ejpmr, 2016,3(1), 142-149



# EUROPEAN JOURNAL OF PHARMACEUTICAL AND MEDICAL RESEARCH

www.ejpmr.com

Review Article ISSN 3294-3211 EJPMR

# **RHEUMATOID ARTHRITIS AND NANOTHERAPEUTICS**

## Mahalakshmi AM, Dr. K. Santhi\* and Ramesh Nidavani

Department of Pharmacology, and Department of Pharmaceutics, JSS College of Pharmacy, JSS University, Sri Shivarathreeshwara Nagara, Mysore-570 015, India.

#### \*Correspondence for Author: Dr. K. Santhi

Department of Pharmacology, and Department of Pharmaceutics, JSS College of Pharmacy, JSS University, Sri Shivarathreeshwara Nagara, Mysore-570 015, India.

Article Received o	on 22/10/2015
--------------------	---------------

Article Revised on 13/11/2015

Article Accepted on 04/12/2015

#### ABSTRACT

Rheumatoid arthritis (RA) is an autoimmune, symmetrical polyarticular disease that affects primarily the diarthrodial joints, which are characterized by chronic inflammation of the synovial joints. Inflammation is the net result of a cascade of highly regulated events propagated upon stimulation, and is the major process through which the body repairs tissue damage and defends itself against foreign materials. Acute inflammation is typically caused by an external chemical, mechanical, or pathogenic influence; while chronic inflammation requires no external stimulus and can cause a range of painful and debilitating symptoms. Histological changes occur with abnormal cell distribution in targeted tissues and indicate localized population of macrophages and lymphocytes, then fibrosis and necrosis. Nanoparticles sized between 1 and 100 nm are already in use including cosmetics, food industry, medicine, electronics, and others. Exposure to nanomaterials induces cell dysfunctions at various levels, such as cell death by oxidative stress, DNA damage, and protein damage. Cell damage produces free radicals including reactive oxygen species (ROSs) and reactive nitrogen species (RNSs). It was also reported that generation of various inflammatory mediators, depends on type of nanoparticles. Present review highlights on the role of various nanoparticles on inflammation including, silica, iron oxide, cerium oxide, yttrium oxide, zinc oxide, silver, nickel oxide and Synthetic hydroxyapatite nanoparticles.

**KEY WORDS:** Inflammation, nanoparticles, proinflammatory mediators, cytokines, rheumatoid arthritis, silica nanoparticles.

# INTRODUCTION

Inflammation is the net result of a cascade of highly regulated events propagated upon stimulation, and is the major process through which the body repairs tissue damage and defends itself against foreign materials. Acute inflammation is typically caused by an external chemical, mechanical, or pathogenic influence; has a relatively short duration (hours to days); and is a necessary protection tool that removes foreign bodies and damaged tissue, preventing further damage. Chronic inflammation requires no external stimulus and can cause a range of painful and debilitating symptoms<sup>1</sup>. Further, such uncontrolled inflammation is often indicative of a more serious, underlying cause whose analysis may be used as a diagnostic marker for a number of conditions, including autoimmune, infectious, neurological, cardiovascular, and metastatic diseases. Histological changes will have occurred with abnormal cell distribution in targeted tissues and indicates localized populations of macrophages and lymphocytes, then fibrosis and necrosis.<sup>[2, 3]</sup>

The application of nanotechnology in industry is rapidly growing, with a worldwide market size estimated to be in excess of US\$1 trillion by the year 2015.<sup>[4]</sup> Despite the quick progress and early acceptance of nanotechnology,

there are lot of research groups revealing, potential for adverse health effects in humans and the environment. Although several research groups have demonstrated that exposure to nanoparticles may affect cellular viability and growth, through inflammatory pathway, and thus, little is known regarding the potential mechanism(s) of toxicity. It is thought that nanoparticles can disrupt and impair normal cellular function through a number of mechanisms.<sup>[5]</sup>

# Role of immune cells in rheumatoid arthritis and other inflammatory diseases

Rheumatoid arthritis is an autoimmune, (RA) symmetrical polyarticular disease that affects primarily the diarthrodial joints, which is characterized by chronic inflammation of the synovial joints.<sup>[6]</sup> There is great heterogeneity in the cells of immune system, most of which originate from hematopoietic stem cells in the fetal liver and the postnatal bone marrow. Immune responses are mediated by a variety of cells and the soluble molecules that these cells secrete. Monocytes belong to class of reticuloendothelial system also named as mononuclear phagocyte system (MPS). Monocytes upon circulation in blood matured and become macrophages. Macrophages are key members of the MPS and play vital roles in inflammation, including

phagocytosis, antigen presentation, and the production of various cytokines and immune regulators.<sup>[7]</sup> Resident macrophages are found in all organs and mainly in connective tissues, involve in the primary immune response of tissues. Upon activation of macrophages, release chemokines that adhere to circulating monocytes, trafficking MPS cells to the inflammation site and generating an immune response. The ability to visualize the migration of MPS cells would contribute to many including common disorders, atherosclerosis, autoimmunity, and major infections.<sup>[8]</sup> The cytokines produced by macrophages not only sequester other immune cells to the tissue, they can also propagate an autoimmune response and inflammation. Cytokines maintain part of a complex regulatory network, and disruption of the balance between pro- and antiinflammatory cytokines has been implicated in the pathogenesis of a number of inflammatory disorders.<sup>[9]</sup> This can be illustrated with tumor necrosis factor-a (TNF- $\alpha$ ), a cytokine implicated to play a key role in RA. Targeting TNF- $\alpha$  as a central product of macrophages has been demonstrated as a powerful approach to treat RA, with many drug treatments utilizing antibodies or receptor fusion proteins to TNF-  $\alpha$ .

