

**MECHANISM, PREVENTION & MANAGEMENT OF POSTERIOR CAPSULAR  
OPACIFICATION- A REVIEW ARTICLE**

<sup>1</sup>Bhavna G., <sup>2</sup>\*Manjunath Natarajan and Dr. Jayashree Dora<sup>3</sup>

<sup>1</sup>Department of Ophthalmology, VIMSAR, Burla, Odisha.

<sup>2</sup>Junior Consultant, Vijaya Nethralaya, Bangalore.

<sup>3</sup>Professor & HOD, The Department of Ophthalmology, VIMSAR, Burla, Odisha.

\*Corresponding Author: Dr. Manjunath Natarajan

Junior Consultant, Vijaya Nethralaya, Bangalore.

Article Received on 04/10/2016

Article Revised on 24/10/2016

Article Accepted on 14/11/2016

**ABSTRACT**

Expectation of patients receiving modern day cataract surgery has become similar to refractive surgery. Over the last few decades, PCO has been the most common visually disabling sequel of cataract surgery and has important medical, social and economic implications. Posterior capsule opacification (PCO) refers to the opacity that develops in the posterior capsule after cataract surgery. In recent years, with better understanding of the mechanism of PCO formation, advancement in technique of surgery, introduction of 360 degrees sharp optic edge IOLs, recognition of the importance of thorough cortical clean-up, there has been a reduction in the incidence of PCO (to < 10%). However, PCO has not yet been eradicated as it has not been possible to totally get rid of regenerative cells in the equatorial lens bow at cellular level, by any interventional method currently known. The present article reviews literature related to the mechanism, prevention and management of PCO, highlighting current concepts and developments in last few years and future endeavours to manage, prevent and eradicate PCO.

**KEYWORDS:** posterior capsule, Soemmering's ring, Elschnig's pearls, lens epithelial cells, Nd:YAG.

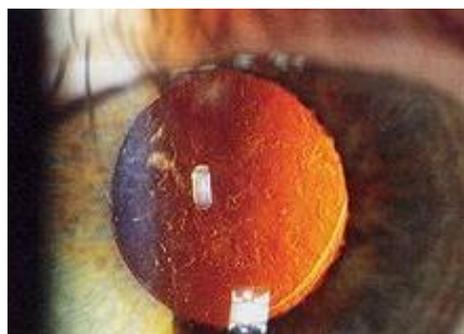
Posterior capsular opacity (PCO) is the most common late onset complication after cataract surgery.<sup>[1]</sup> After cataract & secondary cataract are synonyms for PCO. Incidence of PCO ranges from 5 – 50% in eyes after cataract surgery for senile cataract.<sup>[2,3]</sup> Ignjatovic<sup>[4]</sup> reported a higher incidence in myopes while Vasavada et al<sup>5</sup> reported no significant difference between myopic & normal population. Hayashi et al<sup>[6]</sup> reported higher incidence in diabetic cataract as compared to non-diabetic cataract. Incidence of PCO is decreased over past few decades with advancement in surgical techniques, IOL material & design. Visual axis opacification (VAO) due to PCO is common after cataract surgery & IOL implantation in children & reported to occur in upto 40% of patients even with primary posterior capsulotomy.<sup>[7]</sup> Incidence of VAO has been reported to be much higher in patients without primary posterior capsulotomy. Low recurrence was found in patients with Nd:YAG laser capsulotomy.

**MECHANISM**

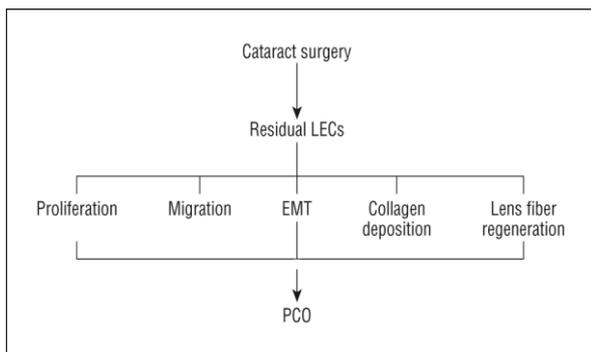
PCO is of 2 types as described by Apple et al.,<sup>[10]</sup> fibrous (Figure 1) & proliferative/pearl (Figure 2). PCO develops mainly by 3 processes- migration, proliferation & differentiation of lens epithelial cells (LECs).



