

ANTIMICROBIAL SCREENING OF CRUDE EXTRACT OF PLUCHEA ARABICA: CHEMICAL COMPONENTS, DRUG LIKENESS, PHYSICOCHEMICAL PROPERTY AND MOLECULAR DOCKING ASSESSMENT

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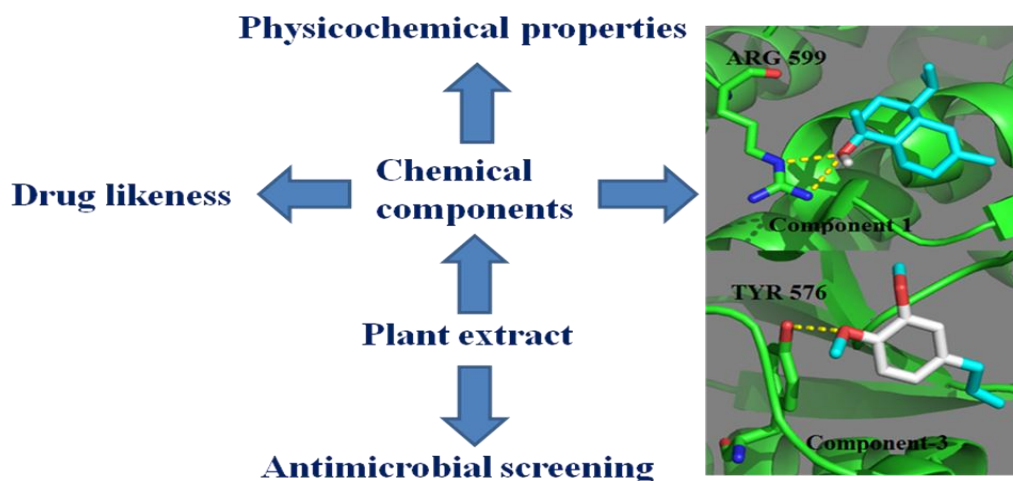
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ABSTRACT

Objective: To explore the antimicrobial efficacy of the crude extract of *Pluchea Arabica*, drug likeness, physicochemical and molecular studies of its components.

Methods: The crude extract was obtained applying soxhelt extraction assembly and the antimicrobial screening was performed using disc diffusion method. The drug likeness and physicochemical were investigated by employing the available software on the web (www.molinspiration.com) and molecular docking studies was carried out by Auto Dock Tools-4.2.



Results: The methanol extract possess significant activity in comparison to the n-hexane extract. All the components following the Lipinski rule, out of all three components showed good bioactivity score in which two (1 & 3) mainly belongs to methanol extract and hydrogen bonding to the receptor also observed with these two component on molecular docking assessment.

Conclusion: The crude extract of *Pluchea Arabica* was screened for antimicrobial evaluation and its components were subjected for calculation of computational parameters and molecular docking studies. Methanol extract of this is found to possess better antimicrobial potential than the n-hexane and the computational studies strongly supported the experimental results.

KEYWORDS: *Pluchea arabica* extract, antimicrobial and computational studies.

INTRODUCTION

Resistance to present antimicrobial drugs has become a major public health concern globally^[1]. The microbial diseases are amplified mainly due to indiscriminate use of antibiotics that leads to the antibiotic resistance^[2-3]. Mutation and genetic exchange systems revoked the elimination of disease and are responsible for antimicrobial resistance^[2-4]. Medicinal plants have been

widely investigated as the rich source of antimicrobial agents. A variety of medicinal plants possess chemical compounds representing different biological and pharmacological potentials. Numerous studies has been carried out on biological and pharmacological potential of plants, such as antimicrobial, antioxidant, antitumor, anti-inflammatory, hypoglycemic, etc.^[5-10]. *Pluchea*, corresponds to the family Asteraceae comprising eighty

species throughout the world mainly in African, American and Asian^[11]. The plant has been found to exhibit the variety of traditional pharmacological applications^[12-19]. The compounds found in the *Pluchea* are as follows- sesquiterpenoids, monoterpenes, lignan glycosides, triterpenoids, flavonoids, lignan glycosides, terpenic glycosides, tertiary bases, a large number of water-soluble quaternary bases, including pluchine 7-O-glucoside. The flavonoids include quercetin, isorhamnetin, hesperidin, a dihydroflavonol-taxilolin 3-arabinoside and an isoflavonoid-formononetin^[20-31]. In a recent study antioxidant properties of *Pluchea arabica* showed the inhibition of DPPH radical at 89-93%. Recently the fresh twigs of *Pluchea arabica* (Boiss) were analyzed for the main components such as δ -cadinol, 9-(1-methylethylidene)-bicyclo[6.1.0]nonane, caryophyllene oxide, methyleugenol and β -caryophyllene^[32]. It is crucial important to discover some new antimicrobial agents with different mode of action and less cytotoxicity considering the medicinally important natural resources which will help to replace the currently available antimicrobials^[33].