Polymorphonuclear (PMN) neutrophils are short lived phagocytes and constitute majority of blood leucocytes and develop from the same early precursors as monocytes and macrophages. Mast cells are important effector cells in IgE-mediated reactions by secreting histamine, chymase, tryptase, leukotrienes (LTs), prostaglandin D2 (PGD2) and several multifunctional cytokines; they include interleukin-6 (IL-6), IL-8, IL-13, TNF-  $\alpha$ , stem cell factor (SCF) and many chemotactic factors.<sup>[10,11]</sup> These cytokines contribute to the late-phase allergic reactions and to allergic inflammation through the recruitment of immune cells into the site of inflammation.<sup>[12, 13]</sup>

# Frontline of nanotherapeutics

Nanotechnology is one of the exponentially developing technologies of the 21st century. Nanoparticles are submicron moieties (diameters ranging from 1 to 100 nm according to the used term, although there are examples of nanoparticles several hundreds of nanometers in size) made of inorganic or organic materials, which have many novel properties compared with the bulk materials. Nanoparticles used for the most part including cosmetics, food industry, medicine, electronics, and others.<sup>[14]</sup> However, because nanoparticles have large surface areas, high chemical reactivity, internal pore volumes, and enhanced cell penetrability, it may induce much more toxic effects.<sup>[15, 16]</sup>

# Dark side of nanotherapeutics

Exposure to nanomaterials induces cell dysfunctions at various levels, such as cell death by oxidative stress, DNA damage, and protein damage. DNA damage is associated with malignant tumors and is closely related to inflammation. For instance, inhalation or intratracheal exposure can cause acute and chronic inflammation to the respiratory tract and pulmonary alveolar space<sup>[17, 18]</sup>, in particular, inflammation causes fibrosis of the lung and pleura and progresses to lung cancer or malignant mesothelioma<sup>[19]</sup> Cell damage produces free radicals including reactive oxygen species (ROSs) and reactive nitrogen species (RNSs). In the ROSs, there are superoxide ions  $(O_2)$ , hydroxyl radicals ( $\cdot OH$ ), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), and singlet oxygen ( $^{1}O_{2}$ ). The  $\cdot OH$  is the most reactive of these ROSs. On the other hand, in the RNSs, there are nitric oxide (NO), nitrosonium ion  $(NO^+)$ , nitrite ion  $(NO_2^-)$ , and peroxynitrite  $(ONOO^-)$ . The ONOO<sup>-</sup> is the most reactive of these RNSs<sup>[20]</sup> Free radicals are produced spontaneously in the energy metabolism of the cell. It was reported that generation of ROSs are depends on type of nanoparticles.

Nanomaterials and asbestos are taken into the body, and free radicals are produced on their surface by inflammatory cells or epithelial cells phagocytising them. The phagocyte cells such as neutrophils and macrophages play a role essential to the host defence to produce superoxides by the active oxygen production enzyme system, such as NADPH oxidase<sup>[21]</sup> Mutations of 8-hydroxydeoxyguanosine (8-OHdG) and 8-nitroguanine (8-NG) are known as DNA damages caused by ROSs and RNSs respectively<sup>[22]</sup> Further accumulation of mutants may induce apoptosis or tumours<sup>[23, 24]</sup> The possible relationship between nanoparticles and inflammation is depicted in figure 1.

## PHARMACOLOGICAL EVIDENCES FOR INFLAMMATION INDUCED BY NANOPARTICLES

## Silica nanoparticles (SiNPs)

Inhalation of higher doses of SiNPs can particularly occur in occupations such as ceramic, mine or foundry workers, dental laboratory technicians, agricultural workers or stone cutters. Amorphous SiNPs are being applied increasingly in industrial manufacturing, high-molecule composite materials, tyre compounds, thermal insulation materials, cosmetics, and food stuffs<sup>[25]</sup> To date SiNPs play an important role in modern technology and nanomedicine. Silicon dioxide exists in either crystalline or amorphous forms<sup>[26]</sup> It is well established that occupational inhalation exposure to crystalline silica causes silicosis. Silica has been considered an ideal nanoparticle for biomedical applications such as gene therapy, drug delivery, biomedical imaging, biosensors, and enzyme immobilization.<sup>[26, 27, 28]</sup>

An association of crystalline silica exposure and silicosis, as well as lung cancer, chronic obstructive pulmonary disease, and pulmonary tuberculosis, have been recently reported.<sup>[29]</sup> The development of nanotechnology has raised new interest in the use of amorphous SiNPs in biomedical, pharmaceutical, and many other industrial applications.<sup>[26, 28]</sup> Ultrafine silica induced oxidative stress and pro-inflammatory responses in macrophages, mice and rats.<sup>[30, 31]</sup> Also, pulmonary

inflammation, emphysema, alveolar hyperinflation, and apoptosis of alveolar and granulomatous cells have been found in animals exposed to silica.<sup>[32, 33]</sup> Lot of works has been done on the toxicity of SiNPs to produce inflammatory mediators. Some are mentioned here.