**Figure 1- Fibrous PCO with posterior capsular wrinkling**



**Figure 2- Proliferative/Pearl PCO**



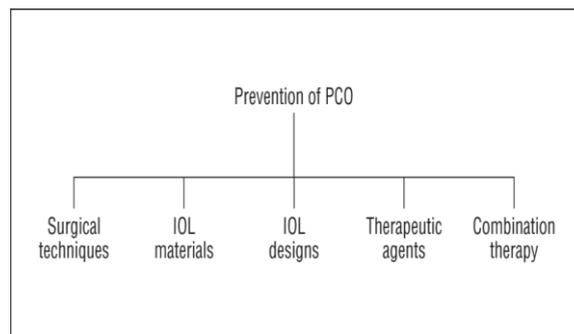
LECs are located on the inner surface of lens capsule at anterior, pre-equatorial & equatorial region. They proliferate at equatorial region throughout life to form lens fibres which are laid down in concentric manner. Fibrosis-type PCO is caused by the proliferation and migration of LECs, which undergo EMT, resulting in fibrous metaplasia and leading to significant visual loss by producing folds and wrinkles in the posterior capsule.<sup>[9]</sup> Pearl type PCO is caused by the LECs located at the equatorial lens region (bladder/Wedl cells)<sup>[8]</sup> causing regeneration of crystallin - expressing lenticular fibers and forming Elschnig pearls and Soemmering ring, responsible for most cases of PCO-related visual loss.<sup>[10,11]</sup> Proliferation occurs most frequently in the first week following surgery. Risk factors that promote proliferation are retained cortical matter, iris pigments & cells from blood due to breakdown of aqueous barrier.<sup>[9]</sup>

Studies show that levels of several cytokines and growth factors increase in aqueous humour influence the behaviour of the remaining LECs after cataract surgery. These factors include transforming growth factor  $\beta$  (TGF- $\beta$ ), fibroblast growth factor 2 (FGF-2), hepatocyte growth factor, interleukins 1 and 6 (IL-1 and IL-6) and epithelial growth factor.<sup>[12]</sup>

Wormstone et al<sup>[13]</sup> & Duncan et al<sup>[14]</sup> have studied LEC growth on human capsular bags in a protein-free medium, which has allowed the autocrine control by individual growth factors to be analyzed. Migration of human LECs plays an important role in the remodeling of the lens capsule<sup>[15,16]</sup> & is associated with matrix metalloproteinase activity in the lens.<sup>[17]</sup> Changes in lens capsule structure during PCO development may include remodeling of the extracellular matrix by matrix metalloproteinases. Abnormal differentiation of LECs forms bladder cells & myofibroblasts and lays down cellular material to form opacification.

## PREVENTION

The modifications which could help in prevention of PCO are related to the surgical technique, IOL & use of therapeutic agents.



### a) Surgical technique related

PCO is predominantly caused by residual LECs in the capsular bag after cataract surgery.<sup>[10,18,19]</sup> Several surgical techniques have been attempted for the removal of these LECs at the time of lens extraction.