Experimental

Extraction

The extraction of the compounds from *Pluchea arabica* was prepared using Soxhlet extraction assembly, methanol and n-hexane solvent. The Fresh twigs of *Pluchea arabica* were grounded and kept inside a thimble loaded into the soxhlet extractor for 15 hr in the solvent (300 mL) with refluxing at the boiling temperature of solvent. After completion dried under vacuum applying Heidolph laborata/Germany. The obtained extract was incubated at 40 °C for 24 hr, after that stored at 4 °C evaluation for antimicrobial activity.

Antimicrobial activity

Antimicrobial activity of the extract was employed against all the gram negative and gram positive pathogens by the reported method. The dilutions were obtained as 3.125, 6.25, 12.5, 25 and 50 μ g/ml. Ciprofloxacin was taken as positive control and Methanol and n-hexane poured disk was used as negative control^[39-47].

Physicochemical properties

The components were subjected for physicochemical properties such as miLogP, TPSA, Natoms, MW, nON, nOHNH, Nviolations, Nrotb, Volume using the online available software (www. molinspiration.com)^[48-54].

Bioactivity score

The components were also subjected for the calculation of the bioactivity score of the components using the same software as for physicochemical properties at (www. molinspiration.com)^[48-54].

Molecular docking studies

The docking analysis for all the component was performed using GlcN-6-P synthase, (PDB: 2VF5)

following the procedure^[55-56]. The structures were made using ChemDraw Ultra 8.0 and were optimized for energy to yield 3D.

RESULTS

Antimicrobial activity

The antimicrobial activity of the extract for both portions methanol and n-hexane was done as the zone of inhibition was mapped with antibiotic zone scale and the findings are reported in the **Table-1, Table-2 and Table-3**.

The phytochemical screening

The reported literature confirmed that the *pluchea* extract has been found to possess

δ -cadinol (26.8%), 9-(1-methylethylidene)-bicyclo[6.1.0]nonane (10.8%), caryophyllene oxide (10.0%), methyleugenol (9.2%) and β -caryophyllene (6.9%) **Figure-1** shows the structures of all the components^[33].

Physicochemical properties^[34-38]

Lipinski's rule of five explained the general criteria for the activity of the drug such as the MW of the drug should not exceed than 500, that, in general, it should not possess more than five OH and NH groups and not ten N and O, the log P must less than 5 and number of violation less than 4. The components were obtained in agreement with Lipinski's rule of five except components three that exhibited the partition coefficient 5.17, the detailed results are reported in **Table 4**.

Bioactivity score^[34-38]

For a drug to be active it is the required condition that it should possess the bioactivity score more than 0.00, if it is up to -0.50 then moderate active and finally if it is less than -0.50 then it lies in the category of inactive drugs. The results for calculated bioactivity score are presented in **Table 5**. The results exhibited that the bioactivity score for component one and three and five are mostly lying under the zone of active and moderately active drugs but for component two and four mostly lying in inactive region, a little moderately active but nothing in active drugs range.

Molecular docking studies

The docking analysis for all the components of *P. Arabica* was performed using GlcN-6-P synthase, (PDB: 2VF5) The results for molecular docking studies are provided in **Figure-2, 3 and 4**, the results exhibited that the hydrogen bonding is found only in two components (component one and component three) of the extracts and both the components belongs to the methanol extract. The component one is forming hydrogen bond with ARG 599 and the component three with TYR 576 residue. The component three was found to exhibit resemblance with the standard drug ciprofloxacin with respect to the H-bond with TYR 576. Rest of the components belonging to n-hexane, have no hydrogen

bonding that provides strong recommendation to the obtained experimental results for antimicrobial assay.

Table: 1. Representing zone of inhibition for methanol extract of *Pluchea arabica* against gram positive and gram negative bacteria.

Compounds (µg/ml)	Effect of methanol extract on Microorganism			
	Gram positive		Gram negative	
	<i>S. aureus</i>	<i>S. epidermidis</i>	<i>P. mirabilis</i>	<i>E. coli</i>
50	19.30±.22	18.50±.34	15.32±.24	15.14±.11
25	17.14±.04	16.25±.22	10.42±.19	13.24±.12
12.5	11.12±.52	07.14±.40	-	11.52±.32
6.25	10.22±.32	-	-	11.08±.14
3.125	09.24±.18	-	-	9.14±.04

Table: 2. Representing zone of inhibition for n-hexane extract of *Pluchea arabica* against gram positive and gram negative bacteria.

