

SILVER NANO – A MAGIC WORLD (A SMALL REVIEW ON BIOMEDICAL APPLICATIONS)Kantipriya Kondala*¹ and Dr. D. Sarvamangala²¹*Research Scholar, Dept. of Biotechnology, GIT, GITAM UNIVERSITY, Visakhapatnam – 530045.²Associate Professor, Dept. of Biotechnology, GIT, GITAM UNIVERSITY, Visakhapatnam – 530045.***Corresponding Author: Kantipriya Kondala**

Research Scholar, Dept. of Biotechnology, GIT, GITAM UNIVERSITY, Visakhapatnam – 530045.

Article Received on 05/03/2017

Article Revised on 25/03/2017

Article Accepted on 15/04/2017

ABSTRACT

Here we report a small review on synthesis of silver nanoparticles by plant materials. Different materials like leaves, flowers, fruits etc. which are ecofriendly were used for the production of silver nanoparticles is became essential. Silver nanoparticles are found to be very effective against bacteria, fungi and viruses. Vast literature is available which shows evidence that silver nanoparticles can directly attack and destroy the cell membrane of bacteria and kills. Depending upon their size, shape, structure and other properties like PH, temperature and organic molecules present in the plant extracts increases the stability of the particle. The components of the plant materials help in the size reduction which increases the surface area of the particle and leads to enhance the antibacterial activity.

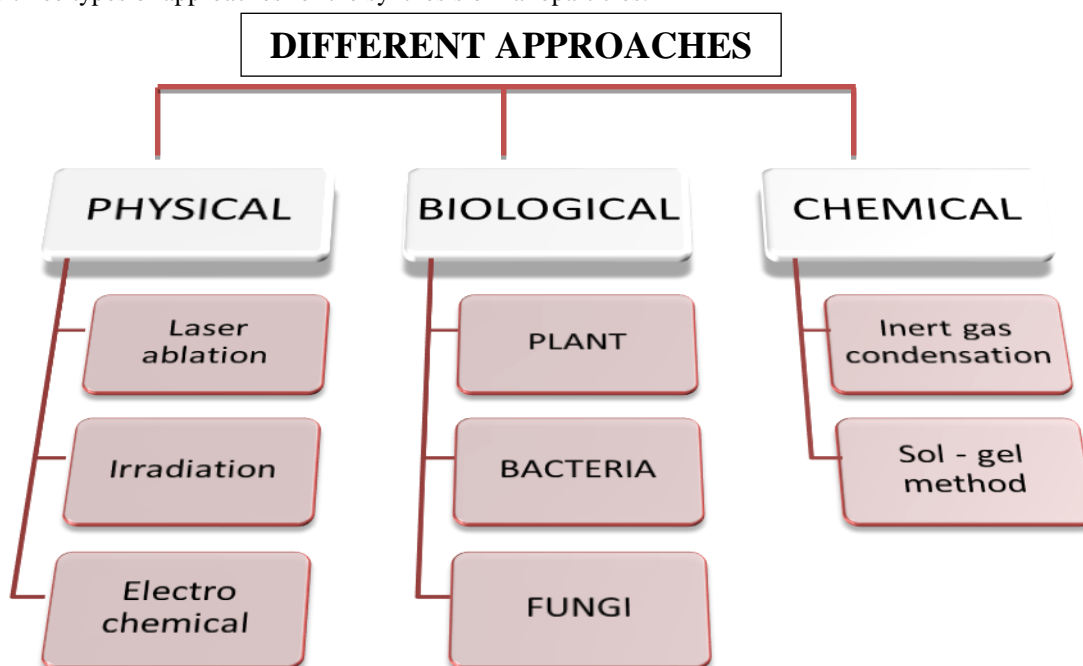
KEYWORDS: Broad spectrum antibacterial, biomedical applications, UV-VIS Spectroscopy, XRD, FTIR.**INTRODUCTION**

Applications of silver nanoparticles have been extended and it is most commonly used in consumer products (Rejeski D 2009).^[1] The broad spectrum activity helps its usage in biomedical applications water and air

purification, food production, cosmetics, clothing, water filters, contraceptives, detergent, sox, shoes and numerous household products (C.Marabio – Jones et. Al 2010).^[2]

Methods for synthesis of nanoparticles

There are three types of approaches for the synthesis of nanoparticles.



Literature survey reveals - biological approaches are single pot, easy to handle and safe method for the production of silver nanoparticles (Table – 1). The biochemical reaction of AgNO_3 with plant extract (Fig. 1) leads to the formation of AgNPs (Tripathy *et al.*, 2010).^[31]

Plant extract + $\text{Ag}^+\text{NO}_3^- \rightarrow \text{AgNPs} + \text{Byproducts}$ (hazardless)