The study hypothesized that direct exposure of human aortic endothelial cells (HAECs) to various nanoparticles induces an inflammatory response. The mechanisms governing the correlation between exposure to nanoparticles and the increased incidence of cardiovascular disease is of particular concern in nanotoxicology related fields. Nanoparticles appear to cross the pulmonary epithelial barrier into the bloodstream, raising the possibility of direct contact with the vascular endothelium.<sup>[34]</sup> And nanoparticle- triggered endothelial dysfunction is hypothesized to be a dominant mechanism in the development of the diseases.<sup>[35]</sup> Acute and chronic inflammation of the endothelium plays a central role in the development of atherosclerosis and other cardiovascular diseases.[36]

There is a growing body of evidence that amorphous SiNPs can cause toxic and inflammatory effects due to their unique physicochemical profile. A concentration-dependent cytotoxicity of SiNPs on endothelial cell line EA.hy926, epithelial cell line A549, and monocyte-macrophages J774 has been reported.<sup>[37, 38]</sup> SiNPs have been reported to induce cytotoxicity and inflammatory responses *in vitro* in a co-culture model of the alveolar–capillary barrier.<sup>[39]</sup> There is evidence that SiNPs can induce impairment of proliferative activity and proinflammatory stimulation of endothelial cells *in vitro*.<sup>[40]</sup>

The measurement of proinflammatory cytokine showed a significant increase of TNF $\alpha$  after the administration of 50 nm SiNPs and IL-1 $\beta$  following the exposure to both 50 nm and 500 nm SiNPs. This finding also confirmed by another study by Nemmar et al. (2013), and they suggested that occurrence of systemic inflammation, which can explain the thrombotic effects of SiNPs.<sup>[41]</sup> The *in vitro* release of IL-1 $\beta$  and TNF $\alpha$  has been reported following exposure to amorphous SiNPs.<sup>[42]</sup>

The study finds, SiNPs shows the placental inflammation and pregnancy complications. SiNPs upregulated the inflammasome component nucleotide-binding oligomerization domain-like receptor (NLR) family pyrin domain-containing 3 (NLRP3) and induced placental inflammation and also ROS's generation, resulting in pregnancy complications. Furthermore, NSinduced pregnancy complications were markedly improved in Nlrp3<sup>-/-</sup> mice but not in component apoptosis-associated speck-like protein containing a caspase recruitment domain (ASC)-deficient (Asc<sup>-/-</sup>) mice, indicating the independence of NLRP3 inflammasomes. Pregnancy complications in Nlrp3<sup>-/-</sup> and Asc<sup>-/-</sup> mice phenotypes were dependent on the balance between IL-1 $\alpha$  and IL-10. SiNPs-induced pregnancy complications were completely prevented by either inhibition of ROS generation or forced expression of IL-10. These findings provide important information about SiNPs-induced placental inflammation and pregnancy complications and the novel pathophysiological roles of NLRP3 and ASC in pregnancy.<sup>[43]</sup>

Kyeongah K and Jong-Seok L (2012), studied SiNPs induces inflammation and also a cytotoxic effect in mouse dendritic cells (DCs). SiNPs decreased the viability of DCs and increased the amount of cell deaths. In addition to the effect on DC differentiation, it induced TNF- $\alpha$  production in DCs and led to inflammatory responses *in vitro* and *in vivo*.<sup>[44]</sup>

The inflammatory and also cytotoxic responses of monodisperse amorphous SiNPs of 30 nm in size on an in vitro coculture model mimicking the alveolar-capillary barrier and compared these to conventional monocultures by using epithelial cell line (lung adenocarcinoma cell line NCI H441, H441) and the endothelial cell line (clone of the angiosarcoma cell line ISO-HAS, ISO-HAS-1) were used in monoculture and in coculture on opposite sides of a filter membrane. The study evaluated, release of proinflammatory mediators like, soluble intracellular cell adhesion molecule-1 (sICCAM-1), IL-6, and IL-8. Additionally cytotoxicity and apoptosis markers were investigated. This experimental work suggested that much more sensitive fashion than the conventional monoculture for the release of inflammatory markers. At concentrations that were 10-100 fold less than the toxic concentrations the apically exposed coculture showed a release of IL-6 and IL-8 to the basolateral side. This may mimic the early inflammatory events that take place in the pulmonary alveoli after SiNPs inhalation.[39] Marzaioli et al (2014), expressed, SiNPs showed a strong proinflammatory effect from broncho-alveolar lavage fluid (BALF) of BALB/c mice, leading to neutrophil and lymphocyte recruitment after intratracheal instillation, showing neutrophils recruitment in the early inflammation stages (up to 1 week after instillation) following intratracheal instillation of SiNPs.<sup>[45]</sup>