Compounds (µg/ml)	Effect of n-Hexane extract on Microorganism			
	Gram positive		Gram negative	
	<i>S. aureus</i>	<i>S. epidermidis</i>	<i>P. mirabilis</i>	<i>E. coli</i>
50	12.30±.22	09±.34	08.32±.20	19.34±.11
25	10.14±.04	8.12±.22	-	9.18±.34
12.5	9.52±.52	-	-	7.24±.04
6.25	08.22±.32	-	-	-
3.125	-	-	-	-

Table: 3. Representing zone of inhibition for ciprofloxacin, methanol and hexane against gram positive and gram negative bacteria.

Microorganism			
	Ciprofloxacin (10 µg/ml)	Methanol	Hexane
<i>S. aureus</i>	21.46 ±.31	-	-
<i>S. epidermidis</i>	22.64±.54	-	-
<i>P. mirabilis</i>	22.24±.30	-	-
<i>E. coli</i>	23.82±.47	-	-

Table: 4. Representing the physicochemical properties of all the components found in *Pluchea arabica* and ciprofloxacin.

Physicochemical property score	Components					
	1	2	3	4	5	Ciprofloxacin
miLogP	4.97	2.41	5.17	5.00	4.14	-0.701
TPSA	20.23	18.47	0.00	0.00	12.53	74.569
Natoms	16	13	15	12	16	24.0
MW	222.37	178.23	204.36	164.29	220.36	331.347
nON	1	2	0	0	1	6
nOHNH	1	0	0	0	0	2
Nviolations	0	0	1	0	0	0
Nrotb	1	4	0	0	0	3
Volume	243.65	179.67	229.95	185.96	234.01	285.46

Table-5: Representing the bioactivity score of all the components found in *Pluchea arabica* and ciprofloxacin.

Bioactivity score	Components					
	1	2	3	4	5	Ciprpfloxacin
GPCR ligand	-0.09	-0.81	-0.34	-0.76	-0.08	0.12
Ion channel modulator	0.05	-0.38	0.28	-0.35	0.14	-0.04
Kinase inhibitor	-0.87	-1.06	-0.78	-1.23	-0.86	-0.07
Nuclear receptor ligand	0.39	-0.80	0.13	-0.63	0.62	-0.19
Protease inhibitor	-0.63	-1.14	-0.60	-1.00	0.00	-0.21
Enzyme inhibitor	0.40	-0.43	0.19	-0.30	0.57	0.28

