

CHANGES IN ELECTROLYTES LEVEL OF *CLARIAS GARIEPINUS* (BURCHELL 1822) EXPOSED TO SUBLETHAL CONCENTRATIONS OF OILFIELD WASTEWATER

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ABSTRACT

The electrolytic changes of twenty-eight adult *Clarias gariepinus* (Burchell, 1822), (mean weight $205 \pm 12.89g$ SD; Mean length; $31.13 \pm 3.82cm$ SD) exposed to sub-lethal concentrations (0, 10, 20, 30, 40, 50 and 60%) of an oilfield wastewater over a period of twenty eight (28) days were studied in a semi-static system. At the end of 28 days, tissue/organ and plasma samples were collected for electrolysis using standard methods. Insignificant ($p \leq 0.05$) fluctuations in electrolytes (sodium, potassium, chloride, calcium and bicarbonates ions) were observed in exposed fishes when compared to the control group. These fluctuations in electrolytes suggest the effort made by the fish to combat the stress imposed by the toxicant. The ions (electrolytes) are involved in regulation of osmotic changes in the fish and can be used to measure long term effect of *C. gariepinus* exposed to oilfield wastewater. The proper treatment of oilfield wastewater prior to discharge into the recipient water body is advocated to reduce ecotoxicological problems.

KEYWORDS: *Clarias gariepinus*, ecotoxicology, electrolytes, oilfield wastewater, organ, tissue.**INTRODUCTION**

Electrolytes are charged minerals found in organisms with enormous functions.^[1] These electrolytes include sodium, potassium, chloride, calcium and bicarbonates. Sodium and potassium are the major cations of the extracellular fluids while chloride and bicarbonates are the major anions of the intracellular fluids.^[2,3] Grouped as divalent and monovalent ions according to functions, the divalent ions (Calcium and Magnesium) are important in neuromuscular excitability, enzymatic reactions and retention of membrane permeability.^[1,4] Accordingly, the monovalent ions (Chloride, Potassium and Sodium) are key in acid to base balance (pH), osmotic pressure as well as neuromuscular excitability.^[5,4] Also, inorganic phosphate is important in energy exchange as it is an important cytoplasmic buffer.^[4,6,7] Electrolytes are useful in redox (electron transfer) processes and as co-factors for enzymes.^[7] Sodium is essential for the activity of many enzymes and has been implicated in the transport of ATP. Sodium and potassium ATPase are located in the cell membrane and are involved in the active transport of Na^+ and K^+ across the cell membrane.^[8]

Clarias gariepinus is a fish of choice in aquaculture due to its hardy nature, ease of larval production in captivity and good market price.^[9,10] It is also wide bred because of their palatability, fecundity, disease resistance and high growth rate.^[11] Further, *Clarias* is available at all seasons, is easy to use in the laboratory and has relative

sensitivity (high level of tolerance) to petroleum products.

Anthropogenic activities have placed the environment at the receiving end of numerous contaminants.^[12] Environmental contaminants are known to alter the levels of electrolytes in fish.^[13,14] In recent years, the concern on the pollution of the environment by produced or formation water has been on the aquatic environments.^[15,16,17] This is due to marked increase in offshore oil operations. A few studies have been carried out on the discharge of produce water or oilfield waste water from onshore operations into terrestrial environments.^[18,19,20]

The potential toxicity of oilfield wastewater on exposed biota is notable, although their actual effect is not clear. The physicochemical properties of oilfield wastewater indicate that it could pose serious threat to biota.^[21] The disposal of such effluents on land during onshore operations could pose a threat to exposed fish in captivity especially fish ponds located around such area. However, not much data is available on the effect of this oilfield waste water on fish and other biota. Fish live in very intimate contact with their environment.^[22,23,24] These indices have been effectively employed in monitoring the responses of the fish to the stressors and evaluating its health status under such adverse conditions. This study therefore aims at evaluating the electrolyte changes of *C.*

gariiepinus exposed to sublethal concentrations of oilfield wastewater.

MATERIALS AND METHODS

Test Organism

Twenty-eight adult (mean weight 205 ± 12.89 g SD; mean length; 31.13 ± 3.82 cm SD) of *C. gariiepinus* were obtained from the African Regional Aquaculture Center (ARAC) at Aluu in Ikwerre Local government area of Rivers State. The *C. gariiepinus* were put into 50 L trough containing borehole water and sealed with a net before transporting to the laboratory for analysis. The fish were then acclimated individually in covered rectangular aquaria for two weeks. The water was changed daily and the aquaria washed with a piece of foam and water. All samples were fed twice with a 35% crude protein diet at 1% biomass daily (8.00 a.m. and 5.00p.m.). Mortality during acclimation was less than one percent. All fish samples were treated in line with recommended scientific procedure.

