

DEVELOPMENT AND VALIDATION OF ANALYTICAL RP-HPLC METHOD FOR THE ESTIMATION OF AZELNIDIPINE IN BULK AND MARKETED PHARMACEUTICAL DOSAGE FORM**Aedirepally Sreeja Reddy^{1*}, Vijaya Kuchana², Pasupuleti Sunitha³ and G. Kalyani⁴**¹Department of Pharmaceutical Analysis, Teegala Krishna Reddy College of Pharmacy, Medbowli, Meerpet (V), Balapur (M), Ranga Reddy (Dist), Hyderabad – 500097, Telangana.²Department of Pharmaceutical Chemistry, Principal and Professor, Teegala Krishna Reddy College of Pharmacy, Medbowli, Meerpet (V), Balapur (M), Ranga Reddy (Dist), Hyderabad – 500097, Telangana.³Department of Pharmaceutical Analysis, Associate Professor, Teegala Krishna Reddy College of Pharmacy, Medbowli, Meerpet (V), Balapur (M), Ranga Reddy (Dist), Hyderabad – 500097, Telangana.⁴Department of Pharmaceutical Chemistry, Associate Professor, Teegala Krishna Reddy College of Pharmacy, Medbowli, Meerpet (V), Balapur (M), Ranga Reddy (Dist), Hyderabad – 500097, Telangana.***Corresponding Author: Aedirepally Sreeja Reddy**

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ABSTRACT

A simple, rapid, specific and accurate reverse phase high performance liquid chromatographic method has been developed for the validated of Azelnidipine in bulk as well as in marketed pharmaceutical dosage form. This separation was performed on a Symmetry ODS C18 (4.6×250mm, 5µm) column with Methanol: Phosphate Buffer (35:65) v/v as mobile phase at a flow rate of 1.0 mL min⁻¹ with UV detection at 235 nm; the constant column temperature was Ambient. The run time under these chromatographic conditions was less than 8 min. The retention time of Azelnidipine was found to be 2.276min. The calibration plot was linear over the concentration range of 6–14 µg mL⁻¹ with limits of detection and quantification values of 1.2 and 3.6 ng mL⁻¹ respectively. The mean % assay of marketed formulation was found to be 99.86%, and % recovery was observed in the range of 98-102%. Relative standard deviation for the precision study was found <2%. The developed method is simple, precise, specific, accurate and rapid, making it suitable for estimation of Azelnidipine in bulk and marketed pharmaceutical dosage form.

KEYWORDS: Azelnidipine, RP-HPLC, Validation, ICH Guidelines.**INTRODUCTION**

Azelnidipine is a dihydropyridine calcium channel blocker. It is marketed by Daiichi-Sankyo pharmaceuticals, Inc. in Japan. It has a gradual onset of action and produces a long-lasting decrease in blood pressure, with only a small increase in heart rate, unlike some other calcium channel blockers.^[1] It is currently being studied for post-ischemic stroke management. Azelnidipine is a vasodilator that induces a gradual decrease in blood pressure in hypertensive patients. Unlike other members of its drug class, Azelnidipine does not induce reflex tachycardia due to vasodilation. This is likely due to the fact that it elicits a gradual fall in blood pressure. Azelnidipine is a vasodilator that induces a gradual decrease in blood pressure in hypertensive patients. Unlike other members of its drug class, Azelnidipine does not induce reflex tachycardia due to vasodilation.^[2] This is likely due to the fact that it elicits a gradual fall in blood pressure. It also exhibits a

prolonged hypotensive effect and has been shown to have a strong anti-arteriosclerotic action in vessels due to its high affinity for vascular tissue and antioxidative activity. Clinical studies have demonstrated that Azelnidipine markedly reduced heart rate and proteinuria in hypertensive patients by inhibiting sympathetic nerve activity.^[3] Azelnidipine has also been confirmed to have cardio-protective, neuroprotective, and anti-atherosclerotic properties, and has also been found to prevent insulin resistance. The IUPAC name of Azelnidipine is 3-O-(1-benzhydrylazetid-3-yl) 5-O-propan-2-yl 2-amino-6-methyl-4-(3-nitro phenyl)-1, 4-dihydro pyridine-3, 5-dicarboxylate.^[4] The Chemical Structure of Azelnidipine is shown in figure-1.