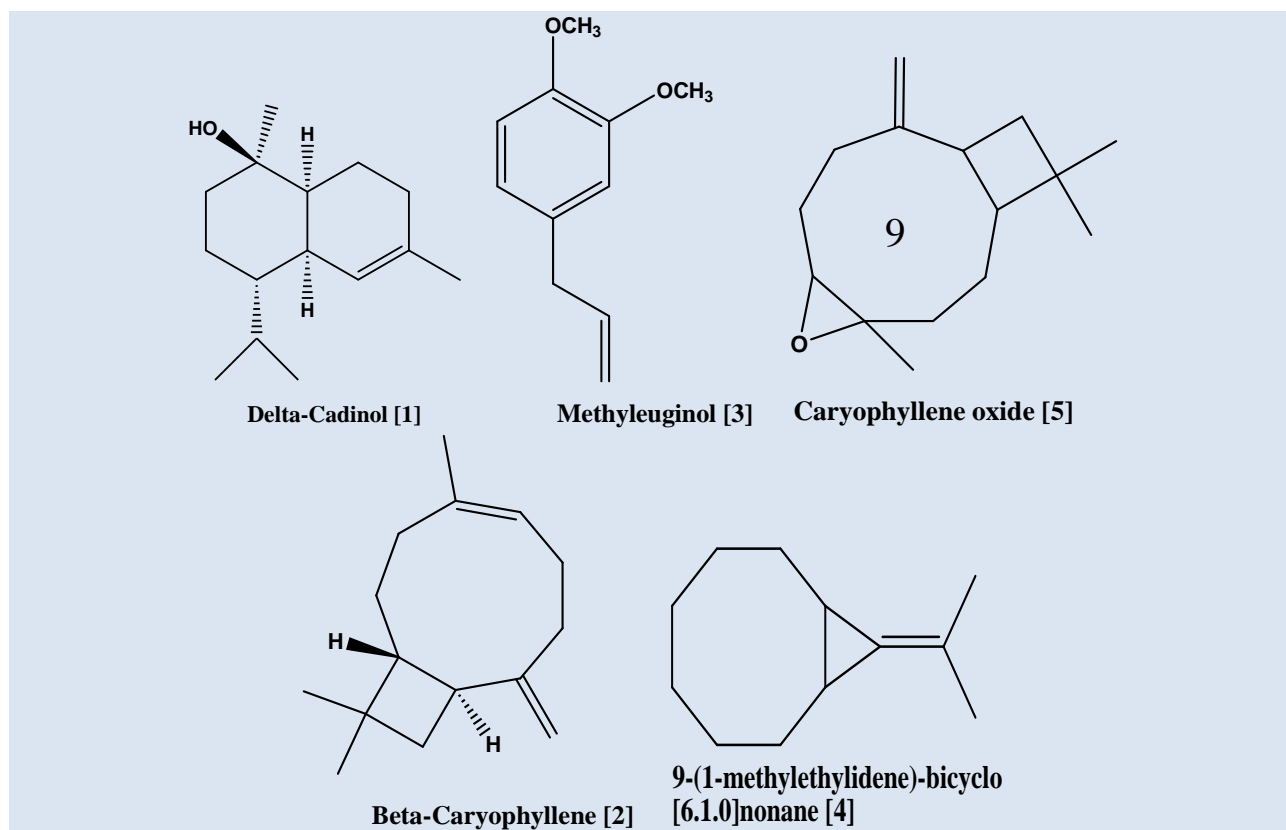
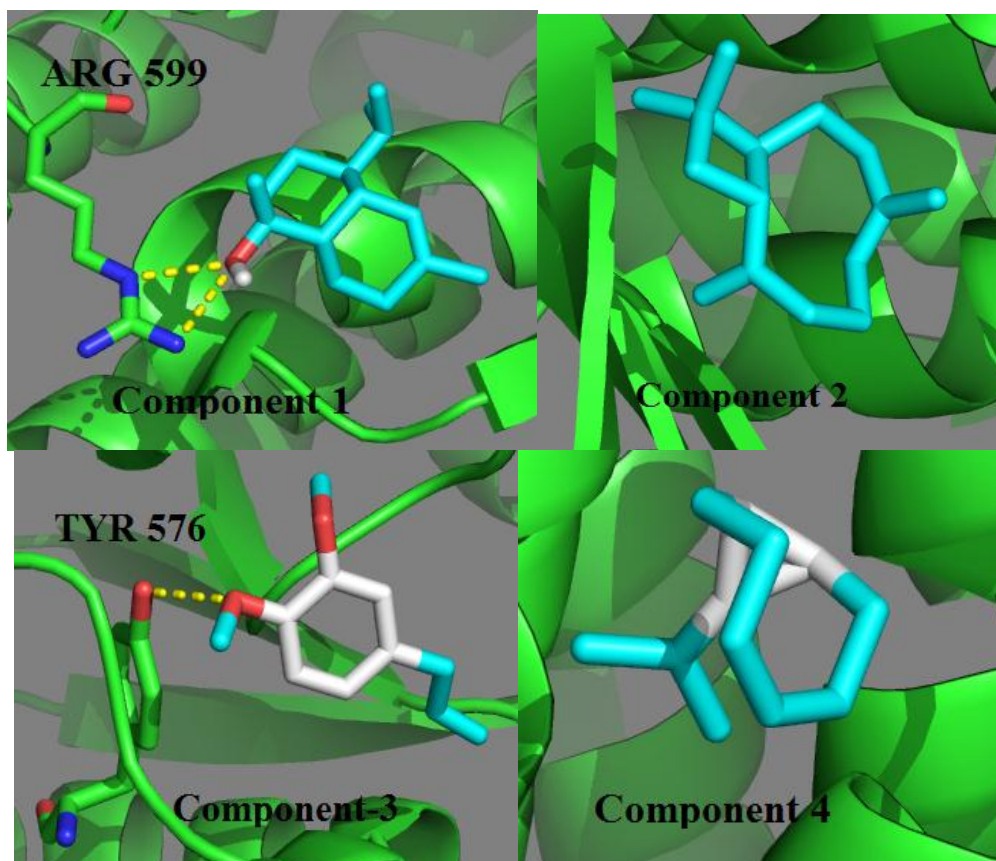


Figure-1- Exhibiting the structures of all the components found in *Pluchea arabica*.