# Iron oxide nanoparticles

Iron oxide nanoparticles ( $Fe_2O_3$  and  $Fe_3O_4$ ) or magnetic nanoparticles are used in important bio applications, including magnetic bioseparation and detection of biological entities (cell, protein, nucleic acids, enzyme, bacterials, virus, etc.), clinic diagnosis and therapy such as magnetic resonance image (MRI) and magnetic fluid hyperthermia (MFH), targeted drug delivery and biological labels.<sup>[46]</sup>

Zhu et al. (2011), studied the proinflammatory action of iron oxide nanoparticles ( $Fe_2O_3$  and  $Fe_3O_4$ ) on HAECs and monocyte, phagocytosis and activation; and also investigated ICAM-1, IL-8, expression, as well as NO and nitric oxide synthase (NOS) activity. In the study, HAECs and U937 cells were exposed to 2, 20, 100 µg/mL of 22nm-Fe<sub>2</sub>O<sub>3</sub> and 43nm-Fe<sub>3</sub>O<sub>4</sub> particles. This work shown that intravascular iron oxide nanoparticles may induce endothelial system inflammation and dysfunction by three ways: first, nanoparticles may escape from phagocytosis that interact directly with the endothelial monolayer; second, nanoparticles are phagocytized by monocytes and then dissolved, thus impact the endothelial cells as free iron ions; and third, nanoparticles are phagocytized by monocytes to provoke oxidative stress responses.<sup>[47]</sup>

Andrea et al. (2007), hypothesized that direct exposure of HAECs to ultrafine particles induces an inflammatory response and that this response depends on particle composition; by incubating HAECs for 1-8 hr with different concentrations (0.001-50 µg/mL) of Fe<sub>2</sub>O<sub>3</sub>, Y<sub>2</sub>O<sub>3</sub>, and ZnO nanoparticles and subsequently measured messenger RNA (mRNA) and protein levels of the three inflammatory markers, namely ICCAM-1, IL-8, and monocyte chemotactic protein-1 (MCP-1). Overall study recommended that, Fe<sub>2</sub>O<sub>3</sub> nanoparticles fail to provoke an inflammatory response in HAECs at any of the concentrations tested; however, Y2O3 and ZnO nanoparticles elicit a pronounced inflammatory response above a threshold concentration of 10µg/mL. This study also suggested that animal studies of the impact of nanoparticles on vascular inflammation should help to find the action of on  $Fe_2O_3$  inflammation.<sup>[48]</sup>

Kennedy IM et al (2009), studied that, incubation of HAECs with  $Fe_2O_3$  (0.001-50 ug/mL) for 1 to 8 hrs. Measured mRNA levels of three markers of inflammation namely, ICCAM-1, IL-8, and MCP-1 using real-time reverse transcriptase-polymerase chain reaction (RT-PCR). Also studied interactions of nanoparticles with HAECs, by using inductively coupled plasma-mass spectrometry (ICP-MS) to measure the concentration of internalized particles. However, overall the study revealed that  $Fe_2O_3$  nanoparticles did not provoke an inflammatory response in HAECs at any of the concentrations tested. These results demonstrate that inflammation in HAECs after acute exposure to metal oxide nanoparticles depends on the concentration and composition of the particles.<sup>[34]</sup>

# Cerium oxide (CeO<sub>2</sub>) nanoparticles

CeO<sub>2</sub> nanoparticles (15-45 nm; 5-40  $\mu$ g/mL) induced oxidative stress and cell death in cultured human lung epithelial cells. CeO<sub>2</sub> nanoparticles have been tested for their ability to serve as free radical scavengers to render protection against chemical, biological and radiological insults that promote the production of free radicals.<sup>[49]</sup>

Kennedy IM et al (2009), studied that, incubation of HAECs with CeO<sub>2</sub> (0.001-50 ug/mL) for 1 to 8 hrs. Measured mRNA levels of three markers of inflammation namely, ICCAM-1, IL-8, and MCP-1 using RT-PCR. Also studied interactions of nanoparticles with HAECs, by using ICP-MS to measure the concentration of internalized particles. Overall CeO<sub>2</sub> particles elicited no response at low concentrations and a weak response

above 10ug/mL. These results demonstrate that inflammation in HAECs after acute exposure to metal oxide nanoparticles depends on the concentration and composition of the particles.<sup>[34]</sup>

# Yttrium oxide (Y<sub>2</sub>O<sub>3</sub>) nanoparticles

The  $Y_2O_3$  nanoparticles are used in a number of different applications, including biological imaging, the material sciences, chemical synthesis of inorganic compounds, manufacturing of plasma televisions, cathode ray display panels, microwave filters, in high-temperature/infraredshielding applications and as additives in paint, plastic, steel, iron, and optics.<sup>[50, 51]</sup>