1. *Continuous curvilinear capsulorhexis (CCC)*-adequately sized<sup>[20]</sup>, circular CCC<sup>[21]</sup> is associated with lesser incidence of PCO.
2. *Cortical cleaving hydrodissection with rotation*-creates gap between lens capsule & cortical matter & hence helps in complete removal of cortical matter. Hydraulic force generated during hydrodissection with rotation helps remove the LECs. Multi quadrant cortical cleaving hydrodissection helps in early & complete removal of epinucleus & cortical matter.<sup>[22]</sup>
3. *Cortical cleanup*- by bimanual irrigation and aspiration in the presence of PCIOL, helps in complete removal of cortical matter especially in areas like sub-incisional part & deep capsular fornices, without disrupting the posterior capsule.<sup>[23]</sup>
4. *Polishing of anterior capsule*- has a role in decreasing fibrotic type of PCO while is less effective for proliferative type.<sup>[24]</sup>
5. *In the bag fixation of IOL*- reduces incidence of central PCO. IOL provides barrier for migration of LECs.<sup>[22]</sup> Incidence of PCO was found to be more in sulcus fixated IOLs as compared to in the bag fixation.<sup>[25]</sup>
6. *Buttonholing of posterior capsule*- posterior CCC with posterior buttonholing of IOL haptic through it can be done to prevent development of PCO. IOL optic prevents migration of LECs into retrolental space and posterior capsule over anterior edges of optic prevents formation of anterior capsular fibrosis.<sup>[26]</sup>
7. *Anterior capsule overlap of IOL optic*- difference in size of CCC causes variable anterior capsular overlap of IOL optic. The size of this CCC has not got any significant effect on severity of PCO.<sup>[27]</sup> The anterior capsular overlap leads to variable incidence of PCO formation with different IOL materials. It remains an important factor for eyes with PMMA IOLs and Silicone IOLs.<sup>[28]</sup> However, it is not a crucial factor in eyes with Acrylic IOL implantation.<sup>[29]</sup>

**b) IOL related**

- IOL design & material-** IOL optic size, edge, angular ion of haptics and material play an important role in preventing PCO. Meacock WR et al reported less PCO with 6mm optic size as compared to 5.5mm optic.<sup>[30]</sup> Sharp-edge optic IOLs made of acrylic and silicone are superior in lowering the rates of PCO and laser capsulotomy.<sup>[31]</sup> Square edge IOL optic was found to be more effective than round edge IOL optic to exert pressure on posterior capsule and reduce PCO formation.<sup>[32]</sup> Angulated IOL haptics also decrease PCO formation by inducing more pressure on posterior capsule.

Surface modifications of PMMA IOLs by carbon and titanium<sup>[33]</sup>, heparin<sup>[34]</sup> & polytetrafluoroethylene<sup>[35]</sup> and of silicon IOLs by oxygen and carbon dioxide plasma<sup>[36]</sup>, a sulfonate and carboxylate group containing polymer<sup>[37]</sup> have been reported to have higher biocompatibility and effectiveness in prevention of PCO. Recently, IOL surface modification by gas plasma<sup>[38]</sup> & polyethylene glycol<sup>[39]</sup> has been shown to influence LEC behavior and to prevent PCO.

Hydrogel IOLs are associated with maximum occurrence of PCO followed by PMMA & silicone. Acrylic IOL is comparatively associated with lesser amount of PCO formation.<sup>[40]</sup>

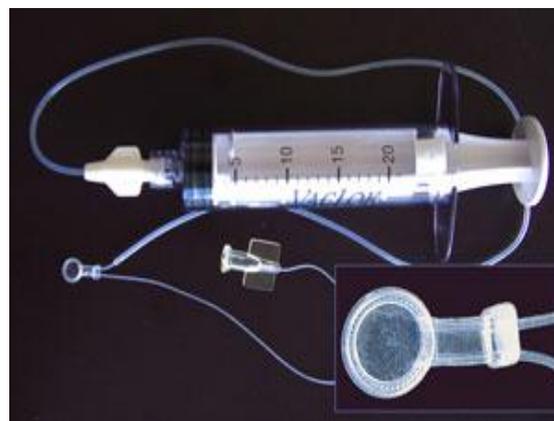
- Single piece versus multiple piece IOL-** there is no difference in PCO development between 3-piece and 1-piece acrylic hydrophobic IOLs.<sup>[41]</sup>

**c) Therapeutic agents**

Many drugs like anti-proliferative, anti-coagulant, anti-inflammatory, anti-adherence & anti-migratory, have been tried to decrease incidence of PCO.

