



**A THEMATIC REVIEW ON POTENTIAL ANTIMICROBIAL POTENTIAL
APPLICATIONS OF BACTERIOCINS FROM LACTIC ACID BACTERIA**

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ABSTRACT

The world-wide emergence of resistant pathogenic strains against the currently available antibiotics and the toxicity associated with them has compelled scientific community to increase their quests in finding other natural antimicrobial alternatives. In this regard recent discoveries on antimicrobial arsenals secreted by bacterial species against competing strains are gaining utmost importance not only because of their antimicrobial but also due to their anti-cancer potential. These natural bacterial arsenals are ribosomally synthesized low-molecular weight, heat stable, membrane active, proteolytically degradable and pore-forming, cationic peptides known as “Bacteriocins”. Owing to their anti-bacterial, anti-viral, anti-fungal, and anti-biofilm potential, bacteriocins seem to be the immediately available promising option for mitigation of AMR (antimicrobial Resistance) crisis and minimization of toxic effects of antibiotics on Host’s microbiome. The non-toxic and Generally Regarded as Safe (GRAS) status of Lactic acid bacteria (LAB) bacteriocins signifies their huge commercialization potential, and research on their biology is currently the most extensively studied subject of biotechnology. Their intrinsic food preservative and therapeutic potential can be enhanced by combinatorial approaches with other antimicrobial substances. However, their potential to be used as AGP (antimicrobial Growth Promoter) in animal production must also be evaluated. The initial half of this review focuses on the significance and biological aspects of LAB bacteriocins, while the latter half addresses the potential antimicrobial aspects of bacteriocins in various sectors.

KEYWORDS: Bacteriocins, Antimicrobial Resistance, Biopreservation.

INTRODUCTION

Since the discovery of penicillin by Alexander-Fleming in 1929 there has been continuous increase in the quests for new antimicrobial compounds from micro-organisms. Penicillin and many other antibiotics are successfully being utilized by medical and veterinary community in treatment against specific diseases caused by micro-organisms. It is a globally accepted fact, that antibiotic’s discovery has considerably improved the quality and expectancy of life. But, unfortunately antimicrobial resistance emerged a few years later since their use was started. This issue has become a global health concern. Its continuous spread among currently sensitive strains is even more alarming. In hospital settings, multidrug and even pan drug resistant strains of both gram-positive and gram-negative bacteria have been observed against the most potent and core antibiotics. Methicillin-resistant *Staphylococcus aureus* (MRSA) and Vancomycin-resistant *Enterococci* (VRE) being the most fatal ones are continuing to increase the morbidity and mortality rates (Simons et al., 2020).

One source of such unremitting increase in AR (Antibiotic Resistance) lies in widespread unrestricted utilization of antibiotics in food producing animals as

antimicrobial growth promoter. This action is potentially transmitting multidrug resistant (MDR) pathogens to humans. In the swine industry, tetracyclines, aminoglycosides, trimethoprim-sulphonamides, and ampicillin resistant enterotoxigenic *Escherichia coli* strains are found. Similarly in poultry industry resistant genes have been found in *Salmonella* serovars, *Enterococcus* spp., and *Clostridium perfringens* collected from feces and ceca of broiler chickens (Lagha et al., 2017). These findings show whenever an antibiotic is added in animal feed for production enhancement purpose it establishes a selective pressure for advent of antibiotic-resistant bacteria. Thus, the utmost need of hour is exploitation of other anti-bacterial substance of biological origin that can be safely used in animal production and against which bacteria won’t develop resistance easily and immediately.

On similar basis the requirement of natural antimicrobial substances for food preservation and safety is inevitable in food industries but the utilization of therapeutic antibiotics against food-borne pathogens and food spoilage bacteria is generally not recommended by FDA to food industries as they can induce antibiotic resistance in Food-borne pathogenic bacteria, decrease the number

of beneficial gut microbiota and can also pose many other serious public health problems (Baltork *et al.*, 2019). The alternative approach used by food industries is addition of antagonistic additives to foods that exhibit both preservative and anti-microbial properties (Chen and Hoover, 2003). Though, these synthetic chemical-preservatives prolong the shelf life of food and aid in delaying/stopping the growth of undesirable food borne microbes, but unfortunately recent findings indicate that their addition in food can cause more harm than benefit as many of them are capable of causing behavioral changes (Gultekin *et al.*, 2013) such as allergies, skin rashes, asthma, urticaria dermatitis and even carcinogenesis in consumers (Sharma, 2015). These facts are raising health concerns among public and consumers are becoming increasingly worried about the impacts of chemical preservatives in food they consume. Therefore, increasing public awareness and demand regarding naturally preserved (biopreserved) foods or foods with no chemical preservative is compelling researchers to develop a novel method of food preservation, entirely different from the traditional ones. Increased number of insurances by dairy plants to prolong shelf-life and preclude deterioration of dairy products is another reason for finding other natural antimicrobial alternatives for biopreservation of foods (Silva *et al.*, 2018).

To overcome all these issues researchers are trying to utilize another class of recently discovered antimicrobial substances i.e. bacteriocins, that cannot only act as potential natural biopreservatives without compromising the organoleptic properties of processed food but can also serve as a best substitute of conventional antibiotics in therapeutics and clinical settings. Their anticancer potential is also being emerging as a novel tool in combating cancer. The main objective of this review is to discuss the general characteristics of bacteriocins, significance of LAB bacteriocins their mode of action and potential applications in various fields. A brief overview on the safety profiles of few bacteriocins has also been presented towards the end of discussion.

GENERAL DESCRIPTION OF BACTERIOCINS

Bacteriocins are ribosomally synthesized low molecular weight extracellular peptides that exhibit bacteriostatic and bactericidal potential against members of closely related species and different strains of similar species. Most of them show narrow range of inhibitory activity. More or less all types of gram-positive and gram-negative bacteria secrete them under stress conditions to kill neighbouring cells when nutrients in environment are depleting, the producer cell remains unaffected due to its simultaneous production of immunity protein against respectively produced bacteriocin. The genes encoding bacteriocins structural proteins, immunity proteins, associated regulatory proteins, transport and modification (in case of Lantibiotics) proteins are located on different genetic determinants in the form of operon clusters. It is proposed by Riley and Gordon (1999) that the primary role of these potent AMP's is to mediate

intra-specific or population-level interactions (Riley and Gordon, 1999). These cationic, hydrophobic peptides kill sensitive strains by either forming pores or by inserting their amphipathic helical structures inside the targets cell's plasma-membrane to cause depolarization of cell membrane and efflux of essential metabolites. The consequent dissipation of PMF (Proton motive force) results in cell death of target bacteria (Da Costa *et al.*, 2019).