Kennedy IM et al (2009), studied that, incubation of HAECs with  $Y_2O_3$  (0.001-50 ug/mL) for 1 to 8 hrs. Measured mRNA levels of three markers of inflammation include, ICCAM-1, IL-8, and MCP-1 using RT-PCR. The study also suggested that, interactions of nanoparticles with HAECs, by using ICP-MS to measure the concentration of internalized particles. Overall  $Y_2O_3$  elicited a pronounced inflammatory response above a threshold concentration of 10ug/mL. These results demonstrate that inflammation in HAECs after acute exposure to metal oxide nanoparticles depends on the concentration and composition of the particles.<sup>[34]</sup>

# Zinc oxide (ZnO) nanoparticles

The biomedical applications of ZnO nanoparticles are biomedical imaging (which includes fluorescence, magnetic resonance, positron emission tomography, as well as dual-modality imaging), drug delivery, gene delivery, and biosensing of a wide array of molecules of interest.<sup>[52]</sup> Kennedy IM et al (2009), studied that, incubation of HAECs with ZnO (0.001-50 ug/mL) for 1 to 8 hrs. Measured mRNA levels of three markers of inflammation namely, ICCAM-1, IL-8, and MCP-1 using RT-PCR. Also studied interactions of nanoparticles with HAECs, by using ICP-MS to measure the concentration of internalized particles. Overall ZnO elicited a pronounced inflammatory response above a threshold concentration of 10ug/mL. These results demonstrate that inflammation in HAECs after acute exposure to metal oxide nanoparticles depends on the concentration and composition of the particles.<sup>[34]</sup>

# Silver nanoparticles (AgNPs)

Currently, silver nanoparticles (AgNPs) are the most common nanoparticles in nanomedicine. According to the Nanotechnology Consumer Products Inventory, AgNPs are currently claimed to be used in more than 400 consumer products.<sup>[53]</sup> AgNPs have antimicrobial activity (along with released ions, binding to sulphur- and phosphorous containing biomolecules such as proteins and DNA) and are used in food packaging material, food supplements, odour-preventing textiles, cosmetics, kitchen utensils, toys, electronics, wound dressings, and room sprays.<sup>[53, 54]</sup> The study hypothesize that small AgNPs will induce more prominent pulmonary inflammation compared to larger, because of the larger deposited dose in the alveoli and the higher dissolution rate. AgNPs of 15 nm and 410 nm were exposed after short-term nose inhalation. All endpoints were determined at 24 hours and 7 days after the last exposure. The inflammatory markers mainly cytokines were estimated in the BALF and also total blood cell counts and cell damage markers. 12 different proinflammatory cytokines were selected: IL-1B, IL-6, TNF-α, IFN-γ (interferon-gama), IL-13, GM-CSF, MCP-1, IL-12p70, IL-18, macrophage inflammatory protein- $1\alpha$  (MIP- $1\alpha$ ), MIP-2 and RANTES (Regulated on Activation, Normal T Cell Expressed and Secreted). Of these cytokines, only IL-1β, MCP-1 and MIP-2 could be measured; all the other cytokines were and stayed below or around the detection level. Both IL-1ß and MCP-1 were significantly increased 24 hours after exposure. The study suggested that inflammation in pulmonary was size-related, that is exposure to 15 nm AgNPs induced moderate pulmonary inflammation at 24 hours after exposure, whereas 410 nm AgNPs did not.<sup>[55]</sup>

#### Synthetic hydroxyapatite nanoparticles (HANPs)

Synthetic hydroxyapatite (HA) (Ca10[PO4]6[OH]2), a typical bioceramic with good osteoconductive and osteoinductive capabilities, has been used clinically for many years.<sup>[56]</sup> Due to their better bioactivity, their

excellent capacity to penetrate cell membranes, and their increased circulation time, HANPs have gradually garnered significant interest in various medical fields, such as bone tissue engineering, cardiovascular graft coating, contrast agent synthesis, drug delivery, and gene therapy.<sup>[57-59]</sup> The study measured the stimulation of mitogen-activated protein kinases (MAPKs) and nuclear factor-kappa B (NF-κB) in human umbilical vein ECs (HUVECs). Four patterns of cytokine release (TNF-α, IL-1β, IL-6, and IL-8) were also estimated. THP-1 cells exposed to HANPs exhibited significant increases in TNF-α (up to 15-fold) and IL-1β (up to twofold) in both monoculture and co-cultures of monocytes.<sup>[60]</sup>

## Nickel oxide (NiO) nanoparticles

The biomedical applications of NiO nanoparticles includes, in battery cathodes, gas materials, photovoltaic devices and others. The NiO nanoparticles toxicity has been evaluated in the human pulmonary epithelial cell lines: BEAS-2B and A549. The nanoparticles, used at the doses of 20, 40, 60, 80, 100  $\mu$ g/ml, induced a significant reduction of cell viability and an increase of apoptotic and necrotic cells at 24h. A significant release of interleukin-6 and-8 was assessed after 24h of treatment, even intracellular ROS increased already at 45 min after exposure. The results obtained evidenced that the cytokines release was dependent on MAPK cascade through the induction of NF-kB pathway.<sup>[61]</sup>



Figure 1: Possible relationship between nanoparticles and inflammation.

