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LIMITATIONS OF NON-STEROIDAL ANTI-INFLAMMATORY DRUGS AND THE UTILITY OF NATURAL PRODUCTS FOR ANTINOCICEPTIVE AND ANTIEXUDATIVE EFFECTS

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ABSTRACT

Nonsteroidal anti-inflammatory drugs (NSAIDs) express anti-inflammatory, analgesic and antipyretic effects, essentially by reversible inhibition of the rate-limiting enzyme-cyclooxygenase (COX), in the biosynthesis of prostaglandins. Among the dominant widely used medications globally are NSAIDs and are indicated for various forms of acute and chronic pain, rheumatoid arthritis, gout and pyrexia. Also some of them (aspirin for instance) are employed for prevention of secondary cardiovascular disorder due to mainly their ability to suppress platelet aggregation. Despite these applications, NSAIDs are known to present limitations in their untoward effects, hence researchers in recent times, have focused their efforts toward various chemical modifications of NSAIDs such as nitro-aspirin, des-methyl (DM)-sulindac, phosphatidylcholine (PC)-aspirin, pegylated phosphoribuprofen, dual COX/LOX inhibitors, Copper (II)/NSAIDs complexes; as well as natural product system of medicine for the discovery of agents that exert efficacy and potency with minimum adverse effects. This study reviewed the mechanisms of action, clinical applications, potential antitumor activity, interactions and limitations of NSAIDs as well as natural products with promising antinociceptive and anti-inflammatory (mainly anti-exudative) activities, utilizing the advantage of the frequent occurance of natural products as complexes of structurally related analogues to be exploited for good efficacy and safety in humans.

KEYWORDS: NSAIDs, limitations, natural products, Chikadoma, antinociceptives, antiexudatives.

1. INTRODUCTION

The first representative of a group of medications that reduce inflammation, relieve pain and fever referred to as Nonsteriodal anti-inflammatory drugs (NSAIDs) is acetyl salicylic acid commonly called by its proprietary/trade name "aspirin" courtesy of the Bayer company. This prototype drug was put into commercial form in 1897 by a German Chemist Felix Hoffmann (1868-1946) and later introduced into clinical practice in 1899. More than 1 billion prescriptions of NSAIDs is estimated throughout the world yearly, and about 30 million persons administer NSAIDs daily. [1]

With more than 40,000 tonnes of "consumption" on yearly basis. [2] However, numerous limitations are associated with NSAIDs, hence this study was aimed at reviewing the utility of natural products for antinociceptive and antiexudative effect with diminished adverse tendencies as against the limitations of NSAIDs.

2. NSAIDs

2.1 Classification

NSAIDs can be classified based on their chemical structure or according to their ability to influence activity of the COX isoenzymes.^[3, 2] Based on their structure, there are the following classes of derivatives: Salicylate, propionic acid, pyrazolone, indoleacetic acid, phenylacetic acid/fenamate, oxicam/enolic acid, and niftyloacetic acid derivatives (Table 1); while based on their influence toward COX isoenzyme, NSAIDs can be classified into non-selective and selective COX-inhibitors (Table 2).

Table 1: Classification of NSAIDs based on their chemical structure.

Group (Derivatives)		Example	
- Salicylate (salicylic acid)	_	Acetysalicylic acid (Aspirin), sodium salicylate, methyl Salicylate.	
- Propionic acid	-	Ibuprofen, ketoprofen, naproxen, fenoprofen, fluribiprofen.	
- Pyrazolone	-	Phenylbutazone, oxyphenbutazone, sulfinpyrazone.	
- Indoleacetic acid	-	Indomethacin, sulindac, tolmetin	
- Fenamate (phenylacetic acid)	-	Diclofenac, meclofenamate, aceclofenac	
- Oxicam (enolic acid)	-	Piroxicam, meloxicam, tenoxicam	
- Niftyloacetic acid	-	Nabumetone	

Table 2: Classification of NSAIDs based on influence upon cyclooxygenase isoenzymes. [3, 2]

Group (inhibitors)		Example
Non-selective COX	-	Piroxicam, acetylsalicylic acid (Aspirin)
Ibuprofen, neproxen		
Selective COX-1	-	Flurbiprofen, ketoprofen
Sective COX-2	_	Coxibs (celecoxib, Etoricoxib, lumiracoxib)
		Diclofenac, mefenamic acid

Table 3: Characteristics of COX-isoenzymes. [4,5]

Parameter	COX-1	COX-2	
Gene size	25 kb	8.6 kb	
Exons	15	10	
Chromosome	9q33.2	1q31.1	
mRNA	2,8 kb	4.1 kb	
Gene expression	Constitutive (ubiquitous)	Constitutive (brain, thymus, gut,	
-		and kidney) and inducible (sites	
		of inflammation, infection,	
Factors promoting expression	-	cytokines, lipopolysaccharide,	
Amino acid composition	599	604	
Molecular weight	68.5 kDa	70 kDa	
Cell localization	Nuclear envelope, endoplasmic	Nuclear envelope, endoplasmic	
	reticulum	reticulum	
Cofactors	1 molecule heme	1 molecule heme	
Acetylated regions	Ser 530	Ser 516	
Substrates	arachidonic acid, aminoacids,	arachidonic acid, aminoacids,	
	γ-linoleic acid	γ-linoleic acid, α- linoleic,	
		eicosapentaenoic acid	

The classification of NSAIDs based on cyclooxygenase isoenzymes into COX-1 and COX-2 is not absolute rather relative since in a concentration dependent manner, all NSAIDs can inhibit both COX-1 and COX-2 activity in vitro. It is the ratio between the 50% inhibitory concentration (1C₅₀) values that has been employed in the determination of the rate of COX-2 selectivity of NSAIDs. Even the most selective COX-2 inhibitors such as lumiracoxib and etoricoxib, can also inhibit COX-1 activity when administered at high doses. [6] Aspirin vice versa, is a prototype non-selective COX inhibitor but at low doses (75-300 mg) it reduces platelet aggregation by predominantly inhibiting more than 70% COX-1 and less than 5% COX-2.^[7] The same aspirin at higher doses (more than 1,200 mg) manifests pronounced anti-inflammatory and analgesic effects by inhibition of both COX-1 and COX-2 isoenzymes. [5]

2.2 Mechanism of action

Reversible suppression of the function of COX is the basis of biological activity of NSAIDs COX, reputed as the rate-limiting enzyme in the biosynthesis of

prostaglandin, is a homodimeric enzyme necessary for the conversion of arachidonic acid to prostaglandine H₂. This is the first step that is crucial for further biosynthesis of prostanoids and thromboxanes). Two isoforms of this protein are known to exist in human namely COX-1 and COX-2. COX-2 is found to be inducible isoform expressed only at sites of inflammation, cancer or infection; while COX-1 is constitutive and expressed in all tissues. COX-1 plays an important role in homeostasis and regulates gastric cytoprotection; thrombosis, platelet aggregation, and kidney function. The concentration of COX-2 can be raised up to 10-50 times in response to mitogenic or oncogenic stimuli. This is equally true in response to inflammatory signals such as TNF-α, 1L-1β, and lipopolysaccharide. [2] Due to untoward cardiovascular and other effects of COX-2 selective inhibitors, subsequent re-evaluation and scrutiny seems to shift from the initial absolute inducible nature and has widely been established that COX-2 is constitutively present in specific discrete sites and maintains vital noninflammatory-related physiological roles.[4] Although

structurally heterogeneous, all members of NSAIDs possess a single common mechanism of action to block prostaglandin synthesis, yet various NSAIDs have other actions that may contribute to differences between the drugs such as neutrophil aggregation and adhesion, cytokins production and cartilage metabolism; superoxide radical production are superoxide scavenging. In addition, data suggesting the central analgesic effects of NSAIDs exist based on various mechanisms of action including glycin-dependent modulation of pain, effect on β-endorphin, modulation of inhibitory fast synaptic currents in lamina I and II of the dorsal horn.[8, 9]

2.3 Clinical application

Based on their main pharmacological and therapeutic effects, the clinical use of NSAIDs can be summarized in three main perspectives. [10, 11]

- i. For Analgesia in both acute and chronic pain situations such as toothache, headache, dysmenorrhea, and post-operative pain. Induction of peripheral analgesia by NSAIDs via inhibition of COX activity and prostaglandin biosynthesis is well documented cum central analgesic effects based on established mechanisms. [9]
- ii. For fever antipyretic effect is based on controlled thermoregulation induced by binding of prostaglandin G_2 (PGE2) to EP_3 receptors on neurons within the preoptic anterior hypothalamic area. In the hypothalamic region, via exposure to exogenous pyrogens (pathogen-associated molecular patterns), cells of the immune system generate and release interleukins, $TNF\alpha$ and other endogenous pyrogens to stimulate the synthesis of PGE_2 . Though the antipyretic activity of NSAIDs is linked to their ability to inhibit prostaglandin synthesis, yet in abnormal elevation of body temperature like malignant hyperthermia and heat stroke, conditions known to be independent of the COX- mediated pathway, NSAIDs have no effect. $^{[12]}$
- **iii. For inflammation** anti-inflammatory especially and mainly anti-exudative action in both acute and chronic inflammatory conditions such as gout and rheumatoid arthritis are part of what NSAIDs are known for.