Test toxicant (oilfield wastewater)

The test toxicant (oilfield wastewater) was collected from Ebocha oilfield in Ogba/Egbema Local Government Area of Rivers State with coordinates N05 27' 40.45' E006 41' 52.14'. The effluent was collected in 50 litres plastic containers on three occasions and transported to the laboratory. These represented different ranges of the discharge at the discharge point.

Optimization Test (Preliminary Investigation)

The range or concentrations of oilfield wastewater that exhibited lethal and sub-lethal effect on the fingerlings and adult of *C. gariiepinus* respectively was determined using standard procedures. Five concentrations (10%, 30%, 50%, 70% and 100%) of the oilfield wastewater were prepared by serial dilution from each effluent sample on a volume to volume (v/v) ratio. The percentage (%) concentration in each test solution was obtained by using the formula below^[25]:

$$\text{Concentration of effluent(\%)} = \frac{V_E}{V_E + V_{DW}} \times 100$$

Where,

V_E = Volume of effluent

V_{DW} = Volume of dilution water

The determined volume of effluent was added to the desired quantity of dilution (borehole) water and vigorously mixed.

The water was not changed for a period of one week. However, the fish were fed twice daily as in the acclimation period. Concentrations that caused death within one week were omitted from the definitive test.^[26] Since the fingerlings did not die after 96 hours of exposure the lethal test for the fingerlings was discontinued.

Sub-lethal test

Based on the results from the optimisation test, 10, 20, 30, 40, 50 and 60% v/v of oilfield wastewater were prepared as the sub-lethal concentrations and used for the study, including a control. Each treatment was set up in four replicates. A fish was then introduced into each of these concentrations contained in an aquarium and incubated for a period of 28 days at room temperature ($30 \pm 2^\circ\text{C}$). Fifteen litres of each prepared concentration was used and fish was fed as in the acclimation period. The test solution was renewed weekly after washing the aquarium with a piece of foam to get fresh toxicant to maintain concentration.

Electrolyte Analysis

After 28 days, blood samples were collected from the fish (behind the anal fin) using a 23G size needle and syringe for electrolyte analysis. Then, the Fish were killed and dissected to obtain samples of the gill, liver, kidney, and muscle tissue. Using pestle and mortar, 0.5g each of the organ/tissues were macerated. To each of the samples, 5ml of deionised water was added, centrifuged at 3000 rpm for 10 minutes. The supernatants were then extracted and stored in plain bottles at -20°C and electrolytes analysed.

The electrolytes analyses were done for Sodium (Na^+), Chloride (Cl^-), Calcium (Ca^{2+}), Potassium (K^+) and Bicarbonates (HCO_3^-) according to Ochei and Kolhatkar.^[4]

Calculations

Absorbance of blank — absorb of sample x conc. of standard
Absorb. of blank — absorbance of standard = concentration of K (mmol/ L).

RESULTS AND DISCUSSION

Environmental pollutants have been known to alter levels of electrolytes in fish^[13,14] and can be used to monitor the responses of the fish to the stressors and evaluating its health status under such adverse conditions.

In this study, significant changes ($p \leq 0.05$) in the electrolytes of *C. gariiepinus* exposed to oilfield wastewater was observed between the control and some treated groups as well as among the treated fish (Table 1). Although no given trend was observed, the value of sodium in plasma ranged between 65.75 ± 8.09 mmol/l at 30% and 142.00 ± 7.48 mmol/l at the control; Chloride ranged between 108.00 ± 16.17 mmol/l at 20% and 134.75 ± 1.50 mmol/l at 40% and Calcium ranged between 6.00 ± 0.00 mmol/l at 50% and 10.50 ± 1.73 mmol/l at 30%. Similarly, Potassium ranged between 6.20 ± 0.48 mmol/l at 20% and 8.55 ± 0.48 mmol/l at 40% while bicarbonates ranged between 3.70 ± 0.00 mmol/l at 30% and 8.48 ± 4.71 mmol/l at 50% of the toxicant.