- Anti-proliferative drugs-** 5-fluorouracil, doxorubicin, daunorubicin, mitomycin-c, octreotide & colchicines were tried in vitro but no outcome was significant.<sup>[42]</sup>
- Anti-coagulant drugs-** Irrigation of eyes with heparin solution (25 IU/ml) before implantation of IOL was significantly associated with less PCO as compared to the non-irritant group.<sup>[43]</sup> Heparin-surface-modified PMMA IOLs were associated with less incidence of PCO formation.<sup>[34]</sup>
- Anti-inflammatory drugs-** reduce release of cytokines and prevent proliferation of LECs.<sup>[44]</sup> Topical diclofenac<sup>[45]</sup> drops were tried in post-operative period without any significant results.
- Anti-adherence & anti-migratory compounds-** prevent migration & adherence of LECs to posterior capsule. These agents include ilomastat (a matrix metalloproteinase inhibitor)<sup>[46]</sup>, RGD peptide<sup>[47]</sup>, mibefradil (Ca-channel inhibitor), EDTA and coating an acrylic IOL surface with MPC polymer.

Maloof et al developed the Perfect Capsule device (Milvella Ltd, Sydney, Australia) (Figure 3), which permits cytotoxic agents to be delivered selectively to the capsular bag, thus selectively targeting residual LECs.<sup>[48]</sup>



**Figure 3- PerfectCapsule device, (Milvella)**

**MANAGEMENT**

PCO involving the visual axis can cause blurring of vision, glare and decrease in visual acuity and contrast sensitivity. Management of PCO has undergone a paradigm shift in strategy and technique. It can be managed by invasive & non-invasive methods.

**1. Non-Invasive methods**

Now, Nd: YAG (Neodymium:yttrium-aluminium-garnet) laser capsulotomy<sup>[49,50,51]</sup> has replaced invasive surgery as the most common treatment modality for PCO management. It was first proposed by Aron-Rosa and Frankhauser in 1980s as an effective treatment for PCO.

**CONTRAINDICATIONS OF Nd:YAG LASER CAPSULOTOMY<sup>[49,50]</sup>****I) Absolute**

- Corneal scarring/edema- inadequate visualisation of target aiming beam
- Glass IOL- chances of fracture of glass IOL
- Inability to maintain stability of eye- risk of inadvertent damage to adjacent intraocular structures.

**II) Relative**

- Active intraocular inflammation- gets aggravated by the procedure
- Cystoid macular edema- due to laser induced breakdown of barrier functions of posterior capsule
- High risk of retinal detachment

**TECHNIQUE<sup>[52,53]</sup>**

- Pre-operative Assessment-** complete ophthalmology all history & examination should be carried out before proceeding
  - Direct ophthalmoscopic visualisation of PCO- most reliable method for assessment
  - Slit lamp biomicroscopy & retro illumination
  - Laser interferometry to assess potential vision

- d) Optical coherence tomography & fundus fluorescein angiography in suspected cases of cystoid macular edema.

### II) Preparation of patient

- Informed consent explaining the nature & consequences of the procedure is taken
- Pupillary dilatation is not required unless the pupil is miotic or surgeon is inexperienced. The visual axis is marked with a single laser spot in the centre before dilatation
- Topical anaesthesia is generally not required unless contact lens is used. Peribulbar anaesthesia is used rarely in cases of nystagmus.

### III) Procedure

- Abraham central contact lens / Peyman lens can be used to stabilise the eye, improve laser beam optics & for accurate focussing
- Minimum amount of laser energy is used to create capsular opening (1-2mJ/pulse)
- Stress lines are seen as wrinkles in the posterior capsule & shots are placed at these stress lines for maximum effect per shot
- Laser shots are given at posterior 150um from a datum point to avoid IOL damage
- Types of opening
  - Cruciate opening*- extends from 12 o'clock to 6 o'clock position. Post procedure glare and increase in vitreous floaters can occur
  - Can opener method*- laser capsulotomy is done along the circumference of the optic. It prevents potential damage to IOL in the visual axis, but the cut capsular fragment might obscure the visual axis
  - Inverted-U method*- capsular fragment remains attached to inferior part of opening. But it is associated with the problem of early visual recovery as time is needed for flap to sink in intravitreal space due to gravity and to get contracted
  - Circular pattern with vitreous strand cutting*- includes the conventional procedure along with cutting of vitreous strands attached to capsular fragment by laser
  - Size of opening*- capsulotomy size should be large enough to cover the pupil under mesopic conditions and avoid glare from edges that occurs during night driving. Small opening is better in eyes with dense membrane that gives excellent optics as well as in eyes at risk of retinal detachment.