Bacteriocins seem to be like antibiotics in their mode of action but they cannot be termed as antibiotics as they restrict their inhibitory activities against strains of species identical with bacteriocin secreting species and more particularly against strains of producer bacterium. While therapeutic antibiotics have broader spectrum of antimicrobial activity and if they occasionally exhibit narrow inhibitory range they never obstruct proliferation of associated strains of same species. Secondly, antibiotics are secondary metabolites while bacteriocins are produced by ribosomes during primary growth phase as small peptides that are often subjected to post-translational modifications. This feature makes them easily amenable for genetic engineering applications. An additional benefit that allow bacteriocins to be used as safe food-preservatives is their biodegradability in Human gastro-intestinal tract by proteolytic digestive enzymes. This feature renders them safe for human consumption (Zacharof and Lovitt, 2012) as well as reduces the risk of pathogenic strains to come in prior contact with them as they don't persist in human bodies and environment. It is an initial point in development of antibiotic resistance and bacteriocins intrinsically stop this from occurring. Their other notable differences with conventional antibiotics include: bacterial species as only source organism; non-toxicity towards eukaryotic cells; broad diversity in terms of molecular size; structure; mode of action and number of target bacteria; high thermal stability; wide range of active pH; strong antimicrobial effect even at nano to micro molar ranges; lacking of properties like odour, colour and taste; strong antibiofilm potential; potential amenability to bio-engineering; pore-formation/inhibition of cell-wall biosynthesis being the prominent modes of action; broad spectrum of applications (not only clinical) and unlike the case of antibiotics, their inability to transfer resistant genes in other bacterial species. This poses least chances for sensitive strains to attain resistance against bacteriocins (Cleveland *et al.*, 2001; Meade *et al.*, 2020; Perez *et al.*, 2014).

BACTERIOCINS FROM LAB

Among gram positive bacteria representatives of group LAB are considered to be of particular importance as they have been used by many populations since centuries for fermenting different food products due to their natural food preserving nature and for their ability of imparting significant texture, aroma and nutritional values to food products. The discovery of novel bacteriocins from Lactic acid bacteria and their exploitation in food

industry is currently the most extensively studied subject of food biotechnology. This group comprises species of *Lactobacillus*, *Lactococcus*, *Leuconostoc*, *Pediococcus*, *Streptococcus*, *Aerococcus*, *Alloiococcus*, *Carnobacterium*, *Dolosigranulum*, *Enterococcus*, *Oenococcus*, *Tetragenococcus*, *Vagococcus* and *Weissella*. The genus *Lactobacillus* with more than 100 species makes the largest LAB group. It is abundantly present in carbohydrate rich environments (Mokoena, 2017).

Members of LAB are known for producing an array of anti-microbial compounds i.e., Organic-acids, Diacetyl, Hydrogen-Peroxide, acetoin and bacteriocins. Due to these defining roles in food safety and food preservation they have attained GRAS status and are regarded as nontoxic/safe for human ingestion. Since the bacteriocins from LAB pose no health risk concerns, they can serve as great alternative to conventional chemical preservatives in dairy products either in their purified semi-purified forms (Silva *et al.*, 2018). Furthermore, most of the bacteriocin producing strains from LAB are normal food-isolates that's why are considered preferably the best source of bacteriocin production. Hence food industry shows particularly high interest in provision of bacteriocins from Lactic acid bacteria (Deegan *et al.*, 2006).

MODE OF ACTION

The cationic nature of bacteriocins is the main factor responsible for cell death of target strains. The anionic surface components like phosphatidylethanolamine (PE), phosphatidylglycerol (PG), lipopolysaccharide (LPS), lipoteichoic acid (LTA) and cardiolipin (CL) on target bacterial cell-surface, serve as main targets for positively charged bacteriocins. The Bacteriocins assume conformations that are suitable for electrostatic binding with target cell surface. The traversing of bacteriocins through lipid bilayer occurs through association of their hydrophobic regions with host cells plasma-membrane (Kumariya *et al.*, 2019). These initial electrostatic interactions lay the foundation of subsequent events. The mechanism of pore formation is evident in cases of both lantibiotics and non-lantibiotics but the manner in which they form differs significantly. For Instance Lantibiotics are known for creating pores in "wedge-like model", whereas class II bacteriocins are known to intensify host's membrane permeability by formation of pores in "barrel stave or carpet manner" (Moll *et al.*, 1999). Above all these facts it is important to note that bacteriostatic and bactericidal potential of any bacteriocin is highly dependent on factors like physiological conditions, amount of bacteriocin being used, Indicator/Target pathogen and environmental factors (Jozala *et al.*, 2015).

Nisin is very potent class I AMP (antimicrobial Peptide) It is effective against sensitive strains even at lower concentrations. The proposed mechanism of its action is that it can stop cell-wall synthesis of sensitive bacterial

strains by binding with lipid II molecule which is the chief transporter of peptidoglycan subunits and moves from cytoplasm to the cell wall. This binding prevents cell wall biosynthesis, thus causing cell death. However, at greater concentrations, the nisin-lipid II docking-complex directs nisin to insert into host's membrane and form pores in their plasma membrane (Perez *et al.*, 2015). This shows Lipid-II acts not only as binding receptor for Nisin but also enable it to exert its renowned targeted pore forming action (Moll *et al.*, 1999). Pore formations that result due to latter case are responsible for leakage of essential ions (K^+ , H^+ Phosphate), cell contents and ATP by disturbance of PMF, pH equilibrium of target cell's plasma membrane (Deegan *et al.*, 2006). Peptidoglycan precursor lipid II acts as docking molecule not only for class I bacteriocins but also for antibiotic vancomycin, although binding-site is different there (Breukink and de Kruijff, 2006).

Like all other LAB Bacteriocins Class IIa bacteriocins are also bactericidal in their modes of action. Their well known representatives like Pediocin PA-1 (Chikindas *et al.*, 1993) Mesentericin Y105 (Maftah *et al.*, 1993) and Bavaricin MN (Kaiser and Montville, 1996) have demonstrated that sensitive bacteria can be killed by permeabilization of cell-membrane of target bacterium through formation of poration-complexes, resulting in subsequent ionic imbalance and leakage of inorganic phosphate. These all factor collectively contribute for dissipation of PMF which can further be partial or total disturbance of transmembrane potential and pH gradient. Only newly discovered Mutacidin shows complete dissipation of transmembrane potential like lantibiotics (Bennik *et al.*, 1998).

Dissimilarly with lantibiotics class IIa Bacteriocins donot seem to cause an ATP leakage from sensitive target strain might be due to smaller pores form by them. The sensitive cell shows depleted ATP concentrations because of its quicker utilization of ATP to restore its original PMF and Protecting cell from ATP exhaustion due to P efflux or due to incompetence of cell to produce ATP due to inorganic Phosphate efflux. pediocin-PA-1-induced ATP reduction and inorganic phosphate efflux were observed by Chen and Montville, 1995 and found that loss in ATP concentration is attributed to cell's ability to regenerate reduced PMF (Chen and Montville, 1995). While Nisin Z shows opposing behavior (Abee *et al.*, 1994). Similarly to lipid II, mannose-phosphotransferase systems (man-PTS) is suspected to serve as receptor molecule for some class II bacteriocins (Jozala *et al.*, 2015).