Fig.1 – Mechanism of synthesis of nanoparticles with silver

Table – 1. Literature on nanoparticle production using green nanotechnology

| S. NO | TITLE | SOURCE/METHOD | PARTICLE SIZE & APPLICATION | YEAR | REF. NO |
|-------|---|-------------------------------------|--|-------|---------|
| 1. | Fabrication of silver nanoparticles and their antimicrobial mechanisms | Silver nitrate + PVP + Formaldehyde | >10 nm Antimicrobial activity | 2006. | [18] |
| 2. | Does the antibacterial activity of silver nanoparticles depend on the shape of the NP? A study of the gram negative bacterium <i>E.coli</i> | | 40 nm Antibacterial activity | 2007. | [19] |
| 3. | Extracellular biosynthesis of silver nanoparticles by some bacteria | Bacteria | 2-100 nm- | 2008. | [20] |
| 4. | Investigation of antibacterial properties of silver NP – loaded poly grafted cotton fabric | Silver nitrate | 12.5 nm Antibacterial activity | 2008. | [21] |
| 5. | Rapid biosynthesis of silver nanoparticles using culture supernatant of bacteria with microwave irradiation | Bacteria | 5-60 nm | 2009. | [22] |
| 6. | Stable silver nanoparticle synthesizing methods and its applications | Chemical method | >5 nm Applications of silver nanoparticles | 2010. | [23] |
| 7. | Extracellular synthesis of silver nanoparticles using leaves of <i>Euphorbia hirta</i> and their antibacterial activities | <i>Euphorbia hirta</i> | 40-50 nm Antibacterial activity | 2010. | [24] |
| 8. | Biomimetic synthesis of silver NP'S by <i>Citrus Limon</i> (lemon) aqueous extract and theoretical prediction of particle size | Lemon | Below 50 nm Particle size prediction | 2010. | [25] |
| 9. | Phytofabrication of silver NP'S using pomegranate seeds | Pomegranate fruit seeds | 30 nm Phytofabrication | 2011. | [26] |
| 10. | Extracellular synthesis of silver nanoparticles using leaf extract of <i>Cassia auriculata</i> | <i>Cassia auriculata</i> | 20-40 nm Antibacterial activity | 2011. | [27] |
| 11. | Extracellular biosynthesis of silver NP'S using the <i>Mycorrhiza mushroom tricholoma crissum</i> (Bark). SACC: Its antimicrobial activity against pathogenic bacteria and fungus, including multidrug resistant plant and human bacteria | Mushroom | 21-91 nm Antimicrobial activity | 2011. | [28] |
| 12. | Antimicrobial activity of silver nanoparticles synthesized by using medicinal plants | <i>Boswellia Shorea Svensonia</i> | Antimicrobial activity | 2011. | [29] |
| 13. | Synthesis of silver nanoparticles using albumin as a reducing agent | Albumin | | 2011. | [30] |
| 14. | Green fruit of Chili (<i>Capsicum annum</i>) synthesizes nanosilver | Chili (<i>Capsicum annum</i>) | 2-6 nm Played the pivotal role in fabrication of AgNP'S | 2011. | [31] |
| 15. | Silver nanoparticles synthesis from <i>Lecanicillium lecenii</i> and evolutionary treatment on cotton fabrics by measuring their improved antibacterial activity with antibiotics against <i>staphylococcus aureus</i> & <i>E.coli</i> | <i>Lecanicillium lecenii</i> | 45-100 nm Prevents pathogen infection. | 2011. | [32] |
| 16. | Extracellular fabrication of silver nanoparticles using <i>pseudomonas aeruginosa</i> and its antimicrobial assay | <i>Pseudomonas aeruginosa</i> | 20–50 nm antimicrobial assay | 2012. | [33] |

| | | | | | |
|-----|--|--|---|-------|------|
| 17. | Silver nanoparticles ecofriendly synthesis by two medicinal plants | <i>Punica granatum</i> <i>Rosa damascena</i> | 21 nm | 2012. | [34] |
| 18. | Biosynthesis of silver nanoparticles from <i>Glycyrrhiza glabra</i> root extract | <i>Glycyrrhiza glabra</i> | 19 nm | 2012. | [35] |
| 19. | Synthesis of silver nanoparticles using <i>Klebsiella pneumonia</i> and its Bio-medical applications | <i>Klebsiella pneumonia</i> | 21±8.9 nm | 2013. | [36] |
| 20. | Synthesis and characterization of silver nanoparticles using <i>cannonball</i> leaves and their cytotoxic activity against MCF – 7 cell line | <i>Cannonball</i> leaves | 5-35 nm Cytotoxic activity | 2013. | [37] |
| 21. | Silver nanoparticles ecofriendly green synthesis by using two medicinal plant extract. | <i>Jasminium grandifolium</i> <i>Cymbopogan citrullus</i> | 206 nm 199 nm | 2013. | [38] |
| 22. | Green synthesis of nanoparticles and its application in treatment of Rheumatoid arthritis. | <i>Night jasmine (nyctanthes arbor tristis)</i> | 46-80 nm Treating Rheumatoid arthritis | 2013. | [39] |
| 23. | Cationic antimicrobial peptides and biogenic silver nanoparticles kill mycobacteria without eliciting DNA damage and cytotoxicity in mouse macrophages. | <i>Cationic antimicrobial peptides</i> <i>Mycobacteria</i> | 177 nm 97.65 nm DNA damage and cytotoxicity | 2013. | [40] |
| 24. | Biological synthesis of silver nanoparticles using <i>Raphanus sativus</i> VAR.LONGIPINNATUS leaf extract and evaluation of their antioxidant and antibacterial activity. | <i>Raphanus sativus</i> | 22 nm antioxidant and antibacterial activity | 2013. | [41] |
| 25. | Leaf extract mediated green synthesis of silver nanoparticles from widely available Indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. | <i>Musa balbisiana</i> <i>Azadirachta indica</i> <i>Ocimum tennflorum</i> <i>Vigna radiate</i> <i>Chick pea</i> | 80.2 nm antimicrobial property and toxicity analysis | 2014. | [42] |
| 26. | One pot light assisted green synthesis, storage and antimicrobial activity of dextran stabilized silver nanoparticles. | <i>AgNO₃+Dextran sol</i> | 50-70 nm antimicrobial activity | 2014. | [43] |
| 27. | Bioreduction based synthesis of silver Nano coats and their application in development of nanoembedded medical fabrics. | <i>Phyllanthus amarus</i> <i>Calotropisgigantea</i> <i>Vitexnegundo</i> <i>Acalyphaindica</i> <i>Cardiospermum helicacabrium</i> <i>Polygala chinensis</i> <i>Ponganiasp</i> <i>Aegle marmelos</i> <i>Vincarosea</i> | - development of nanoembedded medical fabrics | 2014. | [44] |
| 28. | Green synthesis, characterization and antimicrobial properties of silver nanowires by aqueous leaf extract of piper betle. | Piper betle | 40-60 nm antimicrobial properties | 2014. | [45] |
| 29. | Biofabrication and extraction of silver nanoparticles using leaf extract of <i>Azadirachta indica</i> . | <i>Azadirachta indica</i> | 104.8 nm Biofabrication | 2014. | [46] |
| 30. | Bioreduction based synthesis of silver Nano coats and their application in development of nano embedded medical fabrics. | <i>Polygala chinensis</i> | development of nano embedded medical fabrics. | 2014. | [47] |
| 31. | Biogenic synthesis, characterization of silver nanoparticles <i>Tabernaemontana divaricata</i> and <i>Polyanthus tuberosa</i> flower extract and evaluation of their antibacterial activities. | <i>Tabernaemontana divaricata</i> <i>Polyanthus tuberosa</i> | 32 nm 17 nm | 2014. | [48] |
| 32. | Efficacy of green synthesis of silver nanoparticles using flowers of calendula officinalis. | <i>Calendula officinalis</i> | 2-20 nm | 2014. | [49] |
| 33. | Green synthesis and characterization of | <i>Basella alba</i> | 22.6-25 nm | 2014. | [50] |