Table 1: Electrolytes in the Plasma of *C. gariepinus* exposed to different concentrations of an oilfield wastewater after 28 days.

Conc. of Oilfield wastewater (%)	Electrolytes(mmol/l)				
	Na ⁺	Cl ⁻	Ca ²⁺	K ⁺	HCO ³
0	142.00±7.48 ^c	110.50±6.35 ^{ab}	7.50±0.58 ^{ab}	6.48±0.61 ^a	6.50±3.07 ^a
10	84.00±3.65 ^b	122.00±13.86 ^{abc}	9.50±2.89 ^{bc}	6.95±0.52 ^a	6.53±2.98 ^a
20	131.00±4.16 ^c	108.00±16.17 ^a	7.50±2.89 ^{ab}	6.20±0.48 ^a	5.55±0.06 ^a
30	65.75±8.09 ^a	133.50±.58 ^c	10.50±1.73 ^b	6.78±0.51 ^a	3.70±0.00 ^a
40	99.50±9.98 ^c	134.75±1.50 ^c	8.00±1.16 ^{ab}	8.55±0.48 ^c	7.35±2.02 ^a
50	118.75±2.22 ^d	127.00±0.00 ^{bc}	6.00±0.00 ^a	7.43±1.25 ^{ab}	8.48±4.71 ^a
60	112.00±12.96 ^d	127.50±6.35 ^{bc}	6.50±0.58 ^a	7.53±1.03 ^{ab}	5.55±2.14 ^a

*Means with the same superscript in the column are not significantly different at $p \leq 0.05$

Electrolytes in the gills of *C. gariepinus* showed a significant difference ($p \leq 0.05$) at the determined concentrations, although they did not follow any trend (Table 2). Electrolytes in the liver of *C. gariepinus* exposed to the toxicant are as shown table 3. There was a significant difference at $p \leq 0.05$. Generally higher values were recorded for sodium, chloride, calcium and potassium at 60% of the toxicant, but at the other

concentration including the control there was no trend followed. Bicarbonate was highest at 50% while at other concentrations there was no significant difference at $p \leq 0.05$. Similarly, electrolytes in kidney of *C. gariepinus* exposed to the toxicant showed a slight difference ($p \leq 0.05$) at the constituted concentrations although with no defined trend (Table 4).

Table 2: Electrolytes in the gills of *C. gariepinus* exposed to different concentrations of an oilfield wastewater after 28 days.

Conc. of Oilfield wastewater (%)	Electrolytes (mmol/l)				
	Na ⁺	Cl ⁻	Ca ²⁺	K ⁺	HCO ³
0	557.50±68.07 ^a	125.00±16.17 ^a	5.00±0.00 ^a	13.75±3.20 ^a	20.00±5.77 ^a
10	622.50±72.74 ^a	120.00±15.00 ^a	15.00±5.77 ^b	12.50±0.71 ^a	25.00±0.00 ^a
20	982.50±56.79 ^b	138.13±38.42 ^b	5.00±0.00 ^a	14.50±0.58 ^a	36.25±9.47 ^b
30	502.50±12.58 ^a	83.50±0.00 ^a	5.00±0.00 ^a	16.25±1.44 ^a	45.00±5.77 ^b
40	581.25±56.77 ^a	96.75±15.30 ^a	10.00±5.77 ^a	18.75±4.33 ^a	58.75±11.82 ^c
50	535.00±87.37 ^a	110.00±0.00 ^a	5.00±0.00 ^a	15.00±0.00 ^a	22.50±2.89 ^a
60	500.00±8.17 ^a	122.50±14.43 ^a	5.00±0.00 ^a	17.63±5.63 ^a	45.00±5.77 ^b

*Means with the same superscript in the column are not significantly different at $P \geq 0.05$.

This study demonstrated fluctuations in ions of fish exposed to concentrations of oilfield wastewater as compared with the control. Also, test groups showed significant changes in the electrolytes level in the organs studied. However, no remarkable trend was observed in most results. This may be due to disturbances in ionic concentration of the water. Studies have shown the need for strict ionic regulation (homeostasis) in aquatic animals.^[27] There were significant changes ($p \leq 0.05$) in plasma electrolytes in the fish studied except bicarbonates that did not change at the various concentrations of the wastewater. Plasma electrolytes are known to fluctuate in response to various factors.^[28]

These disturbances in homeostasis are induced by pollutants evidenced by altered plasma ion concentration.^[28] Other studies report that the concentrations of monovalent ions in plasma are commonly altered in response to pollutants.^[29,30] The fish in the present study showed fluctuations due to exposure to produced water (a pollutant). Sodium, potassium and calcium are strongly electropositive and not easily oxidized or reduced.^[3] Thus biologically, they have a major role in neutralizing the charge on anions and in maintaining isotonic pressure in natural fluids.^[31] This was observed in this study.