UTILIZATION OF BACTERIOCIN

With the growing population and advancement in science, people need food in terms of quality and quantity. Using bacteriocin as food preservative not only increases the shelf life of food, is also a potent source of eradicating an established infection too, as with the emerging antibiotic resistance put researchers in a stress

condition to find an alternative of antibiotics which are used for therapeutic purpose, and probiotic are found as a safe alternative for the treatment.

Besides this, Multi Drug Resistance (MDR) pathogen, the hurdle of selection of resistant bacteria to antibiotics is a growing problem of the 21st century (Kapil, 2005; Chopra et al, 1998) which is a prime concern as they weekend the ability of antibiotic arsenal to fight with them. To find the possible solution to this growing problem researchers are focusing on the usage of bacteriocin due to its high specific activity even against MDR pathogens. The diversity in the applications of bacteriocin is owing to its recognition of GRAS (Generally Regarded as Safe) substance by the US (United State) of food and FDA (Federal Drug Administration) and the European legislation for pharmaceutical and food industries uses. LAB producing bacteriocin is used as a potential 'Natural-bio preservatives' which reduce chemicals' side effect thus helping in the improvement of human health as these

peptides are active against pathogen (Moreno et al., 2006).

In the current realm of the procession, the research on bacteriocin is, however, gaining importance because of its natural defense system mechanisms ability to compete with the microorganisms in the same environment and hence its widespread potential applications in bio-nonmaterial, veterinity, dairy products (Nisin is used for this purpose which is approved GRAS Bacteriocin), crop management and solutions to problems of systematic infection, oral care, cancer, vaginal infection, skincare, and contraception. The primary function of bacteriocin is food preservation and the continuous development of newer classes of antimicrobial agents has become of emerging importance for medicine (Kumar and Schweiser, 2005; Fisher et al., 2005). Here we will discuss the applications of bacteriocin in detail, which are shown in the form of tabulation as given below in Fig. 1.

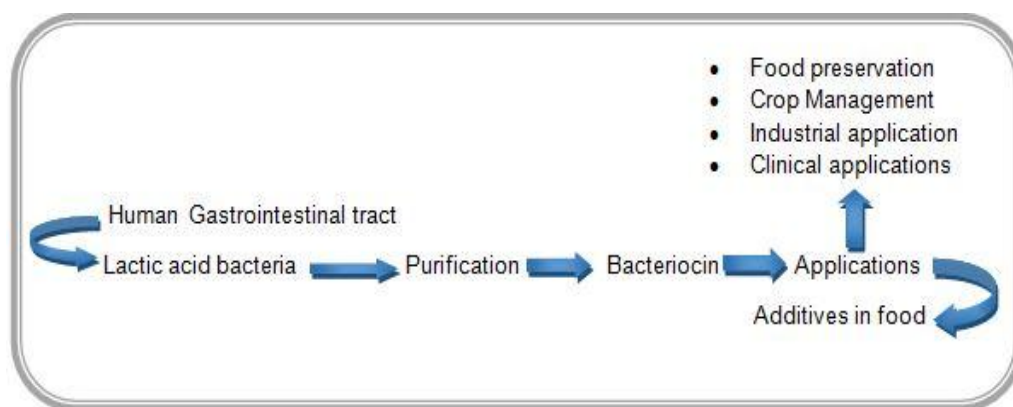


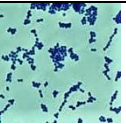


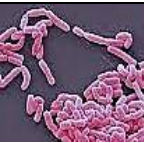


Figure 1: Overview of Applications of Bacteriocin (Evangelin et al., 2015).



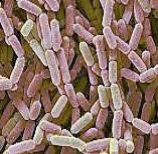


IMPORTANT BACTERIOCIN PRODUCED FROM LAB AND ITS APPLICATION

A vast array of bacteriocin has been produced by lactic Acid Bacteria (LAB) Gram positive bacteria, cationic, heat stable, permeabilizing membrane peptide, amphiphilic, and can work at lower pH; owing to their properties they are exploited for utilization in food preservation as a natural preservative and for therapeutic purpose. The most important bacteriocin 'Nisin' belongs to class-I and Acidocin, Lactacin, Lactococcin, and Pediocin belong to class-II. The bacteriocin produce from LAB can be used in both purified and crude form. Because of a non-toxic effect of LAB bacteriocin, it's been regarded as safe which highlights its importance. LAB producing bacteriocin is used as a potential 'Natural-bio preservatives' which reduce chemicals' side effect and its probiotic properties are used in treatments thus helping in the improvement of human health as these peptides are active against pathogen (Moreno et al., 2006). Therefore, the increasing interest in these compounds has encouraged the researchers for the isolation of LAB producers and the characterization of

many novel peptides (Deraz et al., 2005). In the Table 1 given below, some important bacteriocins produced from LAB are mentioned with description.

Table 1: Comprehensive comparative elaboration of different features of Bacteriocins.

Lactic Acid Bacteria (LAB)	Morphology	Micrograph	Bacteriocin	Source	Characteristics of Bacteriocin	Indicator organism	Potential Application	References
<i>Enterococcus faecium</i>	Gram positive, sometime motile, cocci spherical		Enterocin A and B	Naturally present in the GIT	Class II bacteriocin, 4,8 kDa, 47 amino-acid residues, heat-stable	<i>Listeria monocytogenes</i> , <i>Pediococcus</i> , <i>Enterococcus spp.</i> , VRE	Active against food borne pathogens and have antilisterial effect.	(Lauková and Czikková, 1999)
<i>Lactobacillus acidophilus</i> CH5	Gram positive, fermentative, spiral or straight rod or coccobacillary form		Acidocin B	Member of normal intestinal microflora	Class II bacteriocin, forms high molecular weight aggregates	Inhibiting strains of the genus <i>Lactobacillus</i> , <i>Bacillus</i> , <i>Micrococcus</i> and <i>Corynebacterium</i> Gram-positive bacteria	Used in food industry And kills food spoilage bacteria. Used as probiotic too.	(Chumchalova et al., 1995)
<i>Lactobacillus plantarum</i>	Have rounded end and rod shape, gram positive, occurring singly or in pair		Plantaricin	Fermented food products like kimchi	molecular mass of 4347.8467 Da, wide pH stability, high thermal stability, and surfactants stability	<i>Staphylococcus aureus</i> , <i>E. coli</i> , <i>Streptococcus mutants</i> etc.	Anti listerial cultures for fermented sausages and cooked ham and future application in food preservation	(Olasupo N.A., 1996)
<i>Lactobacillus casei</i>	Non-motile, gram positive rod shape, mesophilic		Lactocin 705	Normally in gut, fermented food yogurt	Class II two-component bacteriocin (33 amino-acids each component), 3,4 kDa,	<i>Listeria monocytogenes</i> , <i>Lactobacillus plantarum</i>	Used for clinical aspects	(Johansson et al., 1993)
<i>Lactobacillus sake</i>	Gram positive, facultative anaerobe, rod shape		lacticin 3147 and lacticin 481	Red meat and fish	Class I bacteriocin, 3,7 kDa, active between pH of 4,5 and 7,5	<i>Lactobacillus</i> , <i>Leuconostoc</i> , <i>Pediococcus</i>	Food industry, veterinary medicine and in treatment of human diseases	(Guinane et al., 2005).
<i>Lactococcus lactis</i> subsp. <i>Lactis</i> ATCC 11454	Spherical or ovoid cells, gram positive, non-motile		Nisin	Dairy fermented product	Class I lantibiotic, 3,5 kDa, 34 amino-acids, commercially available	<i>Listeria monocytogenes</i> , <i>Bacillus cereus</i> & <i>Staphylococcus aureus</i> .	Used in preservation of dairy products e.g. cheese, yogurt and effective against mastitis causing pathogens	(Broadbent et al., 1989)