#### CONCLUSION

Reports on the toxicology of nanomaterials have been increasing recently, but the effect of nanomaterials on the human body is inconclusive, for example, in general, inhaled dusts such as particles and fibrous materials in the lung repeatedly induce inflammation and finally lead to pulmonary fibrosis, respiratory cancer and others. Many factors are likely to be involved in the adjuvant activity of nanoparticles, and a consistent mechanism has not been found. In future it is expected to elucidate what factors are involved for the release of inflammatory mediators in inflammation and other inflammatory diseases induced by various nanoparticles.

## **AUTHORS' STATEMENTS**

The authors declare that there is no conflict of interests about the publication of this review paper.

#### REFERENCES

 Ramesh BN, Mahalakshmi AM, Krishna KL. Update on the diagnosis and management of Reactive arthritis (Reiter's syndrome). Der Pharmacia Sinica, 2015; 6 (2): 12-18.

- Ramesh BN, Mahalakshmi AM, Mallappa S. Vascular permeability and Evans blue dye: a physiological and pharmacological approach. J Applied Pharm Sci. 2014; 4(6): 106-113. http://dx.DOI:10.7324/JAPS.2014.41119.
- Ramesh BN, Mahalakshmi AM. Present clinical approaches to Rheumatoid Arthritis: An ample review. Indo-American J Pharm Sci. 2013; 3(8): 6055-6065.
- Baalousha M, Stolpe B, Lead JR. Flow field-flow fractionation for the analysis and characterization of natural colloids and manufactured nanoparticles in environmental systems: a critical review. J Chromatogr A. 2011; 1218(27): 4078-4103.
- Kirchner C, Liedl T, Kudera S, et al. Cytotoxicity of colloidal CdSe and CdSe/ZnS nanoparticles. Nano Lett. 2005; 5(2): 331-338.
- Mahalakshmi AM, Subhashini V, Ramesh BN, Suresh B. Methotrexate toxicity in Rheumatoid arthritis to shrink on early diagnosis and treatment: A Prospective open label study. World J Pharm Pharm Sci. 2013; 2(6): 5538-5551.
- Fujiwara, N. and Kobayashi, K. Macrophages in inflammation. Curr. Drug Targets Inflamm. Allergy. 2005; 4: 281-286.
- Moghimi SM, Hunter AC, Murray JC. Nanomedicine: current status and future prospects. FASEB J. 2005; 19(3), 311-330.
- 9. McInnes IB, Schett G. Cytokines in the pathogenesis of rheumatoid arthritis. Nat. Rev. Immunol. 2007; 7(6): 429-442.
- 10. Galli SJ. Mast cells and basophils. Curr. Opin. Hematol. 2000; 7: 32-39.
- 11. Mekori YA, Metcalfe DD. Mast cells in innate immunity. Immunol. Rev. 2000; 173: 131-140.
- 12. Wedemeyer J, Tsai M, Galli SJ. Roles of mast cells and basophils in innate and acquired immunity. Curr. Opin. Immunol. 2000; 12, 624-631.
- 13. Theoharides TC, Cochrane DE. Critical role of mast cells in inflammatory diseases and the effect of acute stress. J. Neuroimmunol. 2004; 146: 1–12.
- Chen M, von Mikecz A. Formation of nucleoplasmic protein aggregates impairs nuclear function in response to SiO<sub>2</sub> nanoparticles. Exp. Cell Res. 2005; 305: 51-62.
- 15. Vallhov HS, Gabrielsson M, Strømme A, Scheynius, Garcia-Bennett AE. Mesoporous silica particles induce size dependent effects on human dendritic cells. Nano Lett. 2007; 7: 3576-3582.
- Cho WS, Choi M, Han BS, Cho M, Oh J, Park K et al. Inflammatory mediators induced by intratracheal instillation of ultrafine amorphous silica particles. Toxicol. Lett. 2007; 175: 24-33.
- Roursgaard M, Poulsen SS, Poulsen LK, et al., Time response relationship of nano and micro particle induced lung inflammation. Quartz as reference compound. Human Exp Toxicol; 2010; 29(11): 915-933.
- 18. Park EJ, Yoon J, Choi K, Yi J, Park K. Induction of chronic inflammation in mice treated with titanium

dioxide nanoparticles by intratracheal instillation. Toxicol, 2009; 260(1-3): 37-46.