In neurodegenerative processes, neuro-inflammation is known to play an important role and there are evidences showing that NSAIDs possess the potential to diminish the risk for Alzheimer's disease (AD)^[13] as well as Parkinson's disease ^[14] Amyloid β –protein (A β) accumulates within the brain in Alzheimer's disease, and NSAIDs have been reported to inhibit the activity of gamma-secretase, the enzymes responsible for cutting the transmembrane region of the amyloid β -protein precursor to form the A β . NSAIDs contribute to the removal of plague and debris by modulating the phagocytic activity of microglia. Celecoxib and

aspirin may be helpful in managing the symptoms of schizophrenia and other neuropsychiatric disorders. [17, 18] Since the 1970s, it has been known that acetylsalicylic acid can reduce the risk of pre-eclampsia and Celecoxib in a meta-analysis of randomized, placebocontrolled, double blind trials, appears to be a safe and efficacious treatment in alleviating particularly first-episode schizophrenia and other psychotic symptoms. [20]

Acetylsalicylic acid is also indicated for women undergoing assisted reproductive technology for the purpose of optimizing the chance of live birth. [21] Though the role of aspirin in primary cardiovascular disease appear controversial [22], low-dose regimes are being embraced for prevention of secondary cardiovascular disease on the account of their ability to suppress platelet aggregation [23], but because the enzyme COX-2 is not expressed in these cells selective COX-2 inhibitors do not affect platelet aggregation.

2.4: Anti-Tumour Activity

For at least some categories of cancer such as colorectal, breast and breast cancers, preclinical and clinical investigations have shown a benefit of NSAIDs in reducing their risk. [24, 25] Epidemiological studies have supported that continued use of NSAIDs lead to a significant decline in adenomatous polyps, disease incidence recurrence as well as death from colorectal cancer. [26, 27, 28]

Reports have suggested that an antitumor effect of NSAIDs is pleiotropic and includes groups of mechanisms of action namely: COX-Indpendent pathways; influencing cancer stem cells; and related to suppression of COX activity. [29, 30] German pathologist Rudolf Virchow in 1863 was the first to describe the infiltration of tumours with white blood cells.^[31] Since then till now, it is widely accepted that inflammation (chronic) is essential for tumorigenesis and of course, one of the cancer hallmarks. [32] In 1986 when the relationship existing between prostaglandins and cancer was initially reported^[33], it was established that PGE and PGF production influenced the metastatic and invasive potential of mammalian esophageal carcinoma cells in athymic nude mice. In most tumour tissues, inflammatory mediators and cells can be detected modulating the function of both stromal cells and tumour thereby contributing to the adaptation of a tumourpromoting microenvironment. [34] It has been suggested that increased levels of prostaglandins especially PGE₂ and PGD₂, are involved in signaling pathways important for wide range of cancer induction and progression. [35, 36,

Accordingly, PGE_2 has been published as potent therapeutic target for the treatment of colon cancer^[39] and thromboxane A_2 (TXA₂) has been found to participate in pathogenesis of colorectal cancer.^[40, 41] In addition, TXA₂ is involved in modulation of multiple myeloma and lung cancer cell proliferation^[42, 43]). It is

common knowledge that the best studied molecular target of NSAIDs is cyclooxygenase in which the COX-2 enzyme (inducible cyclooxygenase is of particular interest. This is not surprising because the expression of COX-2 is associated with inflammatory conditions and overexpressed in many solid tumours such as cancers of the prostate, pancreatic, colorectal, mammary gland and lung cancer. [44, 45] COX-2 promotes tumour growth as well as suppresses antitumour immunity. [46] Cancerassociated-fibroblasts, macrophage type 2 cells and cancer cells release COX-2 into the tumour microenvironment. [47]

Epidermal growth factor receptor (EGFR), mitogenactivated protein kinase (MAPK) members and nuclear factor-Kβ are principal upstream modulators for COX-2 cancer cells. The role of constitutive cyclooxygenase (COX-1) in cancer has universally received less attention and COX -2 has been at the forefront as a promising target for cancer therapy. [48] Nonetheless, COX-1 expression has occasionally been detected in several circumstances of cancer, including head and neck cancer, haematological tumours, colorectal, esophageal, breast and cervical cancers. [49] It was in 1995 that COX-1 was first identified as marker of ovarian cancer^[50] and subsequently its overexpression has been proven in various mouse, avian and human models of ovarian cancer. [51, 52, 53] More evidences indicate that both COX-1 and COX-2 isoforms play an important role in cancerogenesis with data that at least in some situations such as serious ovarian carcinoma, COX-1 enzyme may play a pivotal role. [54, 55, 56, 49]

The current situation is that the biological activity of COX-2 inhibitors has been stupendously investigated, while the number of the available COX-1 inhibitors is extremely limited and very small, pitiable part of them have been investigated for antitumour activity[57, 58, 49] (Vitale et al., 2016; Tortorella et al., 2016; pannunzio and coluccia, 2018). In a report, COX-2 has shown to play a significant role in cancer stem cell biology. [59] The antitumour action of aspirin and other NSAIDs as indicated in experimental data, may be due to the selective induction of apoptosis within human intestinal stem cells with changed Writ signaling pathway. [60] COX-independent mechanisms of anti-tumour activity of aspirin-like drugs are not unrelated with their tendency to influence Writ signaling pathway, down-regulation of proto-oncogenes such as c-myc, DNA mismatch repair systems, transcription factors and induction of oxidative stress.[61, 12, 2]

Many researchers in their endeavour to elucidate the antitumour effect of NSAIDs emphasize that the mechanism should also involve DNA to some degree and the processes of transcription and translation. ^[62] But because most NSAIDs at physiological pH are anions, this poses an obstacle in accessing and interacting with the polyanionic strands of DNA. This explains why interaction between NSAIDs and DNA is reported

sparingly by a limited number of investigations. [63] The electrostatic repulsion between the deprotonated negatively charged form of DNA could be surmounted by combining NSAIDs with metal ions. Many studies reported that concomitant administration of copper and NSAIDs exerts a synergetic effect. Meloxicam and Piroxicam can form complexes at physiological pH with CU (II) hence a proposed hypothetical model of interaction between these compounds and DNA, which involves intercalation most likely [64] Following various complexes of NSAIDs compounded and characterized, these compounds have been found to express a more pronounced anti-inflammatory and tumour effects, possess reduced gastrointestinal toxicity compared to individual agents. [65, 66]

2.5 Interactions

The pharmacokinetic or/and pharmacodynamics interaction of drugs concomitantly administered with NSAIDs have been reported, some of which may be clinically significant. NSAIDs can inhibit drug metabolism or interfere with renal excretion of such drugs; displace other agents from their plasma protein binding sites. [67] NSAIDs used concomitantly with other antiplatelets and anticoagulant medications, results to excess risk of upper gastrointestinal bleeding. [68] NSAIDs can during the use of diuretics, ACEIs, and beta adrenergic blockers, adversely influence blood pressure control. Conversely, some treatments are known to influence the rate and extent of aspirin absorption. [69]

2.6 Limitations

Until recently 1999, gastrointestinal toxicity represented the major safety concern of NSAIDs application with symptoms ranging from milder dyspepsia to severe bleeding and ulceration.^[70, 71] Apart from GIT toxicity, the major adverse limitations of NSAIDs affect kidneys, central nervous system, platelet function and cardiovascular system especially with COX-2 inhibitors.