### IV) Postoperative care

Immediately after the procedure, topical anti-glaucoma agents should be administered in the eye to minimise post procedure IOP spike (which is transient in most cases). In cases of advanced glaucoma, oral hyper osmotic agents can be used during and after the procedure.

Topical antibiotics are instilled after the procedure if contact lens is used. Topical steroids and cycloplegics can be given on an individual basis.

### COMPLICATIONS<sup>[51]</sup>

- Rise in IOP- due. To trabecular meshwork block (by debris, inflammatory cells, liquid vitreous) or pupillary block. Peak elevation occurs within 6 hours of capsulotomy which usually returns to baseline within a week.
- Cystoid macular edema(CME) (0.5-2.5%) – due to shockwave damage to vitreous & release of inflammatory mediators.
- IOL pitting/damage (9.4-33%)- degree of damage depends on nature of IOL material, highest damage occurs to silicone IOLs and lowest to acrylic IOLs. Glass IOL might fracture after Nd:YAG capsulotomy. IOL pitting in visual axis can cause degradation of image quality and glare
- Retinal Detachment(RD) (1.6-1.9% cases over 3 years)- It might be due to increased rate of post procedure posterior vitreous detachment. Increased risk of retinal detachment was found in patients with previous history of retinal detachment, axial length > 24mm & lattice degeneration.
- Iritis
- Malignant glaucoma
- Endophthalmitis
- IOL displacement
- Macular hole

### RESULTS

Improvement in visual acuity was reported in most of the eyes with PCO, after Nd:YAG capsulotomy.

Stager et al. evaluated the effectiveness of Nd: YAG laser capsulotomy for the treatment of PCO in children with acrylic IOLs. A total of 51 eyes (70%) maintained a clear visual axis after a single Nd: YAG procedure, 10 eyes (84% cumulative) after two procedures and another 3 eyes (88% cumulative) after three procedures (follow-up period range: 3–92 months; median: 25 months). They concluded that Nd: YAG laser capsulotomy is an acceptable option for the management of PCO in an acrylic IOL implantation in children.<sup>[54]</sup>

In a prospective study on 474 patients with PCO who had Nd: YAG laser capsulotomy, Bhargava et al. found a significant relation between mean total laser energy and complications like IOL pitting, IOP rise, CME and retinal detachment. They concluded that subtype of PCO and IOL fixation significantly influences laser energy required for capsulotomy, whereas IOL biomaterials did not. Rate complications like IOL pitting, uveitis, IOP elevation, RD and CME was significantly more when total laser energy delivered to treatment site was higher.<sup>[55]</sup>

- Invasive methods*- surgical removal of capsular opacity is done in selected cases.

**INDICATIONS**

- I) Visual axis opacification in young children.
- II) Thick PCO.
- III) Cases where Nd:YAG laser capsulotomy is ineffective in clearing visual axis.

**TECHNIQUE**

Posterior capsule can be approached by 2 routes- limbal & pars plana routes.

Surgical removal of PCO using 25 gauge transconjunctival sutureless vitrectomy in children was evaluated by Lam et al. All cases showed significant visual improvement.<sup>[56]</sup>

Pars plana capsulotomy in cases with PCO in which Nd:YAG laser was not successful in clearing the visual axis was performed by Mitra et al and they found success in penetrating the thick pupillary membranes.<sup>[57]</sup>

**COMPLICATIONS<sup>[58]</sup>**

- I) Vitreous loss.
- II) Endophthalmitis.

**CONCLUSION**

PCO is a physiological complication of uneventful cataract surgery. Main culprits are LECs that proliferate & form PCO. Despite advances in our understanding of the mechanism of PCO formation, it remains a significant problem, although there has been a reduction in its incidence. Therefore, research aimed at improving surgical techniques to eliminate almost all LECs from the capsular bag at the time of surgery, optimizing IOL biocompatibility, minimizing postoperative inflammation reaction, and targeting residual LECs by therapeutic agents that have minimal or no effect on other ocular tissues is highly desirable. Biological mechanisms leading to PCO formation have now been revealed and agents to inhibit these signalling systems are currently under evaluation. Nd:YAG laser remains the treatment of choice for managing PCO, though correct technique should be used to prevent complications.

