB-VC (C), or the fusion receptor pair lacking the intracellular domain, TrkBΔICD-VN and TrkBΔICD-VC (D) were treated with (+) or without (-) 10 μg/ml BFA. The cells were fixed, and immunostained with anti-calnexin antibody and then with fluorescently labeled secondary antibody. These figures represent 15 similar images derived from three independent experiments. Bars, 10 μm.

of which lack the intracellular domain (ICD) of TrkB, in the presence of BFA (Figure 2D). This confirms that BFA treatment itself does not induce the Venus reconstitution, and suggests that the intracellular domain plays a crucial role in the spontaneous dimerization of TrkB.

Preformed TrkB homodimers detected by LFC

Since the interaction of the BiFC Venus fragments is irreversible, we cannot determine the ratio of the TrkB dimer and monomer at equilibrium. Therefore, we employed firefly LFC because interaction of the luciferase fragments is reversible [16]. We fused the N-terminal or C-terminal fragment of firefly luciferase to the C-terminus of TrkB, resulting in TrkB-NLuc and TrkB-CLuc, respectively. When HEK 293 cells co-expressing TrkB-NLuc and TrkB-CLuc were incubated with

luciferin in the absence of BDNF, bioluminescence was clearly detected although the bioluminescence level was lower than that of cells expressing pGL3, which encodes a full-length firefly luciferase (Figure 3A). In contrast, bioluminescence was not observed in cells co-expressing TrkBΔICD-NLuc and TrkBΔICD-CLuc, both of which lacked the intracellular domain of TrkB, in cells co-expressing TrkBΔICD-NLuc and TrkB-CLuc, or in cells co-expressing TrkB-NLuc and TrkBΔICD-CLuc. These results demonstrate that TrkB has a preformed homodimeric structure, and its intracellular domain plays a crucial role in the spontaneous dimerization as shown by the BiFC experiment above (Figure 2D). When the cells co-expressing TrkB-NLuc and TrkB-CLuc were treated with 50 or 100 ng/ml BDNF at a final concentration for 10 min at 37°C, the bioluminescence intensity produced



In the present study, we have shown that TrkB exists as a homodimer in living cells before ligand binding, by using three different approaches, chemical cross-linking, BiFC and LFC. The results demonstrate that most, if not all, of the TrkB receptor molecules on the cell surface have homodimeric structures since the addition of

An increasing number of studies demonstrate that many transmembrane receptors, which include receptors previously thought to be activated by ligand-induced

receptor dimerization, exist as preformed, yet inactive, homo- and heterodimers in living cells. These receptors are the aspartate receptor Tar [17], epidermal growth factor receptor (EGFR) family [12,18-21], erythropoietin receptor [22], growth hormone receptor (GHR) [23,24], Toll-like receptor-9 [25], natriuretic peptide receptor A (NPRA) [26], and nerve growth factor receptor TrkB [8]. In the cases of EGFR, GHR, NPRA and Tar, it has been proposed that ligand binding is likely to induce the rotation/twist of the transmembrane domains of the preformed receptor dimers, resulting in rearrangement of the cytoplasmic domains for activation [18,24,26,27]. Therefore, further study of structures of the Trk receptor family will shed light on molecular mechanisms underlying the receptor activation.

Methods

Plasmid construction

BiFC vectors, pBiFC-VN173 and pBiFC-VC155, were described previously [28], and FLAG tags encoded by the vectors were removed as previously described [8]. PCR products that encode a full-length human TrkB protein were amplified using pLNCX-hTrkB [29], a generous gift from Dr. Garrett M. Brodeur (Children's Hospital of Philadelphia, PA), as a template and the following primer pairs: 5'-CCGgaattcACCATGTCTCGTCCTGGATAAGGTG (TrkB-VN173-F) and 5'-GCtctagaGCCTAGAATGTCCAGGTAGAC (TrkB-VN173-R), or 5'-CCGgaattcACCATGTCTCGTCCTGGATAAGGTG (TrkB-VC155-F) and 5'-CCGctcgagAGCCTAGAATGTCCAGGTAGAC (TrkB-VC155-R). The PCR products were inserted between *Eco* RI and *Xba* I sites of pBiFC-VN173, or between *Eco* RI and *Xho* I of pBiFC-VC155, resulting in pTrkB-VN or pTrkB-VC, respectively. Then, a synthetic double-stranded oligonucleotide, 5'-GGTGGTGGTGGT TCTGGTGGTGGTGGTCTGGTGGTGGTGGTCT encoding amino-acid residues GGGSGGGSGGGSGG termed $(G_4S)_3$, were inserted into the unique *Xba* I or *Xho* I site of pTrkB-VN or pTrkB-VC, respectively. Plasmids for firefly (*Photinus pyralis*) LFC were described previously [8]. A DNA fragment encoding TrkB was cut out with *Eco* RI and *Xba* I enzymes from pTrkB-VN, and was then inserted into pBiFC-NLuc and pBiFC-CLuc, resulting in pTrkB-NLuc and pTrkB-CLuc, respectively. The construction of pGFP-TrkB was based on pEGFR-eGFP [30], and a DNA sequence encoding enhanced GFP, which was amplified by PCR using the primers 5'-CCGagatctATGGTGAGCAAGGGCGAGGAGCTGTTCACC (EGFP-F-BglII) and 5'-CCGgaattcCTTGTACAGCTCGTCCATGCCGAGAGTGATCC (EGFP-B-EcoRI), and was then inserted between *Bgl* II and *Eco* RI of pAEMXT-ACPwt-GPI (Covalys Bio-sciences, Witterswil, Switzerland), resulting in pAEMXT/GFP. A DNA