The prevalence in the rate and severity of nephrotoxicity with NSAIDs increase in patients with certain risk factors such as heart failure, diabetes, renal dysfunction and in elderly people. It is important to recognize that in the mammalian kidney, prostaglandins regulate vascular tone as well as salt and water homeostasis playing an important role in hemodynamics^[72], hence NSAIDs increase sodium reabsorption that leads to peripheral edema via the inhibition of prostaglandin (PG₂). The resultant adverse effect ranges from electrolyte retention, reduced glomerular filtration to nephritic syndrome and chronic renal failure; also observed is hyperkalemia. [73, 74]

Selective COX-2 inhibitors have been associated with adverse effects in the cardiovascular system such as elevated blood pressure and risk of atherothrombotic events, mechanism of which is not fully known but the interaction between prostanoids and T cells has been implicated. However, it is not surprising, since COX-2 enzymes generates biologically active molecules that

supports regulation of blood pressure, endothelial thromboresistance, renal haemodynamics as well as in pathophysiology of pain and inflammation such as PGE₂ and PGI₂ [76] In the United States of America and Europe, rofecoxib and valdecoxib (COX-2 inhibitors) have been from the market following withdrawn cardiovascular adverse effects, confirmed in long term placebo-controlled trials.^[77] Decrease in prostaglandin production by NSAIDs can induce alternative leukotriene pathway, leading to bronchoconstriction. NSAIDs- or Aspirin-exacerbated respiratory disease (AERD) comprising asthma, Nasal polyps (referred to as Samter's triad) and aspirin sensitivity are instances of COX inhibition on the respiratory system. Some prostaglandins exhibit important regulatory tendencies on respiratory epithelial cells hence AERD affects approximately 7% of asthmatic patience and 0.3%-0.9% of general population in the USA. $^{[78, 79, 80]}$ Prostaglandins E₂ and D₂ are especially involved in bone remodeling. They do this by mediating the control of osteoblast and osteoclast functions having a mitogenic effect on and stimulations of their osteoclasts biological activity.[81] Prostaglandins behave as paracrine regulators of the bone remodeling cycle^[82]; and NSAIDs should be avoided in high risk patients having been suggested as a risk factor for impairment of bone healing. $^{[83]}$

3. Utility of chemically modified NSAIDs and natural products

Due to the challenges posed by limitations and adverse effects of conventional nonsteroidal anti-inflammatory drugs, a good number of research groups have in recent times, directed attention towards the exploration and discovery of nonconventional chemically modified drugs as well as alternative system of medicine, especially for antinociceptive and antoexudative effects.

3.1 Nonconventional NSAIDs

The design and development of nonconventional NSAIDs usually by various chemical modifications is targeted at strong therapeutic effect with a better safety profile; majority of which are in preclinical and initial stage of clinical trials. [84, 2]

- a. Phosphatidylcholine (PC) aspirin: This is also known as NSAIDs-with-phosphatidylcholine, which rely on the preponderance of PC amongst the gastric phospholipids that form extracellular lining on the mucous gel layer thereby protecting the underlying epithelium from gastric acids.
- b. Nitro-aspirin: This is nitric oxide-releasing-NSAIDs, relying on the fact that nitric oxide (NO) plays a protective role by maintaining gastric mucosal integrity in the gastrointestinal tract through increasing the mucosa secretion as well as mucosal blood flow plus inhibiting neutrophil aggregation. NO exerts beneficial effect on the CVS by inhibiting platelet aggregation and adhesion. At the preclinical level NO has been shown to repair

- NSAID-induced damage while epidemiologic studies have revealed that the use of NO-donating agents with aspirin or NSAIDs leads to increase risk of gastrointestinal bleeding. [86]
- c. Pegylated phosphoribuprofen: This is a pegylatedphospho-NSAIDs noting that polyethylene glycol protects ibuprofen from the hydrolytic mechanism of esterases.
- **d. Des-methyl (DM)-Sulindac**: This represents the esterified/amidated NSAIDs.
- e. Dual COX/LOX inhibitors: Several of these mixed preparations were conceived to overcome some of the limitations presented by NSAIDs during blockade of the arachidonic acid metabolism which can result in generation of proinflammatory mediators -leukotrienes and lipoxins via the lipoxygenase pathway. It should be recognized that leukotrienes and lipoxygenases are potential targets for the management of many pathological conditions including asthma, Alzheimer's disease, cardiovascular disease and various inflammatory diseases. So dual COX/LOX inhibitors are of great importance. [87, 88]
- **f. CU** (**II**)/**NSAIDs mixed preparations:** Examples include meloxicam-and piroxicam/ CU (**II**) complex preparations. [64]

Complexes with NSAIDs increase anti-inflammatory effect and simultaneously reduce gastrointestinal toxicity in comparison to NSAIDs used alone. [64]

3.2 Natural products with antinociceptive and antiexudative effects

Flavonoids, terpenoids, steroids, phenols, glycosides, alkaloids and tannins are among the products of secondary metabolism by plants and they are referred to as natural products. Numerous scientific evidences that buttress successful management of pain and inflammation using natural products abound. In the plant kingdom, there are representative analgesic and anti-inflammatory herbs in almost each family. Lupinus arboreus is of great medicinal value with proven antinociceptive and anti-inflammatory activities. Some other medicinal plants that exert analgesic or/and anti-inflammatory activities are shown in Table 4.

Table 4: Plants with documented analgesic/anti-inflammatory activities. [92]

h documented analgesic/anti-inf		
Plant	Family	Used/studied part Leaf
Agerantum conyzoides Sambucus ebulus	Asteraceae	Rhizome
Caralluna tuberculata	Caprifoliaceae	Flower, leaf
Diodia scandens	Asclepiadacea Rubiaceae	Aerial part
Ficus platyphlla		Bark
Turner ulmifolia	Moraceae Turnderaceae	Leaf
	Crassulaceae	Leaf
Bryophyllum pinnatum Icacina trichantha	Icacinaceae	Tuber
Siderites spp	Lamiaceae	Flowers
Euphorbia royleana	Euphorbiaceae	Latex
Curcuma longa	Zingiberaceae	Rhizome
Pothomorphe peltata	Piperaceae	Leaf
	Rubiaceae	Leaf
Mitracarpus scaber Taxodium distichum	Taxodiaceae	Fruit
		Stem
Moringa oleifera Anthurium cerrocampuse	Moringaceae	Stem
	Araceae	
Centaurea cyamus	Asteraceae	Flowers
Heterotheca inuloides	Asteraceae	Flowers
Dalbergia sissoo	Fabaceae	Leaf
Terminalia ivorensis	Compositae	Leaf
Scutelleriae biocalensis	Scutellaceae	Leaf
Calotropis spp	Asclepiadacea	Flowers
Dicliptera chinensis	Acanthaceae	Aerial part
Tithonia diversifolia	Compositae	Aerial part
Calligonum comosum	Polygonaceae	Aerial part
Agerantum conyzoides	Asteraceae	Leaf
Agerantum conyzoides	Asteraceae	Leaf
Alchornea cordifolia	Euphorbiacea	Leaf
Butea frondosa	Papilionaceae	Leaf
Chasmanthera dependens	Menispermaceae	Leaf
Emilia sonchifolia	Asteraceae	Leaf
Tanacetum parthenium	Asteraceae	Leaf
Syzygium cumini	Myrtaceae	Bark
Premna herbacea	Verbanaceae	Root
Anthurium cerrocampanese	Araceae	Leaf
Culcasia scandens	Araceae	Leaf
Aspilia africana	Compositae	Leaf
Bryophyllum pinnatum	Crassulaceae	Leaf
Entada abyssinica	Mimosaceae	Leaf
Aspilia africana	Compositae	Leaf
Holmskiodia sanguinea	Verbanaceae	Leaf
Opuntia fiscusindica	Cactaceae	Stem
Croton lecheri	Rubiaceae	Leaf
Orbignya phalerata	Aracaceae	Fruit
Rheo spathaceae	Commelinacea	Leaf
Ambrosia artemisiaefolia	Compositae	Leaf
Cissus trifoliate	Vitaceae	Root
Teucrium buxifolium	Lamiaceae	Aerial part
Ficus elastic	Moraceae	Root
Psidium guianense	Myrtaceae	Leaf
Anthurium Cerrocampanese	Araceae	Leaf
Cedrus deodora	Pinaceae	Wood
Anthurium Cerrocampanese	Araceae	Leaf
Aegle marmelos	Rutaceae	Root, bark
Moringa oleifera	Liliaceae	Fresh juice, gel

Adapted with slight modification from: Ohadoma SC. Pharmacological assessment of Lupinus arboreus Sims (Fabaceae) methanol extract and fractions for antinociceptive and anti-inflammatory effects. A Ph.D Thesis, Department of Pharmacology and Toxicology, University of Nigeria, Nsukka, 2014. Available online at UNN Visual Library 2014 p.31.