**REFERENCES**

1. Karahan E, Er D, Kaynak S. An overview of Nd:YAG laser capsulotomy. *Medical Hypothesis, Discovery & Innovation in Ophthalmology*. 2014; 3: 45-50.
2. Dholakia SA, Vasavada AR. Intraoperative performance & longterm outcome of phacoemulsification in age related cataract. *Indian J. Ophthalmol*. 2004; 52: 311.
3. Thompson AM, Sachdev N, Wong T, et al. The Auckland Cataract Study: 2 year postoperative assessment of aspects of clinical, visual, corneal topographic and satisfaction outcomes. *Br. J. Ophthalmol*. 2004; 88: 1042.
4. Ignjatovic, Z. [Secondary cataracts in extreme myopia][serbo-kroatisch]. *Srp Arh Celok Lek*, 1998; 126: 239-41.
5. Vasavada et al. (*J Cataract Refract Surg*, 2009; 35: 15329).
6. Hayashi K, Hayashi H, Nakao F, et al. Posterior capsule opacification after cataract surgery in patients with diabetes mellitus. *Am J Ophthalmol*. 2002; 134: 10-6.
7. Khwaja, W.A., et al. Visual axis opacification in children. *Ophthalmology*, 2011; 118: 224-5.
8. Raj SM, Vasavada AR, Johar SRK, et al. Post-Operative Capsular Opacification: A Review. *International Journal of Biomedical Science: IJBS*. 2007; 3: 237-50.
9. McDonnell PJ, Payel A, Green WR. Comparison of intracapsular and extracapsular cataract surgery. Histopathologic study of eyes obtained postmortem. *Ophthalmology*. 1985; 92: 1208.
10. Apple DJ, Solomon KD, Tetz MR, et al. Posterior capsule opacification. *Surv Ophthalmol*, 1992; 37(2): 73-116.
11. Saika S. Relationship between posterior capsule opacification and intraocular lens biocompatibility. *Prog Retin Eye Res.*, 2004; 23(3): 283-305.
12. Meacock WR, Spalton DJ, Stanford MR. Role of cytokines in the pathogenesis of posterior capsule opacification. *Br J Ophthalmol*, 2000; 84(3): 332-336.
13. Wormstone IM, Liu CS, Rakic JM, Marcantonnio JM, et al. Human lens epithelial cell proliferation in a protein-free medium. *Invest Ophthalmol Vis Sci.*, 1997; 38(2): 396- 404.
14. Duncan G, Wormstone IM, Liu CS, Marcantonio JM. Thapsigargin-coated intraocular lenses inhibit human lens cell growth. *Nat Med*, 1997; 3(9): 1026-1028.
15. Beck R, Nebe B, Guthoff R, Rychly J. Inhibition of lens epithelial cell adhesion by the calcium antagonist Mibefradil correlates with impaired integrin distribution and organization of the cytoskeleton. *Graefes Arch Clin Exp Ophthalmol*, 2001; 239(6): 452- 458.
16. Sugita M, Kato S, Sugita G, et al. T Migration of lens epithelial cells through haptic root of single-piece acrylic-foldable intraocular lens. *Am J Ophthalmol*, 2004; 137(2): 377-379.
17. Wong TT, Daniels JT, Crowston JG, et al. MMP inhibition prevents human lens epithelial cell migration and contraction of the lens capsule. *Br J Ophthalmol*, 2004; 88(7): 868-872.
18. Frezzotti R, Caporossi A, Mastrangelo D, et al. Pathogenesis of posterior capsular opacification, II: histopathological and in vitro culture findings. *J Cataract Refract Surg*, 1990; 16(3): 353-360.
19. Nishi O. Posterior capsule opacification, part 1: experimental investigations. *J Cataract Refract Surg*, 1999; 25(1): 106-117.
20. Ravalico G, Tognetto D, Polambo M, et al. Capsulorhexis size and posterior capsule opacification. *J Cararact Refractive Surgery*, 1996 Jan-Feb; 22(1): 98-103.