sequence encoding human TrkB without its signal peptide (aa 1-31), but with $(G_4S)_3$, was inserted between *Eco* RI and *Xho* I of pAEMXT/GFP, resulting in pGFP-TrkB. To construct various plasmids encoding TrkB lacking its intracellular domain, a DNA fragment encoding the full-length TrkB of the expression plasmid constructs was replaced with a fragment encoding amino-acid residues 1-465 of TrkB. Validity of all the constructs was confirmed by DNA sequencing and Western blotting.

Cell culture

A HEK 293 cell line was purchased from RIKEN cell bank (Cat. No.: RCB 1637; Ibaraki, Japan), and was maintained in Dulbecco's modified Eagle's medium (DMEM; Nacalai Tesque, Kyoto, Japan) supplemented with 10% fetal bovine serum (FBS; Invitrogen). A CHO-K1 cell line purchased from the American Type Culture Collection (ATCC; Manassas, VA) was maintained in Iscove's modified Dulbecco's medium (IMDM; Invitrogen). All media were supplemented with 100 U/ml of penicillin-streptomycin (Nacalai Tesque). A CHO-K1 cell line stably expressing the fusion GFP-TrkB was established by transfecting CHO-K1 cells with the pGFP-TrkB plasmid, followed by selection with 1.0 mg/ml of G418 (Nacalai Tesque). The established cells were treated with (+) or without (-) 10 μ g/ml BFA for 12 h at 37°C.

Chemical crosslinking

CHO cells expressing the fusion construct GFP-TrkB were cultured to a concentration of $\sim 10^7$ cells/dish (10 cm in diameter; Iwaki Asahi Techno Glass, Chiba, Japan) before chemical crosslinking. The cells were serum-starved for 2 h at 37°C, incubated on ice for another 1.0 h, and then incubated in IMDM containing 0.45 M sucrose for 20 min at 37°C. In the presence of 0.45 M sucrose, the cells were treated with or without 100 ng/ml of BDNF (Alomone Labs, Jerusalem, Israel) at a final concentration for 5 min at 37°C. Then, the cells were treated with or without 1.0 mM BS³ (Pierce Biotechnology, Rochford, IL) at a final concentration in Dulbecco's phosphate-buffered saline (DPBS; Invitrogen), which contained 0.45 M sucrose, for 20 min at 37°C. This crosslinking reaction was quenched by adding 40 mM Tris-HCl, pH 8.0, and 0.45 M sucrose at final concentrations, followed by incubation for 15 min at room temperature. After being washed twice with ice-cold DPBS, the cells were lysed with NP-40 lysis buffer (50 mM Tris-HCl, pH 7.4, 250 mM NaCl, 5 mM EDTA, 50 mM NaF, 1.0 mM Na₃VO₄, 1.0% Nonidet P-40) supplemented with a protease inhibitor cocktail (Nacalai Tesque) and a phosphatase inhibitor cocktail (Nacalai Tesque) for 30 min at 4°C. Resulting cell lysates were sonicated twice for 5 s each, and centrifuged at 10,000 rpm for 10 min at 4°C. The supernatants were collected for Western blotting.