Table 3: Electrolytes in the Liver of *C. gariepinus* exposed to different concentrations of an oilfield wastewater after 28 days.

Conc. of Oilfield wastewater (%)	Electrolytes (mmol/l)				
	Na ⁺	Cl ⁻	Ca ²⁺	K ⁺	HCO ³
0	608.75±31.19 ^b	83.50±0.00 ^a	5.00±0.00 ^a	15.75±0.87 ^c	50.00±0.00 ^a
10	567.50±39.69 ^b	69.25±16.46 ^a	7.50±2.89 ^a	17.25±0.96 ^c	40.00±0.00 ^a
20	550.00±8.17 ^b	96.75±15.29 ^a	5.00±0.00 ^a	7.00±1.83 ^a	45.00±5.77 ^a
30	270.00±21.60 ^a	110.00±0.00 ^a	11.25±6.29 ^a	6.50±1.73 ^a	45.00±5.77 ^a
40	608.75±63.03 ^b	83.50±0.00 ^a	7.50±2.89 ^a	13.25±2.02 ^b	62.50±14.43 ^b
50	558.75±26.89 ^b	82.50±31.75 ^a	12.50±8.66 ^a	22.75±2.22 ^d	45.00±5.77 ^a
60	717.50±23.63 ^c	166.50±31.75 ^b	15.00±5.77 ^a	34.00±0.58 ^e	45.00±5.77 ^a

*Means with the same superscript in the column are not significantly different at $p \leq 0.05$

Table 4: Electrolytes in the Kidney of *C. gariepinus* exposed to different concentrations of an oilfield wastewater after 28 days.

Conc. Of Oilfield wastewater (%)	Electrolytes(mmol/ L)				
	Na ⁺	Cl ⁻	Ca ²⁺	K ⁺	HCO ³⁻
0	745.00±72.34 ^c	83.50±0.00 ^a	5.00±0.00 ^a	17.50±1.73 ^b	35.00±5.77 ^a
10	455.00±30.28 ^a	83.50±0.00 ^a	5.00±0.00 ^a	17.00±2.31 ^b	42.50±9.57 ^a
20	650.00±47.61 ^b	83.50±0.00 ^a	12.50±2.89 ^b	16.25±1.44 ^b	67.50±8.66 ^b
30	500.00±23.81 ^a	83.50±0.00 ^a	7.50±2.89 ^a	18.25±0.87 ^b	45.00±5.77 ^a
40	477.50±22.17 ^a	69.25±16.45 ^a	5.00±0.00 ^a	23.25±0.87 ^c	45.00±5.77 ^a
50	446.25±40.29 ^a	83.50±0.00 ^a	7.50±2.89 ^a	16.75±0.87 ^b	40.00±0.00 ^a
60	645.00±42.03 ^b	69.25±16.46 ^a	5.00±0.00 ^a	8.63±1.32 ^a	40.00±11.55 ^a

*Means with the same superscript in the column are not significantly different at $P \leq 0.05$.

In muscle of *C. gariepinus*, significant difference ($p \leq 0.05$) was observed in the value of electrolytes at the constituted concentrations (Table 5). Sodium, chloride and bicarbonate recorded higher values of 792.50±17.08mmol/l, 165.00±0.00mmol/l and

45.00±5.77 mmol/l respectively at 60% while calcium and potassium was the reverse as higher values of 20.00±8.17mmol/l and 46.00±2.94 mmol/l were recorded in the control (0%). The results did not follow any particular trend.

Table 5: Electrolytes in the Muscle of *C. gariepinus* exposed to different concentrations of an oilfield wastewater after 28 days.