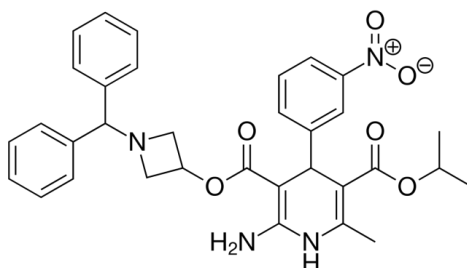


Fig. 1: Chemical Structure of Azelnidipine.

EXPERIMENTAL

Table 1: Instruments Used.

| S.No. | Instruments and Glass wares | Model |
|-------|-----------------------------|--|
| 1 | HPLC | WATERS Alliance 2695 separation module, Software: Empower 2, 996 PDA detector. |
| 2 | pH meter | Lab India |
| 3 | Weighing machine | Sartorius |
| 4 | Volumetric flasks | Borosil |
| 5 | Pipettes and Burettes | Borosil |
| 6 | Beakers | Borosil |
| 7 | Digital ultra sonicator | Labman |

Table 2: Chemicals Used.

| S.No. | Chemical | Brand Names |
|-------|-----------------------------|--------------------|
| 1 | Azelnidipine | Local Market |
| 2 | Water and Methanol for HPLC | LICHROSOLV (MERCK) |
| 3 | Acetonitrile for HPLC | Merck |

HPLC METHOD DEVELOPMENT

Preparation of Standard Solution

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7ml of Methanol and sonicate to dissolve and removal of air completely and make volume up to the mark with the same Methanol.

Further pipette 0.1ml of the above Azelnidipine stock solutions into a 10ml volumetric flask and dilute up to the mark with Methanol.

Procedure

Inject the samples by changing the chromatographic conditions and record the chromatograms, note the conditions of proper peak elution for performing validation parameters as per ICH guidelines.

Mobile Phase Optimization

Initially the mobile phase^[5] tried was Methanol and Methanol: Water with varying proportions. Finally, the mobile phase was optimized to Methanol: Phosphate Buffer in proportion 35:65% v/v.

Optimization of Column

The method was performed with various C18 columns like, X- bridge column, Xterra, and C18 column. Symmetry ODS C18 (4.6 x 250mm, 5 μ m) was found to be ideal as it gave good peak shape and resolution^[6] at 1ml/min flow.

PREPARATION OF BUFFER AND MOBILE PHASE

Preparation of Potassium dihydrogen Phosphate (KH₂PO₄) buffer (pH-3.6)

Dissolve 6.8043 of potassium dihydrogen phosphate in 1000 ml HPLC water and adjust the pH 3.6 with diluted orthophosphoric acid. Filter and sonicate the solution by vacuum filtration^[7] and ultra-sonication.

Preparation of Mobile Phase

Accurately measured 350 ml (35%) of Methanol, 650 ml of Phosphate buffer (65%) were mixed and degassed in digital ultra sonicator for 15 minutes and then filtered through 0.45 μ filter under vacuum filtration.

Diluent Preparation

The Mobile phase was used as the diluent.^[8]

Method Validation Parameters

System Suitability

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Azelnidipine stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

Procedure

The standard solution was injected for five times and measured the area for all five injections in HPLC.^[9] The %RSD for the area of five replicate injections was found to be within the specified limits.

Specificity**Preparation of Standard Solution**

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flask add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Azelnidipine stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Preparation of Sample Solution

Weight 10 mg equivalent weight of Azelnidipine sample into a 10mL clean dry volumetric flask and add about 7mL of Diluent and sonicate to dissolve it completely and make volume up to the mark with the same solvent. Further pipette 0.1ml of Azelnidipine above stock solution into a 10ml volumetric flask and dilute up to the mark with diluent.

Procedure

Inject the three replicate injections of standard and sample solutions and calculate the assay^[10-12] by using formula:

%ASSAY =

$$\frac{\text{Sample area}}{\text{Standard area}} \times \frac{\text{Weight of standard}}{\text{Dilution of standard}} \times \frac{\text{Dilution of sample}}{\text{Weight of sample}} \times \frac{\text{Purity}}{100} \times \frac{\text{Weight of tablet}}{\text{Label claim}} \times 100$$

Linearity and Range

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flask add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Preparation of Level – I (6ppm of Azelnidipine)

Take 0.6ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – II (8ppm of Azelnidipine)

Take 0.8ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – III (10ppm of Azelnidipine)

Take 0.1ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – IV (12ppm of Azelnidipine)

Take 0.12ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – V (14ppm of Azelnidipine)

Take 0.14ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Procedure

Inject each level into the chromatographic system^[13] and measure the peak area.

