



THE EFFECT OF ANTICOAGULANTS, STORAGE TIME AND TEMPERATURE ON LIPOPROTEIN CONCENTRATIONS OF APPARENTLY HEALTHY SUBJECTS IN PORT HARCOURT, NIGERIA.

Ebirien-Agana Samuel Bartimaes^{1*}, Stella Urekweru Ken-Ezihuo² and Theodora Onyema Austin³

¹Senior Lecturer, Department of Medical Laboratory Science, Rivers State University of Science and Technology, P.M.B. 5080, Nkpolu-Oroworukwo, Port Harcourt, Nigeria.

²Lecturer 2, Department of Medical Laboratory Science, Rivers State University of Science and Technology, P.M.B. 5080, Nkpolu-Oroworukwo, Port Harcourt, Nigeria.

³Research Student, Department of Medical Laboratory Science, Rivers State University of Science and Technology, P.M.B. 5080, Nkpolu-Oroworukwo, Port Harcourt, Nigeria.

***Corresponding Author: Dr. Ebirien-Agana Samuel Bartimaes**

Senior Lecturer, Department of Medical Laboratory Science, Rivers State University of Science and Technology, P.M.B. 5080, Nkpolu-Oroworukwo, Port Harcourt, Nigeria.

Article Received on 18/11/2016

Article Revised on 09/12/2016

Article Accepted on 30/12/2016

ABSTRACT

Background: This study was designed to investigate the effect of anticoagulants (lithium heparin and fluoride oxalate) on lipoprotein levels obtained from samples which were analyzed within 2h after collection (control), stored at -4°C for one week (168h), 25°C for one week (168h), -4°C for two weeks (336h) and 25°C for two weeks (336h). 30 apparently healthy subjects ranging between the ages of 23 to 75 years participated in the study. **Materials and Methods:** Standard analytical procedures were used to determine the levels of the lipoproteins in the sample. **Results:** The results of the means \pm SD of the plasma samples in the different anticoagulants stored for both 1 week and 2 weeks at -4°C and 25°C for TC, TG, HDL-C and LDL-C showed significant differences ($p < 0.05$) decrease and deterioration when compared with the controls. There was no significant difference ($p > 0.05$) in the lipoprotein concentrations between lithium heparin and fluoride oxalate controls suggesting that lipoprotein determinations can be performed using fluoride oxalate as anticoagulant if the samples are to be analyzed immediately after collection. **Conclusion:** Anticoagulants, length and storage temperature, affect dramatically the measurement of total cholesterol, triglycerides, high density lipoprotein and low density lipoproteins.

KEYWORDS: Lipoprotein, lithium heparin, fluoride oxalate, anticoagulants, storage, temperature.

INTRODUCTION

Estimation of cardiovascular risk has become the cornerstone of cardiovascular prevention and management and this is achieved by the measurement of body lipids and lipoproteins.^[1] Lipoproteins are complex aggregates of lipids and proteins that render the lipids compatible with the aqueous environment of body fluids and enable their transport throughout the body to tissues where they are required.^[2] There are different types of lipoprotein molecules namely; low-density lipoprotein cholesterol, high-density lipoprotein cholesterol, triglycerides, total cholesterol and chylomicrons. Serum or plasma, free of haemolysis is the ideal specimen for the analysis of lipoproteins.^[3]

An anticoagulant is a substance that prevents the clotting of blood. Anticoagulants can be used endogenously or/and exogenously. The exogenous anticoagulants are compounds that have been developed using several mechanisms of action.^[4] Anticoagulants exert osmotic effects in which water leaves the cells and enters the plasma, thus diluting the plasma and lowering the

concentrations of non-diffusible components. The magnitude of this effect depends on the anticoagulant used and its concentration. Heparin, ethylenediamine tetra-acetate (EDTA) and fluoride oxalate are the mostly used anticoagulants in clinical chemistry.^[5,6] The heparin contained in the heparin tubes serves as blood anticoagulant by deterring the conversion of fibrinogen to fibrin through acceleration of the thrombin neutralizer antithrombin 111. Due to calcium's integral role in the blood coagulation process, oxalate contained in the fluoride oxalate tubes function as anticoagulant by sequestering calcium through the formation of insoluble complexes and chelates of calcium.^[7,8]

It has been reported that improper use of anticoagulant sample tubes, improper storage time and temperature, such as inadequate refrigeration of samples can lead to inaccurate test results.^[9] According to the WHO, MONICA project^[10], isolation of HDL-C should be done on fresh aliquots on the day of blood collection. A study by Shih et al.^[11], recommended that the total cholesterol

and high density lipoprotein-cholesterol levels should be assessed on the day of sample collection.