Conc. of Oilfield wastewater (%)	Electrolytes(mmol/l)				
	Na ⁺	Cl ⁻	Ca ²⁺	K ⁺	HCO ³⁻
0	515.00±69.52 ^c	110.00±0.00 ^c	20.00±8.17 ^b	46.00±2.94 ^c	25.00±5.77 ^a
10	510.00±75.28 ^c	96.75±15.30 ^b	5.00±0.00 ^a	27.75±2.02 ^a	25.00±5.77 ^a
20	332.50±8.66 ^b	110.00±0.00 ^c	5.00±0.00 ^a	36.00±0.58 ^b	25.00±5.77 ^a
30	305.00±5.77 ^{ab}	83.50±0.00 ^a	5.00±0.00 ^a	26.75±0.96 ^a	35.00±17.32 ^a
40	257.50±9.57 ^a	83.50±0.00 ^a	5.00±0.00 ^a	33.50±2.94 ^b	40.00±11.55 ^a
50	597.50±17.08 ^d	83.50±0.00 ^a	16.25±6.29 ^b	29.63±1.44 ^a	45.00±5.77 ^a
60	792.50±17.08 ^e	165.00±0.00 ^b	8.75±4.79 ^a	27.13±1.89 ^a	45.00±5.77 ^a

*Means with the same superscript in the column are not significantly different at $p \leq 0.05$.

Changes in the electrolytes level had no trend in all the parts studied. This could be based on the methods of analysis used and various factors both exogenous and endogenous.^[32] It may be necessary to consider these factors in future studies. Fish in disturbed osmotic environment behaviorally adjust their electrolyte.^[32] This further supports the lack of trend in the results of the present study. The gills play important role in the behavioral ionic regulation of fish. The results on the electrolyte levels of the gills of the test fish show effort to neutralize the effect of pollutants as in other studies.^[9,10] This is also true for the kidneys. This further demonstrates the hardy nature of *C. gariepinus*.

The lack of trend in the electrolytes analysed in these organs may be due to stress induced degeneration of these organs.^[33,32] These studies report that exposure to toxicants could lead to degeneration. Kidney degeneration has been implicated for reabsorption.^[33,34,29] This may explain uneven trends in calcium, phosphates and other electrolytes.

Similarly, fluctuations were observed in the chloride and bicarbonates levels in the tissues of exposed fishes when compared to the control group. Gabriel *et al.*^[14] noted that levels of electrolytes in organs and tissue of *Heterobranchus bidorsalis* exposed to concentrations of

cypermethrin were either higher or lower than the control value. This is a reflection of changes in the fluxes of the electrolytes as a result of cypermethrin toxicosis. The presence of oilfield wastewater may be responsible for the fluctuations observed in this study.

Further, the fluctuations in electrolytes analysed could be linked to heart stress. Reduction of Na⁺ ion level may affect the heart functions and cause neurotoxic damage to the central nervous system (CNS) of fish.^[35] According to Cox^[36] cypermethrin alters nerve impulse travels along nerves of vertebrates and other animals. The nerves become momentarily permeable to sodium atoms, thus allowing it to flow into nerves. It also delays the closing of the gate that allows the sodium flow.^[37] This eventually results in multiple nerve impulses instead of the usual single. In turn, these impulses cause the nerve to release the neurotransmitter, acetylcholine and then stimulate other nerves.^[14] Multiple impulses may result when fish are exposed to toxicants.

Further, the lack in trend observed in this study could indicate osmoregulation in the fish tested. Electrolyte concentrations have been used as an indicator of fishes' ability to osmo-regulate.^[38] This is often compromised with stress, disease or gill lesions that increase gill permeability to ions or lateral line imbalance and

hormonal disorder.^[39] Electrolytes have been shown to play a central role in gaseous exchange and intercompartmental water balance.^[4] Hence elevated or low levels may result in hyper or hypo functions of tissues or organs of fish.^[40,14]

CONCLUSION

The current study observed fluctuations in electrolytes in the fish exposed to sublethal concentrations of oilfield wastewater. This indicates the effort made by the fish to combat the stress imposed by the toxicant. The ions (electrolytes) involved in regulation of osmotic changes in the fish can be used to measure long term effect of *C. gariepinus* exposed to oilfield wastewater. The proper treatment of oilfield wastewater prior to discharge into water body is advocated to reduce ecotoxicological problems.