<i>Lactococcus lactis</i>	Gram positive, ovoid or spherical, occur in pairs or short chain, gram +ve		Lacticin 3147	Milk, cheese, egg etc	Class I two-component lantibiotic, 4,2 kDa, heat-stable, active under acid and physiological pH	<i>Clostridium sp.</i> , <i>Listeria monocytogenes</i> , <i>Staphylococcus aureus</i> , <i>Enterococcus faecalis</i> , <i>Propionibacterium acne</i> , <i>Streptococcus mutans</i> ,	Antimicrobial activity against food born pathogen and food spoilage bacteria	(McAuliffe et al., 1998).
<i>Leuconostoc gelidum</i>	Gram positive, ovoid cocci, without cytochrome		Leucocin A	Isolated from plant matter, kefir etc	Class II bacteriocin, 3,9 kDa, 37 amino-acids, stable at low pH values, even after heating (100°C for 20 min)	<i>Lactobacillus</i> , <i>Enterococcus faecalis</i> , <i>Listeria monocytogenes</i> .	Control of listeria in vacuum and modified atmosphere stored meat products	(Saucier et al., 2005)
<i>Lactobacillus rhamnosus</i>	Rod shaped appear in chain, Short gram positive strain		Rhamnosin	Grape peel	1427 Da - 602.6 Da, broad range of activity and inhibit MRB	Gram-positive bacteria, <i>Listeria monocytogenes</i> , <i>Staphylococcus aureus</i>	Inhibition of mold and psychrotrophes in cottage cheese	(Makhal et al., 2015)
<i>Pediococcus acidilactici</i>	Gram positive, cocci, in pairs or tetrad form		Pediocin F	Found in plants and milk.	Class II bacteriocin, 4,5 kDa, sensitive to proteolytic enzymes.	<i>E. coli</i> Other Gram positive bacteria	Application in food, avoid food spoilage	(Schöbitz et al., 2006).
<i>Pediococcus pentosaceus</i>	Non-motile, gram positive, spherical, gram positive		Pediocin A	soy sauce mash or moromi	Class II bacteriocin, 2,7 kDa, sensitive to proteolytic enzymes and heat stable.	<i>Lactobacillus</i> , <i>Lactococcus</i> , <i>Leuconostoc</i> , <i>Pediococcus</i> , <i>Staphylococcus</i> , <i>Enterococcus</i>	Active against food borne pathogen	(Cintas et al., 1998)

ACTIVITY OF BACTERIOCIN IN ANTIMICROBIAL COATING FILMS AND NANOTECHNOLOGY

The standard of food quality is being increased over time, along with the concern to reduce waste in the environment, which encourages the research of edible coating having antimicrobial additives. The most broadly investigated biopolymers i.e. ‘Hydrocolloids’ (proteins and polysaccharides) used in edible coatings and films applied to cheese. These biopolymers are used to modify the atmosphere of surrounding of foods, and make a barrier between the food and the environment, without changing organoleptic and nutritional properties it helps to improve the safety, quality, and functionality of food products (Han, 2003; Valdés *et al.*, 2017). Using bacteriocin in coating films inhibits the growth of unwanted microorganisms like nisin/chitosan combination gives effective antilisterial action (Guo *et al.*, 2014; Muriel-Galet *et al.*, 2012) when applied on the covered surface of the food. Stability against proteolytic enzymes is provided, by immobilizing bacteriocin via covalent linkage in the packaging system (Bali *et al.*, 2014; Al-Mathkhury *et al.*, 2011).

Besides this, through nanoliposomes, nanoemulsions, nanoparticles, and nanofibers we can obtain the

bacteriocin nano-capsules which have possible applications in the food industry and medical field. Nisin, pedocin and subtilisin including in nanoemulsions have been tested in combination with carvacrol, curcumin, and cymene against *L. monocytogenes*, *E. coli*, *S. typhimurium* and *Candida lusitanae* (Zhang *et al.*, 2014; Ndoti-Nembe *et al.*, 2013). In deep wounds infected with *S. aureus* can be treated with nanofibers with ethylene oxide and poly (D, L-lactic acid) including nisin or plantaricin (Dicks *et al.*, 2011; Brand *et al.*, 2013). In dairy products, to increase the shelf life the bacteriocins and/or bacteriocin-producing LABs are incorporated in coatings and films. The food packed with coatings and films containing antimicrobial metabolites synthesized by LABs are responsible for the inhibition of pathogenic microorganisms (CaoHoang *et al.*, 2010; da Silva Malheiros *et al.*, 2010; Aguayo *et al.*, 2016) or having a viable count of LAB in the film. Without bacteriocin coating the dairy products spoiled immediately as compared with bacteriocin application on films and coatings. Bacteriocins along with antimicrobial films and coating are used in dairy products which are responsible for the inhibition of pathogenic microbes as described in the form of flow sheet diagram given below in Fig. 2.

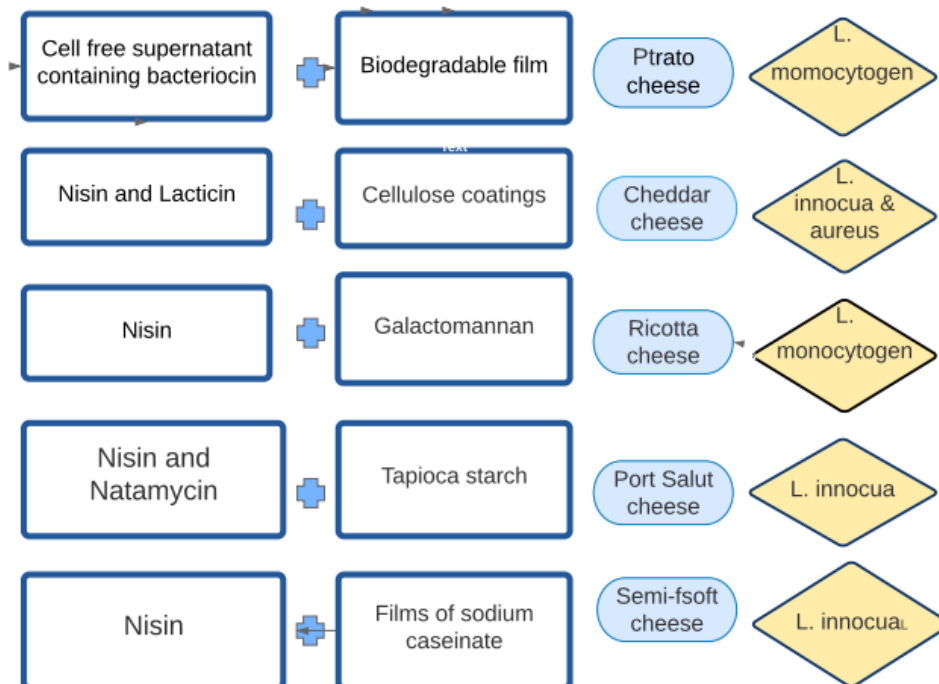


Figure 2: Combination of bacteriocins with Antimicrobial films and coating, inhibiting pathogens (Silva *et al.*, 2018).