3.2.1 Flavonoids with antinociceptive and antiinflammatory activities

Flavonoids are one of the largest classes of naturally-occurring polyphenolic compounds. This group of plant pigments is largely responsible for the colors of many fruits and flowers, and over 4,000 flavonoid compounds have been characterized and classified according to chemical structure. The word "flavonoid" comes from the Latin flavus which means yellow; however some flavonoids are red, blue, purple or white. The word "flavonoids" compounds in which the two C6 groups are substituted benzene rings, and the C3 is an aliphatic chain which contains a pyran ring. Flavonoids occur as O- or C-glycone.

Numerous medicinal plants contain therapeutic amounts of flavonoids, which are used to treat disorders of the peripheral circulation^[95], to lower blood pressure^[97], to improve aquaresis^[98], as anti-inflammatory, antispasmodic^[98] and anti-allergic agents.^[95] The many pharmacological effects of flavonoids are linked to their ability to act as strong antioxidants and free radical scavengers, to chelate metals, and to interact with enzymes, adenosine receptors, and biomembrane.^[99, 95] Some flavonoids also possess antimicrobial activity.^[100]

The flavonoid glycoside, chrysoeriol 7-0-b-D glucopyranosyl (2 1) – D-apiofuranoside isolated from Dalbergia volubilis exhibited anti-inflammatory activity. A flavonoid from Hedychium spicalum showed a significant activity with less ulcerogenic index than phenylbutazone. Two new flavanone glycosides, diinsininol and diinsinin from rhizomes of Sacropthyte piriei (Balanophoraceae), showed IC values of prostaglandin synthesis inhibition 9-20 μ m and 13-14 μ m respectively and in the inhibition of platelet-activating-factor-induced exocytosis, IC values of 49 and 39 μ m, respectively. $^{[103]}$

Three flavonoids identified as (i) 4' hydroxy3', 5' liprenylisoflavanone (ii) 4' hydroxy 6, 3', 5' triprenylisoflavano-ne (iii) 3, 9-dihydroxy-2, 0 diprenylprterocarpene (erycrystagallin)^[104] were isolated from a methanol extract of bark of the Samoan medicinal plant *Erythrina variegate* (Leguminosae). The compounds were shown to possess phospholipase A₂ (PLA₂) inhibitory activity. The compounds-Dicadalenol, Caryolane-1, 9 b-diol and quercetin isolated from aerial parts were identified as the active principle of Boswellia resin, inhibiting the key enzyme in leukotrine biosynthesis, 5-lipooxygenase (5-LOX). Of the boswellic acids characterized, 3-0-acetyl-11-keto-b- boswellic acid

(AKBA) proved to be the most potent inhibitor of 5-LOX. [105]

A new diterpenoid, tolypodial has been established from the terrestrial Cyanobacteria tolypothrix nodosa (HT-58-2). [106] Tolypodiol and its monoacetate derivative show potent anti-inflammatory activity in mouse ear oedema assay. [106] The antipyretic and anti-inflammatory activity of a new sesquiterpene, spartidienedione isolated from Psila spartioides (Asteraceae) were evaluated in rabbits and guinea pigs. [107] At a dose of 25 mg/kg, this substance showed anti-inflammatory activity and antipyretic activity. Two new sesquiterpene cyclopentenones, dysidenones A, B and a new sesquiterpene aminoquinone, dysidine, all containing the same rearranged drimane skeletone have been isolated along with bolinaquinone from Sponge dysidea species. [108] Bolinaquinone, dysidine and a 1:1 mixture of dysidenone A and B significantly inhibited human synovial phospholiphase A₂ (PLA_2) concentration. [108]

The oleoresin fraction of Commiphora mukul exhibited significant anti-arthritic and anti-inflammatory activities^[109, 110] (Satyavati et al., 1969; Shukla, 1986). Methanol leaf extract of Chikadoma exhibited significant anti-arthritis activity. [111] Chikadoma is the common Nigeria nomenclature for Lupinus arboreus, derived from a certain lead researcher Dr. Chika Ohadoma who conducted extensively pioneering novelty work on the plant. [112] A steroidal compound isolated from C. mukul displayed a significant activity which is dose dependent and more potent than the resin fraction present in C. mukul^[110] (Shukla, 1986). β- sitosterol isolated from Cyperus rotendus possessed potent anti-inflammatory activity against carrageenan and cotton pellet-induced oedema in rats and was comparable to hydrocortisone and oxyphenbutazon^[113] (Singh 1970). Quercetin, quercetin 3-0-rhamnoside (quercitrin) and quercetin 3-0rutinoside (rutin) from 80% MeOH extract of leaves of Morinda morindoides (Rubiaceae) also showed similar inhibition of classical pathway complement system^[114] (Climanaga, 1995).

The dichloromethane extract of the aerial parts of Tanacetum microphyllum (Compositae) yielded two antiinflammatory flavanoids: 5, 7, 3' trihydroxy-3, 6, 4'trimethoxy flavone (Centaureidin) and 5, 3'-dihydroxy-4'-methoxy-7- carbomethoxyflavonl^[115] (Abad et al., 1993). Three flavonoids, 7-0 methylaromadendrin, rhamnocitrin, and 3-0 acetylpadmatin, isolated from Inula viscose (Asteraceae) dichloromethane extract were shown to have 12-0 tetradecanoylphorbol-13-acetate induced oedema inhibitory activity in mice. [116] Some other flavonoid subtypes, eg. Coumarins and xanthones, have been shown to inhibit the COX-1/COX- 2 catalyzed prostaglandin (PG) biosynthesis in vitro. [117, 118, 119] The coumarin Calophylolide from the nuts of Calophyllum inophyllum (Clusiaceae) effectively reduced the increased permeability induced by the chemical

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mediators involved in inflammation, like histamine, serotonin and bradykinin. The ED₅₀ was found to be 144.1, 250 and 135.5 mg/kg p.o., respectively against mediators.[120] Four coumarins, methoxycoumarin (herniarin), 6, 7-dihydroxycoumarin 6-methoxy-7-glucosidyl (aescuetin), coumarin (Scopolin), 6-hydroxy-7-methoxycoumarin and (Scopoletin), were isolated from the ethanol extract of the flower tops of *Santolina oblongifolia* Boiss. (Compositae). The isolated compounds showed marked activity as inhibitors of eicosanoidrelease from ionophore stimulated mouse peritoneal macrophages. [121] Magniferin, a xanthon C-glucoside from Canscora decussatta, mangostin and related compounds from mangostana^[122] and xanthones Calophyllum scohyllum and Mesua ferrea are shown to have anti-inflammatory activity.[123]

3.2.2 Terpenoids and steroids with antinociceptive and anti-inflammatory effects

Several triterpenoids and steroids have been isolated from extracts of plants with anti-inflammatory effect. In most instances, the anti-inflammatory activity of such extracts has been attributed to the isolated compounds.

The isolation of seven novels naturally occurring triterpene alchohols from non-saponifiable lipids of the seeds of Camellia japonica and Camellia sasanqua has been reported. The anti-inflammatory activity of these compounds, tirucall-5, 7, 24-triene-3b-ol, lemmaphylla-7, 21-diene-3b-ol, isoeuphol, isotirucallol, (24R)-24, 25-epoxybutrospermol and its 24S-epimer, and isoaglaoil, was studied in mouse ear edema model. Lupeol was isolated from the hexane stem bark extract of Crataeva religiosa Forst. (Caparidaceae) as the antiinflammatory component. In an extensive antiinflammatory studies, lupeol exhibited anti-inflammatory effect in a variety of acute and chronic anti-inflammatory test models in rat and mice. [125] Lupeol ester, lupeol, lineolate, obtained by esterification of lupeol with linleoyl chloride was shown to exhibit antiarthritic activity greater than that of lupeol.