Plot a graph of peak area versus concentration (on X-axis concentration and on Y-axis Peak area) and calculate the correlation coefficient.^[14]

PRECISION**Repeatability****Preparation of Azelnidipine Product Solution for Precision**

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flask add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Azelnidipine stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Procedure

The standard solution was injected for six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits.

Intermediate Precision

To evaluate the intermediate precision^[15] (also known as Ruggedness) of the method, Precision was performed on different days by maintaining same conditions.

Procedure**Analyst 1**

The standard solution was injected for six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits.

Analyst 2

The standard solution was injected for six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits.^[16]

Accuracy**For Preparation of 50% Standard Stock Solution**

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flask add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.05ml of the above Azelnidipine stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

For Preparation of 100% Standard Stock Solution

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Azelnidipine stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

For Preparation of 150% Standard Stock Solution

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.15ml of the above Azelnidipine stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

Procedure

Inject the Three replicate injections of individual concentrations (50%, 100%, 150%) were made under the optimized conditions.^[17] Recorded the chromatograms and measured the peak responses. Calculate the Amount found and Amount added for Azelnidipine and calculate the individual recovery and mean recovery values.

Limit of Detection and Limit of Quantification (LOD & LOQ)

Preparation of 0.95µg/ml Solution (For LOD)

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution).

Further pipette 0.0095ml of the above Azelnidipine stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Preparation of 2.9µg/ml Solution (For LOQ)

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

RESULTS AND DISCUSSION

Optimization of Analytical Method

Optimized Chromatographic Conditions

| | | |
|--------------------|---|--|
| Mobile phase ratio | : | Methanol: Phosphate Buffer (35:65) V/V |
| Column | : | Symmetry ODS C18 (4.6×250mm, 5µm) |
| Column temperature | : | Ambient |

Further pipette 0.029ml of the above Azelnidipine stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Robustness

The analysis was performed in different conditions to find the variability of test results.^[18] The following conditions are checked for variation of results.

For Preparation of Standard Solution

Accurately weigh and transfer 10 mg of Azelnidipine working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Azelnidipine stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

Effect of Variation of Flow Conditions

The sample was analyzed at 0.9 ml/min and 1.1 ml/min instead of 1ml/min, remaining conditions are same. 10µl of the above sample was injected and chromatograms were recorded.

Effect of Variation of Mobile Phase Organic Composition

The sample was analyzed by variation of mobile phase i.e. Methanol: Phosphate Buffer was taken in the ratio and 40:60, 30:70 instead (35:65), remaining conditions^[19] are same. 10µl of the above sample was injected and chromatograms were recorded.

| | | |
|------------------|---|---------|
| Wavelength | : | 235nm |
| Flow rate | : | 1ml/min |
| Injection volume | : | 10µl |
| Run time | : | 8min |

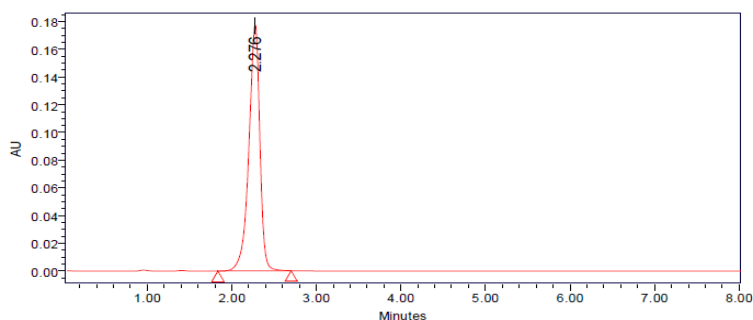


Fig. 2: Optimized Chromatographic Condition.

Method Validation

Method validation^[20] is the process used to confirm that the analytical procedure employed for a specific test is suitable for its intended use. Results from method

validation can be used to judge the quality, reliability and consistency of analytical results; it is an integral part of any good analytical practice.