| | | | | | |
|-----|--|---|---|-------|------|
| | silver NP's from aqueous extract of <i>Basella alba</i> and their in-vitro antioxidant potentials. | | antioxidant potentials | | |
| 34. | Green synthesis of plant mediated silver NP's and evaluation of their antimicrobial activities. | <i>Morinda tintoria</i> <i>Michelia champaca</i> | < 100 nm antimicrobial activities | 2014. | [51] |
| 35. | Green synthesis of silver NP's from marigold flower and its synergistic antimicrobial potential. | <i>Marigold flower</i> | 10-90 nm synergistic antimicrobial potential | 2014. | [52] |
| 36. | Biosynthesis of silver NP's using flower extracts of <i>Catharanthus roseus</i> and evaluation of its antibacterial efficacy. | <i>Catharanthus roseus</i> | 11-15 nm antibacterial efficacy | 2014. | [53] |
| 37. | Biosynthesis and characterization of silver nanoparticles using <i>Ficus benghalensis</i> leaf extract. | <i>Ficus benghalensis</i> | 10-50 nm | 2014. | [54] |
| 38. | Synthesis and characterization of silver nanoparticles using <i>Tabernaemontana Divaricata</i> and its cytotoxic activity against MCF – 7 cell line. | <i>Tabernaemontana Divaricata</i> (common garden plant) | 22.85 nm cytotoxic activity | 2014. | [55] |
| 39. | Leaf extract mediated green synthesis of silver NP's from widely available Indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. | <i>Seeds of Moong bean and Chick pea</i> | 80.2 nm antimicrobial property and toxicity analysis | 2014. | [56] |
| 40. | Syntheses of silver nanoparticles using the vegetable extract of <i>Raphanus sativus</i> (Radish) and assessment of their antimicrobial activity. | <i>Raphanus sativus (Radish)</i> | 30-60 nm antimicrobial activity | 2015. | [57] |
| 41. | Green synthesis of silver NPs using <i>Brassica oleracea</i> (Cauliflower) and <i>Brassica oleracea capitata</i> (Cabbage) and the analysis of antimicrobial activity. | <i>Brassica oleracea (Cauliflower)</i> <i>Brassica oleracea capitata (Cabbage)</i> | 36-42 nm antimicrobial activity | 2015. | [58] |
| 42. | Biological synthesis of silver nanoparticles using <i>Colocassia</i> extract and their antimicrobial activity. | <i>Colocassia</i> | 10-30 nm antimicrobial activity | 2015. | [59] |
| 43. | Evaluation of bactericidal and fungicidal properties of silver NP's fabricated using <i>Jasminium sambac</i> . | <i>Jasminium sambac</i> | 14-17 nm bactericidal and fungicidal properties | 2015. | [60] |
| 44. | Biosynthesis, characterization and antimicrobial applications of silver nanoparticles. | <i>Brevibacterium frigoritolerans DC2</i> | 97 nm antimicrobial applications | 2015. | [61] |
| 45. | Synthesis of silver nanoparticles using <i>Muntingia calabura</i> leaf extract. | <i>Muntingia calabura</i> | - Screening of secondary metabolites | 2015. | [62] |
| 46. | Antibacterial and anticancer activity of silver nanoparticles synthesized from <i>Cynodon dactylon</i> leaf extract. | <i>Cynodon dactylon</i> | - Antibacterial and anticancer activity | 2015. | [63] |
| 47. | Bactericidal paper trays doped with silver nanoparticles for egg storing applications. | <i>Gelatin chitosan</i> | 37.35 nm Antibacterial activity | 2016. | [64] |
| 48. | Green Synthesis and Characterization of Silver Nanoparticles Using <i>Mimusops elengi</i> Flower Extract and Its Synergistic Antimicrobial Potential. | <i>Mimusops elengi</i> | - Antibacterial agent | 2016. | [65] |
| 49. | Green Synthesis of Silver Nanoparticles Using Apple Extract and Its Antibacterial Properties. | Apple Extract | Antibacterial Properties | 2016. | [66] |
| 50. | Rapid green synthesis of silver nanoparticles by aqueous extract of seeds of <i>Nyctanthes arbor-tristis</i> . | <i>Nyctanthes arbor-tristis</i> | Biomedical applications | 2016. | [67] |