Western blotting

Cell lysates, 10 µg each as the amount of total protein per lane, were loaded to NuPAGE 4–12% Bis-Tris gels (Invitrogen), followed by electrophoresis using XCell SureLock Mini-Cell (Invitrogen) and transferred onto Hybond-P PVDF membrane (Amersham Biosciences, Little Chalfont, UK) using a mini Trans-Blot electrophoretic transfer cell (Bio-Rad). The membrane was blocked overnight at 4°C in TBST (10 mM Tris-HCl, pH7.6, 0.9% NaCl, 0.1% Tween 20) supplemented with 5% skimmed milk (Wako Pure Chemical, Osaka, Japan) or in TBST supplemented with 2% ECL advance blocking agent (GE Healthcare, Little Chalfont, UK), and was then incubated with the following primary antibody in blocking buffer for 2 h at room temperature: anti-TrkB monoclonal antibody (mAb) (1:500 dilution; Novus Biologicals, Littleton, CO), anti-phospho-TrkA (Tyr674/675)/TrkB (Tyr706/707) (C50F3) rabbit mAb (1:1000 dilution; Cell Signaling Technology, Tokyo, Japan), or mouse mAb against α -tubulin (1:2000 dilution; Sigma-Aldrich Japan, Tokyo). The membrane was washed three times with TBST, and was then incubated with sheep anti-mouse or donkey anti-rabbit secondary antibody conjugated with horseradish peroxidase (1:10000 dilution; GE Healthcare) in the same blocking mixture for 90 min at room temperature. After being washed four times with TBST, the membrane was reacted with an ECL advance Western blotting detection kit (GE Healthcare). Protein bands were visualized using a lumino-image analyzer (LAS3000; Fujifilm, Tokyo, Japan).

Immunofluorescent staining

Immunofluorescent staining was performed as previously described [12]. HEK 293 cells ($\sim 2 \times 10^4$ cells/well) were seeded into an eight-well Lab-Tek II chamber slide (Nalge Nunc International, Naperville, IL), and then transfected with or without expression plasmids of interest. After washing twice with ice-cold $1 \times$ PBS, the cells were fixed with methanol-acetone (1:1) for 10 min at -20°C . After being blocked with a mixture of Blocking One-P (Nakalai Tesque) and 2% ECL Advance Blocking Agent (GE Healthcare) and 50 mM NaF for 1.0 h at room temperature, the cells were incubated with the following primary antibodies diluted in blocking buffer for 1.0 h at room temperature: rabbit mAb against phospho-TrkA (Tyr490)/TrkB (Tyr516) (Cell Signaling Technology) and rabbit anti-calnexin antibody (Sigma). After washed three times with TBST, the cells were incubated with the following secondary antibodies diluted in blocking buffer for 1.0 h at room temperature: Alexa Fluor 633-conjugated goat anti-rabbit IgG (H+L) antibody (Molecular Probes, Eugene, OR). After washed three times with TBST, the cells were mounted in Vectashield (Vector Laboratories, Burlingame, CA) and observed under an Axiovert 200 M inverted

microscope equipped with an LSM 510 META Ver 3.5 scan head (Carl Zeiss, Jena, Germany), using 488 nm (argon) and 633 nm (HeNe) (Titanium:Sapphire from Mai Tai; Spectra-Physics, Mountain View, CA) laser lines for excitation of Venus and Alexa Fluor 633, respectively.

Firefly LFC

HEK 293 cells were plated in a 96-well black polystyrene borosilicate glass microplate (Iwaki) at a concentration of $\sim 8,000$ cells/well 12 h before transfection. Then the cells were transfected with 50 ng of the control vector pGL3 (Promega, Madison, WI), which encoded a full-length luciferase, or with 100 ng each of expression plasmid constructs by using 0.1 µl of Plus Reagent (Invitrogen) and 0.25 µl of Lipofectamin LTX Reagent (Invitrogen) in 20 µl of Opti-MEM I reduced serum medium (Invitrogen). Forty hours after transfection, the cells were serum-starved for 3 h, washed once with DPBS, and then incubated in 100 µl of DPBS, which contained 0.15 mg/ml D-luciferin (Promega) as well as 0 ng/ml, 50 ng/ml or 100 ng/ml BDNF at a final concentration, for 10 min at room temperature. After being incubated in 50 µl DPBS containing 0.45 M sucrose for 10 min at room temperature, alternatively, the transfected cells were incubated with 50 µl DPBS, which contained 0.45 M sucrose and 0.15 mg/ml D-luciferin at final concentrations, in the presence or absence of 100 ng/ml BDNF at a final concentration at 37°C . Then, bioluminescence intensity was measured by using a LAS 3000 or a microplate luminometer, LB 960 Centro (Berthold Japan, Osaka). Data are presented as the mean \pm S.D. of triplicate measurements. Statistical significance was assessed using Student's *t*-test. The *P* values less than 0.01 were assessed to be statistically significant.