REFERENCES

- Hasona NA, Elasbali A. (Evaluation of electrolytes imbalance and dyslipidemia in diabetic patients). *Medical Sciences*, 2016; 4(2): 7.
- Bhanu AP. (Cypermethrin induced alterations in Electrolytes in the blood and tissues of *Cyrius carpio*). *International Journal of Technical Research and Applications*, 2016; 4(3): 248-250.
- Adeoye A. A text book of medical laboratory practice 1st edition., 2007; 238.
- Teila MA. (Serum electrolyte changes in West African dwarf (WAD) sheep with single or concurrent (Bebesia ovis and Trypanosome congolense) infections. *African Journal of Biological Research*), 2005; 8: 63 - 65.
- Ochei JO, Kolhatkar AA. *Medical Laboratory Science: Theory and Practice* Tata McGraw-Hill Publishing Company Limited, New York, 2008; 637-745.
- Aurbach GD, Marx SJ, Spigel AM. Parathyroid hormone calcitonin and calciferols. In, Wilson JD, Foster DW. *Williams textbook of endocrinology*. W.B. Saunders Company Philadelphia, 1985; 1137-1217.
- Mohanty BK, Mishra BN. (Effects of mercurial drugs (Kajyoli) on albino rat blood). *Journal of Environmental Biology*, 1983; 4(4): 201 – 206.
- Ragama B, Chapatwala KD, Vaishnav DD, Desai D. (Changes in ATPase activity in tissues of rat fed on cadmium). *Journal of Environmental Biology*, 1981; 2(1): 1 – 9.
- Efuntoye MO, Omotosho OO, Ashidi JS. (Prevalence of methicillin-resistant *Staphylococcus aureus* and coagulase negative *Staphylococci* among male students in a private tertiary institution and their enterotoxin-producing potential). *Asian Journal of Pharmaceutical Health Care and Sciences*, 2012; 2: 231-234.
- Herald SE. *Living Fishes of the World*, 1st Edition. Doubleday and Company Inc., New York, 1971; 126.
- Ikpi G, Offem B. (Bacterial infection of mudfish *Clarias gariepinus* (Siluriformes Clariidae) fingerlings in tropical nursery ponds). *Revista de Biología Tropical*, 2011; 59: 751-759.
- Gillings MR., Gaze, WH, Pruden A, Smalla K, Tiedje J M, Zhu YG. (Using the class 1 integron-integrase gene as a proxy for anthropogenic pollution). *The ISME Journal*, 2015; 9(6): 1269.
- Pracheil BM, Adams SM, Bevelhimer M S, Fortner AM, Greeley MS, Murphy CA, Peterson MJ. (Relating fish health and reproductive metrics to contaminant bioaccumulation at the Tennessee Valley Authority Kingston coal ash spill site). *Ecotoxicology*, 2016; 25(6): 1136-1149.
- Gabriel UU, Jack IR, Edori OS, Egobueze E. (Electrolytes of selected tissues of *Heterobranchus bidorsalis* treated with sublethal levels of cypermethrin). *Ethiopian Journal of Environmental Studies and Management*, 2009; 2(3): 83 – 87.
- Obire O, Amusan FO. (The Environmental Impact of Oilfield Formation Water on a Freshwater stream in Nigeria). *Journal of Applied Science and Environmental Management*, 2003; 7(1): 61 - 66.
- Wills J. A survey of offshore oilfield drilling wastes and disposal techniques to reduce the ecological impact of sea dumping, the effect of discharges of produced waters. *Ekologičeaya Vahktan Sakhalina (Sakhalina Environmental Watch)* 25th May, Sakhalina, London, 2000; 1-5.
- Somerville HJ, Benneth D, Davenport JN, Holt MS, Lynes A, Mahieu A, Mccourt B, Parker JG, Stephenson RR, Watkinson RJ, Wilkinson TG. (Environmental Effect of Produced Water from North Sea Oil Operations). *Marine Pollution Bulletin*, 1987; 18(10): 549-558.
- Obire O, Wemedo SA. (Seasonal effect on the Bacterial and Fungal population of an oilfield wastewater polluted soil in Nigeria). *Journal of Applied Science and Environmental Management*, 2002; 6(2): 17 - 21.
- Obire O Wemedo SA. (The effect of oilfield Wastewater on the microbial population of a soil in Nigeria). *Niger Delta Biologia*, 1996; 1(1): 77 - 85.
- Odeigah PGC, Nuruddeen O, Amund OO. (Genotoxicity of oilfield wastewater in Nigeria). *Hereditas*, 1997; 126: 161 - 167.
- Akani NP, Ugbomeh AO, Gabriel UU. (Evaluation of metabolites of *Clarias gariepinus* exposed to sublethal concentrations of oilfield wastewater). *International Journal of Ecotoxicology and Ecobiology*, 2017; 2(4): 150-157.
- Akani NP, Gabriel UU. (Enzymatic Responses of *Clarias gariepinus* (Burchell, 1822) exposed to sublethal concentrations of an oilfield wastewater). *Journal of Applied Science and Biotechnology*, 2016; 4(05): 026 – 032.
- Akani NP, Obire O. (Mycological flora of *Clarias gariepinus* exposed to an oilfield wastewater in Nigeria). *International Journal of Microbiology and Mycology*, 2015; 3(1): 1 – 9.