Various plant parts were used to synthesize silver nanoparticles (Fig. 2) such as stem, root, fruit, seed, callus, leaves and flower (Palaniselvam Kuppusamy *et al.*, 2014).^[4]

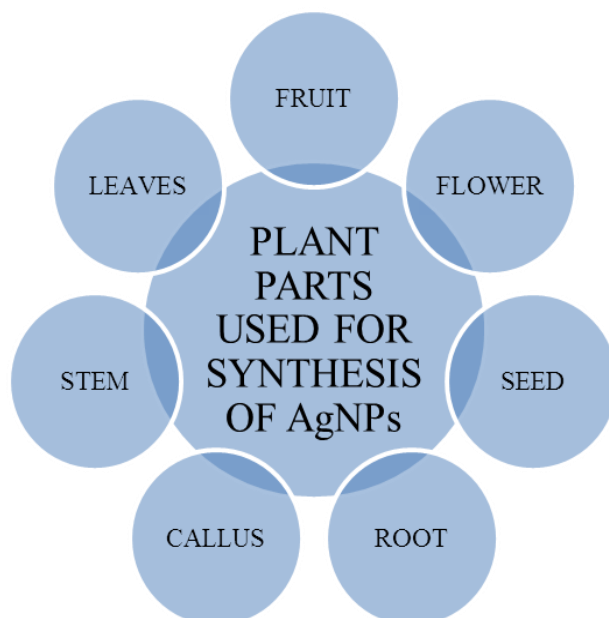


Fig. 2 - Various plant parts used for the production of AgNP

CHARACTERIZATION

Various methods are employed to know the size, shape, structure of surface and other characteristics. Mainly used techniques are UV – VIS Spectroscopy, XRD, EDS, SEM, AFM, FTIR and TEM.

UV- VISIBLE Spectroscopy of silver nanoparticles shows an absorption peak is usually in between 400 and 450 nm (Devika R *et al.* 2012, Poinern G E J *et al.*

2013).^[5,6] The elemental composition of nanoparticles can be determined by EDS whereas XRD technique identifies size, structure and orientation of crystal (Klug H P *et al.* 1974, Barrett C S *et al.* 1986).^[7, 8] FTIR Spectroscopy used to identify the functional groups and surface chemistry (Poinern G E J *et al.* 2014).^[9] SEM and TEM techniques reveal the morphology and size of the stabled nanoparticles.

Silver nanoparticles – Applications

There are several applications for silver nanoparticles. The major applications are mentioned in the fig. 3.

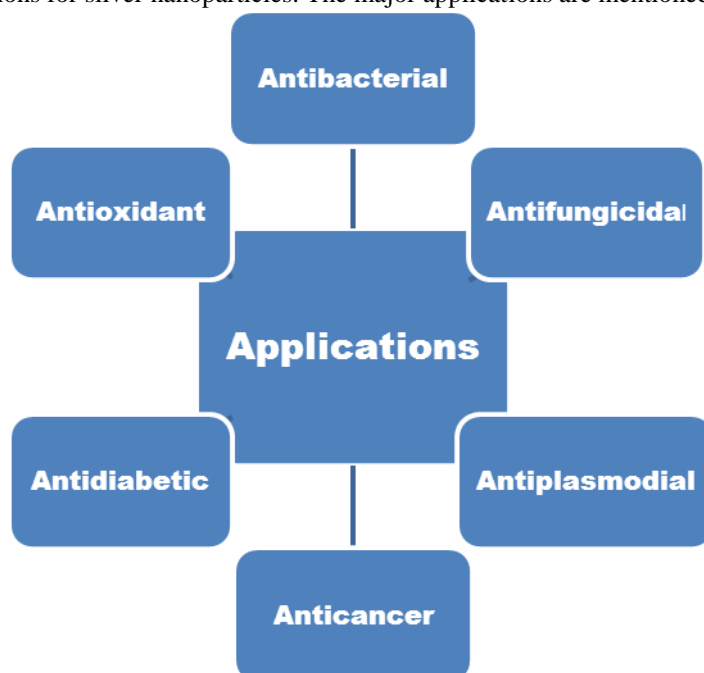


Fig. 3 – Major applications of silver nanoparticles

Applications

1. Antibacterial activity: The double action of AgNPs eventually breaks the cell wall and disturbs the mechanism of protein synthesis in the bacteria (Sondi and Salopek–sondi, 2014).^[10]

Silver nanoparticles exhibited good antibacterial properties against *Staphylococcus aureus* and *Pseudomonas aeruginosa* (D. Sarvamangala et al. 2014).^[11]

2. Antifungicidal activity: Plant mediated AgNPs damage the intercellular components of fungus and destroy the cell function (Logeswari et al. 2012).^[12]
3. Antiplasmodial activity: Synthesis of silver nanoparticles from plant extract has been used to suppress the number of malarial productions.

