



**VIRTUAL SCREENING AND DOCKING STUDIES OF SYNTHESIZED CHALCONES: A
POTENT ANTI- CANCER DRUGS**

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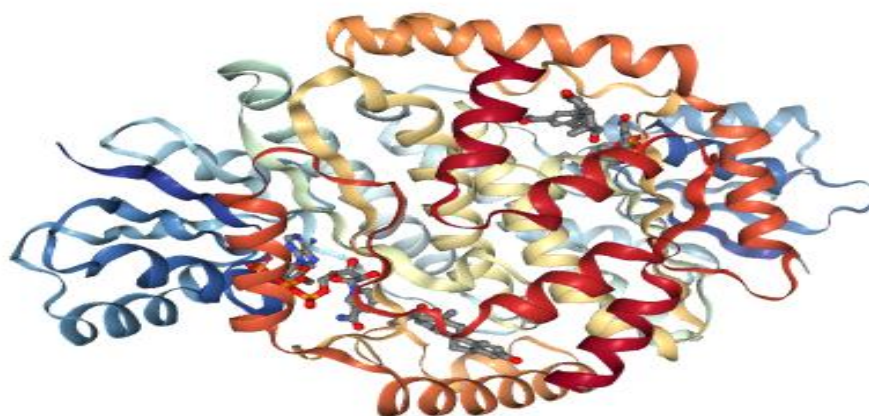
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ABSTRACT

A novel series of chalcones are characterized by processing an enone moiety between two aromatic rings. The chalcone-like agents, during which the covalent bond of the enone system is embedded with in indole ring. Statistically significant structure-based qualitative structure activity relationship models were generated and validated through acceptable predictive ability to support internal and external set of compounds. Screening of most valuable drug among of pre-synthesized drug on the idea of binding efficiency to focus on receptor was administered by docking view. Molecular docking programme Glide iGEMDock was went to determine binding feasibility of seven analogues of chalcones. The comparison of docking parameters showed, quite 5 analogues are better ligand of 3HB5. The binding of chalcones to 3HB5 is mediated by both hydrogen bonding, hydrophobic and polar interactions. Our result suggest that chalcones analogues are promising lead compounds for the event of anti-cancer drugs (mainly the breast cancer).

KEYWORDS: Chalcones, Docking, Binary And Ternary Of Novel Inhibitor Of 17 Beta-HSD, Structure Activity Relationship, Ligand, Indole, Igem Docking.



3D View of binary and ternary crystal structures of of novel inhibitor of 17 beta-HSD type 1 : lead compound for breast cancer.

INTRODUCTION

Chalcones are an important class of natural compounds. Chalcones, or 1, 3-diaryl-2-propen-1-ones, belong to the flavonoid family. they're precursors for flavanoid biosynthesis, which play ecological role in regard to plant color. Chemically they contains open-chain

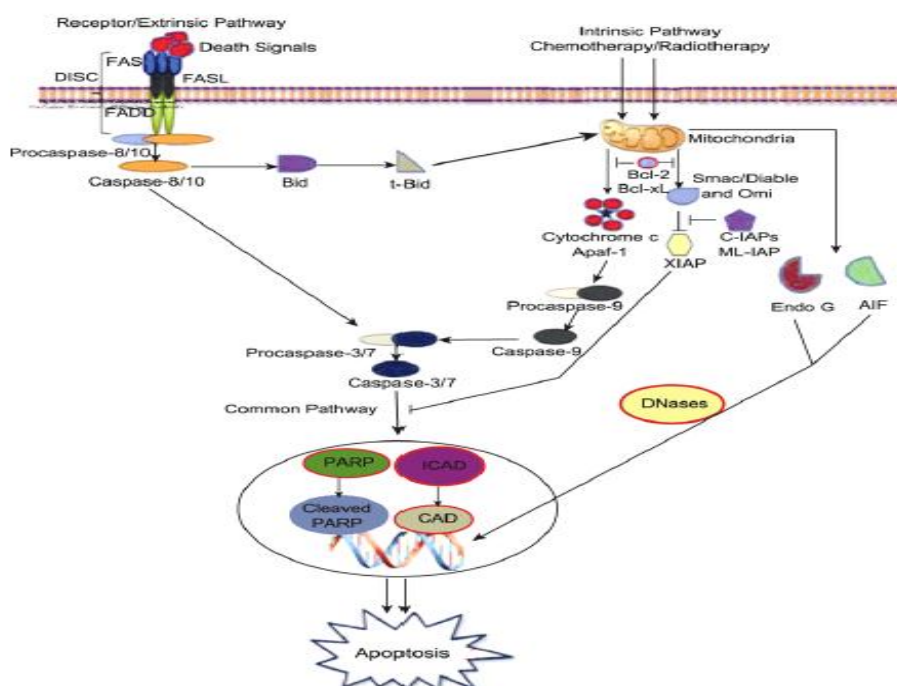
flavanoids during which the aromatic ring join by a three-carbon α , β unsaturated carbonyl system.^[1,2]

