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Ammonia volatilization from urea at different levels of zeolite

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Ammonia (NH₃) loss due to hydrolysis of urea can be substantial. When applied to the soil, up to 50% NH₃ volatilizes. This decreases urea-N use efficiency. The objective of this study was to minimize ammonia loss from urea using different levels of clinoptilolite zeolite. Six treatments were evaluated; (1) soil alone, (2) 2.60 g granular urea, (3) 2.60 g liquid urea, (4) 2.60 g liquid urea + 4 g of zeolite, (5) 2.60 g liquid urea + 8 g of zeolite and (6) 2.60 g liquid urea + 12 g of zeolite. The incubation study was conducted using close-dynamic air flow system method to measure the amount of NH₃ released. At the end of this study, soils samples were analyzed for pH, exchangeable ammonium (NH₄⁺) and available nitrate (NO₃⁻) using standard procedures. When urea was applied in liquid form, NH₃ loss was reduced when compared with granular urea, but not significantly different among the treatments with and without zeolite inclusion. Increase in soil pH was observed upon addition of higher amount of zeolite when compared with urea alone. High NH₄⁺ content and low NH₃ loss for T5 directly showed that high cation exchange capacity (CEC) of the amended soil due to inclusion of zeolite was responsible for retention of more NH₄⁺ in the soil even though the soil pH was higher when compared with T1 and T2. Liquid urea mixed with zeolite reduced NH₃ loss and increased NH₄⁺ retention in the soil.

Key words: Ammonia volatilization, granular urea, liquid urea, zeolite.

INTRODUCTION

When urea is applied to soil, urea hydrolysis occurs because urea's enzyme breaks down urea molecule into NH_4^+ (Mengel and Kirkby, 1987) resulting in up to 50% loss of urea in the form of NH_3 (Sommer et al., 2004). This loss is associated with application method, soil factors and environmental conditions (Sharpley et al., 1998). Many approaches and additives have been used by researchers to reduce NH_3 loss from urea (Voss, 1984; Gould et al., 1986).

Combination of acidic and high CEC materials showed reduction in NH₃ loss (Ahmed et al., 2006, 2008, 2010) because urea hydrolysis is reduced when pH of soil is less than 5.5 (Fan and Mackenzie, 1993) and retention of NH_4^+ by the cation- exchange sites. Application of

clinoptilolite zeolite has been effective in reducing NH_3 volatilization in alkaline, coarse-textured soil (Lewis et al., 1984). This depends on the amounts of zeolite used (Ahmed et al., 2008). This is because of its high CEC and great affinity for NH_4^+ . The small channels of zeolites have been reported to reduce nitrification by microbes and protect NH_4^+ physically (Ming and Dixon, 1986; Ferguson and Pepper, 1987).

When liquid urea is used, NH₃ loss may be reduced, because it penetrates into the soil layer. For instance, NH₃ loss decreased when urea was used in fertigation (Papadopoulos, 2000). Furthermore, NH₃ loss from broadcasting urea could be reduced if the amount of rainfall is sufficient to disperse urea and the reaction product indirectly reduce alkaline condition to soil (Harper et al., 1983). Therefore, this study was carried out to minimize ammonia loss from urea using different levels of clinoptilolite zeolite.

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MATERIALS AND METHODS

The soil used in this experiment was Tipik Tualemkuts (Bekenu, Series), collected from an undisturbed area at Universiti Putra Malaysia, Bintulu Sarawak Campus, Malaysia. Samples were collected at 0 to 20 cm soil depth, air dried, meshed and sieved to pass a 2 mm sieve. The zeolite used was in powder form.

The soil and zeolite pH were determined in a 1:2.5 soil:distilled water suspension and 1 M potassium chloride (KCI) using a glass electrode (Peech, 1965). Soil texture was determined using the hydrometer method (Tan, 2005). Soil CEC was determined by the leaching method with 1 N ammonium acetate followed with steam distillation technique (Bremner, 1965) and CEC of clinoptilolite zeolite was determined using the CsCI method (Ming and Dixon, 1986). Exchangeable K, Ca and Mg were extracted using the leaching method (Cottenie, 1980) and determined by atomic absorption spectrometry (AAS). Inorganic N (NH₄⁺ and NO₃⁻) was determined using the method of Keeney and Nelson (1982). Total N of the soil was determined by the method described by Bremner (1965).