As part of the laboratory accreditation process and to address the pre-analytical requirements of ISO 15189^[12] it was necessary to investigate the effect of different pre-analytical storage conditions of blood samples. In literature, the stability of numerous analytes following prolonged contact of serum with cells has been described.^[13,14] To our knowledge, no studies have been conducted on the investigation of the stability of lipoproteins at different laboratory temperatures, storage times and in different anticoagulant tubes in Nigeria. This study was therefore designed to provide information on:

- A. The effect of storage times (delays before analysis) (2h, 1 week-168hr), and 2 weeks- 336hr),
- B. The effect of different storage temperatures (4°C, +/-2 or room temperatures (RT) 25°C, +/-2),
- C. The effect of choice of anticoagulant tubes (lithium heparin, and fluoride oxalate) and
- D. The effect of analysis within 2hr of storage versus delays before analysis on the levels of total cholesterol (TC), triglycerides (TG), high density lipoprotein cholesterol (HDL-C) and low density lipoprotein cholesterol (LDL-C) in blood samples collected from apparently normal subjects in Port Harcourt, Nigeria.

MATERIALS AND METHODS

Apparently 30 healthy male volunteers at the University of Port Harcourt teaching Hospital (UPTH), Port Harcourt were recruited into the after approval by the University of Port Harcourt Teaching Hospital (UPTH) Local Ethics Committee. Whole blood samples (10ml each) were collected through clean venepuncture after a 12 hour fast into well-sealed screw-capped sample bottles which helped to prevent oxidation of the unsaturated lipids. 2.5ml of this were dispensed into commonly used fluoride oxalate and lithium heparin anticoagulant sample bottles in Nigeria and centrifuged at 3000rpm for 5 minutes immediately after collection. Fresh plasma supernatants were analyzed at room temperature (25°C +/-2) within 2hr after centrifugation and the samples were then stored at room (25°C +/-2) for 1 week (168hr) and 2 weeks (336hr) and freezer temperatures (-4°C +/-2) for 1 week (168hr) and 2 weeks (336hr) respectively before being analyzed for total cholesterol (TC), triglycerides (TG), high density lipoprotein cholesterol (HDL-C) and low density lipoprotein cholesterol (LDL-C) respectively.

The enzymatic procedure for total cholesterol determination in serum based upon the Trinder^[15] method as modified by the Centers for Disease Control and Prevention was used. The method of Lopes-Virella *et al*^[16] for the determination of high-density cholesterol in serum was employed. The Friedewald *et al.*^[17]

equation was used to calculate the LDL-cholesterol in mmol/L.

$$\text{LDL-C (mmol/L)} = \text{Total Cholesterol} - \text{TG}/2.2 - \text{HDL-C}$$

The data obtained from the study were analyzed using the GraphPad InStat Version 3.10, 12 bit for Windows. The analyses performed included the computation of the means and standard deviation. Comparison of the means was done using the one-way analysis of variance (ANOVA). The Dunnett's comparison test was used to verify significant differences between the means of the parameters at the 2hr storage at 25°C and those stored at the various storage temperatures and times. The differences in mean were considered significant at $p < 0.05$.