24. Wilson RW, Taylor EW. (The physiological responses of freshwater rainbow trout, *Onchorynchus mykiss* during acute exposure). *Journal of Comparative Physiology*, 1993; 163b: 38-47.
25. FAO. (Meeting on the toxicity and bioaccumulation of selected substances in marine organisms). FAO Fisheries Report, Rovinj, Yugoslavia, Nov.FIR/R334, 1984; 334: 5 – 9.
26. Gurure RM. (Influence of two organo-chloride pesticides, thiodan and lindane on survival of fingerlings of *O. niloticus* and *T. zilli*). African Regional Aquaculture Centre Working Paper ARAC/WP.6/87, 1978.
27. Takei Y. (Comparative physiology of body fluid regulation in vertebrates with special reference to thirst regulation). *The Japanese journal of physiology*, 2000; 50(2): 171-186.
28. Nilsson S, Holmgren S. *Fish physiology, recent advances* British library cataloguing in publication data. Croom Helm, 51 Washington Street, Dover, New Hampshire 03820, USA. 0-7099-1837-2, 1986.
29. Larsson A, Bengtsson BE, Haux C. (Disturbed ion balance in flounder; *platichthys flesus* L. exposed to sublethal levels of cadmium). *Aquatic Toxicology*, 1981; 1: 19-35.
30. Larsson A, Haux C, Sjobeck ML. (Fish physiology and metal pollution, results and experiences from laboratory and field studies). *Ecotoxicology and Environmental Safety*, 1985; 9: 250-281.
31. Sukling CJ. *Enzyme chemistry, impact and application*. 2nd. Ed. Printed in Great Britain by T.J. press Ltd, Padstow, Cornwall, 1990; 250 - 251.
32. Kopp R, Mares J, Lang S, Brabec T, Zikova A. (Assessment of ranges plasma indices in rainbow trout (*Oncorhynchus mykiss*) reared under conditions of intensive aquaculture). *Acta Universitatis Agriculturae et Silviculturae Mendelienae Brusensis*, 2011; 24(6): 181-188.
33. Srivastav AK, Rai R, Suzuki N, Mishra D, Srivastav SK. (Effects of lead on the plasma electrolytes of a freshwater fish, *Heteropneustes fossilis*). *International Aquatic Research*, 2013; 5(4): 1-7.
34. Oluwole SF. (Effects of garlic on some haematological and biochemical parameters). *African Journal of Biomedical Research*, 2001; 4: 139 – 141.
35. Zimmer T, Haufe V, Blechschmidt S. (Voltage-gated sodium channels in the mammalian heart). *Global Cardiology Science and Practice*, 2014; 4: 58.
36. Cox C. (Cypermethrine). *Journal of Pesticide Reform*, 1996; 16(2): 15 – 20.
37. Vijverberg HPM, Van den Bercken J. (Neurotoxicological effect and mode of action of Pyrethroid insecticides). *Critical Reviews in Toxicology*, 1990; 21: 105 - 120.
38. Alam MS, Watanabe WO, Myers AR, Resek TC, Carroll PM, Skrabal SA. (Effects of dietary salt supplementation on growth, body composition, tissue electrolytes and gill and intestinal Na⁺/K⁺ ATPase activities of black sea bass reared at low salinity). *Aquaculture*, 2015; 446: 250-258.
39. McDonald DG, Milligan CI. *Chemical properties of blood*. In, Hoar, W. S. Randall, D. J. and Farrell, A. P. (Eds.), *Fish Physiology 12B, The Cardiovascular system*, 1992; 56 – 135.
40. Finco DR. *Kidney function*. In: *Clinical biochemistry of domestic animals*. 4th Ed. J.J. Kanekoo Ed., Academic Press Inc. San Diego, California, 1989; 496-537.