The triterpenoids of the oleanene and ursine series were found to be active against carrageenan induced oedema, formaldehyde-induced arthritis in rats. It has been suggested that the anti-inflammatory activity of the triterpenoids of the oedema series vary with the polarity of compounds which is enhanced by the number of hydroxyl groups in the molecule. [126] Oleanolic acid 3-βglucoside isolated from the seeds of Randia dumertorum (25-500mg kg, p.o) showed a significant antiinflammatory activity in the exudative and proliferative phases of inflammation in rats. [127] Salai guggal, the oleogum of *Boswellia serrata*, has been shown to possess anti-inflammatory and anti-arthritic activities. It was shown to be effective in controlled clinical trials in arthritic patients. Its activity may be due to the boswellic acids present in the olegum.[128]

Two new triterpene saponins having phospholipase inhibitory activity were isolated from methanol extract of the leaves of *Myrsine australis*. [104] These are 3- 0{-β-Dxylopyranosy-(1®2)-0-β-D-glucopyranosyl-(1®4) {0-β-D-glucopyrnosyl- (1 \mathbb{R}^2) {- α -L-arabinosyl} 16 α hydroxy-13 β , 28-epoxyoleanane and 3 β -0-{- β -Drhamnopyranosyl}-16-α-hydroxy-13,28epoxyoleanane. Both compounds showed IC50 values of high significant, versus phorbol 42-myristate 13-acetate stimulated phospholipase in human promylocytic lukemic (HI-60)^[104] cells Two oleane type triterpene saponin, zanhasaponins A and B and the cyclitol pinitol isolated from the methanol extract of root bark of Zanha africana (Sapindaceae) were active as inhibitors of phospholipase A₂ [116] Pentacyclic triterpenes from the 11-keto-boswellic acid series has stabilizing effect, inhibition of leucocytes migration and antipyretic activity. [129] The steroid, - spinasterol, obtained from the stem-bark of Symplocos spicata showed a significant activity against acute inflammation induced by carrageenan in rats and was more potent than phenylbutazone but less potent than betamethasone. [130] Six steroidal saponins were isolated from Anemarrhena asphodeloides Bunge (Liliaceae), a traditional Chinese medicine, and named anemarrhenasaponin I (An-I), anemarrhenasaponin Ia (An-Ia), timosaponin B-I (TB-I), timosaponin B-II (TB-II), timosaponin B-III (TB-III), and timosaponin A-III (TA-III). All these compounds provoked remarkable inhibiting effect on platelet aggregation. [131] Steroids consist of an essentially lipophilic (or hydrophobic, cyclopentanoperhydrophenanthrene nucleus modified on the periphery of the nucleus or on the side chain by the addition of hydrophilic (or lipophobic, polar) groups.

CONCLUSION

For lead optimization through derivatization/semi synthesis, the challenge of any drug discovery effect is to identify and develop compounds with properties that are predictive of good efficacy and safety in humans. In this regard, organic synthesis plays a pivotal role. Once lead services with some desirable profiles are identified, the compounds can progress to lead optimization, entailing structural modifications with the goal of achieving optimal efficacy and pharmacokinetic/pharmacodynamics properties.

The frequent occurrence of natural products as complexes of structurally related analogues can be exploited by the natural-product investigator as a guide for initial Structure-activity Relationship (SAR) experiments. In this regard, even simple synthetic modifications, as those obtained through "shotgun" transformations (key functional groups required for biological activity can be identified by allowing the parent compound to react with derivatizing reagents, such as alkyl halides, anhydrides, acyl halides) can be instrumental in leading to an optimized semisynthetic analogue. The knowledge gained through understanding the natural SAR and the shotgun approach can provide

an early foundation on which an overall synthetic strategy could be developed.

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Authors Contributions

OSC conceived and designed the study.

OSC, APA, OCE, OEP and MHU gathered materials and data.

OSC wrote the paper.

OSC, APA, OCE, OEP and MHU (All the authors) read and approved the paper.

REFERENCES

- 1. Ascherio A, Schwarzschild MA. The epidemiology of Parkinson's disease: risk factors and prevention. Lancet Neurol. 2016; 15: 1257–1272.
- 2. Rigas B, Tsioulias J. The evolving role of nonsteroidal anti-inflammatory drugs in colon cancer prevention: a cause for optimism. Journal of Pharmacological and Experimental Therapeutics, 2015; 353(1): 2-8.
- 3. Ohadoma SC. Clinical and natural product pharmacology made easy. 2nd ed., Nigeria; Reviewed Publishers, 2017; p. 282-298.
- Kirkby NS, Chan MV, Zaiss AK, Garcia-Vaz E, Jiao J, Berglund LM, Verdu EF, Ahmetaj-Shala B, Wallace JL, Herschman HR, Gomez MF, Mitchell JA. Systematic study of constitutive cyclooxygenase-2 expression: Role of NF-κB and NFAT transcriptional pathways. Proc Natl Acad Sci USA, 2016; 113(2): 434-439.
- 5. Ornelas A, Zacharias-Millward N, Menter DG, Davis JS, Lichtenberger L, Hawke D, Hawk E, Vilar E, Bhattacharya P, Millward S. Beyond. COX-1: the effects of aspirin on platelet biology and potential mechanisms of chemoprevention. Cancer Metastasis Rev, 2017; 36(2): 289-303.
- Patrono C. Cardiovascular effects of cyclooxygenase-2 inhibitors: a mechanistic and clinical perspective. Br J Clin Pharmacol, 2016; 82(4): 957-64.
- Dovizio M, Bruno A, Tacconelli S, Patrignani P. Mode of action of aspirin as a chemo-preventive agent. Recent Results Cancer Res, 2013; 191: 39-65.
- Luan YH, Wang D, Yu Q, Chai XQ. Action of β-endorphin and non-steroidal anti-inflammatory drugs, and the possible effects of non-steroidal anti-inflammatory drugs on β-endorphin. J Clin Anesth, 2017; 37: 123-128.
- 9. Vuilleumier PH, Schliessbach J, Curatolo M. Current evidence for central analgesic effects of NSAIDs: an overview of the literature. Minerva Anestesiol, 2018; 84(7): 865-870.
- 10. Ho KY, Gwee KA, Cheng YK, YoonKH, Hee HT, Omar AR. Nonsteroidal anti-inflammatory drugs in

- chronic pain: implications of new data for clinical practice. J Pain Res, 2018; 11: 1937-1948.
- Ghlichloo I, Gerriets V. Nonsteroidal Antiinflammatory Drugs (NSAIDs) StatPearls Publishing, 2019.
- 12. Osafo N, Agyare C, Obiri DD, Antwi AO. Mechanism of Action of Nonsteroidal Anti-Inflammatory Drugs, Nonsteroidal Anti-Inflammatory Drugs, Ali Gamal Ahmed Al-kaf, IntechOpen, 2017; DOI: 10.5772/68090.
- Benito-León J, Contador I, Vega S, Villarejo-Galende A, Bermejo-Pareja F. Non-steroidal antiinflammatory drugs use in older adults decreases risk of Alzheimer's disease mortality, PLoS One, 2019: 14(9): e0222505.
- 14. Etminan M, Suissa S. NSAID use and the risk of Parkinson's disease. Curr Drug Saf1 2006; 223–225.
- 15. Cudaback E, Jorstad NL, Yang Y, Montine TJ, Keene CD. Therapeutic implications of the prostaglandin pathway in Alzheimer's disease. Biochem Pharmacol88, 2014; 565–572
- Koenigsknecht-Talboo J, Landreth GE. Microglial phagocytosis induced by fibrillar beta-amyloid and IgGs are differentially regulated by proinflammatory cytokines J Neurosci, 2005; 25: 8240– 8249.
- 17. Sethi R, Gómez-Coronado N, Walker AJ, Robertson OD, Agustini B, Berk M, Dodd S. Neurobiology and Therapeutic Potential of Cyclooxygenase-2 (COX-2) Inhibitors for Inflammation in Neuropsychiatric Disorders, Front Psychiatry, 2019; 10: 605.
- Müller N. COX-2 Inhibitors Aspirin, and Other Potential Anti-Inflammatory Treatments for Psychiatric Disorders Front Psychiatry, 2019; 10: 375.
- Mirabito Colafella KM, Neuman RI, Visser W, Danser AHJ, Versmissen J. Aspirin for the prevention and treatment of pre-eclampsia: A matter of COX-1 and/or COX-2 inhibition? Basic Clin Pharmacol Toxicol doi: 2019; 10.1111/bcpt.13308.
- 20. Zheng W, Cai DB, Yang XH, Ungvari GS, Ng CH, Müller N, Ning YP, Xiang YT. Adjunctive celecoxib for schizophrenia: A metaanalysis of randomized, double-blind, placebo-controlled trials. J Psychiatr Res, 2017; 92: 139-146.
- 21. Siristatidis CS, Basios G, Pergialiotis V, Vogiatzi P. Aspirin for in vitro fertilisation. Cochrane Database Syst Rev, 2016; 3: 11.
- 22. Raber I, McCarthy CP, Vaduganathan M, Bhatt DL, Wood DA, Cleland JGF, Blumenthal RS, McEvoy JW. The rise and fall of aspirin in the primary prevention of cardiovascular disease Lancet, 2019; 3 93(10186): 2155-2167.
- 23. Ansa BE, Hoffman Z, Lewis N, Savoy C, Hickson A, Stone R, Johnson T. Aspirin Use among Adults with Cardiovascular Disease in the United States: Implications for an Intervention Approach J Clin Med, 2019; 8(2): 264.
- Chan AT, Arber N, Burn J, Chia WK, Elwood P, Hull MA, Logan RF, Rothwell PM, Schrör K, Baron