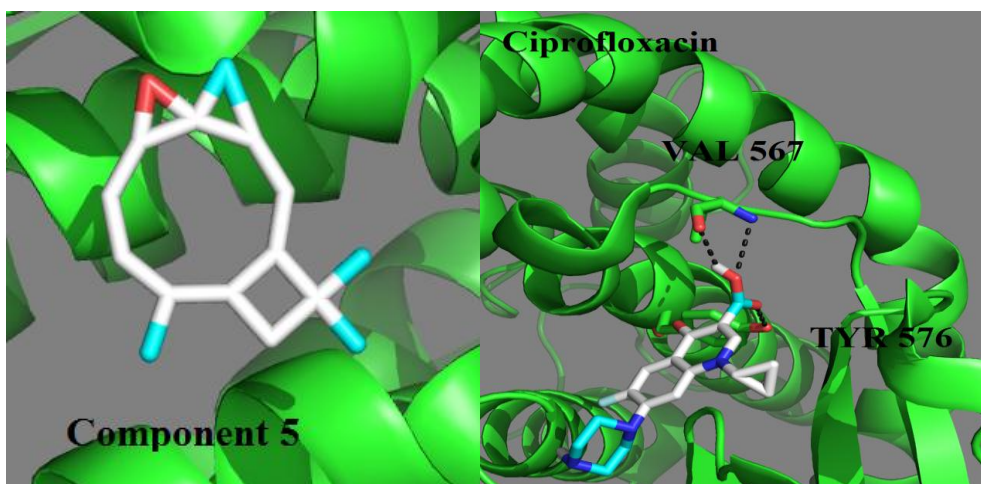


Figure-2- Representing the docked images of the synthesized compounds with the GlcN-6-P-synthase.

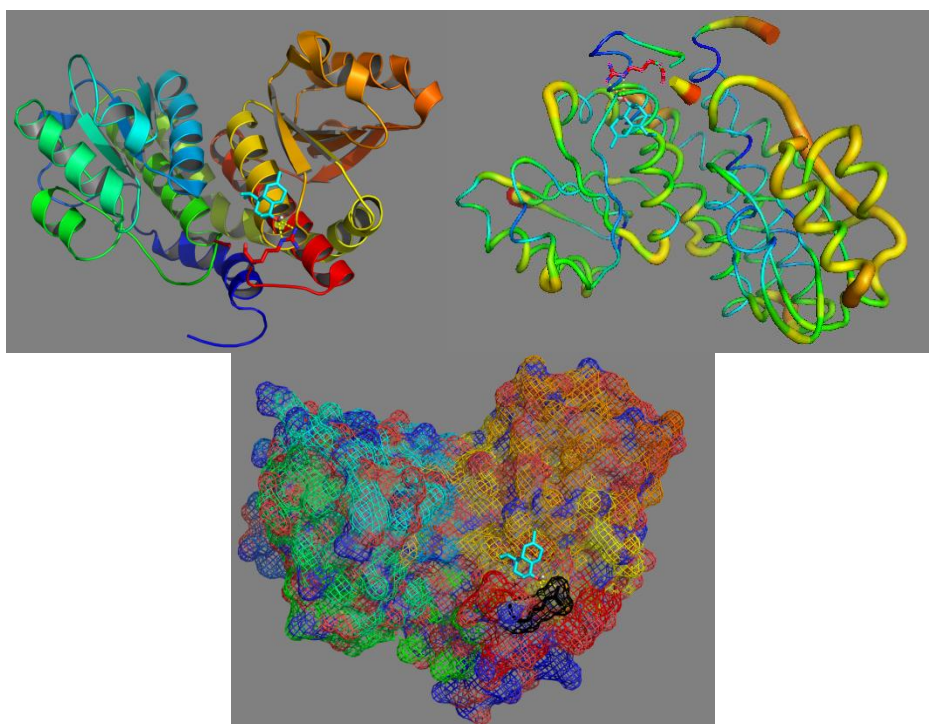
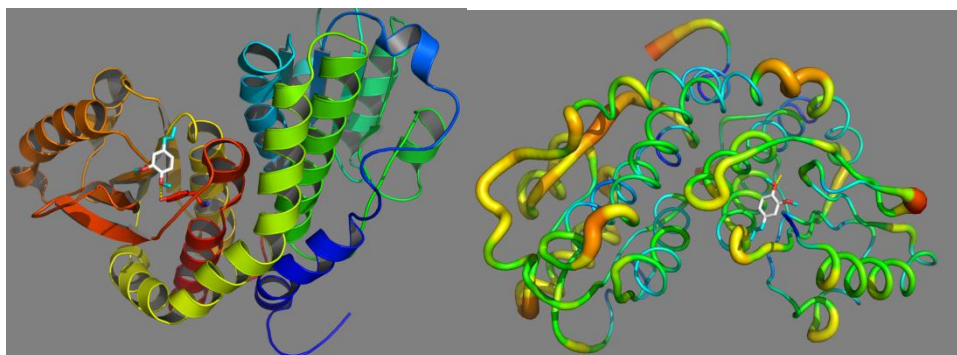


Figure-3- Exhibiting the different binding modes for component -1



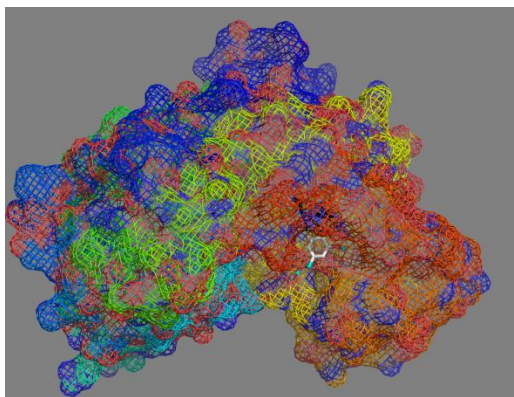


Figure-4- Exhibiting the different binding modes for component-3.