Anti-cancer agents are usually evaluated for his or her ability to induce apoptosis. Indoles are verified to inhibit proliferation, expansion and invasion of human cancer cells.^[3,4,5-7] Many mechanisms of apoptosis stimulation

of indole derivatives, I3C and DIM, were reported for, (a): down-regulation of anti-apoptotic gene products like Bcl-2 (B-cell lymphoma 2) and Bcl-XL (B-cell leukemia-extra large), (b): down-regulation of the inhibitor of apoptosis proteins, e.g. CIAPs, X-chromosome linked inhibitor of apoptosis protein (XIAP) and survival, (c): up-regulation of pro-apoptotic factors like Bax gene, (d): liberation of mitochondrial cytochrome c additionally to stimulating of caspase-9 and caspase-3[8], and (e): inhibition of the NF- κ B signaling pathway.^[9-14] an enormous number of diverse mechanisms of apoptosis induction by indoles have also been reported^[15-19] Figure 1 demonstrates the extrinsic and thus the intrinsic pathways of apoptosis (programmed cell death). The Extrinsic Route: within the extrinsic pathway, signal molecules identified as ligands, which are released by the immune system's natural killer cells possess the Fas ligand (FasL) on their exterior to connect to transmembrane death receptors on the target cell. After the binding of the death ligand to the death receptor the target cell triggers multiple receptors to aggregate together on the surface of the target cell. The aggregation of these receptors recruits an adaptor protein mentioned as Fas-associated death domain protein (FADD) on the cytoplasmic side of the receptors. FADD, in turn, recruits Caspase-8. Caspase-8 will then be activated and may be now able to directly activate caspase-3 and caspase-7. The activation of caspase-3 will initiate the degradation of the cells.^[20] The Intrinsic Route: The intrinsic pathway is triggered by cellular strain, particularly mitochondrial stress caused by factors like DNA damage from chemotherapy or UV exposure. Upon delivery of the strain signal, the pro-apoptotic proteins within the cytoplasm (Bcl-2-like protein 4 (BAX) and BAX-like

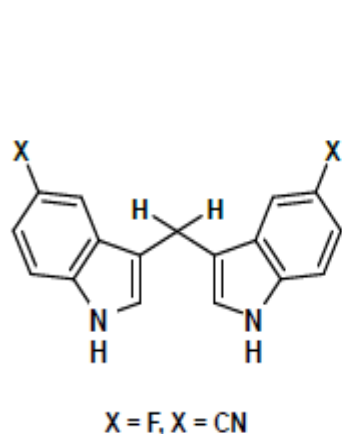
Bcl-2 homology domain 3 protein (BID)) bind to the outer membrane of the mitochondria to signal the discharge of the inside content.

The interaction between the pro-apoptotic (BAX and BID) and the antiapoptotic proteins (Bcl-2) on the surface of the mitochondria is thought to be important in the formation of the PT pores in the mitochondria, and hence, the release of cytochrome c and the intramembrane content from the mitochondria. Following the release, cytochrome c forms a multi protein complex known as apoptosome which consists of cytochrome c, Apaf-1, procaspase-9 and ATP. Following its formation, the complex will activate caspase-9. The activated caspase-9 will then turn the procaspase-3 and procaspase-7 into active caspase-3 and active caspase-7. These activated proteins initiate cell degradation or cell death. Besides the release of cytochrome c from the intramembrane space, the intramembrane also releases Smac/Diablo proteins to inhibit the inhibitor of apoptosis (IAP). IAP is a protein family which consists of 8-human derivatives. Their function is to stop apoptotic cell death by binding to caspase-3, caspase-7 and caspase-9 and inhibit them, the schematic representation of these pathways are shown in Figure 1.^[21]

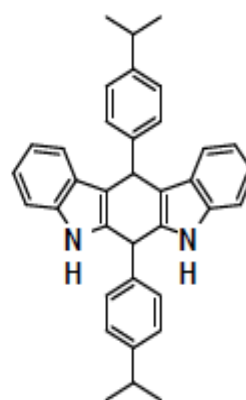


“Fig. 1”: Intrinsic and extrinsic pathways leading to apoptosis [available from <https://innspubnet.files.wordpress.com/2015/04/mitochondrial-pathway.jpg>].