System Suitability

Table 3: Results of System Suitability for Azelnidipine.

| S.No. | Peak Name | RT | Area (µV*sec) | Height (µV) | USP Plate Count | USP Tailing |
|-----------|--------------|-------|---------------|-------------|-----------------|-------------|
| 1 | Azelnidipine | 2.277 | 1652847 | 185647 | 6589 | 1.24 |
| 2 | Azelnidipine | 2.277 | 1653658 | 186254 | 6587 | 1.26 |
| 3 | Azelnidipine | 2.267 | 1654521 | 185475 | 6584 | 1.28 |
| 4 | Azelnidipine | 2.265 | 1653564 | 186594 | 6582 | 1.29 |
| 5 | Azelnidipine | 2.277 | 1658745 | 185684 | 6895 | 1.24 |
| Mean | | | 1654667 | | | |
| Std. Dev. | | | 2355.764 | | | |
| % RSD | | | 0.142371 | | | |

Specificity

The ICH documents define specificity as the ability to assess unequivocally the analyte in the presence of components that may be expected to be present, such as impurities, degradation products, and matrix components.

Analytical method was tested for specificity²¹ to measure accurately quantitates Azelnidipine in drug product.

%ASSAY =

$$\frac{\text{Sample area}}{\text{Standard area}} \times \frac{\text{Weight of standard}}{\text{Dilution of standard}} \times \frac{\text{Dilution of sample}}{\text{Weight of sample}} \times \frac{\text{Purity}}{100} \times \frac{\text{Weight of tablet}}{\text{Label claim}} \times 100 = 99.40\%$$

The % purity of Azelnidipine in pharmaceutical dosage form was found to be 99.40%.

Linearity

Table 4: Data for Linearity of Azelnidipine.

| Concentration µg/ml | Average Peak Area |
|---------------------|-------------------|
| 6 | 1078475 |
| 8 | 1461129 |
| 10 | 1808358 |
| 12 | 2211573 |
| 14 | 2593778 |

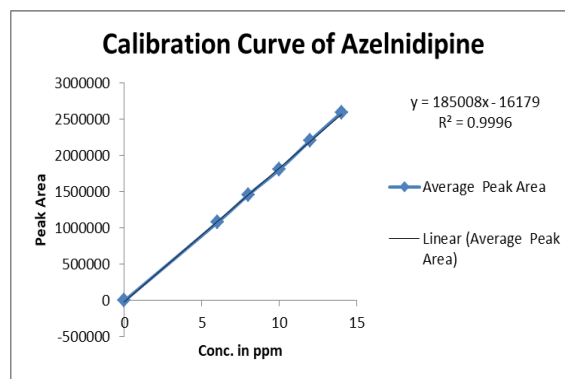


Fig. 3: Linearity Curve of Azelnidipine.

Linearity Plot

The plot of Concentration (x) versus the Average Peak Area (y) data of Azelnidipine is a straight line.

$$Y = mx + c$$

Slope (m) = 18500

Intercept (c) = 16179

Correlation Coefficient (r) = 0.999

Validation Criteria: The response linearity²² is verified if the Correlation Coefficient is 0.99 or greater.

Conclusion: Correlation Coefficient (r) is 0.99, and the intercept is 0.16179. These values meet the validation criteria.

Precision

The precision^[23] of an analytical procedure expresses the closeness of agreement (degree of scatter) between a series of measurements obtained from multiple sampling

of the same homogeneous sample under the prescribed conditions.

Repeatability

Obtained Six (6) replicates of 100% accuracy solution as per experimental conditions. Recorded the peak areas and calculated % RSD.

Table 5: Results of Repeatability for Azelnidipine.

| S. No | Peak Name | Retention time | Area ($\mu\text{V}\cdot\text{sec}$) | Height (μV) | USP Plate Count | USP Tailing |
|-----------------|--------------|----------------|---------------------------------------|--------------------------|-----------------|-------------|
| 1 | Azelnidipine | 2.293 | 1658954 | 186958 | 1.26 | 6785 |
| 2 | Azelnidipine | 2.276 | 1658745 | 187548 | 1.27 | 6854 |
| 3 | Azelnidipine | 2.286 | 1659865 | 189854 | 1.26 | 6852 |
| 4 | Azelnidipine | 2.277 | 1653254 | 186985 | 1.25 | 6784 |
| 5 | Azelnidipine | 2.280 | 1654781 | 189542 | 1.24 | 6895 |
| 6 | Azelnidipine | 2.293 | 1661324 | 187586 | 1.28 | 6965 |
| Mean | | | 1657821 | | | |
| Std. Dev | | | 3120.433 | | | |
| %RSD | | | 0.188225 | | | |