On the other hand, via using ‘Nanotechnology’ the bacteriocin nano-capsules are utilized in food and have medical applications as well. In the Fig. 3, a combination

of nano-capsules and bacteriocins are mentioned which are responsible for inhibiting pathogens.

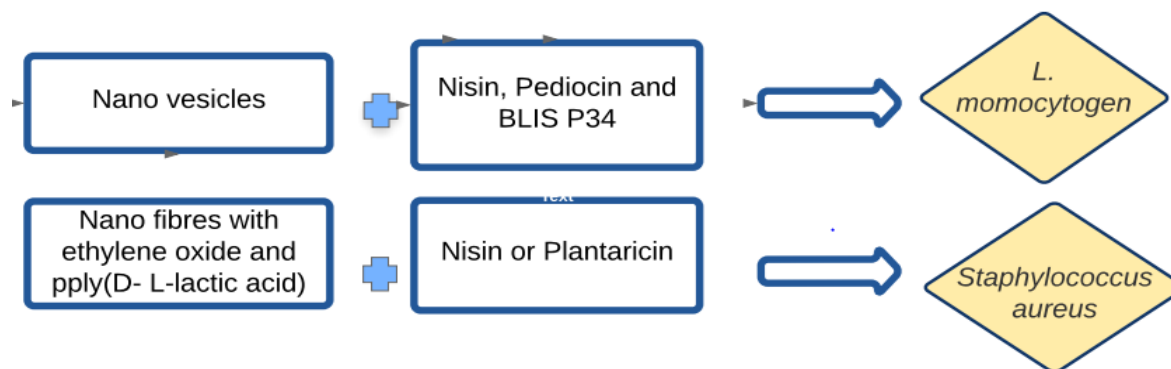


Figure 3: Combination of Nano capsules with bacteriocin, inhibiting pathogens
(Silva *et al.*, 2018).

CLINICAL APPLICATIONS

The first and foremost requirement of practical utilization of Bacteriocins is in hospital settings where appearance of Multi-drug Resistant pathogens is most evident and many nosocomial infections due to such MDR strains are becoming a global threat to human health. With the alarming condition of bacterial resistance, alternative of antibiotic was required for the betterment of human health. Number of bacteriocin is used as probiotic in replacement of antibiotic which play crucial role in boosting our defense mechanism (Immunity System). The notable resistant pathogens are *Staphylococcus aureus*, *Enterococci*, *Pneumococci*, *A.baumannii*, *Citrobacter freundii*, *E.coli*, *Klebsiella pneumoniae* and *Proteus spp.* The most important ones Vancomycin-Resistant- *Enterococci* and Multi- Drug-Resistant *Staphylococcus aureus* are recently been found to be sensitive against lactacin 3147 and Nisin A. Both stated bacteriocins are also important in inhibiting liver, spleen and kidneys-pathogens. Other infectious and resistant strains of micro-organisms have also been found to be controlled by bacteriocins NAI-107, microbisporicin and Mutacin B-Ny266 (Ahmad *et al.*, 2017).

Toxicity profile of Bacteriocins

Bacteriocins are generally considered as safe alternatives of conventional antibiotics in literature and very few studies depict their consequent side-effects. No Doubt they are promising candidates to be used as substitute for antibiotics but here we will highlight finding of few studies that were meant for predicting the therapeutic efficacy and toxicity associated with certain Bacteriocins. The highly cationic nature of bacteriocins cannot be neglected as far as their safety in animal and human is concerned as they can equally be interactive with cell-membrane's phosphatidylethanolamine (PE) component as Teichoic Acids of target-Bacterial cell-walls. PE in mammalian cells serves as substrate for inflammatory responses mediator Phosphholipase-A2. The confiscation of PE by cinnamycin and duramycin might be the cause of Immune Modulation reactions via indirect inactivation of PL-A2. Another potential

problem speculated with Binding of bacteriocins with PE is their deposition inside the cell-membrane causing change in membrane's bio-physical properties and altered ion-channels (Dicks *et al.*, 2018). Cancer cells being negatively charged are also the best target for bacteriocin preferential attack. Such verdicts have been obtained from studies on head and neck squamous-cell-carcinoma-cell lines where Nisin was found to effectively perform apoptosis and cell-cycle arrest of cancerous cells selectively without implicating any toxicity on human consumption (Dicks *et al.*, 2018).The preclinical invivo and in-vitro trial findings have showed novel-circular bacteriocin AS-48 non-toxic as no loss of cell-viability at therapeutic concentration was found (Cebrián *et al.*, 2019).

The in-vitro cytotoxicity, of Peptide-p34 was tested in different eukaryotic cells and was found to be toxic to VERO cells (Vaucher *et al.*, 2010). The crude-supernatant of AMP- LR14 from *Lactobacillus plantarum* Strain LR/14 showed delayed life cycle of fruit fly at 10mg/ml conc. While at 15 m-g/ml conc. exerted a strong larvicidal consequence. Ingestion of said peptide by fruit flies also resulted in profound loss in weight and size (Gupta *et al.*, 2014). Sometime bacreriocinogenic strain from Gram-positive group of Bacteria act as virulence Factors. A two peptide enterococcal Lantibiotic named Cytolysin has been observed for its broad spectrum of activity which it exert not only against Gram-positive bacteria but it is also lethal against eukaryotic cell-lines such as Human-Bovine, Horse Erythrocytes, Retinal-cells and also causes acute terminal outcomes in Human Infections. It is believed that its anti-eukaryotic activities might have been selectively adapted for Eukaryotic cell-predation in soil and water ecologies. Also, another non-lanthionine-containing peptide secreted by some group A streptococci named Haemolysin, and Streptolysin S, are found to cause invasive infections (Cotter *et al.*, 2005).Keeping in view these findings practical utilization of bacteriocins require accurate and safe clinical trials and approval from regulatory authorities.