List of abbreviations

BDNF: Brain-derived neurotrophic factor; BFA: Brefeldin A; BiFC: bimolecular fluorescence complementation; DPBS: Dulbecco's phosphate-buffered saline; EGFR: epidermal growth factor receptor; ER: endoplasmic reticulum; GFP: green fluorescent protein; ICD: intracellular domain; LFC: luciferase fragment complementation; mAb: monoclonal antibody; NT: neurotrophin; Trk: tropomyosin-related kinase.

Acknowledgements

We thank Garrett M. Brodeur for providing us with pLNCH/hTrkB, David Piwnica-Worms for FRB-NLuc and CLUC-FKBP. We also thank the laboratory members for their critical readings of the manuscript.

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Authors' contributions

The project was initiated by J.S. and I.N.M. All the experiments were designed by J.S. and I.N.M., and were carried out by J.S. The manuscript was written by J.S. and I.N.M. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 8 November 2011 Accepted: 24 January 2012

Published: 24 January 2012

References

- Chao MV: Neurotrophins and their receptors: a convergence point for many signalling pathways. *Nat Rev Neurosci* 2003, **4**(4):299-309.
- Huang EJ, Reichardt LF: Trk receptors: roles in neuronal signal transduction. *Annu Rev Biochem* 2003, **72**:609-642.
- Kaplan DR, Miller FD: Neurotrophin signal transduction in the nervous system. *Curr Opin Neurobiol* 2000, **10**(3):381-391.
- Simi A, Ibáñez CF: Assembly and activation of neurotrophic factor receptor complexes. *Dev Neurobiol* 2010, **70**(5):323-331.
- Brodeur GM, Minturn JE, Ho R, Simpson AM, Iyer R, Varela CR, Light JE, Kolla V, Evans AE: Trk receptor expression and inhibition in neuroblastomas. *Clin Cancer Res* 2009, **15**(10):3244-3250.
- McDonald NQ, Lapatto R, Murray-Rust J, Gunning J, Wlodawer A, Blundell TL: New protein fold revealed by a 2.3-Å resolution crystal structure of nerve growth factor. *Nature* 1991, **354**(6352):411-414.
- Radziejewski C, Robinson RC, DiStefano PS, Taylor JW: Dimeric structure and conformational stability of brain-derived neurotrophic factor and neurotrophin-3. *Biochemistry* 1992, **31**(18):4431-4436.
- Shen J, Maruyama IN: Nerve growth factor receptor TrkA exists as a preformed, yet inactive, dimer in living cells. *FEBS Lett* 2011, **585**(2):295-299.
- Daukas G, Zigmond SH: Inhibition of receptor-mediated but not fluid-phase endocytosis in polymorphonuclear leukocytes. *J Cell Biol* 1985, **101**(5 Pt 1):1673-1679.
- Heuser JA, Anderson RG: Hypertonic media inhibit receptor-mediated endocytosis by blocking clathrin-coated pit formation. *J Cell Biol* 1989, **108**(2):389-400.
- Hu CD, Chinenov Y, Kerppola TK: Visualization of interactions among bZIP and Rel family proteins in living cells using bimolecular fluorescence complementation. *Mol Cell* 2002, **9**(4):789-798.
- Tao RH, Maruyama IN: All EGF(ErbB) receptors have preformed homo- and heterodimeric structures in living cells. *J Cell Sci* 2008, **121**(Pt 19):3207-3217.
- Nagai T, Ibatani K, Park ES, Kubota M, Mikoshiba K, Miyawaki A: A variant of yellow fluorescent protein with fast and efficient maturation for cell-biological applications. *Nat Biotechnol* 2002, **20**(1):87-90.
- Fujiwara T, Oda K, Yokota S, Takatsuki A, Ikehara Y: Brefeldin A causes disassembly of the Golgi complex and accumulation of secretory proteins in the endoplasmic reticulum. *J Biol Chem* 1988, **263**(34):18545-18552.
- Wada I, Rindress D, Cameron PH, Ou WJ, Doherty JJ, Louvard D, Bell AW, Dignard D, Thomas DY, Bergeron JJ: SSR alpha and associated calnexin are major calcium binding proteins of the endoplasmic reticulum membrane. *J Biol Chem* 1991, **266**(29):19599-19610.