Nowadays, clinical association of human sex organ cancers requests new chemotherapeutics. In recent times, tons of hard works are done to arrange antiproliferative signaling pathway of indole-3-carbinol and its foremost indole metabolite 3, 3'-diindolylmethane (DIM).^[22-28] While DIM significantly reduces the occurrence of impulsive and carcinogen induced mammary tumor establishment (Figure 2).^[29-31] It also exhibits unpleasant promoting action in convinced investigation procedure.^[32,33] As a result, the choice was to seem for novel effective chemotherapeutics amongst 3, 3'-diindolylmethane derivatives. Moreover, the X-ray studies of 5, 5'-dimethoxy-3,3'-methanediyl-bis-indole^[34] revealed its 'butterfly' conformation, which is analogous to the



one proposed earlier for inhibitors of HIV-1 polymerase, sharing the mode of action of nevirapine.^[35] Other diindolylmethane derivatives and their corresponding tetrahydroindolocarbazoles are synthesized and screened for anti-cancer activity during which two compounds indicated were significantly more sensitive for several neoplastic cell lines like their GI50 values. The very best antiproliferative activity recorded for the carbazole derivatives during a nanomolar scale towards the three certain cancers cell lines: non-small lung cell NCI-H460 with GI = 616 nmol/L, ovarian cancer cell line OVCAR-4 with GI = 562 nmol/L and breast neoplastic cell line MCF7 with GI = 930 nmol/L (Figure 2).^[36]

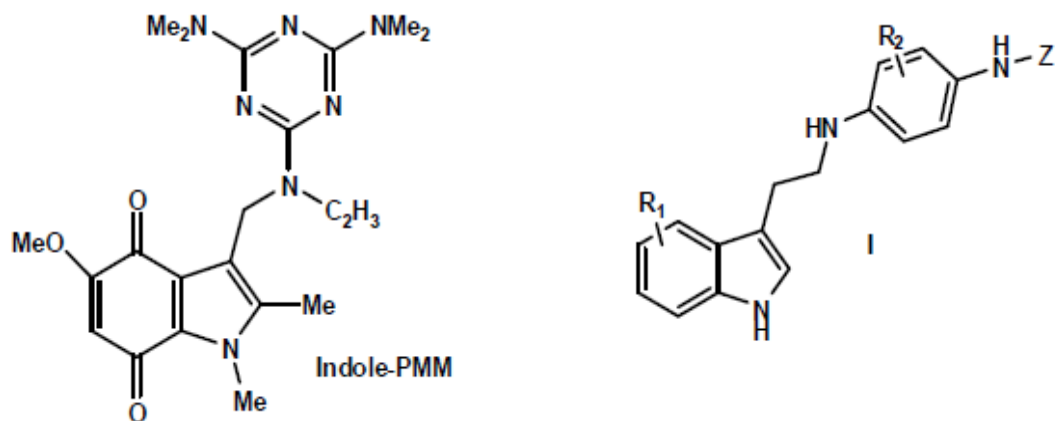


Carbazole derivative

“Fig. 2”: 3, 3'-diindolylmethane derivatives and tetrahydro-indolocarbazoles.

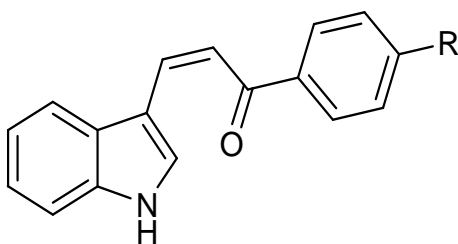
Dorota *et al.*^[37,38] in 2005 synthesized the disubstituted diindolylmethanes fluoro and cyano derivatives which decrease the expansion of MCF7 (breast), NCI-H460 (lung) and SF-268(NCS) cells, considerably 5,5'-difluoro-3,3'-methanediyl-bis-indole and 5,5'-dicyano-3,3'-methanediyl-bis-indole were tested against the MCF7 (breast), NCI-H460 (lung) and SF-268 (CNS) tumor cell lines. The results are reported because the proportion of growth of the tested cells to untested control cells (Figure 3). F-derivative at concentration 1.10–4.00 mol/L reduces the expansion of MCF7, NCI-H460, and SF-268 cell lines to 1 / 4, 0% and 2%, whereas the CN derivative at concentration 5.10–5.00 mol/L to 4%, 1% and 9%, respectively. Both compounds are extremely cytotoxic in vitro towards those tumor lines. Their cytotoxicity indicates that they could be motivating as prospective antitumoral chemotherapeutics.^[37,38] Indoles (I3C and DIM or its derivatives) are revealed to induce apoptosis in breast^[39-45], squamous cell carcinoma^[46], cholangiocarcinoma^[47], colon^[48-51], cervical^[52], ovarian^[53], pancreatic^[54,55] and prostate^[56-59] cancer cells. Many other indole derivatives that were reported as active anti-cancer agents as follow: the potential prodrug (1,2-dimethyl-3-(*N*-(4,6-bis(dimethylamino)-1,3,5-triazin-2-yl)-*N*-trideuteriomethylaminomethyl)-5-methoxyindole-4,7-dione), pentamethylmelamine (PMM) in which the labeled pentamethylmelamine is attached to an indole-4,7-dione moiety has attracted

much interest as an anti-tumor agent for over 35 years (Figure 3). It entered clinics in the 1970s for the treatment of ovarian carcinoma but difficulties were encountered, as it was insoluble in water and thus is difficult to formulate. However, it has recently been recognized as a second-line treatment for ovarian carcinoma.^[60-67,68-70]



“Fig. 3”): Structure of prodrug indole-PMM derivative and tryptamine derivatives.