Silver nanoparticles synthesizes rapidly by using the plant medicinal plants *Andrographis paniculata* against the malarial parasites *Plasmodium falciparum* (C. Panneerselvam et al. 2011).^[13]

4. Anticancer studies: Synthesized silver nanoparticles using *Abelmoschus esculentus* (L.) pulp extract showed that the IC₅₀ dose of AgNPs leads to the increase in intracellular reactive oxygen species and significantly diminished mitochondrial membrane potential, indicating the effective involvement of apoptosis in cell death. (Md. Masud Rahaman Mollick. et al. 2015).^[14]
5. Antidiabetic activity: Plant mediated AgNPs contains TAP (Terpenoids in *Andrographis Paniculata* leaf) shows good Antidiabetic activity which has been investigated by glucose oxidase method (S. Kavitha et al. 2014).^[15]
6. Antioxidant activity: Antioxidant activities are attributed to the phenolic contents in plants probably due to their redox properties, which allow them to act as reducing agents, hydrogen donors and singlet oxygen quenchers (S. T. Chang et al. 2001).^[16]

The total phenolic compounds and total flavonoids were higher in plant-AgNPs compared to the plant extract alone. Plant-AgNPs showed a higher antioxidant and antimicrobial activity compared to *C.murale* leaf extract alone or silver nitrate (Mohamed S. Abdel-Aziz et al. 2014).^[17]

CONCLUSION

In the last two decades, applications of plant derived silver nanoparticles were extensively studied. Biosynthesis of plant mediated AgNPs is a simple, easy to handle and ecofriendly method with controlled side effects. Applications of silver nanoparticles derived from plant material are tremendous in various fields such as medical diagnosis, pharmaceuticals, cosmetics, medical treatment, electronics, optics and other commercial products.

In future there is a vast scope in this field of Nano science and more things to be explored. Few of them are:

- The active phytochemical compound which helps in the reduction of silver ions to silver nanoparticles is to be explored.

- A particular route of functional mechanism on different pathogenic microorganisms and virus to be explored.
- Implementation of large scale production and action of silver nanoparticles on their target sites have to be studied in depth.
- New pharmaceutical formulations are to be explored from these plant mediated AgNPs i.e., the process in which plant mediated silver nanoparticles including the active drug combined to produce a final medical product.

REFERENCES

1. Rajeski D. Nanotechnology and consumer products. http://www.nanotechproject.org/publications/archive/nanotechnology_consumer_products/. Accessed 22 February 2010.
2. Marmbio-Jones C, Hoek E M V. A review of the antibacterial effects of silver nanoparticles and potential implications for human health and the environment. *Journal of nanoparticle research*. 2010; 12: 1531-1551.
3. Tripathy A, Raichur A M, Chandrasekaran N, Prathna T C, Mukherjee A. process variables in biomimetic synthesis of silver nanoparticles by aqueous extract of *Azadirachta indica* leaves. *Journal of nanoparticle research*. 2010; 12: 237-246.
4. Palaniselvam Kuppusamy, Mashitah M, Yusoff, Gaanty Pragas Maniam, Natanamurugaraj Govindan. Biosynthesis of metallic nanoparticles using plant derivatives and their new avenues in pharmacological applications – An updated report. *Saudi pharmaceutical journal*. 2014; 24: 473-484.
5. Devika R, Elumalai S, Manikandan E, Eswaramoorthy D. Biosynthesis of silver nanoparticles using fungus *Pleurotus ostreatus* and their antimicrobial activity. *Open Access Science Report*. 2012; 1: 1-5.
6. Klug H P, Alexander L E. X-ray diffraction procedures for poly-crystallite and amorphous materials. 2nd ed.; Wiley: New york, NY, USA, 1974.
7. Poinern G E J, Shah M, Chapman P, Fawcett D. Green biosynthesis of silver nanotubes using the leaf extracts from *Eucalyptus macrocarpa*. *Nano Bull*. 2013; 2: 1-7.
8. Barrett C S, Cohen J B, Faber J, Jenkins R, Leyden D E, Russ J C, Predeck P K. *Advances in X-ray analysis*. Plenum Press: New york, NY, USA, 1986; volume 29.
9. Poinern G E J. *A laboratory course in Nanoscience and Nanotechnology*, 1st ed.; CRC Press Taylor & Francis: Boca Raton, FL, USA, 2014.
10. Sondi I, Salopek-Sondi B. Silver nanoparticles as antimicrobial agent: a case study on *E.coli* as a model for Gram-negative bacteria. *Journal of colloid interface science*. 2004; 275: 177-182.
11. Sarvamangala D, Kantipriya Kondala, Murthy U S N, Narasingarao B, Sivakumar N. Green synthesis of AgNPs using *Alternanthera sessilis* leaf extract (A