Bacteriocin used as Probiotic for treatment

Lactobacilli which are a bacteriocin producing organism, present naturally in the human gut as natural micro flora can be used as a probiotic to influence the gut ecology. Hence in most stomach problems, we are advised to take yogurt for the good working of our natural micro flora to combat harmful bacteria. The gut micro-organisms provide health benefits including inactivation of carcinogenic compound's potential, activate our defense mechanism, and responsible for the reduction of serum cholesterol. Bacteriocin helps to compete with potentially pathogenic gut micro flora and also enhances the ability of colonization organisms. Hence, FAO/WHO (2002) recommended on the careful evaluation of probiotics in food. As probiotics, are live

microorganisms that, when applied in the adequate quantity, have a health benefit on the host (Sanders, 2008; Schrezenmeir and De Vrese, 2001). In the pharmaceutical shops of Pakistan most common probiotics used are GUTCARE, BIFLORA®, HIFLORA™, ECOTEC™, AMYBACT® and LAKTY^K.

Intestinal Infection

Several bacteriocin are used as a probiotic in the replacement of antibiotics which plays a vital role in boosting our defense mechanism (Immunity System) (López-Cuellar *et al.*, 2016). In the Table 2 names of some bacteriocins are mentioned which are used as probiotics as alternative of antibiotics for the treatment of Gastro Intestinal Tract (GIT) infections.

Table 2: Comparison of common antibiotics and bacteriocins for antimicrobial potential.

Antibiotic used for treatment	Alternative Bacteriocin Used As (Probiotic)	Producing strain	Bioactivity	Potential application	References
Carbapenems	Plantaricin 423	<i>Enterococcus mundtii</i>	Antimicrobial agent	GIT infections	(Kumarasamy <i>et al.</i> , 2010) (Dreyer, 2018; Van Zyl, 2018)
Cyclic lipopeptide	Piscicolin 126	<i>Carnobacterium piscicola</i>	Antimicrobial agent	GIT infections	(Coates <i>et al.</i> , 2011) (Ingham <i>et al.</i> , 2003)
Metronidazole	Pediocin PA-1	<i>Pediococcus acidilactici</i>	Antimicrobial agent	GIT infections	(Persky and Brandt, 2000) (Cintas <i>et al.</i> , 1998; Dabour <i>et al.</i> , 2009)
Penicillin	Peptide ST4SA	<i>Enterococcus mundtii</i>	Antimicrobial agent	GIT infections	(Tan and Tatsumura, 2015) (Knoetze <i>et al.</i> , 2008; Dreyer, 2018; Van Zyl, 2018)

CONCLUSION

This review highlights the antimicrobial activity of bacteriocin and its utilization in the field of both pharmaceutical and food products. Owing to abundant diversity and successful incorporation make it an ideal candidate for researchers. With the growing problems of antibiotic resistance, food shortage and increasing Multi-Drug Resistance (MDR) in the 21st century encourages the researcher to study about the natural product such that bacteriocin which shows promising applications in the food industry in term of increasing shelf life of food and preserve quality. LAB plays a vital role in the application of bacteriocin as they are generally regarded as safe (GRAS) and approved by the FDA. Bacteriocins have inhibitory action against food borne pathogens like *L. monocytogenes*, but they are not antibiotics. Bacteriocin is different from antibiotics in its synthesis and mode of action. Furthermore, unlike antibiotic resistance, bacteriocin resistance is not usually determined genetically and organisms which show resistance to antibiotics are not cross-resistant generally with bacteriocins. Therefore, bacteriocin could be considered as potential alternative to antibiotic. In clinical aspects bacteriocin used as a probiotic which show antimicrobial activity not only in bacteria but has

effective action against virus and fungal infection. The bacteriocin showcases hidden potential which motivates the scientist for new inventions and continuing the search for more novel bacteriocin with dedicated and promising applications.

REFERENCES

1. Simons A, Alhanout K, Duval, R.E. Bacteriocins, antimicrobial peptides from bacterial origin: Overview of their biology and their impact against multidrug-resistant bacteria. *Microorganisms*, 2020; 8(5): 639.
2. Ben Lagha A, Haas B, Gottschalk M, Grenier D. Antimicrobial potential of bacteriocins in poultry and swine production. *Vet. Res*, 2017; 48: 22.
3. Baltork FH, Hosseini H, Aliabadi SS, Torbati MA, Alizadeh AM, Alizadeh M, Drug Resistance and the Prevention Strategies in Food Borne Bacteria: An Update Review. *Adv Pharm Bull*, 2019; 9(3): 335-347.
4. Chen H, Hoover DG. Bacteriocins and their Food Applications. *Compr Rev Food Sci Food Saf*, 2003; 2(3): 82-100.