MATERIALS AND METHODS



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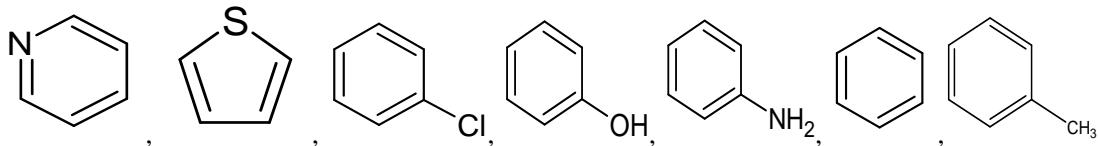


Table 1: Interaction of chalcone A1-A7 with 3HB5 (Total energy, logP, Vander Waals energy, H-bonding energy) and with different amino acids.

LIGAND	LOG P	TOTAL ENERGY	VDW	H-BOND	V-M-GLY-148	V-S-PRO-150	V-M-VAL-276	V-M-PHE-284	H-S-ASN-90	V-M-GLY-9	V-S-SER-11	V-S-ARG-37	H-S-ARG-132	V-S-ARG-76	V-M-VAL-79	H-M-THR-3	H-S-ARG-83	HS-ASP-85	H-M-SER-134	V-S-THR-3
A1	3.68	-77.37	-73.54	-3.83	-7.4	-9.6	-7.9	-4.5	0	0	0	0	0	0	0	0	0	0	0	0
A2	2.61	-79.05	-79.05	0	-4.2	-5.9	-5.4	0	0	0	0	0	0	0	0	0	0	0	0	0
A3	4.46	-79.55	-73.5	-6.05	0	0	0	0	-3.5	-8.1	-4.2	4.4	0	0	0	0	0	0	0	0
A4	3.30	-84.98	-70.69	-14.29	0	0	0	0	0	0	0	0	-3.5	-6.7	-4.6	0	0	0	0	0
A5	2.86	-83.62	-79.49	-4.12	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
A6	3.78	-76.53	-68.53	-8.01	0	0	0	0	0	0	0	0	0	0	0	0	0	-3.5	-3.2	-6.6
A7	4.23	-83.59	-79.26	-4.33	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

A1: (Z)-3-(1*H*-indol-3-yl)-1-(thiophen-2-yl) prop-2-en-1-one.

A2: (Z)-3-(1*H*-indol-3-yl)-1-(pyridin-2-yl) prop-2-en-1-one.

A3: (Z)-1-(4-chlorophenyl)-3-(1*H*-indol-3-yl) prop-2-en-1-one.

A4: (Z)-1-(4-hydroxyphenyl)-3-(1*H*-indol-3-yl) prop-2-en-1-one.

A5: (Z)-1-(4-aminophenyl)-3-(1*H*-indol-3-yl) prop-2-en-1-one.

A6: (Z)-3-(1*H*-indol-3-yl)-1-phenylprop-2-en-1-one.

A7: (Z)-3-(1*H*-indol-3-yl)-1-*p*-tolylprop-2-en-1-one.

EXPERIMENTAL

Compound selection and ligand preparation

All the compounds (7 compounds) were drawn on Chem Draw as in Table 1 and were optimized using MOPAC (a computational tool) using AMI calculation and closed shell restricted with RMS gradient.

Protein preparation: Bioinformatics is seen as emerging field with the potential to significantly improve how drugs are found, brought to the clinical trials and eventually released to the marketplace. The crystal structure of 3HB5 having PDB ID: 1MVT was taken (<http://www.rcsb.org/>). The protein was prepared using Molegro Molecular viewer 2.5 and the same was used for the virtual drug screening.

Docking studies: Herein, iGEMDock was used for the drug screening as in table 1. iGEMDock is a molecular docking tool and generates diversity of chalcone conformations from different seed with high temperature molecular dynamics. Then, it orients the chalcone conformations within the defined protein active site by translating the center of the surfactant. Each orientation is subjected to simulated annealing molecular dynamics and sorted according to the interaction energy. IGEMDOCK energy function consists of electrostatic, steric, and hydrogen-bonding potentials. Steric and hydrogen bonding potentials use a linear model. There are four main steps which are used here. Parameters used for drug screening in iGEMDOCK were as followed: initial step sizes ($\sigma=0.8$ and $\psi=0.2$), family competition length ($L=2$), population size ($N=200$), and recombination probability ($pc=0.3$). Optimization is set to generate 70 iterations for which it generated 1200 solutions in one generation process and if exceeded then it terminated after 84,000 solutions.