RESULTS

The means \pm SD (mmol/L) of the total cholesterol (TC), triglycerides (TG), high density lipoprotein (HDL) and low density lipoprotein (LDL) of the samples collected into lithium heparin anticoagulant and analyzed within 2hr (control), stored at -4°C for 168 hours (1 week), 25°C for 168 hours (1 week), -4°C for 366 hours (2 weeks) and 25°C for 366 hours (2 weeks) is shown in table 1. The mean of the total cholesterol concentration decreased significantly ($p < 0.05$, $F = 54.00$) from 4.78 ± 1.13 mmol/L at within 2hr at 25°C of storage time and temperature to 1.88 ± 0.86 mmol/L at 336hr (2 weeks) at 25°C. Similarly, significant decrease in means were also observed in the concentrations of triglycerides ($p < 0.05$, $F = 98.41$) and high density lipoprotein ($p < 0.05$, $F = 9.33$) from 1.41 ± 0.28 mmol/L and 1.69 ± 0.70 mmol/L at within 2hr at 25°C of storage time and temperature to 0.53 ± 0.15 mmol/L and 1.11 ± 0.59 mmol/L at 336hr (2 weeks) at 25°C respectively. The LDL concentration also showed similar significant ($p < 0.05$, $F = 49.74$) decrease from 2.45 ± 0.61 mmol/L at within 2hr at 25°C of storage time and temperature to 0.51 ± 0.47 mmol/L at 336hr (2 weeks) at 25°C respectively.

The effect of storage temperatures and time on the total cholesterol, triglycerides, high density lipoprotein and low density lipoprotein concentrations in the samples collected in lithium heparin was investigated using the Dunnett's multiple comparison test. This involved the comparison of the lipoprotein means from the experimental temperatures and time of storage against the mean of the control samples. The result is shown in table 2. The result showed that significant differences ($p < 0.05$) exist between the means of samples stored at 25°C for 2hr (control) and those stored at the other experimental storage temperatures and times for total cholesterol, triglycerides, high density lipoprotein and low density lipoprotein. However, for the data obtained for the high-density lipoprotein and low density lipoprotein, the comparison of the means of the control sample against the means of the samples stored at -4°C for 168hr did not show significant difference ($p > 0.05$). Graphical representation of these is shown in fig. 1,2,3,4.

Table 1: Mean \pm SD of lipoprotein concentrations stored lithium heparin anticoagulant at various storage times and temperatures

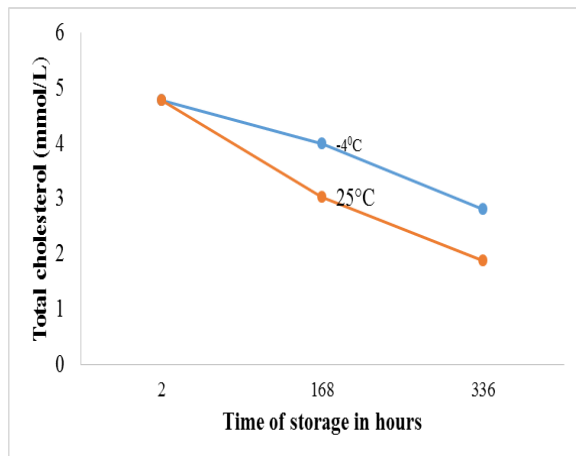
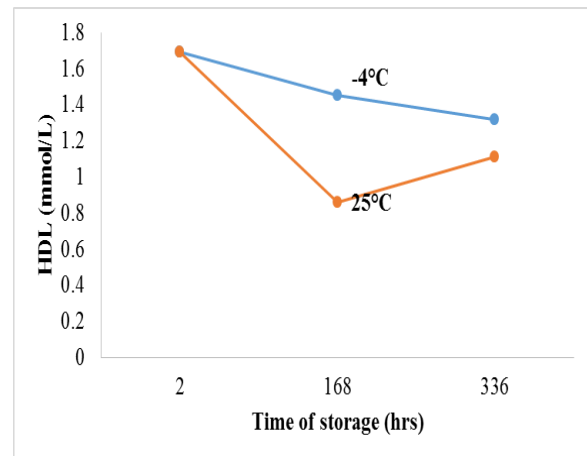
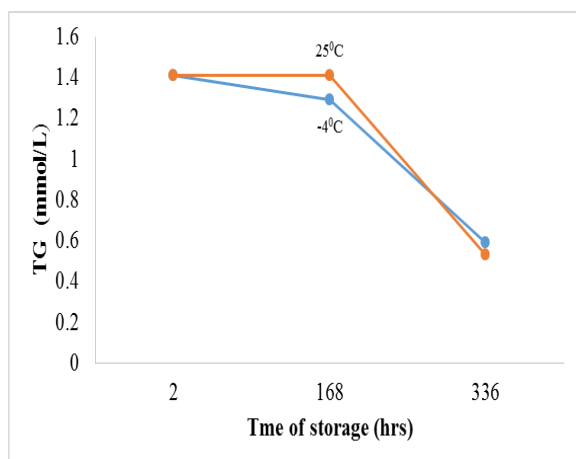
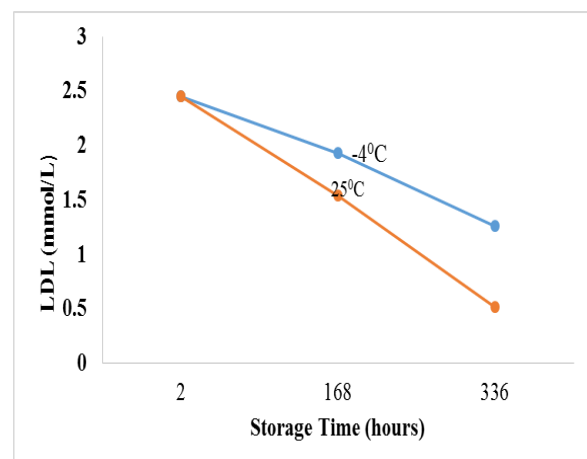
Storage Times/Temperatures	Lipoproteins			
	TC (mmol/L)	TG (mmol/L)	HDL (mmol/L)	LDL (mmol/L)
2hr at 25 ⁰ C (Control)	4.78 \pm 1.13	1.41 \pm 0.28	1.69 \pm 0.70	2.45 \pm 0.61
168hr at -4 ⁰ C	3.99 \pm 0.67	1.29 \pm 0.29	1.45 \pm 0.46	1.93 \pm 0.48
168hr at 25 ⁰ C	3.03 \pm 0.84	1.41 \pm 0.30	0.86 \pm 0.57	1.54 \pm 0.78
336hr at -4 ⁰ C	2.80 \pm 0.57	0.59 \pm 0.17	1.32 \pm 0.49	1.26 \pm 0.41
336hr at 25 ⁰ C	1.88 \pm 0.86	0.53 \pm 0.15	1.11 \pm 0.59	0.51 \pm 0.47
P-value	p<0.05, F=54.00	p<0.05, F=98.41	p<0.05, F=9.33	p<0.05, F=49.74