- Ardies CM. Inflammation as cause for scar cancers of the lung. Integ Cancer Therapies. 2003; 2(3): 238-246.
- Reiter RJ, Tan D, Manchester LC, Qi W. Biochemical reactivity of melatonin with reactive oxygen and nitrogen species: a review of the evidence. Cell Biochem Biophysics. 2001; 34(2): 237-256.
- Manke A, Wang L, Rojanasakul Y.Mechanisms of nanoparticle-induced oxidative stress and toxicity. Bio Med Res Int. 2013; Article ID 942916:1-15. http://dx.doi.org/10.1155/2013/942916.
- 22. Kaneko K, Akuta T, Sawa T, et al. Mutagenicity of 8-nitroguanosine, a product of nitrative nucleoside modification by reactive nitrogen oxides, in mammalian cells. Cancer Let, 2008; 262(2): 239-247.
- 23. Yoo K, Yoon C, Kwon D, et al. Titanium dioxide induces apoptotic cell death through reactive oxygen species-mediated Fas upregulation and Bax activation. Intern J Nanomed. 2012; 7: 1203-1214.
- 24. Morishige T, Yoshioka Y, Tanabe A, et al., Titanium dioxide induces different levels of IL-1 $\beta$  production dependent on its particle characteristics through caspase-1 activation mediated by reactive oxygen species and cathepsin B. Biochem Biophysical Res Communications. 2010; 392(2): 160-165.
- 25. Greenberg MI, Waksman J, Curtis J: Silicosis: a review. Dis Mon 2007; 53: 394-416.
- 26. Calvert GM, Rice FL, Boiano JM, Sheehy JW, Sanderson WT. Occupational silica exposure and risk of various diseases: an analysis using death certificates from 27 states of the United States. Occup Environ Med. 2003; 60(2): 122-129.
- 27. Huang DM, Chung TH, Hung Y, Lu F, Wu SH, Mou CY et al. Internalization of mesoporous silica nanoparticles induces transient but not sufficient osteogenic signals in human mesenchymal stem cells. Toxicol. Appl. Pharmacol. 2008; 231: 208-215.
- Borm PJ, Robbins D, Haubold S, Kuhlbusch T, Fissan H, Donaldson K, et al. The potential risks of nanomaterials: a review carried out for ECETOC. Part Fibre Toxicol 2006; 3:11.
- 29. Napierska D, Thomassen LC, Lison D, Martens JA, Hoet PH. The nanosilica hazard: another variable entity. Part Fibre Toxicol. 2010; 7(1):39.
- Park EJ, Park K. Oxidative stress and proinflammatory responses induced by silica nanoparticles in vivo and in vitro. Toxicol. Lett. 2009; 184: 18-25.
- 31. Fubini B, Hubbard A. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis. Free Radic. Biol. Med. 2003; 34: 1507-1516.
- 32. Warheit DB, McHugh TA, Hartsky MA. Differential pulmonary responses in rats inhaling crystalline,

colloidal or amorphous silica dusts. Scand. J. Work Environ. Health 1991; 21(Suppl 2): 19-21.

- 33. Leigh J, Wang H, Bonin A, Peters M, Ruan X. Silica-induced apoptosis in alveolar and granulomatous cells in vivo. Environ. Health. Perspect. 1997; 105(Suppl 5): 1241-1245.
- Kennedy IM, Wilson D, Barakat AI. Uptake and inflammatory effects of nanoparticles in a human vascular endothelial cell line. Res Rep Health Eff Inst. 2009; 136: 3-32.
- 35. Zhu MT, Wang B, Wang Y, Yuan L, Wang HJ, Wang M, et al. Endothelial dysfunction and inflammation induced by iron oxide nanoparticle exposure: Risk factors for early atherosclerosis. Toxicol Lett. 2011; 203(2): 162-171. http://dx:doi:10.1016/j.toxlet.2011.03.021.
- Libby P. 2002. Inflammation in atherosclerosis. Nature. 420:868–874.
- Napierska D, Thomassen LC, Rabolli V, et al. Sizedependent cytotoxicity of monodisperse silica nanoparticles in human endothelial cells. Small. 2009; 5(7): 846-853.
- Lison D, Thomassen LC, Rabolli V, et al. Nominal and effective dosimetry of silica nanoparticles in cytotoxicity assays. Toxicol Sci. 2008; 104(1): 155-162.
- 39. Kasper J, Maria IH, Christoph B, Michael M, Roland S, Christine P, Ronald EU, James CK. Inflammatory and cytotoxic responses of an alveolar-capillary coculture model to silica nanoparticles: Comparison with conventional monocultures. Part Fibre Toxicol. 2011; 8 (6): 1-16.
- 40. Peters K, Unger RE, Kirkpatrick CJ, Gatti AM, Monari E. Effects of nano-scaled particles on endothelial cell function in vitro: studies on viability, proliferation and inflammation. J Mater Sci Mater Med. 2004; 15(4): 321-325.
- 41. Nemmar A, Holme JA, Rosas I, Schwarze PE, Alfaro-Moreno E. Recent advances in particulate matter and nanoparticle toxicology: a review of the in vivo and in vitro studies. Biomed Res Int. 2013; 2013:279371. http://dx:doi:10.1155/2013/279371.
- 42. Sandberg WJ, Låg M, Holme JA, et al. Comparison of non-crystalline silica nanoparticles in IL-1beta release from macrophages. Part Fibre Toxicol. 2012; 9:32.
- 43. Shirasuna K, Usui F, Karasawa T, Kimura H, Kawashima A, Mizukami H, et al. Nanosilicainduced placental inflammation and pregnancy complications: Different roles of the inflammasome components NLRP3 and ASC. Nanotoxicol. 2014 Sep; 11: 1-14.
- 44. Kyeongah K and Jong-Seok L. Induction of Functional Changes of Dendritic Cells by Silica Nanoparticles. Immune Network. 2012; 12(3): 104-112.
- 45. Marzaioli V, Juan AAP, Ingrid W, Georg L, Martin W, Robert L et al. Surface modifications of silica nanoparticles are crucial for their inert versus proinflammatoryand immunomodulatory properties.