- Natural source for ocular therapy). International journal Innovative Research in Science, Engineering and Technology. 2014; 3(7).
12. Logeswari P, Silambarasan S, Abraham J. Synthesis of silver nanoparticles using plant extracts and analysis of their antimicrobial activity. Journal of Saudi chemical society. 2012; 4: 23-45.
 13. Panneerselvam C, Ponaruselvam S and Murugan K. Potential anti – plasmodial activity of synthesized silver nanoparticle using *Andrographis paniculata* nees (Acanthaceae). Archives of Applied Science Research. 2011; 3(6). 208-217.
 14. Md. Masud Rahaman Mollick, Dipak Rana, Sandeep Kumar Dash, Sourav Chattopadyay, Biplab Bhowmick, Dipanwita Maity, Dibyendu Mondal, Sutanuka Pattanayak, Somenath Roy, Mukut Chakraborty, Dipankar Chattopadyay. Studies on green synthesized silver nanoparticles using *Abelmoschus esculentus* (L.) pulp extract having anticancer (Invitro) and antimicrobial applications. Arabian journal of chemistry. 2015. [http://dx.doi.org/10.1016/j.arabjc.2015.04.033].
 15. Kavitha S, Dr Dhamodharan M, Satish Kannan. Synthesis, characterization and anti-diabetic, antibacterial activity of silver nanoparticle in terpenoids for *Andrographis paniculata* (AG-TAP). International journal of scientific engineering and technology research. 2014; 3(47): 9467-9473.
 16. Chang S T, Wu J H, Wang S Y, Kang P L, Shyur L F. Antioxidant activity of extracts from *Acacia confuse* bark and heart wood. Journal of agriculture food chemistry. 2001; 49: 3420-3424.
 17. Mohamed S, Abdel- Aziz, Mohamed S Shaheen, Aziza A. EL – Neekeety, Mosaad A. Abdel – Wahhab. Antioxidant and antibacterial activity of silver nanoparticles biosynthesized using *Chenopodium murale* leaf extract. Journal of Saudi chemical society. 2014; 18(4): 356-363. [http://dx.doi.org/10.1016/j.jscs.2013.09.011].
 18. H.Y Song, K.K Ko, I.H Oh and B.T Lee. Fabrication of Silver Nanoparticles and Their Antimicrobial Mechanisms. European Cells and Materials. 2006; 11(1): 58.
 19. Sukdeb Pal, Yu Kyung Tak and Joon Myong Song. Does the Antibacterial Activity of Silver Nanoparticles Depend on the Shape of the Nanoparticle? A Study of the Gram-Negative Bacterium *Escherichia coli*. Applied and environmental microbiology. 2007; 73(6): 1712-1720. [Doi: 10.1128/AEM.02218-06].
 20. S. Minaeian, A. R. Shahverdi, A. S. Nohi and H. R. Shahverdi. Extracellular biosynthesis of silver nanoparticles by some bacteria. J. Sci. I. A. U (JSIAU). 2008; 17(66): 1-4.
 21. Gupta P, Bajpai M and Bajpai S K. Investigation of antibacterial properties of silver nanoparticle – loaded poly (acrylamide-co-itaconic acid) – Grafted cotton fabric. The journal of cotton science. 2008; 12: 280-286.
 22. N. Saifuddin, C. W. Wong and A. A. Nur Yasumira. Rapid Biosynthesis of Silver Nanoparticles Using Culture Supernatant of Bacteria with Microwave Irradiation. E-Journal of Chemistry. 2009; 6(1): 61-70.
 23. Thirumalai Arasu V, Prabhu D and Soniya M. Stable Silver Nanoparticle Synthesizing methods and its Applications. Journal of bioscience research. 2010; 1(4): 259-270.
 24. EK.Elumalai, T.N.V.K.V.Prasad, J.Hemachandran, S.Viviyam Therasa, T.Thirumalai and E David. Extracellular synthesis of silver nanoparticles using leaves of *Euphorbia hirta* and their antibacterial activities. Journal of pharmaceutical sciences and research. 2010; 2(9): 549-554.
 25. Prathna TC Chandrasekaran N, Raichur AM and Mukherjee A. Biomimetic synthesis of silver nanoparticles by Citrus limon (lemon) aqueous extract and theoretical prediction of particle size. Colloids Surf B Biointerfaces. 2010; 82(1): 152-159. [DOI: 10.1016/j.colsurfb.2010.08.036].
 26. Shalini Chauhan, Mukesh Kumar Upadhyay, Narayan Rishi and Sushma Rishi. Phytofabrication of silver nanoparticles using pomegranate fruit seeds. International Journal of Nanomaterials and Biostructures. 2011; 1(2): 17-21.
 27. C. Udayasoorian, K. Vinoth Kumar and R. M. Jayabalakrishnan. Extracellular Synthesis of Silver Nanoparticles using Leaf Extract of *Cassia Auriculata*. Digest Journal of Nanomaterials and Biostructures. 2011; 6(1): 279-283.
 28. Sarmistha Ray, Swadesh Sarkar and Surekha Kundu. Extracellular Biosynthesis Of Silver Nanoparticles using the Mycorrhizal Mushroom *Tricholoma Crassum* (Berk.) Sacc: Its Antimicrobial activity against Pathogenic Bacteria and Fungus, Including Multidrug Resistant Plant and Human Bacteria. Digest Journal of Nanomaterials and Biostructures. 2011; 6(3): 1289-1299.
 29. N. Savithramma, M. Linga Rao, K. Rukmini and P. Suvarnalatha Devi. Antimicrobial activity of Silver Nanoparticles synthesized by using Medicinal Plants. International Journal of ChemTech Research. 2011; 3(3): 1394-1402.
 30. Elpidio Morales – Sanchez, Jesus Guajardo – Pacheco, Maria Noriega – Trevino, Cristina Quintero – Gonzalez, Martha C ompean – Jasso, Francisco Lopez – Salinas, Jesus Gonzalez – Hernandez, Facundo Ruiz. Synthesis of Silver Nanoparticles using Albumin as a Reducing agent. Materials Sciences and Applications. 2011; 2: 578-581.
 31. Anal K. Jha, K. Prasad. Green Fruit of Chilli (*Capsicum annum* L.) Synthesizes Nano silver. Digest Journal of Nanomaterials and Biostructures. 2011; 6(4): 1717-1723.
 32. S. Karthick Raja Namasivayam and Avimanyu. Silver nanoparticles synthesis from *Lecanicillium Lecanii* and Evolutionary Treatment on cotton Fabrics by measuring their improved antibacterial