DISCUSSION

The crude extract of the plant *pluchea arabica* was obtained and subjected for antibacterial study. The results for antibacterial activity exhibited that methanol extract represented better antibacterial potential than n-hexane. Targeting the phytochemical screening were achieved on the basis of available literature which confirms the presence of components δ -cadinol and methyleugenol mainly in methanol extract^[32-33]. The elevated bioactivity might be because of the composition of methanol extract. Though to confirm the results obtained by phytochemical and antibacterial analysis we carried out the virtual screening of all the components present in the extract of *Pluchea Arabica*, calculated the physicochemical parameters and bioactivity score and found that results strongly supported the studies. To find out more clear picture regarding the study molecular docking studies were also carried out which provided strong recommendation that the antibacterial potential of methanol extract of *pluchea Arabica* is having better antibacterial potential than the n-hexane extract as the formation of hydrogen bond to the receptor is only observed in case of the components belongs to methanol extract.

CONCLUSION

The extraction of plant *Pluchea arabica* was performed and subjected for biological potential. The obtained results showed that the methanol extract of the plant contained better antimicrobial potential than the n-hexane extract. On the basis of available literature the plant components of *P. Arabica*, aimed for virtual screening to calculate the bioactivity, physicochemical and docking properties. The computational findings were found in strong recommendation to experimental results. Our study, the screening for antimicrobial potential in addition to the computational studies can be used as lead finding for the development of antimicrobial agents from natural sources.

Conflict of Interest

There is no conflict of interests.

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REFERENCES

1. Avila M, Said N, Ojcius D M. The book reopened on infectious diseases. *Microb. Infect.* 2008; 10: 942–947.
2. Triyana SY. Antibiotic resistance of pathogenic bacteria. *KESPHA* 2009; 1: 92–94.
3. Kumar VP, Chauhan, NS, Padh H, Rajani M. Search for antibacterial and antifungal agents from selected Indian medicinal plants. *J. Ethnopharmacol.* 2006; 107: 182–188.
4. Kin-Ying W, Paritala V, Kishore KC, Arifullah M. Phytochemical screening and antimicrobial potentials of *Borreria* sps (Rubiaceae), *Journal of King Saud University-Science.* 2015; 27: 302–311.
5. Kane JH, Finlay AC, Sobin BA. Antimicrobial agents from natural sources. *Ann. NY Acad. Sci.* 1950; 53: 226–228.
6. Gyawali R, Salam AI. Natural products as antimicrobial agents. *Food Control* 2014; 46: 412–29.
7. Krishnaiah D, Sarbatly R, Nithyanandam R. A review of the antioxidant potential of medicinal plant species. *Food Bioprod Process.* 2011; 89: 217–33.
8. Luo Y, Cai Q, Sun M, Corke H. Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sci.* 2004; 74: 2157–84.
9. Kumar S, Bajwa BS, Singh K, Kalia AN. Anti e inflammatory activity of herbal plants: a review. *Int J Adv Pharm Biol Chem.* 2013; 2: 272–81.
10. Atta-Ur-Rahman, Zaman K. Medicinal plants with hypoglycemic activity. *J Etnopharmacol* 1989; 26: 1–55.
11. Kirtikar KR, Basu BD. *Indian Medicinal Plants.* International Book Distributor, Dehradun. 1975; 2: 1344–1345.
12. Anderberg Asteraceae AA,. In: Bremer, Kare (Ed.), *Cladistics & Classification.* Timber Press, Portland, Oregon. 1994; 292–303.