Docking studies not only provide an understanding of the binding mode of the ligands but are also employed to validate homology models. The molecular models for 3HB5 were generated by Molegro Molecular Viewer 2.5 docking module. Among the docked poses for each ligand, one with the highest dock score was chosen as the final conformation. Docking allows screening a database of compounds and calculating the strongest binders based on various scoring functions. It explores ways in which two molecules, such as drugs and an enzyme, fit together and docks to each other well. The molecule may bind to receptor and modify their function. The interaction of drug and receptor complex was identified via docking and their relative stabilities were evaluated using molecular dynamics and also evaluated their binding affinities using free energy simulations.

RESULTS AND DISCUSSION

Docking studies are used extensively in drug discovery such as in the prediction of ligand-receptor complex structures and also to rank the ligand molecules based upon the binding energies of the corresponding ligand-enzyme complex. The objective of our docking study is

to elucidate the potential interaction mode of the chalcones with 3HB5. A general conclusion derived from these docking results is that the side chain of the VAL and TYR forms hydrogen bonding with the chalcone. The details mentioned in Table 1. Interestingly, this interaction is almost conserved with all studied inhibitors. The docking calculations provided us with a general static picture of the most energetically favourable binding orientation of inhibitors to the enzyme. To obtain further insight into the dynamic changes of the docked inhibitors within the enzyme active site pocket over time, the lowest energy docked complex of the most active inhibitor, 7, was subjected to unconstrained MD simulations. The standard drug Lichochalone A and the logP value of A3 and Lichochalone A are found similar.

CONCLUSION

The continued development of novel anti-cancer chemotherapies, particularly those aimed toward new pathways, is important for the successful treatment of cancer as resistance to presently utilized drugs becomes more widespread. Unlike its human host, the cancer parasite doesn't possess a salvage pathway for indole and must believe de novo biosynthesis for its metabolic needs.

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REFERENCES

1. B. A. Bohm, in the flavonoids (Eds.: J. B. Harborne T. J. Mary, H. Mabry), Chapman and Hall, London, 1975; 442-504.
2. Z. Nowakowska, Eur. J. Med. Chem., 2007; 42: 125-137.
3. Ahmad A, Sakr WA, Rahman KW. Mechanisms and therapeutic implications of cell death induction by indole compounds. Cancers (Basel), 2011; 3(4): 2955-2974.
4. Fischer E, Jourdan F. Ueber die Hydrazine der Brenztraubensäure. Berichte der deutschen chemischen Gesellschaft, 1883; 16(2): 2241-2245.
5. Sarkar FH, Li Y. Harnessing the fruits of nature for the development of multi-targeted cancer therapeutics. Can-cer Treat Rev., 2009; 35(7): 597-607.
6. Sarkar FH, Li Y, Wang Z, et al. Cellular signaling perturbation by natural products. Cell Signal, 2009; 21(11): 1541-1547.
7. Moiseeva EP, Almeida GM, Jones GD, et al. Extended treatment with physiologic concentrations of dietary phytochemicals results in altered gene expression, reduced growth, and apoptosis of cancer cells. Mol Cancer Ther., 2007; 6(7): 3071-3079.