- activity with antibiotics against *Staphylococcus Aureus* (ATCC 29213) and *E. coli* (ATCC 25922) Strains. *International Journal of Pharmacy and Pharmaceutical Sciences*, 2011; 3(4): 190-195.
33. Goldie Oza, Sunil pandey, Ritu Shah, Madhuri Sharon. Extracellular Fabrication of Silver Nanoparticles using *Pseudomonas aeruginosa* and its Antimicrobial Assay. *Advances in Applied Science Research*. 2012; 3(3): 1776-1783.
 34. Mousa Solgi and Mina Taghizadeh. Silver Nanoparticles Ecofriendly Synthesis by Two Medicinal Plants. *International Journal of Nanomaterials and Biostructures*. 2012; 2(4): 60-64.
 35. S. Dinesh, S. Karthikeyan and P. Arumugam. Biosynthesis of silver nanoparticles from *Glycyrrhiza glabra* root extract. *Archives of Applied Science Research*. 2012; 4(1): 178-187.
 36. Amar Ratan, Ekta Gupta and R. Ragunathan. Synthesis of Silver Nanoparticles using *Klebsiella Pneumonia* and its Bio-medical Applications. *International Journal of enhanced Research in Science Technology & Engineering*. 2013; 2(3): 1-7.
 37. Preetha Devaraj, Prachi Kumari, Chirom Aarti and Arun Renganathan. Synthesis and Characterization of Silver Nanoparticles using cannonball leaves and their cytotoxic activity against MCF – 7 cell line. 2013; 1-5.
 38. Rashmi Dwivedi. Silver Nanoparticles ecofriendly Green synthesis by using Two Medicinal Plant extract. *International Journal of Biotechnology and Research (IJBTR)*. 2013; 3(4): 61-68.
 39. Dr. P. Arumugam, Imrankhan Khansahib, Sarkarvyas Suriyanaarayanan. Green Synthesis of Nano-particles and its Application in Treatment of Rheumatoid Arthritis. *International Journal of Computing Algorithm*. 2013; 2: 450-457.
 40. Soumitra Mohanty, Prajna Jena, Ranjit Mehta, Rashmirekha Pati, Birendranath Banerjee, Satish Patil, Avinash Sonawane. Cationic Antimicrobial Peptides and Biogenic Silver Nanoparticles Kill *Mycobacteria* without Eliciting DNA Damage and Cytotoxicity in Mouse Macrophages. *Antimicrobial Agents and Chemotherapy*. 2013; 57(8): 3688-3698.
 41. Rama Koyyati, Veerababu Nagati, Ramchander Merugu and PratapRudra Manthurpadigya. Biological Synthesis of Silver Nanoparticles using *Raphanus Sativus* VAR. *Longipinnatus* leaf extract and Evaluation of their Antioxidant and Antibacterial activity. *International Journal of Medicine and Pharmaceutical Sciences (IJMPS)*. 2013; 3(4): 89-100.
 42. Priya Banerjee, Mantosh Satapathy, Aniruddha Mukhopahayay and Papita Das. Leaf extract mediated green synthesis of silver nanoparticles from widely available Indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. *Bio resources and Bioprocessing*. 2014; 1(3): 1-10.
 43. Muhammad Ajaz Hussain, Abdulla Shah, Ibrahim Jantan, Muhammad Raza Shah, Riaz Ahmed and Syed Nasir Abbas Bukhari. One pot light assisted green synthesis, storage and antimicrobial activity of dextran stabilized silver nanoparticles. *Journal of Nano biotechnology*. 2014; 53(12): 1-6.
 44. M. Srividhya, C. Mohanapriya, C. A. Akilan, K. Kathivelan, A. P. Subitha and M. A. Sundaramahalingam. Bioreduction based synthesis of silver nanocoats and their applications in development of nano embedded medical fabrics. *International journal of pharmaceutical sciences review and research*. 2014; 27(2): 210-215.
 45. R. K. Bhanisana Devi, H. N. K. Sarma, W. Radhapiyari and Ch. Brajakishore. Green synthesis, characterization and antimicrobial properties of silver nanowires by aqueous leaf extract of piperbetle. *International journal of pharmaceutical sciences review and research*. 2014; 26(1): 309-313.
 46. Sultana S. Biofabrication and extraction of silver nanoparticles using leaf extract of *azadirachta indica*. E-Thesis submitted to NIT Rourkela. Department of Life Science National Institute of Technology Rourkela-769008, Orissa, India. 2014; 1-43.
 47. M. Srividhya, C. Mohanapriya, C.A.Akilan, K.Kathirvelan, A.P.Subitha and M.A. Sundaramahalingam. Bioreduction Based Synthesis of Silver Nanocoats and their Application in Development of Nano Embedded Medical Fabrics. *International Journal of Pharmaceutical Sciences Review and Research*. 2014; 27(2): 210-215.
 48. Valli. G, Perlina. R and Anusuya. M. Biogenic synthesis, characterization of silver nanoparticles using *Tabernaemontana Divaricata* and *Polianthus Tuberosa* flower extract and evaluation of their antibacterial activities. *International journal of research in applied science and engineering technology (IJRASET)*. 2014; 2(8): 200-206.
 49. J. Chidambaram, K. Saritha, R. Maheswari and M. Syed Muzammil. Efficacy of green synthesis of silver nanoparticles using flowers of *Calendula officinalis*. 2014; 3(2): 773-777.
 50. K. L. Niraimathi, R. Lavanya, V. Sudha and P. Brindha. Green synthesis and characterization of silver nanoparticles from aqueous extract of *Basella alba* and their in-vitro antioxidant potentials. *International journal of pharmacy and pharmaceutical sciences*. 2014; 6(10): 393-396.
 51. Umadevi Parimi and Karteek Rao Amperayani. Green Synthesis of Plant Mediated Silver Nano Particles and Evaluation of their Antimicrobial Activities. *American journal of pharmatech research*. 2014; 4(3): 673-679.
 52. Hemali Padalia, Pooja Moteriya and Sumitra Chanda. Green synthesis of silver nanoparticles from marigold flower and its synergistic antimicrobial potential. *Arabian Journal of Chemistry*. 2014; 8(5): 732-741. [<http://dx.doi.org/10.1016/j.arabjc.2014.11.015>].
 53. D. R Manisha, Jahnvi Alwala, Karunakar Rao Kundle, M. P. Rudra. Biosynthesis of silver