Note: TC represents total cholesterol, TG = triglycerides, HDL = high density lipoprotein and LDL = low density lipoprotein.

Table 2: Results of Dunnett's multiple comparison test for means of lipoproteins in lithium heparin anticoagulant

Storage Times/Temperatures	Lipoproteins							
	TC		TG		HDL		LDL	
	p-value	q-value	p-value	q-value	p-value	q-value	p-value	q-value
Control. vs storage for 168hr at -4 ⁰ C	p<0.05	3.87	p<0.05	7.69	NS	1.65	NS	2.32
Control vs. storage for 168hr at 25 ⁰ C	p<0.05	6.46	p<0.05	9.72	p<0.05	5.65	p<0.05	3.05
Control vs. storage for 336hr at -4 ⁰ C	p<0.05	9.76	p<0.05	10.31	p<0.05	2.57	p<0.05	7.33
Control vs storage for 336hr at 25 ⁰ C	p<0.05	12.87	p<0.05	11.90	p<0.05	3.96	p<0.05	10.47

NS represents not significant

**Fig. 1: Effect of time of storage and temperature on TC concentration in lithium heparin****Figure 3: Effect of time of storage and temperature on HDL concentration in lithium heparin****Figure 2: Effect of time of storage and temperature on TG concentration in lithium heparin****Figure 4: Effect of time of storage and temperature on LDL concentration in lithium heparin**