Intermediate Precision

Analyst-1

Table 6: Results of Intermediate Precision for Azelnidipine.

| S.No. | Peak Name | RT | Area ($\mu\text{V}\cdot\text{sec}$) | Height (μV) | USP Plate Count | USP Tailing |
|------------------|--------------|-------|---------------------------------------|--------------------------|-----------------|-------------|
| 1 | Azelnidipine | 2.274 | 1678541 | 186589 | 6587 | 1.26 |
| 2 | Azelnidipine | 2.258 | 1685985 | 186598 | 6321 | 1.26 |
| 3 | Azelnidipine | 2.267 | 1685745 | 186985 | 6385 | 1.25 |
| 4 | Azelnidipine | 2.270 | 1685987 | 187854 | 6580 | 1.26 |
| 5 | Azelnidipine | 2.264 | 1698526 | 187549 | 6721 | 1.27 |
| 6 | Azelnidipine | 2.265 | 1685943 | 186598 | 6637 | 1.26 |
| Mean | | | 1686788 | | | |
| Std. Dev. | | | 6463.466 | | | |
| % RSD | | | 0.383182 | | | |

Analyst 2

Table 7: Results of Intermediate Precision Analyst 2 for Azelnidipine.

| S.No. | Peak Name | RT | Area ($\mu\text{V}\cdot\text{sec}$) | Height (μV) | USPP late count | USP Tailing |
|------------------|--------------|-------|---------------------------------------|--------------------------|-----------------|-------------|
| 1 | Azelnidipine | 2.277 | 1665847 | 167481 | 6854 | 1.25 |
| 2 | Azelnidipine | 2.255 | 1658989 | 167854 | 6785 | 1.26 |
| 3 | Azelnidipine | 2.265 | 1659845 | 167895 | 6854 | 1.24 |
| 4 | Azelnidipine | 2.255 | 1665964 | 167854 | 6895 | 1.26 |
| 5 | Azelnidipine | 2.253 | 1659863 | 168585 | 6459 | 1.25 |
| 6 | Azelnidipine | 2.252 | 1665986 | 167859 | 6456 | 1.26 |
| Mean | | | 1662749 | | | |
| Std. Dev. | | | 3501.766 | | | |
| % RSD | | | 0.210601 | | | |

Accuracy: Accuracy at different concentrations (50%, 100%, and 150%) was prepared and the % recovery²⁴ was calculated.

Table 8: The Accuracy Results for Azelnidipine.

| % Concentration (at specification Level) | Area | Amount Added (ppm) | Amount Found (ppm) | % Recovery | Mean Recovery |
|--|----------|--------------------|--------------------|------------|---------------|
| 50% | 109068.3 | 5 | 5.021 | 100.420% | 100.72% |
| 100% | 202187 | 10 | 10.054 | 100.540% | |
| 150% | 297032.3 | 15 | 15.181 | 101.206% | |

Limit of Detection

The detection limit^[25] of an individual analytical procedure is the lowest amount of analyte in a sample which can be detected but not necessarily quantitated as an exact value.

$$\text{LOD} = 3.3 \times \sigma / s$$

Where

σ = Standard deviation of the response

S = Slope of the calibration curve

Result

= 0.95 μ g/ml

Quantitation Limit

The quantitation limit^[26] of an individual analytical procedure is the lowest amount of analyte in a sample which can be quantitatively determined.

$$\text{LOQ} = 10 \times \sigma / S$$

Where

σ = Standard deviation of the response

S = Slope of the calibration curve

Table 10: Results for Robustness.

| Parameter Used for Sample Analysis | Peak Area | Retention Time | Theoretical Plates | Tailing Factor |
|------------------------------------|-----------|----------------|--------------------|----------------|
| Actual Flow rate of 1.0 mL/min | 1658242 | 2.312 | 6569 | 1.24 |
| Less Flow rate of 0.9 mL/min | 1854215 | 2.458 | 6865 | 1.35 |
| More Flow rate of 1.1 mL/min | 1758468 | 2.032 | 6254 | 1.32 |