13. Mandeel Q, Taha A. Assessment of *in vitro*. Antifungal Activities of Various Extracts of Indigenous Bahraini Medicinal Plants, *Pharmaceutical Biology*, 2005, 43(4): 340.
14. Sittiwet C. In vitro Antimicrobial Activity of *Pluchea indica* Aqueous Extract: The Urinary Tract Infection Treatment, *Journal of Pharmacology and Toxicology*, 2009; 4(2): 87.
15. Anonymous. The Ayurvedic Pharmacopoeia of India. Vol 3, Ministry of Health and Family Welfare, Department of Health, Govt. of India, New Delhi, 1989; 163-165.
16. Chaturvedi GN, Singh RH. Experimental studies on the antiarthritic effect of certain indigenous DRUGS *Indian J. Med. Res.*, 1965; 53: 71.
17. Farnsworth NR, Bunyaphatsara N. Thai Medicinal Plants. Prachachon Co: Bangkok. 1992; pp. 200–201.
18. Khare CP. *Indian Medicinal Plants: An Illustrated Dictionary*. Springer-Verlag Berlin/Heidelberg. 2007; p. 500.
19. Nadkarni AK. *Indian Materia Medica*. Popular Prakashan, Bombay. 1976; 242.
20. Ahmad VU, Fizza KZ, Khan MA, Farooqui TA. Sesquiterpenes from *Pluchea arguta*. *Phytochemistry*, 1991; 30: 689.
21. Ahmad VU, Sultana A, Fizza KZ. Two new terpenoids from *Pluchea arguta*. *Naturforsch* 45B, 385–388, *Naturforsch*, 1990; 45: 385.
22. Ahmed AA, Melek FR, Mabry TJ. Flavonoids of *Cotula cinerea*. *Journal of Natural Products*. 1987; 50: 311.
23. Chakravarty AK, Mukhopadhyay S. New thiophene derivative from *Pluchea indica*. *Indian Journal of Chemistry*. 1994; 33: 978.
24. Chawla AS, Kaith BS, Handa SS, DK Kulshreshtha; RC Srimal. A lignan from *Vitex negundo* seeds. *Fitoterapia*, 1991; 62: 441.
25. Chiang MT, Bittner M, Silva M, Watson WH, Sammes PG. A new sesquiterpene from *Pluchea chingoyo*. *Phytochemistry*, 1979; 18: 2033.
26. GS Dixit; RP Tewari. Chemical constituents of *Pluchea lanceolata*. *Sacitra Ayurveda*, 1991; 43: 841.
27. Inderjit; KMM Dakshini. Investigations on some aspects of chemical ecology of cogongrass *Imperata cylindrical*. *J. Chem. Ecol.*, 1991; 17: 1585.
28. Inderjit; KMM Dakshini. Interface potential of *pulchella lanceolata* (Asteraceae) on characteristics of four soils and mustard and tomato growth. *J. Chem. Ecol.* 1992; 18: 713.
29. Uchiyama T, Miyase T, Ueno A, Usmanhani K. Terpenic glycosides from *Pluchea indica* *Phytochemistry*. 1989; 28: 3369.
30. DN Prasad; SK Bhattacharya; PK Das. A study of antiinflammatory activity of some indigenous drugs in albino rats. *Indian J. Med. Res*, 1966; 54: 582.
31. Uchiyama T, Miyase T, Ueno A, Usmanhani K. Terpene and lignan glycosides from *Pluchea indica* *Phytochemistry*, 1991; 30: 655.
32. Fakhr Eldin O. Suliman, Majekodunmi O. Fatope, Salim H. Al-Saidi, Salma M. Z. Al-Kindy and Ruchi G. Marwah, Composition and antimicrobial activity of the essential oil of *Pluchea arabica* from Oman, *Flavour and Fragrance Journal*. 2006; 21(3): 469-471.
33. Marwah RG, Fatope MO, Mahrooqi RA, Varma GB, Abadi HA, Al- Burtamani SKS, Antioxidant capacity of some edible and wound healing plants in Oman. *Food Chemistry*. 2007; 101: 465.
34. C.A. Lipinski, F. Lombardo, B.W. Dominy, P.J. Feeney, Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev*. 1997; 23(1-3): 3-26.
35. Molinspiration Cheminformatics. Nova ulica, SK-900 26 Slovensky Grob, Slovak Republic. [Online] Available from: <http://www.molinspiration.com> [Accessed on 3rd July, 2012].
36. Verma A. Lead finding from *Phyllanthus debelis* with hepatoprotective potentials. *Asian Pac J Trop Biomed*. 2012; 2: S1735–S1737.
37. Alodeani E A, Arshad M, Izhari MA. Anti-uropathogenic activity, drug likeness, physicochemical and molecular docking assessment of (E)-N'-(substituted-benzylidene)-2-(quinolin-8-ylloxy) acetohydrazide. *Asian Pacific Journal of Tropical Biomedicine*. 2015; 5(8): 676-683.
38. Alodeani E A, Arshad M, Izhari MA. Antileishmanial screening, physicochemical properties drug likeness of pyrazole carbaldehyde derivatives. *Asian Pac. J. Health Sci.*, 2015; 2(2): 41-47.
39. Ho PL, Yung RW, Tsang DN, Que TL, Ho M, Seto WH, et al. Increasing resistance of *Streptococcus pneumoniae* to fluoroquinolones: results of a Hong Kong multicentre study in 2000. *J Antimicrob Chemother* 2001; 48: 659-65.
40. Mohammad Arshad, Abdul Roouf Bhat, Kwon Kang Hoi, Inho Choi, Fareeda Athar, Synthesis, characterization and antibacterial screening of some novel 1,2,4-triazine derivatives. <http://dx.doi.org/10.1016/j.ccl.2016.12.037>
41. Abdul Kareema, Laxmi, Mohammad Arshad and Nahid Nishat. Herbo-Mineral based Schiff base ligand and its metal complexes: synthesis, characterization, catalytic potential and its biological applications. *Journal of Photochemistry and Photobiology B: Biology*. 2016; 160: 163-171.
42. Noor e Iram, Mohd Shoeb Khan, Reshma Jolly, Mohammad Arshad, Mahboob Alam, Parvez Alam, Rizwan Hasan Khan, Farha Firdaus, Interaction mode of polycarbazole-titanium dioxide nanocomposite with DNA: Molecular docking simulation and in-vitro antimicrobial study. *Journal of Photochemistry and Photobiology B: Biology*. 2015; 153: 20-32.
43. Rani Bushra, M. Shahadat, Meraj Alam Khan, R. Adnan, Mohammad Arshad, M. Rafatullah and Mu.