5. Gultekin F, Doguc DL, Vatansev H, Taysi E. The Effects of Food and Food Additives on Behaviors. *Int J Health Nutr*, 2013; 4(1): 21-32.
6. Sharma, D.S. Food Preservatives and their harmful effects. *IJSRP*, 2015; 5(4).
7. Silva CCG, Silva SPM, Ribeiro SC. Application of bacteriocins and protective cultures in dairy food preservation. *Frontiers in Microbiol*, 2018; 9: 7-11.
8. Riley M.A, Gordon D.M. The ecological role of bacteriocins in bacterial competition. *Trends Microbiol*, 1999; 7: 129–133.
9. Da Costa RJ, Voloski FLS, Mondadori RG, Duval EH, Fiorentini AM. Preservation of Meat Products with Bacteriocins Produced by Lactic Acid Bacteria Isolated from Meat. *J Food Qual*, 2019.
10. Zacharof MP, Lovitt RW. Bacteriocins Produced by Lactic Acid Bacteria A Review Article. *APCBEE Procedia*, 2012; 2: 50–56.
11. Cleveland J., Montville, T.J., Nes, I.F., Chikindas, M.L. Bacteriocins: Safe, natural antimicrobials for food preservation. *Int. J. Food Microbiol*, 2001; 71(1): 1–20.
12. Meade E., Slattery M.A., Garvey M. Bacteriocins, potent antimicrobial peptides and the fight against multi drug resistant species: Resistance is futile? *Antibiotics (Basel)*, 2020; 9(1): 32.
13. Perez RH, Zendo T, Sonomoto K. Novel bacteriocins from lactic acid bacteria (LAB): Various structures and applications. *Microb Cell Fact*, 2014; 13: 1–13.
14. Mokoena, M.P. Lactic acid bacteria and their bacteriocins: Classification, biosynthesis and applications against uropathogens: A mini-review. *Molecules*, 2017; 22(8): 1255.
15. Deegan LH, Cotter PD, Hill C, Ros P. Bacteriocins: Biological tools for biopreservation and shelf-life extension. *Int. Dairy J*, 2006; 16(9): 1058-1071.
16. Kumariya R, Garsa AK, Rajput YS, Sood SK, Akhtar N, Patel S. Bacteriocins: Classification, synthesis, mechanism of action and resistance development in food spoilage causing bacteria. *Microb Pathog*, 2019; 128: 171–177.
17. Moll GN, Konings WN, Driessen AJM. Bacteriocins: Mechanism of membrane insertion and pore formation. *Antonie van Leeuwenhoek*, 1999; 76: 185–198.
18. Jozala AF, Novaes LCL, Junior AP. Nisin. In: Bobbarala V. (ed.). *Concepts, Compounds and the Alternatives of Antibacterials*, Croatia; IntechOpen: 2015, pp. 103-119.
19. Perez R, Perez MT, Elegado F. Bacteriocins from Lactic Acid Bacteria: A Review of Biosynthesis, Mode of Action, Fermentative Production, Uses, and Prospects. *Int. J. Philipp Sci Technol*, 2015; 8(2): 61–67.
20. Breukink, E., Kruijff, BD. Lipid II as a target for antibiotics. *Nat. Rev. Drug Discov*, 2006; 5(4): 321–332.
21. Chikindas M.L., Garcia-Garcera M.J., Driessen A.J.M., Ledebouer A.M., Nissen- Meyer, J., Nes, I.F., Abee T, Konings W.N, Venema G. Pediocin PA-1, a bacteriocin from *Pediococcus acidilactici* PAC1.0, forms hydrophilic pores in the cytoplasmic membrane of target cells. *Appl. Environ. Microbiol*, 1993; 59(11): 3577–3584.
22. Maftah A, Renault D, Vignoles C, Hechard Y, Bressollier P, Ratinaud MH, Cenatiempo Y, Julien R. Membrane permeabilization of *Listeria monocytogenes* and mitochondria by the bacteriocin mesentericin Y105. *J Bacteriol*, 1993; 175(10): 3232–3235.
23. Kaiser A.L., Montville T.J. Purification of the bacteriocin bavaricin MN and characterization of its mode of action against *Listeria monocytogenes* scott A cells and lipid vesicles. *Appl. Environ. Microbiol*, 1996; 62(12): 4529–4535.
24. Bennik M.H.J., Vanloo B., Bresseur R., Gorris, L.G.M., Smid, E.J. A novel bacteriocin with a YGNGV motif from vegetable-associated *Enterococcus mundtii*: Full characterization and interaction with target organisms. *Biochim. Biophys. Acta – Biomembr*, 1998; 1373(1): 47–58.
25. Chen Y., Montville T.J. Efflux of ions and ATP depletion induced by pediocin PA-1 are concomitant with cell death in *Listeria monocytogenes* Scott A. *J. Appl. Bacteriol*, 1995; 79(6): 684–690.
26. Abee T, Rombouts FM., Hugenholtz J, Guihard G, Letellier, L. Mode of action of nisin Z against *Listeria monocytogenes* scott A grown at high and low temperatures. *Appl. Environ. Microbiol*, 1994; 60(6): 1962–1968.
27. Kapil A. The challenge of antibiotic resistance: need to contemplate. *Indian J Med*, 2005; 121(2): 83-91.
28. Chopra I, Hodgson J, Metcalf B, Poste G. The search for antimicrobial agents effective against bacteria resistant to multiple antibiotics. *Antimicrob Agents Chemoter*, 1998; 41(3): 497-503.
29. Moreno I, Lerayer ASL, Baldini VLS, de F Leitão MF. Characterization of bacteriocins produced by *Lactococcus lactis* strains. *Braz J Microbiol*, 2006; 31(3): 184-192.
30. Kumar A, Schweiser HP. Bacterial resistance to antibiotics: active efflux and reduced uptake. *Adv Drug Deliv*, 2005; 57(10): 1486-1513.
31. Fisher JF, Meroueh SO, Mobashery S. Bacterial resistance to beta-lactam antibiotics: compelling opportunism, compelling opportunity. *Chem Rev*, 2005; 105(2): 395-424.
32. Evangelin Y, Venkateswarulu TC, John Babu D, Kasturi K. Bacteriocins from Lactic Acid Bacteria and Its Potential Applications. *Int J Pharm Sci*, 2015; 32: 306-309.
33. Deraz SF, Karlsson EN, Hedstrom M, Andersson MM, Mattiasson B. Purification and characterisation of acidocin D20079, a bacteriocin produced by *Lactobacillus acidophilus* DSM 20079. *J Biotechnol*, 2005; 117(4): 43-354.
34. Lauková A, Czikková S. The use of enterocin CCM 4231 in soy milk to control the growth of *Listeria*