Stability Studies

The specificity of the method can be demonstrated by applying stress conditions using acid, alkaline, peroxide, thermal, UV, water degradations. The sample was

Result

= 2.9 μ g/ml

Table 9: Results of LOD and LOQ.

| | |
|-----|-------------|
| LOD | 0.95925344 |
| LOQ | 2.906828605 |

SE of Intercept = Excel Function (Data Analysis \rightarrow Regression)

SD of Intercept = SE of Intercept * \sqrt{N}

LOD = 3.3 * (SD of Intercept/Slope)

LOQ = 10 * (SD of Intercept/Slope)

Robustness: The robustness was performed for the flow rate variations from 0.9 ml/min to 1.1ml/min and mobile phase ratio variation from more organic phase to less organic phase ratio for Azelnidipine. The method is robust^[27] only in less flow condition. The standard of Azelnidipine was injected by changing the conditions of chromatography. There was no significant change in the parameters like resolution, tailing factor, asymmetric factor, and plate count.

exposed to these conditions the main peak of the drug was studied for peak purity that indicating the method effectively separated the degradation products^[28] from the pure active ingredient.

Table 11: Results of Forced Degradation Studies for Azelnidipine.

| S.No. | Stress Condition | Peak Area | % of Degraded Amount | % of Active Amount | Total % of Amount |
|-------|------------------|------------|----------------------|--------------------|-------------------|
| 1 | Standard | 1658242 | 0 | 100% | 100% |
| 2 | Acidic | 1331734.15 | 19.69 | 80.31 | 100% |
| 3 | Basic | 1594233.85 | 3.86 | 96.14 | 100% |
| 4 | Oxidative | 1394747.34 | 15.89 | 84.11 | 100% |
| 5 | Thermal | 1575827.37 | 4.97 | 95.03 | 100% |
| 6 | Photolytic | 1345331.73 | 18.87 | 81.13 | 100% |
| 7 | Water | 1360090.08 | 17.98 | 82.02 | 100% |

SUMMARY AND CONCLUSION

The analytical method was developed by studying different parameters. First of all, maximum absorbance was found to be at 235nm and the peak purity was excellent. Injection volume was selected to be 10 μ l which gave a good peak area. The column used for study was Symmetry ODS C₁₈ (4.6 \times 250mm, 5 μ m) because it was giving good peak. Ambient temperature was found to be suitable for the nature of drug solution. The flow rate was fixed at 1.0ml/min because of good peak area and satisfactory retention time. Mobile phase is Methanol: Phosphate Buffer pH-3.6 in the ratio of 35:65 v/v was fixed due to good symmetrical peak. So this mobile phase was used for the proposed study. Methanol

was selected because of maximum extraction sonication time was fixed to be 10min at which all the drug particles were completely soluble and showed good recovery. Run time was selected to be 8min because analyze gave peak around 2.276 and also to reduce the total run time. The percent recovery was found to be 98.0-102 was linear and precise over the same range. Both system and method precision was found to be accurate and well within range. The analytical method was found linearity over the range of 6-14ppm of the Azelnidipine target concentration. The analytical passed both robustness and ruggedness tests. On both cases, relative standard deviation was well satisfactory.