Similarly, the mean \pm SD (mmol/L) of the total cholesterol (TC), triglycerides (TG), high density lipoprotein (HDL) and low density lipoprotein (LDL) of the samples collected and stored in fluoride oxalate anticoagulant and analyzed within 2hr (control), stored at -4°C for 168 hours (1 week), 25°C for 168 hours (1 week), -4°C for 366 hours (2 weeks) and 25° for 366 hours (2 weeks) before analysis is shown in table 3. The table also showed significant ($p < 0.05$) progressive decrease in the means of total cholesterol, triglycerides, high density lipoprotein and low density lipoproteins from 2hr storage time to 338hr and 25°C and -4°C respectively. The effect of storage temperatures and time on the total cholesterol, triglycerides, high density lipoprotein and low density lipoprotein concentrations on the samples collected in fluoride oxalate was also investigated using the Dunnett's multiple comparison test (table 4). The result showed that significant differences ($p < 0.05$) exist between the means of samples stored at 25°C for 2hr (control) in fluoride oxalate and those stored at the other experimental storage temperatures and times for total cholesterol, triglycerides, high density lipoprotein and low density lipoprotein. However, with respect to high-density lipoprotein and low density lipoprotein, the comparison of the mean of the control sample against the mean of the samples stored at -4°C for 168hr did not show significant difference ($p > 0.05$). Graphical representation of these is shown in fig. 5,6,7,8. The effect of the anticoagulants (lithium heparin and fluoride oxalate) on lipoprotein concentrations in the control samples which were stored for 2 hours at 25°C did not show any significant difference ($p > 0.05$) (table 5) although the means of the samples stored in lithium heparin anticoagulant were higher than that obtained for samples stored in fluoride oxalate.

DISCUSSION

Laboratory evaluation of patients is an integral part of diagnostic procedure. Although serum is used for most assays, plasma is a useful alternative due to its rapid processing time.^[18] Plasma which was obtained with an appropriate anticoagulant has been reported to be an equally valid sample and in certain conditions preferable to serum.^[19] Anticoagulants used to preserve analytes may interfere with other analyte determinations when using plasma^[5] and changes as little as possible in the concentration of substances to be measured has been reported to occur before the analytical processes.^[20] Results obtained from this study show that storage temperature and duration affects the concentration of TC, TG, HDL and LDL in the samples stored in both lithium heparin and fluoride oxalate anticoagulants. The length of time of storage also has effects on the levels of the analytes obtained in the study. The concentration of the analytes deteriorated considerably as the duration of storage was increased. The samples stored in lithium heparin show a percentage decrease of 60.67% for TC, 62.41% for TG, 34.32% for HDL and 79.18% for LDL while the percentage decrease in analyte concentration

stored in fluoride oxalate show a percentage decrease of 65.97% for TC, 60.00% for TG, 36.42% for HDL and 86.30% for LDL respectively (table 1 and 3). The finding in this study does agree with the result of Ignatius *et al.*^[21] in respect of TC and HDL who observed increase in these parameters even after 16 days (384 hours) of storage. Prolonged power failure, a very common experience in the developing countries has been reported to affect sample storage at various temperatures (for example samples stored at refrigerator and freezer temperatures).^[21] In a related study, storage time and temperature were also reported to affect measurable concentration of progesterone in the blood of zebu cows.^[22] In effect, storage time considerably affected the concentration of the lipoproteins in the sample.

At room temperature (25°C) and at 2-hour storage, the concentration of the analytes in the fluoride oxalate anticoagulants decreased appreciably when compared with the levels in the lithium heparin anticoagulants (table 5). Lithium heparin has been generally recommended as the most suitable anticoagulant for plasma biochemical measurements.^[23,6] Although serum and heparinized plasma specimens are considered equivalent for many assays, differences in results between these two sample types have been reported for several chemistry analytes.^[18] Oxalate inhibit several enzymes, such as amylase, lactate dehydrogenase, and acid and alkaline phosphatase.^[24] Fluoride may also interfere with electrolyte measurements by altering cell permeability and promoting haemolysis by red blood cell ATP with subsequent potassium efflux.^[24] Results of our study showed that all the means of the measured parameters were not significantly different ($p > 0.05$) among the sample types stored at 25°C for 2hours in the lithium heparin and fluoride although the values obtained were reduced in the fluoride oxalate samples. The study by Nwangwu *et al.*^[25] reported that lithium heparin has a stabilizing effect on blood sample stored for 2 hours and may not likely cause alteration in the concentrations of most biochemical parameters measured in humans. The results of this study on lipoprotein concentrations as the measured parameters in human plasma stored for 2 h in lithium heparin thus agrees with the findings of Nwangwu *et al.*^[25]

In conclusion, anticoagulants, length and storage temperature, affect dramatically the measurement of total cholesterol, triglycerides, high density lipoprotein and low density lipoproteins. In experimental planning, it is advisable to take into account the effect of these factors, in addition to sample analysis and reported interpretations of these data in literature.