- monocytogenes* and *Staphylococcus aureus*. J Appl Microbiol, 1999; 87: 182-182.
35. Chumchalova J, Josephsen J, Plockova M. Characterization of acidocin CH5, a saccharolytic sensitive bacteriocin of *Lactobacillus acidophilus* CH5. Chem Microbiol Technol Lebensm, 1995; 17: 145-150.
 36. Olasupo NA. Inhibition of *Listeria monocytogenes* by plantaricin NA, an antibacterial substance from *Lactobacillus plantarum*. Folia Microbiol, 1996; 43(2): 151-155.
 37. Johansson ML, Molin G, Jeppsson B, Nobaek S, Ahnér S, Bengmark S. Administration of different *Lactobacillus* strains in fermented oatmeal soup: In vivo colonization of human intestinal mucosa and effect on the indigenous flora. Appl Environ Microbiol, 1993; 59(1): 15-20.
 38. Guinane CM, Cotter DP, Hill C, Ross PR. Microbial solutions to microbial problems; lactococcal bacteriocins for the control of undesirable biota in food. J Appl Microbiol, 2005; 98(6): 1316-1323.
 39. Broadbent JR, Chou C, Guillies K, Kondo KJ. Nisin inhibits several gram-positive, mastitis-causing pathogens. J Dairy Sci, 1989; 72(12): 3342-3345.
 40. McAuliffe O, Ryan PM, Ross PR, Hill C, Breeuwer P, Abee T. Lacticin 3147, a broad-spectrum bacteriocin which selectively dissipates the membrane potential. Appl Environ Microbiol, 1998; 64(2): 439-445.
 41. Saucier L, Champagne CP. Immobilized cell technology and meat processing. In: Nedović V, Willaert R, editors. Applications of cell immobilisation technologies. 8B. Springer; Focus on Biotech, 2005; 8: 337-353.
 42. Makhmal S, Kanawjia SK, Gigi A. Effect of microGARD on keeping quality of direct acidified Cottage cheese. J Food Sci Technol, 2015; 52(2): 936-943.
 43. Schöbitz RP, Bórquez AP, Costa EM, Ciampi RL, Brito SC. Bacteriocin like substance production by *Carnobacterium piscicola* in a continuous system with three culture broths. Study of antagonism against *Listeria monocytogenes* in vacuum packaged salmon. Braz J Microbiol, 2006; 37(1): 52-57.
 44. Cintas L, Casaus P, Fernández M, Hernández P. Comparative antimicrobial activity of enterocin L50, pediocin PA-1, nisin A and lactocin S against spoilage and foodborne pathogenic bacteria. Food Microbiol, 1998; 15(3): 289-298.
 45. Han JH. "Antimicrobial food packaging," in *Novel Food Packaging Techniques*, ed. R. Ahvenainen (Cambridge: Woodhead Publishing), 2003; 50-70.
 46. Valdés A, Ramos M, Beltrán A, Jiménez A, Garrigós MC. State of the art of antimicrobial edible coatings for food packaging applications. Coatings, 2017; 7(4): 56-59.
 47. Guo M, Jin TZ, Wang L. Antimicrobial films and coatings for inactivation of *Listeria innocua* on ready-to-eat deli turkey meat. Food Control, 2014; 40(1): 64-70.
 48. Muriel-Galet V, Lopez-Carballo G, Gavara R. Antimicrobial food packaging film based on the release of LAE from EVOH. Int J Food Microbiol, 2012; 157(2): 239-244.
 49. Bali V, Panesar PS, Bera MB. Potential of immobilization technology in bacteriocin production and antimicrobial packaging. Food Res Int, 2014; 30(6): 244-263.
 50. Al-Mathkhury HJ, Ali AS, Ghafil JA. Antagonistic effect of bacteriocin against urinary catheter associated *Pseudomonas aeruginosa* biofilm. N Am J Med Sci, 2011; 3(8): 367-370.
 51. Zhang Z, Vriesekoop F, Yuan Q. Effects of nisin on the antimicrobial activity of d-limonene and its nanoemulsion. Food Chem, 2014; 150: 307-312.
 52. Ndoti-Nembe A, Vu KD, Doucet N. Effect of combination of essential oils and bacteriocins on the efficacy of gamma radiation against *Salmonella Typhimurium* and *Listeria monocytogenes*. Int J Radiat Bio, 2013; 89(10): 794-800.
 53. Dicks LMT, Heunis TDJ, Van Staden DA. Medical and personal care applications of bacteriocins produced by *lactic acid bacteria*. In: Drider D, Rebuffat S, editors. Prokaryotic antimicrobial peptides. From genes to applications. New York (NY): Springer-Verlag, 2011; 391-421.
 54. Brand AM, Smith C, Dicks LMT. The effects of continuous in vivo administration of nisin on *Staphylococcus aureus* infection and immune response in mice. Probiotics Antimicro. Prot, 2013; 5(4): 279-286.
 55. Cao-Hoang L, Chaîne A, Grégoire L, Waché Y. Potential of nisin-incorporated sodium caseinate films to control *Listeria* in artificially contaminated cheese. Food Microbiol, 2010; 27(7): 940-944.
 56. da Silva MP, Daroit DJ, Brandelli A. Food applications of liposome-encapsulated antimicrobial peptides. Trends Food Sci Technol, 2010; 21(6): 284-292.
 57. Aguayo MDCL, Burgos MJG, Pulido RP, Gálvez A, López RL. Effect of different activated coatings containing enterocin AS-48 against *Listeria monocytogenes* on apple cubes. Innov Food Sci Emerg Technol, 2016; 35: 177-183.
 58. Ahmad V, Khan MS, Jamal QMS, Alzohairy MA, Al Karaawi MA, Siddiqui MU. Antimicrobial potential of bacteriocins: in therapy, agriculture and food preservation. Int J Antimicrob Agents, 2017; 49(1): 1-11.
 59. Dicks LMT, Dreyer L, Smith C, van Staden AD. A review: The fate of bacteriocins in the human gastrointestinal tract: Do they cross the gut-blood barrier? Front Microbiol, 2018; 9: 1-14.
 60. Cebrián R, Rodríguez-Cabezas ME, Martín-Escobano R, Rubiño S, Garrido-Barros M, Montalbán-López M, Rosales MJ, Sánchez-Moreno M, Valdivia E, Martínez-Bueno M, Marín C, Gálvez J, Maqueda M. Preclinical studies of toxicity and safety of the AS-48 bacteriocin. J Adv Res, 2019; 20: 129-139.

61. Vaucher RA, Teixeira ML, Brandelli A. Investigation of the cytotoxicity of antimicrobial peptide P40 on eukaryotic cells. *Curr Microbiol*, 2010; 60(1): 1–5.
62. Gupta R, Sarkar S, Srivastava S. In vivo Toxicity Assessment of Antimicrobial Peptides (AMPs LR14) Derived from *Lactobacillus plantarum* Strain LR/14 in *Drosophila melanogaster*. *Probiotics Antimicrob Proteins*, 2014; 61(1): 59–67.
63. Cotter PD, Hill C, Ross PR. Bacteriocins: developing innate immunity for food. *Nat Rev Microbiol*, 2005; 3(10): 777–788.
64. Sanders ME. Probiotics: Definition, Sources, Selection, and Uses. *Clin Infect Disea*, 2008; 46(2): 58-61.
65. Schrezenmeir J, De Vrese, M. Probiotics, prebiotics, and symbiotics – approaching a definition. *Am J Clin Nutr*, 2001; 73(2): 361-364.
66. López-Cuellar MR, Rodríguez-Hernández AI, Chavarría-Hernández N. LAB bacteriocin applications in the last decade. *Biotechnol & Biotechnol Equip*, 2016; 1-12.
67. Kumarasamy KK, Toleman MA, Walsh TR, Bagaria J, Butt F, Balakrishnan R. Emergence of a new antibiotic resistance mechanism in India, Pakistan, and the UK: a molecular, biological and epidemiological study. *Lancet Infect Dis*, 2010; 10(9): 597-602.
68. Dreyer L. The Ability of Antimicrobial Peptides to Migrate Across the Gastrointestinal Epithelial and Vascular Endothelial Barriers. M.Sc. thesis, Stellenbosch University, Stellenbosch, 2018.
69. Van Zyl W. Gastrointestinal Persistence of the Probiotic Bacteria *Lactobacillus plantarum* 423 and *Enterococcus mundtii* ST4SA, and Their Antilisterial Activity. Ph.D. Dissertation, Stellenbosch University, Stellenbosch, 2018.
70. Coates ARM, Halls G, Hu Y. Novel classes of antibiotics or more of the same? *Br J Pharmacol*, 2011; 163: 184-194.
71. Ingham A, Ford M, Moore RJ, Tizard M. The bacteriocin piscicolin 126 retains antilisterial activity in vivo. *J Antimicrob Chemother*, 2003; 51: 1365-1371.
72. Persky SE, Brandt LJ. Treatment of recurrent *Clostridium difficile*-associated diarrhea by administration of donated stool directly through a colonoscopy. *Am J Gastroenterol*, 2000; 95(1): 3283-3285.
73. Dabour N, Zihler A, Kheadr E, Lacroix C, Fliss I. In vivo study on the effectiveness of pediocin PA-1 and *Pediococcus acidilactici* UL5 at inhibiting *Listeria monocytogenes*. *Int J Food Microbiol*, 2009; 133(3): 225-233.
74. Tan SY, Tatsumura Y. Alexander Fleming (1881-1955): discoverer of penicillin. *Singapore Med J*, 2015; 56(7): 366-367.
75. Knoetze H, Todorov SD, Dicks LMT. A class IIa peptide from *Enterococcus mundtii* inhibits bacteria associated with otitis media. *Int J Antimicrob Agents*, 2008; 31(3): 228-234.