Int J Nanomed. 2014; 9: 2815-2832. http://dx:doi:10.2147/IJN.S57396.

 Gupta AK, Gupta M. Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. Biomaterials. 2005; 26(18); 3995.

http://dx:doi:10.1016/j.biomaterials.2004.10.012 .

- 47. Zhu MT, Wang B, Wang Y, Yuan L, Wang HJ, Wang M, et al. Endothelial dysfunction and inflammation induced by iron oxide nanoparticle exposure: Risk factors for early atherosclerosis. Toxicol Lett. 2011; 203(2): 162-171. http://dx:doi:10.1016/j.toxlet.2011.03.021.
- 48. Andrea G, Bing G, Rama SK, John CR, Ian MK, Abdul IB, Induction of Inflammation in Vascular Endothelial Cells by Metal Oxide Nanoparticles: Effect of Particle Composition. Environmental Health Persp. 2007; 115 (3): 403-409.
- 49. Park EJ, Choi J, Park YK, et al. Oxidative stress induced by cerium oxide nanoparticles in cultured BEAS-2B cells. Toxicol 2008; 245: 90-100.
- Andelman T, Gordonov S, Busto G, Moghe PV, Riman RE. Synthesis and Cytotoxicity of Y<sub>2</sub>O<sub>3</sub> Nanoparticles of Various Morphologies. Nanoscale Res Let. 2009; 5(2): 263-273.
- Xu C. Synthesis of Nanometer-sized Yttrium oxide Particles in Diisooctyl Sodium Sulphosuccinate (AOT)/Isooctane Reverse Micelle Solution. Blacksburg: Virginia Polytechnic Institute; 1999; 1-84.
- 52. Zhang Y, Nayak TR, Hong H, Cai W. Biomedical Applications of Zinc Oxide Nanomaterials. Cur Mol Med. 2013; 13(10): 1633-1644.
- 53. Nanotechnologies PoE: Consumer Products Inventory. In Retrieved [March, 2015], from http://www.nanotechproject.org/cpi.
- 54. Wijnhoven SWP, Peijnenburg WJGM, Herberts CA, Hagens WI, Oomen AG, Heugens EHW, et al. Nano-silver - a review of available data and knowledge gaps in human and environmental risk assessment. Nanotoxicol 2009; 3: 109-138.
- 55. Braakhuis HM, Ilse G, Petra K, John AFB, Flemming RC, Paul HBF, et al. Particle size dependent deposition and pulmonary inflammation after short-term inhalation of silver nanoparticles. Part Fibre Toxicol. 2014; 11: 49.
- Aulakh TS, Jayasekera N, Kuiper JH, Richardson JB. Long-term clinical outcomes following the use of synthetic hydroxyapatite and bone graft in impaction in revision hip arthroplasty. Biomaterials. 2009; 30(9): 1732–1738.
- 57. Zhou H, Lee J. Nanoscale hydroxyapatite particles for bone tissue engineering. Acta Biomater. 2011; 7(7): 2769-2781.
- Kadono H, Furuzono T, Masuda M, et al. In vivo evaluation of hydroxyapatite nanocoating on polyester artificial vascular grafts and possibility as soft-tissue compatible material. ASAIO J. 2010; 56(1): 61-66.

- 59. Ashokan A, Chandran P, Sadanandan AR, et al. Development and haematotoxicological evaluation of doped hydroxyapatite based multimodal nanocontrast agent for near-infrared, magnetic resonance and X-ray contrast imaging. Nanotoxicol. 2012; 6(6): 652-666.
- Xin L, Jiao S. Potential proinflammatory effects of hydroxyapatite nanoparticles on endothelial cells in a monocyte–endothelial cell coculture model. Int J Nanomed. 2014; 9:1261-1273. http://dx.doi.org/10.2147/IJN.S56298.
- 61. Capasso L, Camatini M, Gualtieri M. Nickel oxide nanoparticles induce inflammation and genotoxic effect in lung epithelial cells. Toxicol Lett. 2014 Apr 7; 226(1): 28-34. http://dx:doi:10.1016/j.toxlet.2014.01.040.