Table 3: Mean ±SD of Lipoprotein Concentrations Stored Fluoride Oxalate Anticoagulant at Various Storage Times and Temperatures

Storage Times/Temperatures	Lipoproteins			
	TC (mmol/L)	TG (mmol/L)	HDL (mmol/L)	LDL (mmol/L)
2hr at 25 ⁰ C	4.32±1.04	1.30±0.30	1.51±0.77	2.19±0.62
168hr at -4 ⁰ C	3.46±0.70	0.67±0.30	0.94±0.60	1.68±0.71
168hr at 25 ⁰ C	2.89±0.92	0.67±0.30	0.94±0.60	1.62±0.71
336hr at -4 ⁰ C	2.16±0.68	0.63±0.20	1.01±0.51	0.87±0.63
336hr at 25 ⁰ C	1.47±0.89	0.52±0.20	0.96±0.51	0.30±0.82
P-value	p<0.05	p<0.05	p<0.05	p<0.05

Note: TC represents total cholesterol, TG = triglycerides, HDL = high density lipoprotein and LDL = low density lipoprotein.

Table 4: Results of Dunnett’s multiple comparison test for means of lipoproteins in fluoride oxalate anticoagulant

Storage Times/Temperatures	Lipoproteins							
	TC		TG		HDL		LDL	
	p-value	q-value	p-value	q-value	p-value	q-value	p-value	q-value
Control. vs storage for 168hr at -4 ⁰ C	p<0.05	3.86	p<0.05	7.70	NS	1.70	NS	2.32
Control vs. storage for 168hr at 25 ⁰ C	p<0.05	6.46	p<0.05	9.72	p<0.05	3.56	p<0.05	4.05
Control vs. storage for 336hr at -4 ⁰ C	p<0.05	9.75	p<0.05	10.31	p<0.05	3.15	p<0.05	6.33
Control vs storage for 336hr at 25 ⁰ C	p<0.05	12.80	p<0.05	11.90	p<0.05	3.45	p<0.05	9.47

NS represents not significant

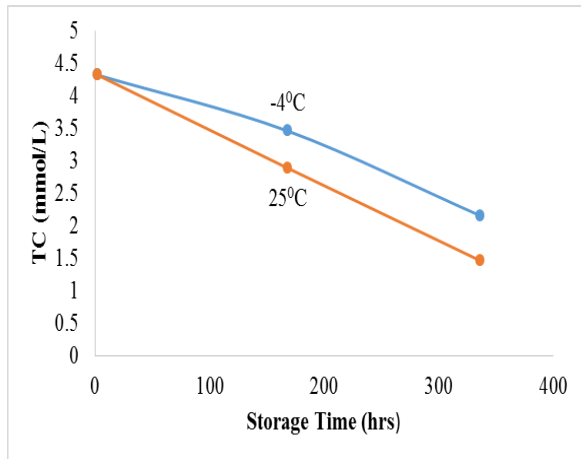


Figure 5: Effect of Time of Storage and Temperature on TC Concentration in Fluoride Oxalate

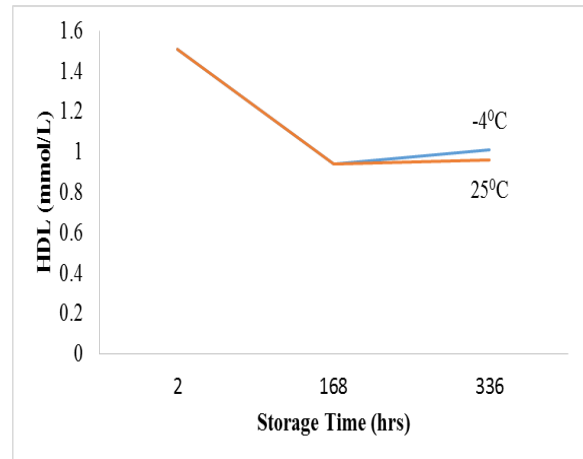


Figure 7: Effect of Time of Storage and Temperature on HDL Concentration in Fluoride Oxalate