- JA. Aspirin in the chemoprevention of colorectal neoplasia: an overview Cancer Prev Res (Phila), 2012; 5: 164–178.
- 25. Bosetti C, Rosato V, Gallus S, Cuzick J, Vecchia La C Aspirin and cancer risk a quantitative review to 2011. Ann Oncol, 2012; 23: 1403–1415.
- 26. Garcia-Albeniz X, Chan AT.Aspirin for the prevention of colorectal cancer Best Pract Res ClinGastroenterol, 2011; 25: 461–472.
- 27. Rayburn ER, Ezell SJ, Zhang R. Anti-Inflammatory Agents for Cancer Therapy Mol Cell Pharmacol, 2009; 1: 29–43.
- 28. Rostom A, Dubé C, Lewin G, Tsertsvadze A, Barrowman N, Code C, Sampson M, Moher D, U.S. Preventive Services Task Force. Nonsteroidal antiinflammatory drugs and cyclooxygenase-2 inhibitors for primary prevention of colorectal cancer: a systematic review prepared for the U.S. Preventive Services Task Force Ann Intern Med, 2007; 146: 376–389.
- 29. Stolfi C, Simone VDe, Pallone F, Monteleone G. Mechanisms of action of non-steroidal anti-inflammatory drugs (NSAIDs) and mesalazine in the chemoprevention of colorectal cancer Int J MolSci, 2013; 14: 17972–17985.
- 30. Schrör K. Pharmacology and cellular/molecular mechanisms of action of aspirin and non-aspirin NSAIDs in colorectal cancer Best Pract Res ClinGastroenterol, 2011; 25: 473–484.
- 31. Benito-León J, Contador I, Vega S, Villarejo-Galende A, Bermejo-Pareja F. Non-steroidal anti-inflammatory drugs use in older adults decreases risk of Alzheimer's disease mortality PLoS One, 2019; 14(9): e0222505.
- 32. Hanahan D, Weinberg R.A. The hall marks of cancer Cell, 2000; 100: 57-70.
- 33. Botha JH, Bobinson KM, Ramchurren N, Reddi K, Norman RJ. Human esophageal carcinoma cell lines: Prostaglandin production, biological properties, and behavior in nude mice J Natl Cancer Inst, 1986; 76: 1053–1056.
- 34. Coussens LM, Zitvogel L, Palucka AK. Neutralizing tumor promoting chronic inflammation: A magic bullet? Science, 2013; 339: 286–291.
- 35. Kalinski P. Regulation of immune responses by prostaglandin E2. J Immunol, 2012; 188: 21–28.
- 36. Oshima H, Oshima M. The inflammatory network in the gastrointestinal tumor microenvironment: Lessons from mouse models. J Gastroenterol, 2012; 47: 97–106.
- 37. Sha W, Brüne B, Weigert A. The multi-faceted roles of prostaglandin E2 in cancer-infiltrating mononuclear phagocyte biology. Immunobiology, 2012; 217: 1225–1232.
- 38. Nakanishi M, Rosenberg DW. Multifaceted roles of PGE2 in inflammation and cancer Semin Immunopathol, 2013; 35: 123–137.
- 39. Karpisheh V, Nikkhoo A, Hojjat-Farsangi M, Namdar A, Azizi G, Ghalamfarsa G, Sabz G, Yousefi M, Yousefi B, Jadidi-Niaragh F.

- Prostaglandin E2 as a potent therapeutic target for treatment of colon cancer. Prostaglandins Other Lipid Mediat, 2019; 144: 106338.
- 40. Dovizio M, Tacconelli S, Ricciotti E, Bruno A, Maier TJ, Anzellotti P, Di Francesco L, Sala P, Signoroni S, Bertario L, Dixon DA, Lawson JA, Steinhilber D, FitzGerald GA, Patrignani P. Effects of celecoxib on prostanoid biosyntesis and circulating angiogenesis proteins in familial adenomatous polyposis. J Pharmacol Exp Ther, 2012; 341: 242–250.
- 41. Li H, Liu K, Boardman LA, Zhao Y, Wang L, Sheng Y, Oi N, Limburg PJ, Bode AM, Dong. Circulating prostaglandin biosynthesis in colorectal cancer and potential clinical significance. EBioMedicine, 2015; 2: 165–171.
- 42. Liu Q, Tao B, Liu G, Chen G, Zhu Q, Yu Y, Yu Y, Xiong H. Thromboxane A2 receptor inhibition suppresses multiple myeloma cell proliferation by inducing p38/c-Jun N-terminal Kinase (JNK) Mitogen-activated Protein Kinase (MAPK)-mediated G2/M progression delay and cell apoptosis. J Biol Chem, 2016; 291: 4779–4792.
- 43. Li X, Tai HH (). Activation of thromboxane A receptors induces orphan nuclear receptor Nurr1 expression and stimulates cell proliferation in human lung cancer cells. Carcinogenesis, 2009; 30: 1606–1613.
- 44. Wang D, Dubois RN. Eicosanoids and cancer. Nat Rev Cancer, 2010; 10: 181–193.
- 45. Liu B, Qu L, Yan S.I. Cyclooxygenase-2 promotes tumor growth and suppresses tumor immunity. Cancer Cell International, 2015; 15
- 46. Patel CK, Sen DJ. COX-1 and COX-2 inhibitors: Current status and future prospects over COX-3 inhibitors Int J Drug Dev Res, 2009; 1: 136–145.
- 47. HashemiGoradel N, Najafi M, Salehi E, Farhood B, Mortezaee K. Cyclooxygenase-2 in cancer: A review. J Cell Physiol, 2019; 234(5): 5683-5699.
- 48. Yu T, Lao X, Zheng H. Influencing COX-2 Activity by COX Related Pathways in Inflammation and Cancer, Mini Rev. Med. Chem, 2016; 16(15): 1230-1243.
- 49. Pannunzio A, Coluccia M. Cyclooxygenase-1 (COX-1) and COX-1 Inhibitors in Cancer: A Review of Oncology and Medicinal Chemistry Literature. Pharmaceuticals (Basel), 2018; 11(4): pii E101.
- 50. Lee G, Ng HT. Clinical evaluations of a new ovarian cancer marker, COX-1. Int J Gynaecol Obstet, 1995; 49: S27–S32.
- 51. Eilati E, Pan L, Bahr JM, Hales DB. Age dependent increase in prostaglandin pathway coincides with onset of ovarian cancer in laying hens. Prostaglandins Leukot Essent Fat Acids, 2012; 87: 177–184.
- 52. Hales DB, Zhuge Y, Lagman JA, Ansenberger K, Mahon C, Barua A, Luborsky JL, Bahr JM. Cyclooxygenases expression and distribution in the normal ovary and their role in ovarian cancer in the