21. Sinha R, Shekhar H, Sharma N, Titiyal JS, Vajpayee RB. Posterior capsular opacification: A review. *Indian J Ophthalmol*, 2013; 61: 371-6.
22. Peng Q, Visessook N, Apple DJ et al. Surgical prevention of posterior capsule opacification, part 3: intraocular lens optic barrier effect as a second line of defense. *J Cataract Refract Surg*, 2000; 26(2): 198-213.
23. Fine IH. Cortical cleaving hydrodissection. *J Cataract Refract Surg*, 1992; 18(5): 508-512.
24. Mathey CF, Kohnen TB, Ensikat HJ, et al. Polishing methods for the lens capsule: histology and scanning electron microscopy. *J Cataract Refract Surg*, 1994; 20(1): 64-69.
25. Tan DT, Chee SP. Early central posterior capsular fibrosis in sulcus fixated biconvex intraocular lenses. *J Cataract Refract Surg*. 1993; 19: 471.
26. Peng Q, Visessook N, Apple DJ, et al. Surgical prevention of posterior capsule opacification, part 3: intraocular lens optic barrier effect as a second line of defense. *J Cataract Refract Surg*, 2000; 26(2): 198-213.
27. Hayashi Y, Kato S, Fukushima H, et al. Relationship between anterior capsule contraction and posterior capsule opacification after cataract surgery in patients with diabetes mellitus. *J Cataract Refract Surg*. 2004; 30: 1517.
28. Daynes T, Spencer TS, Doan K, et al. Three-year clinical comparison of 3-piece Acrysof and SI-40 silicone intraocular lenses. *J Cataract Refract Surg*. 2002; 28: 1124.
29. Vasavada AR, Raj SM. Anterior capsule relationship of the Acrysof Intraocular Lens optic and posterior capsule opacification. A prospective randomised clinical trial. *Ophthalmology*. 2004; 111: 886.
30. Meacock WR, Spalton DJ, Boyce JF, et al. Effect of optic size on posterior capsular opacification: 5.5mm versus 6mm Acrysof intraocular lenses. *J Cataract Refract Surg*. 2001; 27: 1194.
31. Cheng JW, Wei RL, Cai JP et al. Efficacy of different intraocular lens materials and optic edge designs in preventing posterior capsular opacification: a meta-analysis. *Am J Ophthalmol*, 2007; 143(3): 428- 436.
32. Boyce JF, Bhermi GS, Spalton DJ, et al. Mathematical modelling of forces between an intraocular lens and the capsule. *J Cataract Refract Surg*. 2002; 28: 1853.
33. Yuan Z, Sun H, Yuan J. A 1-year study on carbon, titanium surface-modified intraocular lens in rabbit eyes. *Graefes Arch Clin Exp Ophthalmol*, 2004; 242(12): 1008-1013.
34. Larsson R, Selén G, Björklund H, Fagerholm P. Intraocular PMMA lenses modified with surface-immobilized heparin: evaluation of biocompatibility in vitro and in vivo. *Biomaterials*, 1989; 10(8): 511-516.
35. Werner L, Legeais JM, Nagel MD, Renard G. Evaluation of teflon-coated intraocular lenses in an organ culture method. *J Biomed Mater Res*, 1999; 46(3): 347-354.
36. Hettlich HJ, Otterbach F, Mittermayer C, Kaufmann R, et al. Plasma-induced surface modifications on silicone intraocular lenses: chemical analysis and in vitro characterization. *Biomaterials*, 1991; 12(5): 521-524.
37. Yammine P, Pavon-Djavid G, Helary G. Surface modification of silicone intraocular implants to inhibit cell proliferation. *Biomacromolecules*, 2005; 6(5): 2630-2637.
38. Yuen C, Williams R, Batterbury M, et al. Modification of the surface properties of a lens material to influence posterior capsular opacification. *Clin Experiment Ophthalmol*, 2006; 34(6): 568-574.
39. Bozukova D, Pagnoulle C, De Pauw-Gillet MC et al. Improved performances of intraocular lenses by poly (ethylene glycol) chemical coatings. *Biomacromolecules*, 2007; 8(8): 2379-2387.
40. Hollick EJ, Spalton DJ, Ursell PG, Meacock WR, Barman SA, Boyce JF. Posterior capsular opacification with hydrogel, polymethylmethacrylate and silicone intraocular lenses: two-year results of a randomized prospective trial. *Am J Ophthalmol*, 2000; 129(5): 577-584.
41. Zemaitiene R, Jasinskas V, Auffarth GU. Influence of three-piece and single-piece designs of two sharp-edge optic hydrophobic acrylic intraocular lenses on the prevention of posterior capsule opacification: a prospective, randomised, long-term clinical trial. *Br J Ophthalmol*, 2007; 91(5): 644-648.
42. Nishi O. Posterior capsule opacification, part 1: experimental investigations. *J Cataract Refract Surg*, 1999; 25(1): 106-117.
43. Wang et al. (*Zhonghua Yan Ke Za Zhi*, 1994; 30: 4057).
44. Chandler HL, Barden CALu P, Kusewitt DF, Colitz CM. Prevention of posterior capsular opacification through cyclooxygenase-2 inhibition. *Mol Vis.*, 2007; 13:677-691.
45. Cortina P, Gómez-Lechón MJ, Navea A, Menezo JL, Terencio MC, Diaz-Llopis. Diclofenac sodium and cyclosporin A inhibit human lens epithelial cell proliferation in culture. *Graefes Arch Clin Exp Ophthalmol*, 1997; 235(3): 180-185.
46. Wong TT, Daniels JT, Crowston JG, Khaw PT. MMP inhibition prevents human lens epithelial cell migration and contraction of the lens capsule. *Br J Ophthalmol*, 2004; 88(7): 868-872.
47. Awasthi N, Wagner BJ. Suppression of human lens epithelial cell proliferation by proteasome inhibition, a potential defense against posterior capsular opacification. *Invest Ophthalmol Vis Sci.*, 2006; 47(10): 4482-4489.
48. Maloof A, Neilson G, Milverton EJ, Pandey SK. Selective and specific targeting of lens epithelial cells during cataract surgery using sealed-capsule irrigation. *J Cataract Refract Surg*, 2003; 29(8): 1566-1568.