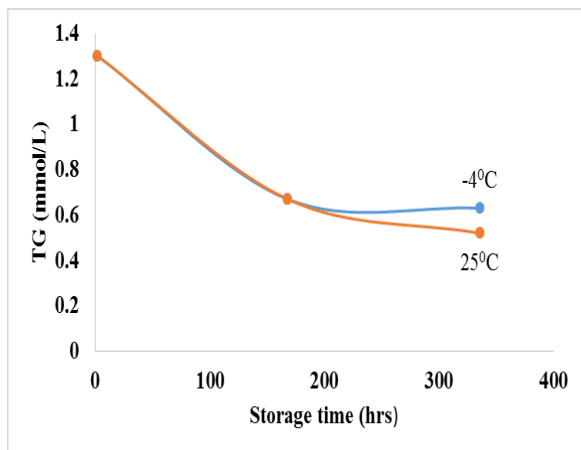


Figure 6: Effect of Time of Storage and Temperature on TG Concentration in Fluoride Oxalate

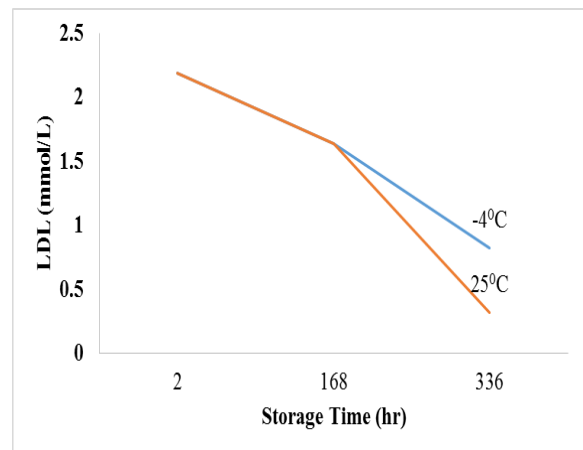


Figure 8: Effect of time of storage and Temperature on LDL Concentration in Fluoride Oxalate

Table 5: Mean \pm SD of lipoprotein concentrations in plasma stored for 2hr at 25⁰C in lithium heparin and fluoride oxalate anticoagulants

Anticoagulants	Lipoproteins			
	TC (mmol/L)	TG (mmol/L)	HDL (mmol/L)	LDL (mmol/L)
Lithium heparin	4.78 \pm 1.13	1.42 \pm 0.28	1.69 \pm 0.70	2.45 \pm 0.61
Fluoride oxalate	4.32 \pm 1.04	1.30 \pm 0.31	1.51 \pm 0.77	2.19 \pm 0.62
p-value	ns	ns	ns	ns

Note: TC represents total cholesterol, TG= triglycerides, HDL= high density lipoprotein, LDL= low density lipoprotein.