- Naushad, Preparation of Polyaniline based Nanocomposite material and their Environmental Applications. 2014, *Int. J. Env. Sci. Technol.*
44. Mohammad Arshad, Tazeem, A R Bhat, Fareeda Athar, Heterocyclic Azoles and their biological application as antimicrobials. *Journal of Natural Science, Biology and Medicine*, 2011; 2(2): 1-156.
 45. Tazeem, Mohammad Arshad, A R Bhat, Fareeda Athar, Synthesis, characterization of heterocyclic compounds and their application as antibacterial therapeutic agents. *Journal of Natural Science, Biology and Medicine*, 2011; 2(2): 1-156.
 46. Tuhfa, Mohammad Younus Wani, Mohammad Arshad, Tazeem, Fareeda Athar, Chalcone scaffold: Synthesis, modification, characterization, molecular properties and screening against microbes. *Journal of Natural Science, Biology and Medicine*, 2011; 2(2): 1-156.
 47. Alodeani E A, Arshad M, Izhari M A: Anti-uropathogenic activity, drug likeness, physicochemical and molecular docking assessment of (E)-N'-(substitutedbenzylidene)-2-(quinolin-8-yloxy) acetohydrazide. *Asian Pac J Trop Biomed* 2015; 5(8): 676-683.
 48. Nova ulica: Molinspiration cheminformatics [homepage on the internet], SK-900 26 Slovensky Grob, Slovak Republic [cited 2012 July3], Available from <http://www.molinspiration.com>
 49. Verma, A: Lead finding from *Phyllanthus debelis* with hepatoprotective potentials *Asian Pacific Journal of Tropical Biomedicine* 2012; S1735-S1737.
 50. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ: Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv. Drug Delivery Rev.* 1997; 23(1-3): 3-25.
 51. Alodeani E A, Arshad M, Izhari M A: Anti-uropathogenic activity, drug likeness, physicochemical and molecular docking assessment of (E)-N'-(substitutedbenzylidene)-2-(quinolin-8-yloxy) acetohydrazide. *Asian Pac J Trop Biomed* 2015; 5(8): 676-683.
 52. Alodeani E A, Arshad M, Izhari M A: Antileishmanial screening, physicochemical properties and drug likeness of pyrazole carbaldehyde derivatives: *Asian Pac. J. Health Sci.* 2015; 2(2): 41-47.
 53. Alodeani E A, Arshad M, Izhari M A: Antileishmanial activity and computational studies of some hydrazone derivatives possessing quinoline nucleus. *European Journal of Pharma and medical research.* 2015; 2(7): 324-328.
 54. Alodeani E A, Arshad M, Izhari M A: Drug likeness and physicochemical properties evaluation of the alkaloids found in black pepper: piperine, piperidine, piperettine and piperanine. *European Journal of Pharma and medical research* 2015; 2(6): 296-301.
 55. Morris GM, Goodsell DS, Halliday RS, Huey R, Hart WE, Belew RK, et al. Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *J Comput Chem* 1998; 19: 1639-62.
 56. Moulleron S, Badet-Denisot MA, Golinelli-Pimpaneau B. Ordering of C-terminal loop and glutaminase domains of glucosamine-6-phosphate synthase promotes sugar ring opening and formation of the ammonia channel. *J Mol Biol.* 2008; 377(4): 1174-85.