- Luker KE, Smith MC, Luker GD, Gammon ST, Pivnicka-Worms H, Pivnicka-Worms D: Kinetics of regulated protein-protein interactions revealed with firefly luciferase complementation imaging in cells and living animals. *Proc Natl Acad Sci USA* 2004, **101**(33):12288-12293.
- Milligan DL, Koshland DE Jr: Site-directed cross-linking. Establishing the dimeric structure of the aspartate receptor of bacterial chemotaxis. *J Biol Chem* 1988, **263**(13):6268-6275.
- Moriki T, Maruyama H, Maruyama IN: Activation of preformed EGF receptor dimers by ligand-induced rotation of the transmembrane domain. *J Mol Biol* 2001, **311**(5):1011-1026.
- Yu X, Sharma KD, Takahashi T, Iwamoto R, Mekada E: Ligand-independent dimer formation of epidermal growth factor receptor (EGFR) is a step separable from ligand-induced EGFR signaling. *Mol Biol Cell* 2002, **13**(7):2547-2557.
- Clayton AH, Walker F, Orchard SG, Henderson C, Fuchs D, Rothacker J, Nice EC, Burgess AW: Ligand-induced dimer-tetramer transition during the activation of the cell surface epidermal growth factor receptor-A multidimensional microscopy analysis. *J Biol Chem* 2005, **280**(34):30392-30399.
- Liu P, Sudhaharan T, Koh RML, Hwang LC, Ahmed S, Maryama IN, Wohland T: Investigation of the dimerization of proteins from the epidermal growth factor receptor family by single wavelength fluorescence cross-correlation spectroscopy. *Biophys J* 2007, **93**(2):684-698.
- Constantinescu SN, Keren T, Socolovsky M, Nam H, Henis YI, Lodish HF: Ligand-independent oligomerization of cell-surface erythropoietin receptor is mediated by the transmembrane domain. *Proc Natl Acad Sci USA* 2001, **98**(8):4379-4384.
- Gent J, van Kerkhof P, Roza M, Bu G, Strous GJ: Ligand-independent growth hormone receptor dimerization occurs in the endoplasmic reticulum and is required for ubiquitin system-dependent endocytosis. *Proc Natl Acad Sci USA* 2002, **99**(15):9858-9863.
- Brown RJ, Adams JJ, Pelekanos RA, Wan Y, McKinstry WJ, Palethorpe K, Seeber RM, Monks TA, Eidne KA, Parker MW, Waters MJ: Model for growth hormone receptor activation based on subunit rotation within a receptor dimer. *Nat Struct Mol Biol* 2005, **12**(9):814-821.
- Latz E, Verma A, Visintin A, Gong M, Sirois CM, Klein DC, Monks BG, McKnight CJ, Lamphier MS, Duprex WP, Espevik T, Golenbock DT: Ligand-induced conformational changes allosterically activate Toll-like receptor 9. *Nat Immunol* 2007, **8**(7):772-779.
- Parat M, Blanchet J, De Léan A: Role of juxtamembrane and transmembrane domains in the mechanism of natriuretic peptide receptor A activation. *Biochemistry* 2010, **49**(22):4601-4610.
- Maruyama IN, Mikawa YG, Maruyama HI: A model for transmembrane signalling by the aspartate receptor based on random-cassette mutagenesis and site-directed disulfide cross-linking. *J Mol Biol* 1995, **253**(4):530-546.
- Shyu YJ, Liu H, Deng X, Hu CD: Identification of new fluorescent protein fragments for bimolecular fluorescence complementation analysis under physiological conditions. *BioTechniques* 2006, **40**(1):61-66.
- Nakagawara A, Liu X-G, Ikegaki N, White PS, Yamashiro DJ, Nycum LM, Biegel JA, Brodeur GM: Cloning and chromosomal localization of the human Trk-B tyrosine kinase receptor gene (NTRK2). *Genomics* 1995, **25**(2):538-546.
- Miyagi H, Maruyama IN: Analysis of ligand-receptor interaction on the surface of living cells by fluorescence correlation spectroscopy. *Open Spectrosc J* 2010, **4**:28-31.

doi:10.1186/1750-2187-7-2

Cite this article as: Shen and Maruyama: Brain-derived neurotrophic factor receptor TrkB exists as a preformed dimer in living cells. *Journal of Molecular Signaling* 2012 **7**:2.

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