8. Cover CM, Hsieh SJ, Tran SH, et al. Indole-3-carbinol inhibits the expression of cyclin-dependent kinase-6 and induces a G1 cell cycle arrest of human breast cancer cells independent of estrogen receptor signaling. *J Biol Chem.*, 1998; 273(7): 3838–3847.
9. Rahman KW, Sarkar FH. Inhibition of nuclear translocation of nuclear factor- κ B contributes to 3,3'-diindolylmethane-induced apoptosis in breast cancer cells. *Cancer Res.*, 2005; 65(1): 364–371.
10. Sarkar FH, Li Y. Indole-3-carbinol and prostate cancer. *J Nutr.*, 2004; 134(Suppl 12): 3493S–3498S.
11. Wang Z, Yu BW, Rahman KM, et al. Induction of growth arrest and apoptosis in human breast cancer cells by 3,3'-diindolylmethane is associated with induction and nuclear localization of p27kip. *Mol Cancer Ther.*, 2008; 7(2): 341–349.
12. Sarkar FH, Li Y, Wang Z, et al. NF- κ B signaling pathway and its therapeutic implications in human diseases. *Int Rev Immunol*, 2008; 27(5): 293–319.
13. Kong D, Li Y, Wang Z, et al. Inhibition of angiogenesis and invasion by 3,3'-diindolylmethane is mediated by the nuclear factor- κ B downstream target genes MMP-9 and uPA that regulated bioavailability of vascular endothelial growth factor in prostate cancer. *Cancer Res.*, 2007; 67(7): 3310–3319.
14. Bhuiyan MM, Li Y, Banerjee S, et al. Down-regulation of androgen receptor by 3,3'-diindolylmethane contributes to inhibition of cell proliferation and induction of apoptosis in both hormone-sensitive LNCaP and insensitive C4-2B prostate cancer cells. *Cancer Res.*, 2006; 66(20): 10064–10072.
15. Ahmad A, Sakr WA, Rahman KM. Role of nuclear factor- κ B signaling in anticancer properties of indole compounds. *J Exp Clin Med*, 2011; 3(2): 55–62.
16. Rahman KM, Banerjee S, Ali S, et al. 3, 3'-Diindolyl-methane enhances taxotere-induced apoptosis in hormone-refractory prostate cancer cells through survivin down-regulation. *Cancer Res.*, 2009; 69(10): 4468–4475.
17. Bhatnagar, N, Li X, Chen Y, et al. 3,3'-diindolylmethane enhances the efficacy of butyrate in colon cancer prevention through down-regulation of survivin. *Cancer Prev Res.*, 2009; 2(6): 581–589.
18. Ahmad A, Kong D, Sarkar SH, et al. Inactivation of uPA and its receptor uPAR by 3,3'-diindolylmethane (DIM) leads to the inhibition of prostate cancer cell growth and migration. *J Cell Biochem*, 2009; 107(3): 516–527.
19. Ahmad A, Kong D, Wang Z, et al. Down-regulation of uPA and uPAR by 3,3'-diindolylmethane contributes to the inhibition of cell growth and migration of breast cancer cells. *J Cell Biochem*, 2009; 108(4): 916–925.
20. Li Y, Wang Z, Kong D, et al. Regulation of FOXO3a/ beta-catenin/GSK-3beta signaling by 3,3'-diindolyl-methane contributes to inhibition of cell proliferation and induction of apoptosis in prostate cancer cells. *J Biol Chem.*, 2007; 282(29): 21542–21550.
21. Gupta MK, Qin RY. Mechanism and its regulation of tumor-induced angiogenesis. *World J Gastroenterol*, 2003; 9(6): 1144–1155.
22. Bradfield CA, Bjeldanes LF. Effect of dietary indole-3-carbinol on intestinal and hepatic monooxygenase, glutathione S-transferase and epoxide hydrolase activities in the rat. *Food Chem Toxicol*, 1984; 22(12): 977–982.
23. Bradlow HL, Sepkovic DW, Telang NT, et al. Indole-3-carbinol. A novel approach to breast cancer prevention. *Ann N Y Acad Sci.*, 1995; 768: 180–200.
24. Chen I, Safe S, Bjeldanes L. Indole-3-carbinol and diindolylmethane as aryl hydrocarbon (Ah) receptor agonists and antagonists in T47D human breast cancer cells. *Biochem Pharmacol*, 1996; 51(8): 1069–1076.
25. Vang O, Jensen MB, Astrup H. Induction of cytochrome P450IA1 in rat colon and liver by indole-3-carbinol and 5, 6-benzoflavone. *Carcinogenesis*, 1990; 11(8): 1259–1263.
26. Hong C, Kim HA, Firestone GL, et al. 3,3'-Diindolyl-methane (DIM) induces a G(1) cell cycle arrest in human breast cancer cells that is accompanied by Sp1-mediated activation of p21(WAF1/CIP1) expression. *Carcinogenesis*, 2002; 23(8): 1297–1305.
27. Firestone GL, Bjeldanes LF. Indole-3-carbinol and 3,3'-diindolylmethane antiproliferative signaling pathways control cell-cycle gene transcription in human breast cancer cells by regulating promoter-Sp1 transcription factor interactions. *J Nutr.*, 2003; 133(Suppl 7): 2448S–2455S.
28. Grubbs CJ, Steele VE, Casebolt T, et al. Chemoprevention of chemically-induced mammary carcinogenesis by indole-3-carbinol. *Anticancer Res.*, 1995; 15(3): 709–116.
29. Chinni SR, Li Y, Upadhyay S, et al. Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest and apoptosis in prostate cancer cells. *Oncogene*, 2001; 20(23): 2927–2936.
30. Chen DZ, Qi M, Auburn KJ, et al. Indole-3-carbinol and diindolylmethane induce apoptosis of human cervical cancer cells and in murine HPV16-transgenic preneoplastic cervical epithelium. *J Nutr.*, 2001; 131(12): 3294–3302.
31. Dashwood RH, Fong AT, Williams DE, et al. Promotion of aflatoxin B1 carcinogenesis by the natural tumor modulator indole-3-carbinol: influence of dose, duration, and intermittent exposure on indole-3-carbinol promotional potency. *Cancer Res.*, 1991; 51(9): 2362–2365.
32. Kim DJ, Han BS, Ahn B, et al. Enhancement by indole-3-carbinol of liver and thyroid gland neoplastic development in a rat medium-term