REFERENCES

- Burtis, CA, Ashwood, R, Aldrich, JE. Tietz Fundamentals of Clinical Chemistry, 5th Edn, .W.B. Saunders and Co., 2001; 780-94.
- Rodenburg KW, Van der Horst DJ. Lipoprotein-mediated lipid transport in insects: analogy to the mammalian lipid carrier system and novel concepts for the functioning of LDL receptor family members. *Biochim Biophys Acta*, Sep 5, 2005; 1736(1): 10-29.
- Myers, GL, Kimberly, MM, Smith, SJ. A reference method laboratory network for cholesterol: A model for standardization and improvement of Clinical laboratory measurements. *Clin. Chem.*, 2000; 46: 1762-72.
- Schrezenmeier, H. Anticoagulant-induced pseudo-thrombocytopenia and pseudo-leucocytosis. *Thromb. Haemost.*, 1995; 73: 566-13.
- Sevastos N, Theodossiades G, Efstathiou S, Papatheodoridis GV, Manesis E, Archimandritis AJ. Pseudo-hyperkalemia in serum: the phenomenon and its clinical magnitude. *J Lab Clin Med.*, 2006; 147: 139-44. <http://dx.doi.org/10.1016/j.lab.2005.11.008>.
- Young DS, Bermes EW, Haverstick DM. Specimen collection and processing. In: Burtis CA, Ashwood ER, Bruns DE, eds. *Tietz Textbook of Clinical Chemistry and Molecular Diagnostics*. 4th Ed. St. Louis: Elsevier Saunders; 2006.
- Horton HR, Moran, LA, Ochs RS, Rawn JD, Scrimgeour KG. *Principles of Biochemistry*, 2nd Edition. Prentice-Hall, Upper Saddle River, NJ, 1996; 801.
- Sacks DB, 1999. Carbohydrates. In: Burtis, C.A., Ashwood, E.R.(Eds.), *Tietz Textbook of Clinical Chemistry*, 3rd edn. Saunders, Philadelphia, PA, USA, 1999; 750-08.
- Evans K, Mitcheson J, Laker M. Effect of storage at -4 C and - 20 on lipid, lipoprotein and apolipoprotein concentrations. *J Clin Chem.*, 1995; 41: 392-6.
- Tunstall-Pedoe H, for the WHO MONICA Project. The World Health Organization MONICA Project (Monitoring Trends and Pajak. Determinants in Cardiovascular Disease): a major international collaboration. *Journal of Clinical Epidemiology*, 1988, 41: 105-114. PMID: 3335877. MONICA Publication 2.
- Shih WJ, Bachorik PS, Haga, JA, Myers GL, Stein EA. Estimating the long-term effects of storage at -70⁰ C on cholesterol, triglyceride, and HDL-cholesterol measurements in stored sera. *Clin. Chem.*, 2000; 46: 351-64.
- ISO 15189. *Medical laboratories- particular requirements for quality and competence*, 2007.
- Heins M, Heil W, Withold W. Storage of serum and whole blood samples. Effect of time and temperature on 22 serum analytes. *Eur J Clin Chem Biochem*, 1995; 33: 231-8.
- Rehak NN, Chlang BT. Storage of whole blood. Effect of temperature on the measured concentrations of analytes in serum. *Clin Chem.*, 1988; 34(10): 211-214.
- Trinder P. Determination of glucose in blood using glucose Oxidase with an alternative oxygen receptor. *J Analyt Clin Biochem*, 1969; 6: 24.
- Lopez- Virella MF, Stone P, Eltis S, Colwell JA (1977). Cholesterol determination in HDL separated by three different methods. *Clin Chem.*, 1977; 23: 822-84.
- Friedewald MT, Lopes VF, Levy RT. Estimation of the concentration of low-density lipoprotein cholesterol in plasma without the use of ultracentrifugation. *Chem Chem*, 1972; 18: 499-02.
- Ladenson JH, Tsai LMB, Michael JM. Serum versus heparinized plasma for eighteen common chemistry tests. *Am J Clin Pathol*, 1974; 62: 545-52.
- Young DS, Bermes EW. Specimen collection and processing: sources of biological variation. In: C.A. Burtis and A.R. Ashwood, Editors, *Tietz Textbook of Clinical Chemistry* (3rd ed.), W.B. Saunders, Philadelphia, 1999; 4272.
- Guder WG. 2001. The quality of diagnostic samples, *Blood Gas News*, 10: 18-24.
- Ignatius, MC, Emeka NE, Ebele IJ, Chinelo MV, Fidelis EE, Silas UA, The effect of sample storage total and HDL cholesterol assays. *Curr Res J Bio Sci.*, 2009; 1(2): 1-5.
- Inns JH, Cecchini G, Mattoni, M. The effect of anticoagulant, storage time and time and temperature, and sodium azide on blood progesterone concentrations. *International Livestock Centre for Africa*, 1988; 1-11.
- Mohri M, Rezapoor H. Effects of heparin, citrate, and EDTA on plasma biochemistry of sheep: comparison with serum, *Res. Vet. Sci.*, 2009; 86: 111-4.

24. Narayanan S. The preanalytic phase: an important component of laboratory medicine. *Am J Clin Pathol*, 2000; 113: 429-452.
25. <http://dx.doi.org/10.1309/CONM-Q7R0-LL2E-B3UY>.
26. Nwagwu CO, Spencer JJ, Sunday OK, Erifeta OG, Asuk AA, Uhunmwangho SE and Jenevieve O. Comparative stabilizing effects of some anticoagulants on fasting blood glucose of diabetics and non-diabetics, determined by spectrophotometry (glucose oxidase). *Asisn J Med. Sc.*, 2011; 3(6): 234-236.