BIBLIOGRAPHY

1. <https://go.drugbank.com/drugs/DB09230>
2. <https://pubchem.ncbi.nlm.nih.gov/compound/Azelnidipine>
3. <https://en.wikipedia.org/wiki/Azelnidipine>
4. <https://medicaldialogues.in/generics/azelnidipine-2722518>
5. Dr. Kealey and P.J Haines, Analytical Chemistry, 1st edition, Bios Publisher, 2002; P1-7.
6. A. Braith Wait and F. J. Smith, Chromatographic Methods, 5th edition, Kluwer Academic Publisher, 1996; 1-2.
7. Andrea Weston and Phyllisr. Brown, HPLC Principle and Practice, 1st edition, Academic press, 1997; 24-37.
8. Yuri Kazakevich and Rosario Lobrutto, HPLC for Pharmaceutical Scientists, 1st edition, Wiley Interscience A John Wiley & Sons, Inc., Publication, 2007; 15-23.
9. Chromatography, (online). URL:<http://en.wikipedia.org/wiki/Chromatography>.
10. Meyer V.R. Practical High-Performance Liquid Chromatography, 4th Ed. England, John Wiley & Sons Ltd, 2004; 7-8.
11. Sahajwalla CG a new drug development, Vol 141, Marcel Dekker Inc., New York, 2004; 421-426.
12. Introduction to Column. (Online), URL:http://amitpatel745.topcities.com/index_files/study/column_care.pdf
13. Detectors used in HPLC (online) URL:http://wiki.answers.com/Q/What_detectors_are_used_in_HPLC
14. Detectors (online), URL:http://hplc.chem.shu.edu/NEW/HPLC_Book/Detectors/det_uvda.html
15. Detectors (online), URL:http://www.dionex.com/enus/webdocs/64842-31644-02_PDA-100.pdf
16. Detectors (online), URL:<http://www.ncbi.nlm.nih.gov/pubmed/8867705>
17. Detectors (online), URL:<http://www.chem.agilent.com/Library/applications/59643559.pdf>
18. Detectors (online), URL:<http://hplc.chem.shu.edu/new/hplcbook/detector>
19. Draft ICH Guidelines on Validation of Analytical Procedures Definitions and terminology. Federal Register, vol 60. IFPMA, Switzerland, 1995; 1126.
20. Code Q2B, Validation of Analytical Procedures; Methodology. ICH Harmonized Tripartite Guidelines, Geneva, Switzerland, 1996; 1- 8.
21. Introduction to analytical method validation (online), available from: URL: <http://www.standardbase.hu/tech/HPLC%20validation%20PE.pdf>.
22. Data elements required for assay validation, (online) available from: URL: <http://www.labcompliance.com/tutorial/methods/default.asp>.
23. Snyder LR practical HPLC method development, 2nd edition. John Wiley and sons, New York, 1997; 180-182.
24. Skoog D A, West D M, Holler FJ: Introduction of analytical chemistry. Sounder college of publishing, Harcourt Brace college publishers, 1994; 1-5.
25. Sharma B K, Instrumental method of chemical analysis Meerut, 1999; 175-203.
26. Breaux J and Jones K: Understanding and implementing efficient analytical method development and validation. Journal of Pharmaceutical Technology, 2003; 5: 110-114.
27. Willard, H. y. Merritt L.L, Dean J.A and Settle F.A "Instrumental Methods of Analysis" 7th edition CBS publisher and distributors, New Delhi, 1991; 436-439.
28. ICH Q2A, "Validation of Analytical Methods, Definitions and Terminology", ICH Harmonized tripartite guideline, 1999.
29. Dr. Sushil D. Patil^{1*}, Dr. Rishikesh S Bachhav², Dr. Pavan B. Udavant³, Dr. Sapana P. Ahirrao⁴, Dr. Deepak S. bhambere⁴, Development and Validation of Stability Indicating RP-HPLC Method for Azelnidipine for bulk drug, Nat. Volatiles & Essent. Oils, 2021; 8(5): 11151-11157.
30. Megha G. Gore * and Pratap S. Dabhade, RP-HPLC Method Development and Validation of Azelnidipine, Gore and Dabhade, IJPSR, 2016; 7(12): 5111-5114.
31. D. Prabhakar^{1*}, J. Sreekanth², K.N. Jayaveera³, Method Development and Validation of Azelnidipine by RP-HPLC, International Journal of ChemTech Research, 2017; 10(10): 418-423.
32. Sneha Ubale*, Dr. M.S.Kalshetti, Bhavana Habib, Jyoti Mittha, Sagar Adlinge, Development and Validation of RP-HPLC Method for Quantification of Azelnidipine in Tablet, International Journal of Creative Research Thoughts (IJCRT), © 2021 IJCRT, July, 2021; 9(7): f797-f802.
33. *Snehal D. Jadhav, Prachi B. Lokhande, Vaibhav L. Narwade, Mahananda V. Ghodke, Rutuja S. Desai and Prerna R. Mote, Method Development & Validation of Stability Indicating Rp-Hplc Method for Simultaneous Estimation for Azelnidipine & Telmisartan in Bulk & Pharmaceutical Dosage Form, World Journal of Pharmaceutical and Medical Research, wjpmr, 2022; 8(3): 216-222.