- multiorgan carcinogenesis model. *Carcinogenesis*, 1997; 18(2): 377–381.
33. Maciejewska D, Niemyjska M, Wolska I, et al. Synthesis, spectroscopic studies and crystal structure of 5,5'- Dimethoxy-3,3'-methanediyl-bis-indole as the inhibitor of cell proliferation of human tumors. *Z Naturforsch*, 2004; 59b: 1137–1142.
34. Hannongbua S, Prasithichokekul S, Pungpo P. Conformational analysis of nevirapine, a non-nucleoside HIV-1 re-verse transcriptase inhibitor, based on quantum mechanical calculations. *J Comput Alded Mol Des.*, 2001; 15(11): 997–1004.
35. Kim DJ, Han BS, Ahn B, et al. Enhancement by indole-3-carbinol of liver and thyroid gland neoplastic development in a rat medium-term multiorgan carcinogenesis model. *Carcinogenesis*, 1997; 18(2): 377–381.
36. El Sayed MT, Ahmed KM, Mahmoud K, et al. Synthesis, cytostatic evaluation and structure activity relationships of novel bis-indolylmethanes and their corresponding tetrahydroindolocarbazoles. *Eur J Med Chem.*, 2015; 90: 845–859.
37. Mousavi SH, Moallem SA, Mehri S, et al. Improvement of cytotoxic and apoptogenic properties of crocin in cancer cell lines by its nanoliposomal form. *Pharm Biol.*, 2011; 49(10): 1039–1045.
38. Hong C, Firestone GL, Bjeldanes LF. Bcl-2 family-mediated apoptotic effects of 3,3'-diindolylmethane (DIM) in human breast cancer cells. *Biochem Pharmacol*, 2002; 63(6): 1085–1097.
39. Hong C, Kim HA, Firestone GL, et al. 3,3'-Diindolylmethane (DIM) induces a G(1) cell cycle arrest in man breast cancer cells that is accompanied by Sp1-mediated activation of p21(WAF1/CIP1) expression. *Carcinogenesis*, 2002; 23(8): 1297–1305.
40. Rahman KM, Aranha O, Glazyrin A, et al. Translocation of Bax to mitochondria induces apoptotic cell death in indole-3-carbinol (I3C) treated breast cancer cells. *Onco-gene*, 2000; 19(50): 5764–5771.
41. Rahman KM, Aranha O, Sarkar FH. Indole-3-carbinol (I3C) induces apoptosis in tumorigenic but not in nontumorigenic breast epithelial cells. *Nutr Cancer*, 2003; 45(1): 101–112.
42. Rahman KM, Li Y, Sarkar FH. Inactivation of akt and NF-kappaB play important roles during indole-3-carbinol-induced apoptosis in breast cancer cells. *Nutr Cancer*, 2004; 48(1): 84–94.
43. Rahman KW, Sarkar FH. Inhibition of nuclear translocation of nuclear factor- κ B contributes to 3, 3'- diindolylmethane-induced apoptosis in breast cancer cells. *Cancer Res.*, 2005; 65(1): 364–371.
44. Rahman KW, Li Y, Wang Z, et al. Gene expression profiling revealed survivin as a target of 3,3'-diindolyl-methane-induced cell growth inhibition and apoptosis in breast cancer cells. *Cancer Res.*, 2006; 66(9): 4952– 4960.
45. Ali S, Varghese L, Pereira L, et al. Sensitization of squa-mous cell carcinoma to cisplatin induced killing by natural agents. *Cance Lett.*, 2009; 278(2): 201–209.
46. Chen Y, Xu J, Jhala N, et al. Fas-mediated apoptosis in cholangiocarcinoma cells is enhanced by 3,3'-diindolylmethane through inhibition of AKT signaling and FLICE-like inhibitory protein. *Am J Pathol*, 2006; 169(5): 1833–1842.
47. Pappa G, Lichtenberg M, Iori R, et al. Comparison of growth inhibition profiles and mechanisms of apoptosis induction in human colon cancer cell lines by isothiocyanates and indoles from Brassicaceae. *Mutat Res.*, 2006; 599(1-2): 76–87.
48. Kim EJ, Park SY, Shin HK, et al. Activation of caspase-8 contributes to 3, 3'-Diindolylmethane-induced apoptosis in colon cancer cells. *J Nutr.*, 2007; 137(1): 31–36.
49. Frydoonfar HR, McGrath DR, Spigelman AD. Inhibition of proliferation of a colon cancer cell line by indole-3-carbinol. *Colorectal Dis.*, 2002; 4(3): 205–207.
50. Suzui M, Inamine M, Kaneshiro T, et al. Indole-3-carbinol inhibits the growth of human colon carcinoma cells but enhances the tumor multiplicity and volume of azoxymethane-induced rat colon carcinogenesis. *Int J Oncol*, 2005; 27(5): 1391–1399.
51. Savino JA 3rd, Evans JF, Rabinowitz D, et al. Multiple, disparate roles for calcium signaling in apoptosis of human prostate and cervical cancer cells exposed to diindolylmethane. *Mol Cancer Ther.*, 2006; 5(3): 556–563.
52. Stresser DM, Williams DE, Griffin DA, et al. Mechanisms of tumor modulation by indole-3-carbinol. Disposition and excretion in male Fischer 344 rats. *Drug Metab Dispos*, 1995; 23(9): 965–975.
53. Abdelrahim M, Newman K, Vanderlaag K, et al. 3, 3'-diindolylmethane (DIM) and its derivatives induce apoptosis in pancreatic cancer cells through endoplasmic reticulum stress-dependent upregulation of DR5. *Carcinogenesis*, 2006; 27(4): 717–728.
54. Banerjee S, Wang Z, Kong D, et al. 3, 3'-Diindolylmethane enhances chemosensitivity of multiple chemotherapeutic agents in pancreatic cancer. *Cancer Res.*, 2009; 69(13): 5592–5600.
55. Frydoonfar HR, McGrath DR, Spigelman AD. The effect of indole-3-carbinol and sulforaphane on a prostate cancer cell line. *ANZ J Surg*, 2003; 73(3): 154–156.
56. Savino JA 3rd, Evans JF, Rabinowitz D, et al. Multiple, disparate roles for calcium signaling in apoptosis of human prostate and cervical cancer cells exposed to diindolylmethane. *Mol Cancer Ther.*, 2006; 5(3): 556–563.
57. Chinni SR, Li Y, Upadhyay S, et al. Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest and apoptosis in prostate cancer cells. *Oncogene*, 2001; 20(23): 2927–2936.