<u>www.ejpmr.com</u> 95

- domestic hen (Gallus domesticus) Endocrine, 2008; 33: 235–244.
- 53. Urick ME, Johnson PA. Cyclooxygenase 1 and 2 mRNA and protein expression in the Gallus domesticus model of ovarian cancer. Gynecol Oncol, 2006; 103: 673–678.
- 54. Tiano HF, Loftin CD, Akunda J, Lee CA, Spalding J, Sessoms A, Dunson DB, Rogan EG, Morham SG, Smart RC, Langenbach R). Deficiency of either cyclooxygenase (COX)-1 or COX-2 alters epidermal differentiation and reduces mouse skin tumorigenesis. Cancer Res, 2002; 62: 3395–3401.
- 55. Kino Y, Kojima F, Kiguchi K, Igarashi R, Ishizuka B, Kawai S. Prostaglandin E2 production in ovarian cancer cell lines is regulated by cyclooxygenase-1, not cyclooxygenase-2. Prostaglandins Leukot Essent Fat Acids, 2005; 73: 103–111.
- 56. Lau MT, Wong AS, Leung PC. Gonadotropins induce tumor cell migration and invasion by increasing cyclooxygenases expression and prostaglandin E(2) production in human ovarian cancer cells Endocrinology, 2010; 151: 2985–2993.
- Vitale P, Panella A, Scilimati A, Perrone MG. COX-1 inhibitors: Beyond structure toward therapy Med Res Rev, 2016; 36: 641–671.
- Tortorella MD, Zhang Y, Talley J. Desirable Properties for 3rd Generation Cyclooxygenase-2 Inhibitors. Mini Rev Med Chem, 2016; 16: 1284–1289.
- Pang LY, Hurst E.A, Argyle D.J. Cyclooxygenase A Role in Cancer Stem Cell Survival and Repopulation of Cancer Cells during Therapy. Stem Cells Int, 2016: 2048731.
- 60. Qiu W, X. Wang, Leibowitz B, Liu H, Barker N, Okada N, Oue N, Yasui W, Clevers H, Schoen R.E, et al. Chemoprevention by nonsteroidal anti-inflammatory drugs eliminates oncogenic intestinal stem cells via SMAC-dependent apoptosis. Proc. Natl. Acad. Sci. USA, 2010; 107: 20027–20032.
- 61. Gunaydin C, Bilge SS). Effects of Nonsteroidal Anti-Inflammatory Drugs at the Molecular Level. Eurasian J Med, 2018; 50(2): 116-121.
- Subbaramaiah K, Dannenberg A.J). Cyclooxygenase
 a molecular target for cancer prevention and treatment. Trends Pharmacol. Sci, 2003; 24(2): 96–102
- 63. Neault J.F, Naoui M, Manfait M, Tajmir-Riahi H.A. Aspirin-DNA interaction studied by FTIR and laser Raman difference spectroscopy. FEBS Lett, 1996; 382: 26–30
- 64. Subbaramaiah K, Dannenberg A.J). Cyclooxygenase 2: a molecular target for cancer prevention and treatment. Trends Pharmacol. Sci, 2003; 24(2): 96–102.
- 65. Puranik R, Bao S, Bonin AM, Kaur R, Weder JE, Casbolt L, Hambley TW, Lay PA, Barter PJ, Rye KA. A novel class of copper(II)- and zinc(II)-bound non-steroidal anti-inflammatory drugs that inhibits acute inflammation in vivo Cell Bio sci, 2016; 6: 9.

- 66. Bonin AM, Yáñez JA, Fukuda C, Teng XW, Dillon CT, Hambley TW, Lay PA, Davies NM. Inhibition of experimental colorectal cancer and reduction in renal and gastrointestinal toxicities by copperindomethacin in rats. Cancer Chemother Pharmacol, 2010; 66(4): 755-64.
- 67. Verbeeck RK. Pharmacokinetic drug interactions with nonsteroidal anti-inflammatory drugs. Clin Pharmacokinet, 1990; 19: 44–66.
- 68. Tielleman T1, Bujanda D1, Cryer B2). Epidemiology and Risk Factors for Upper Gastrointestinal Bleeding. GastrointestEndoscClin N Am, 2015; 25(3): 415-28.
- 69. Whelton A. Nephrotoxicity of nonsteroidal antiinflammatory drugs: physiologic foundations and clinical implications Am J Med, 1999; 106(5B): 13S-24S.
- 70. Zavodovsky BV Sivordova LE. Cardiovascular safety of nonsteroidal anti-inflammatory drugs in chronic inflammatory rheumatic diseases. Ter Arkh, 2018; 90(8): 101-106.
- 71. Walker C, Biasucci LM. Cardiovascular safety of non-steroidal anti-inflammatory drugs revisited. Postgrad Med, 2018; 130(1): 55-71.
- 72. Smith WL (). Prostanoid biosynthesis and mechanisms of action Am J Physiol, 1992; 263(2 Pt 2): F181-91.
- 73. Mérida E, Praga M (). NSAIDs and Nephrotic Syndrome Clin J Am Soc Nephrol, 2019; 14(9): 1280-1282.
- 74. Bakhriansyah M, Souverein PC, van den Hoogen MWF, de Boer A, Klungel OH. Risk of Nephrotic Syndrome for Non-Steroidal Anti-Inflammatory Drug Users. Clin J Am Soc Nephrol, 2019; 14(9): 1355-1362.
- 75. Khan S, Andrews KL, Chin-Dusting JPF. Cyclo-Oxygenase (COX) Inhibitors and Cardiovascular Risk: Are Non-Steroidal Anti-Inflammatory Drugs Really Anti-Inflammatory? Int J Mol Sci, 2019; 20(17): E4262.
- 76. Dovizio M, Alberti S, Sacco A, GuillemLlobat P, Schiavone S, Maier TJ, Steinhilber D, Patrignani P. Novel insights into the regulation of cyclooxygenase2 expression by plateletcancer cell crosstalk. Biochem Soc Trans, 2015; 43: 707–714.
- 77. Sun SX, Lee KY, Bertram CT, Goldstein JL (). Withdrawal of COX- 2 selective inhibitors rofecoxib and valdecoxib: impact on NSAID and gastroprotective drug prescribing and utilization. Curr Med Res Opin, 2007; 23(8): 1859-66.
- 78. Kim SD, Cho KS. Samter's Triad: State of the Art. ClinExpOtorhinolaryngol, 2018; (2): 71-80.
- Laidlaw TM. Pathogenesis of NSAID-induced reactions in aspirin-exacerbated respiratory disease. World J Otorhinolaryngol Head Neck Surg 2018; 4(3): 162-168.
- 80. Li KL, Lee AY, Abuzeid WM. Aspirin Exacerbated Respiratory Disease: Epidemiology, Pathophysiology, and Management Med Sci (Basel), 2019; 7(3): pii: E45.

- 81. Hadjidakis DJ, Androulakis II. Bone remodeling Ann N Y AcadSci, 2006; 1092: 385-96.
- 82. Kenkre JS, Bassett J. The bone remodeling cycle. Ann ClinBiochem, 2018; 55(3): 308-327.
- 83. Pountos I, Georgouli T, Calori GM, Giannoudis PV. Do nonsteroidal anti-inflammatory drugs affect bone healing? A critical analysis. Scientific World Journal, 2012; 606404.
- 84. Rao PP, Kabir SN, Mohamed T. Nonsteroidal Anti-Inflammatory Drugs (NSAIDs): Progress in Small Molecule Drug Development. Pharmaceuticals, 2010; 3(5): 1530-1549.
- 85. Mitchell JA, Warner TD. COX isoforms in the cardiovascular system: Understanding the activities of non-steroidal anti-inflammatory drugs Nat Rev Drug Discov, 2006; 5: 75–85.
- 86. Lanas A. Role of nitric oxide in the gastrointestinal tract. Arthritis Res Ther, 2008; 10(2).
- 87. Colazzo F, Gelosa P, Tremoli E, Sironi L, Castiglioni L. Role of the Cysteinyl Leukotrienes in the Pathogenesis and Progression of Cardiovascular Diseases. Mediators Inflamm, 2017; 2432958.
- 88. Haeggström JZ. Leukotriene biosynthetic enzymes as therapeutic targets. J Clin Invest, 2018; 128(7): 2680-2690.
- 89. Harborne, J. B. (ed). The Flavonoids: Advances in research since 1986. New York: Chapman and Hall 1994.
- Okoli, C. O., Akah P. A., Nwafor, S. V. (). Antiinflammatory activity of plants. J. Nat. Remed, 2003; 3: 1-30.
- 91. Ohadoma SC, Akah PA, Enye JC. Pharmacological assessment of the antinociceptive anti-inflammatory effects of the methanol extract and fractions of *Lupinus arboreus*. Asian J. SCi. Technol, 2015; 6(8): 1663-1668
- 92. Ohadoma SC. Pharmacological assessment of *Lupinus arboreus* Sims (Fabaceae) methanol extract and fractions for antinociceptive and anti-inflammatory effects. A P.hD. Thesis, Department of Pharmacology and Toxicity, University of Nigeria, Nsukka 2014. Available online at University of Nigeria, Nsukka, Visual Library, 2014; 31.
- 93. Geissman, T. A., Crout, D. H. G. Organic chemistry of secondary plant metabolism. San Francisco: Freeman, Cooper & Company, 1969; 183-230.
- 94. Murray, M. T. Encyclopedia of nutritional supplements. California: prima publishing, 1996; 320-331.
- 95. Mills, S., Bone, K. Principles and practice of phytotherapy- Modern Herbal Medicine. New York: Churchhill Livingstone, 2000; Pp. 31-34.
- 96. Robbinson, T. The organic constituents of higher plants- their chemistry and relationships. 6th ed. North Amherst: Cordus press, 1991; Pp. 187-217
- 97. Blumenthal, M. The ABC Clinical Guide to Herbs. Austin: American Botanical Council, 2003; 239.
- 98. Robbers, J. E., Tyler, V. E. Tyler's herbs of choice-The therapeutic use of phytomedicinals.