49. Belcher CD 3<sup>rd</sup>, Mainster MA, Buzney SM (1983). Current status of neodymium: YAG laser photodisrupters in Ophthalmology: Part I. *Ann Ophthalmol*, 15: 997-999.
50. Belcher CD 3<sup>rd</sup>, Mainster MA, Buzney SM (1983). Current status of neodymium: YAG laser photodisrupters in Ophthalmology: Part II. *Ann Ophthalmol*, 15: 1097-1099.
51. Chambliss WS(1985). Neodymium:YAG laser posterior capsulotomy results and complications. *J Am Intraocul Implant Soc.*, 11: 31-33.
52. Steinert, Roger F. "Nd: YAG laser posterior capsulotomy". American Academy of Ophthalmology, One network, 2013.
53. Min Jk, An JH, Yim JH. A new technique for Nd:YAG laser posterior capsulotomy. *International Journal of Ophthalmology*. 2014; 7: 345-349.
54. Stager DR Jr, Wang X, Weakley DR Jr, et al. (2006) The effectiveness of Nd:YAG laser capsulotomy for treatment of posterior capsular opacification in children with acrylic intraocular lenses. *J AAPOS*, 10: 159-163.
55. Bhargava R, Kumar P, Phogat H, et al., (2014) Analysis of Neodymium: Yttrium-Aluminium Garnet Laser Capsulotomy Energy Levels For Posterior Capsule Opacification. *J Ophthalmic Vision & Research*.
56. Lam et al. *Clin Experiment Ophthalmol*, 2005; 33: 4958.
57. Mitra et al. (*Ophthalmic Surg Lasers Imaging*, 2003; 34: 32731.
58. Shah GR, Gills JP, Durham DG, et al. Three thousand YAG lasers in posterior capsulotomies: an analysis of complications and comparison to polishing and surgical discission. *Ophthalmic Surg*. 1986; 17: 473-7.