58. Le HT, Schaldach CM, Firestone GL, et al. Plant-derived 3,3'-Diindolylmethane is a strong androgen antagonist in human prostate cancer cells. *J Biol Chem*, 2003; 278(23): 21136–21145.
59. Hahn DA. Hexamethylmelamine and pentamethylmelamine: an update. *Drug Intel Clin Pharm*, 1983; 17(6): 418–424.
60. El Sayed M. Development of novel indolyl-derived bio-logically active compounds [Ph.D]. Martin Luther Uni-versity; 2013.
61. El Sayed M. Development of novel indolyl-derived bio-logically active compounds. Saarbrücken: LAP LAMBERT Academic Publishing, 2015.
62. Wikipedia contributors. Indole alkaloid [Internet]. Wik-imedia. [cited 1 May 2015]. Available from https://en.wikipedia.org/wiki/Indole_alkaloid.
63. El Sayed M, Mahmoud K, Hilgeroth A. Glacial acetic acid as an efficient catalyst for simple synthesis of dindolylmethanes. *Curr Chem Lett.*, 2014; 3(1): 7–14.
64. El Sayed MT, Ahmed KM, Mahmoud K, et al. Synthe-sis, cytostatic evaluation and structure activity relation-ships of novel bis-indolylmethanes and their correspon-ding tetrahydroindolocarbazoles. *Eur J Med Chem.*, 2015; 90: 845–859.
65. El Sayed MT, Mahmoud K, Ahmed KM, et al. First oxi-dized tetraindoles with antimicrobial evaluation and structure activity relationship. *J Harmon Res Pharm*, 2014; 3(4): 167–176.
66. El Sayed MT, Mahmoud K, Ahmed KM, et al. Novel aspects of domino reaction of indoles with homo- phthalaldehyde and tere-phthalaldehyde. *Glob J Sci Front Res.*, 2014; 14(6): 15–22.
67. El Sayed MT, Mahmoud K, Hilgeroth A, et al. Synthesis of novel indolo-spirocyclic compounds. *J Heterocyclic Chem.*, 2015.
68. Foster BJ, Clagett-Carr K, Hoth D, et al. Pentamethyl-melamine: review of an aqueous analog of hexame-thylmelamine. *Cancer Treat Rep.*, 1986; 70(3): 383–389.
69. Ferrer S, Naughton DP, Threadgill MD. Labelled com-pounds of interest as antitumour agents–VIII. Synthesis of 2H-isotopomers of pentamethylmelamine and of a po-tential prodrug thereof. *J Label Compd Radiopharm*, 2002; 45(6): 479–484.
70. Janssen Pharmaceutica. Indole derivatives as anticancer agents. United States; 20110294846, 2011.