- Binghamton, New York: Haworth Herbal Press, 2000; Pp. 69, 89.
- Ohadoma SC, Eban LK. Antioxidant and free radical scavenger effects of methanol leaf extract of *Lupinus arboreus*. Euro. J Biomed. Pharm Sci, 2018; 5(1) 70-73.
- 100.Harborne, J. B., Williams, C.A. Advances in flavonoid research since 1992. J. Phytochem. 2000; 55: 481-504.
- 101. Hye, H. K., Gafur, M. A. "Anti-inflammatory and anti-arthritic activity of a substance isolated from *Dalbergia volubilis*. Ind. J. Med. Res, 1975; 163: 93-100.
- 102.Srimal, R. C., Sharma, S. C., Tandon, J. S. Antiinflammatory and other pharmacological effects of *Hedtchium spicatum* (Buch-Hem). Ind. J. Pharmacol, 1984; 16: 143-147.
- 103.Ogundaini, A. Isolation of two anti-inflammatroy biflavanoid from *Sacrophyte pieriei*. J. Nat. Prod, 1996; 59(6): 587-590.
- 104.Hedge, V. R. Phospholipase A2 inhibitors from an *Erythrina species* from Samoa. J. Nat. Prod, 1997; 60: 537-539
- 105.Stephan, S. Workup dependent formation of lipooxygenase inhibitory boswellic acid analogues .J. Nat. Prod, 2000; 63(8): 1058-1061.
- 106. Jiang, L. L. Two new prinilated three benzoxepin derivates as Cyclooxygenase, inhibitors from *Perilla frutescens* var. Acuta. J. Nat. Prod, 2000; 63(3): 403-405.
- 107. Delporte, C. Anti-inflammatory and antipyretic activities of Spartidienedione isolated from *Psila spartiodes*. Int. J. Pharmacog, 1996; 34(3): 179-183.
- 108.Clelia, G.. New sesquiterpene derivatives from the Sponge dysidea species with a selective inhibitor profile against human phospolipase A2 and other leucocyte functions. J. Nat. Prod, 2001; 64(5): 612-615.
- 109. Satyavati, G. V., Dwarkanath, C., Tripathi, S. N. Experimental studies on the Hypocholesterolemic effect of *Commiphora mukul* Engl (Guggul). Ind. J. Med. Res, 1969; 57: 1950-1962.
- 110.Shukla, B. A comparative study of guggal and *Yograji guggal* on immune responses in rabbits. Indian Drugs, 1986; 23: 335-337.
- 111.Ohadoma SC, Lawal BAS, Chukwu LC. Antiarthritic activity of methanol leaf extract of Chikadoma in complete Freud's Adjust- induced arthritic rats. British J Med. Health Sci, 2019; 1(2): 43-46.
- 112.Ohadoma SC. Scientific basis for the therapeutic use of *Lipunus arboreus*. Euro. J Pharm Med. Res, 2018; 5(3): 30-34.
- 113.Singh, N. A pharmacological study of *Cyperus rotendus*. Ind. J. Med. Res. 1970; 58: 103-109.
- 114. Climanaga K. In vitro anticomplementary activity of constituents from *Morinda nordindoides*: J. Nat. Prod, 1995; 58(3): 371-278.
- 115.Abad, M.J., Bermejo, P., Villar, A. Antiinflammatory activity of two flavanoids from

- Nacetum macrophyllum. J. Nat. Prod, 1993; 56(7): 1164-1167.
- 116.Selvador, M. A glycosyl analogue of diacyclglycerol and other anti-inflammatory constituents from *Inula viscose*. J. Nat. Prod, 1999; 62(4): 601-604.
- 117.Moroney, M. A., Alcaraz, M. J., Forder, R. A., Carey, F., Hoult, J. R. Selectivity of neutrophil 5lipoxygenase and cyclo-oxygenase inhibition by an anti-inflammatory flavonoid glycoside and related aglycone flavonoids. J. Pharm. Pharmacol, 1988; 40: 787-92.
- 118. Kavimani, S., Mounissamy, V. M., Gunasegaran, R. Analgesic and anti-inflammatory activities of Hispudulin isolated from *Helichrysum bracteatum*. Indian Drugs, 2000; 37: 582-584.
- 119.Jachak, S. M. Natural Products: potential source of COX Inhibitors. Clin. Res. Immunol. Pharm. Sci. (CRIPS), 2001; 2: 12-15.
- 120.Bhalla, T. N. Calophyllolide, a new nonsteroidal anti-inflammatory agent. Ind. J. Med. Res, 1980; 72: 762-765.
- 121. Silvan, A. M. Anti-inflammatory activity coumarins from *Santolina obngiofolia*. J. Nat. Prod, 1996; 59(12): 1183-1185
- 122.Shankarnarayan, D., Gopalakrishnan C., Kameshwaran, L. Pharmacological profile of Mangostin and its derivative. Arch. Int. Pharmacodyn, 1979; 239: 257-269.
- 123.Gopalkrishnan, C. Anti-inflammatory and C. N. S. depressant activities of xanthones from *Calophyllum inophyllum* and *Musa ferrea*. Ind. J. Pharmacol, 1980; 12: 181- 191.
- 124.Akihisa, T., Yasukawa, K., Kimura, Y., Yamanouchi, S., Tamura, T. Sasanquol, a 3,4-secotriterpene alcohol from sasanqua oil, and its anti-inflammatory effect J. Phytochem, 1998; 48: 301-305.
- 125.Singh, R. K., Pandey, B.L. Further study of anti-inflammatory effects of *Abies pindrow*. J. phytother. Res, 1997; 11: 535-537.
- 126.Bhargava, K. P. Anti-inflammatory activity of saponins and other natural. Products. Ind. J. Med. Res, 1970; 58: 724-730.
- 127.Ghosh, D., Thejomoothy, P., Veluchamy, G. Antiinflammatory and analgesic activities of oleanolic acid 3-/3-Glucoside (RGD-1) from *Randia dumetorum* (Rubiaceae). Ind. J. Pharmacol, 1983; 15: 331-342.
- 128. Singh, G. B., Singh, B., Atal, C. K. Assessment of total boswellic acids for anti-inflammatory activity. Ind. J. Pharmacol, 1984; 14: 103.
- 129.Okoli, C. O., Akah, P. A. Mechanisms of antiinflammatory activity of the leave extracts of *Culcasia scandens* P. Beauv (Araceae). Pharmacol. Biochem. Behav, 2004; 79: 473-481.
- 130. Froton, M. H. Pharmacological investigations on aspinasterol isolated from *Simplocos spicata*. Ind. J. Pharmacol, 1983; 15: 197-201.
- 131.Zhang, J., Meng, Z., Zhang, M., Ma D., Xu, S. Kodama, H. Effect of six steroidal saponins isolated

from *Anemarrhenae rhizome* on platelet aggregation and hemolysis in human blood. Clin. Chim. Acta, 1999; 289: 79-88.