- nanoparticles using flower extracts of *Catharanthus roseus* and evaluation of its antibacterial efficacy. *World journal of pharmacy and pharmaceutical sciences*. 2014; 3(5): 877-885.
54. Kantrao Saware, Balaji Sawle, Basav Raja Salimath, Kamala Jayanthi, Venkateswaraman Abbaraju. Biosynthesis and characterization of silver nanoparticles using *Ficus benghalensis* leaf extract. *International journal of research in engineering and technology*. 2014; 3(5): 867-874.
55. Preetha Devaraj, Renganathan Arun, Chirom Aarti, Prachi Kumari. Synthesis and characterization of silver nanoparticles using *Tabernaemontana Divaricata* and its cytotoxic activity against MCF-7 cell line. *International journal of pharmacy and pharmaceutical sciences*. 2014; 6(8): 86-90.
56. Priya Banerjee, Mantosh Satapathy, Aniruddha Mukhopahayay and Papita Das. Leaf extract mediated green synthesis of silver nanoparticles from widely available Indian plants: Synthesis, characterization, antimicrobial property and toxicity analysis. *Bioresources and bioprocessing*. 2014; 1(3): 1-10.
57. R. Tamileswari, M. Haniff Nisha, Sr. S. Jesurani, S. Kanagesan, M. Hashim, S. Catherine and P. Alexander. Synthesis of silver nanoparticles using the vegetable extract of *Raphanus sativus* (radish) and assessment of their antibacterial activity. *International journal of advanced technology in engineering and science*. 2015; 3(5): 207-212.
58. R. Tamileswari, M. Haniff Nisha and Sr. S. Jesurani. Green Synthesis of Silver Nanoparticles using *Brassica Oleracea* (cauliflower) and *Brassica Oleracea Capitata* (Cabbage) and the Analysis of Antimicrobial activity. 2015; 4(4): 1071-1074.
59. Boruah Himangshu, Talukdar Binita, Parveen Assma, Goswami Gunajit, Barooah Madhumita and Boro Robin Chandra. Biological synthesis of silver nanoparticles using *Colocassia* extract and their antimicrobial activity. *Research journal of biotechnology*. 2015; 10(2): 22-27.
60. Babu Vaishnavi, Ganesan Rameshkumar, Thangavel Rajagopal and Ponmanickam. Evaluation of bactericidal and fungicidal properties of silver nanoparticles fabricated using *Jasminium sambac* (L.). *Global journal of biotechnology & biochemistry*. 2015; 10(1): 23-31.
61. Priyanka Singh, Yeon Ju Kim, Hina Singh, Chao Wang, Kyu Hyon Hwang, Mohamed El – Agamy Farh, Deok Chun Yang. Biosynthesis, characterization and antimicrobial applications of silver nanoparticles. *International journal of Nanomedicine*. 2015; 10: 2567-2577.
62. B. Kannamba, P. Nagaraju and S. Lavanya. Synthesis of silver nanoparticles using *Muntingia calabura* leaf extract. *International journal of engineering research online*. 2015; 3(S1): 42-46.
63. S. Supraja and P. Arumugam. Antibacterial and anticancer activity of silver nanoparticles synthesized from *Cynodon dactylon* leaf extract. *Journal of academia and industrial research (JAIR)*. 2015; 3(12): 629-631.
64. Kaliyaperumal Viswanathan, M Latha Mala Priyadarshini, Karuppasamy Nirmala, Muthusamy Raman and Gopal Dhinakar Raj. Bactericidal paper trays doped with silver nanoparticles for egg storing applications. *Material science*. 2016; 39(3): 819-826.
65. J. Jeyasundari, P. Shanmuga Praba, Y. Brightson Arul Jacob, S. Rajendran and K. Kaleeswari. Green Synthesis and Characterization of Silver Nanoparticles Using *Mimusops elengi* Flower Extract and Its Synergistic Antimicrobial Potential. *American chemical science journal*. 2016; 12(3): 1-11.
66. Zainal Abidin Ali, Rosiyah Yahya, Shamala Devi Sekaran and R. Puteh. Green Synthesis of Silver Nanoparticles Using Apple Extract and Its Antibacterial Properties. *Advances in materials science and engineering*. 2016; 1-6. [<http://dx.doi.org/10.1155/2016/4102196>].
67. Shibani Basu, Priyankar Maji and Jhuma Ganguly. Rapid green synthesis of silver nanoparticles by aqueous extract of seeds of *Nyctanthes arbor-tristis*. *Applied naoscience*. 2016; 6(1): 1-5.