Treatments evaluated per 250 g of soil were:

(i) T0 = Soil alone

(ii) T1 = 2.60 g granular urea (iii) T2 = 2.60 g liquid urea (iv) T3 = 2.60 g liquid urea + 4 g of zeolite (v) T4 = 2.60 g liquid urea + 8 g of zeolite (vi) T5 = 2.60 g liquid urea + 12 g of zeolite

The amount of zeolite used was because previous study showed reduction of NH_4^+ , leaching was 66% by adding 8 g of natural zeolite kg⁻¹ of sandy soil (Zwingmann et al., 2009). The amounts of urea used were calculated from the N fertilizer recommendations for pepper (Sadanandan, 2000). Soil (250 g) was well mixed with different levels of zeolite and was wetted with distilled water for granular and for liquid fertilizer using mixture of urea and distilled water to get the 60% of soil field capacity. For granular urea treatment, urea was applied at the surface of the soil. Daily ammonia loss from the soil was measured using a close-dynamic air flow system method until the loss was reduced to 1% of urea added (Siva et al., 1999; Ahmed et al., 2006).

The incubation chamber was kept at room temperature and air passed through the chambers at a rate of 3.5 L/min. This rate of air flow, corresponding to 8.5 volume exchanges per minute and it was maintained throughout the incubation period using a Gilmont flow meter (Gilmont Instrument, Great Neck, New York) to measure and adjust the air flow.

At the end of this study, soils samples were analyzed for pH, NH_4^+ and NO_3^- contents. Treatments were arranged in complete randomized design (CRD) with three replications. Analysis of variance (ANOVA) was used to test for treatment effects while means were compared using Duncan's test using Statistical Analysis System (SAS Version 9.2).

RESULTS AND DISCUSSION

The selected physical and chemical characteristics of the soil used in this study are presented in Table 1. The soil texture was sandy clay loam. The soil was acidic and total N was consistent with that reported by Paramananthan (2000). The pH of zeolite was high as expected.

All the treatments with urea (Figure 1) showed NH_3 loss except for soil alone (T0). Volatilization loss rapidly

 Table 1. Selected chemical and physical characteristics of soil and zeolite.

| Property | Soil | Zeolite |
|--|-------|---------|
| pH _{Water} | 4.11 | 7.90 |
| PH _{KCI} | 3.53 | 6.64 |
| Exchangeable K ⁺ (cmoL kg ⁻¹) | 8.20 | 21.53 |
| Exchangeable Ca ²⁺ (cmoL kg ⁻¹) | 1.92 | 39.47 |
| Exchangeable Mg ²⁺ (cmoL kg ⁻¹) | 2.33 | 26.30 |
| Bulk density (g cm ⁻³) | 1.09 | nd |
| CEC (cmoL kg ⁻¹) | 7.13 | 100.20 |
| Total nitrogen (%) | 0.06 | nd |
| Clay (%) | 26.35 | nd |
| Sand (%) | 46.47 | nd |
| Silt (%) | 27.18 | nd |

CEC, cation exchange capacity; nd, not determined.

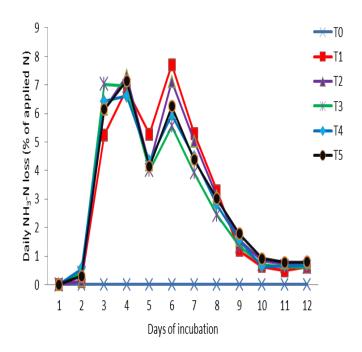


Figure 1. Daily losses of ammonia from urea fertilizer over 12 days of incubation.

occurred in liquid urea treatments (T2, T3, T4 and T5) and the maximum NH_3 loss occurred at 3 and 4 days after incubation while that of T1 occurred at day 6. This was because the dissolved urea reacted with soil water to form NH_4^+ . As soil surface rapidly dried due to air velocity in the chamber, NH_3 from urea treatment decreased at day 5 of incubation. Ammonia loss decreased when soil water was insufficient for the necessary chemical reactions and it increased at day 6 after the second addition of water. Similar results have been reported by Raison et al. (1978) and Cabrera et al. (2005), where NH_3

 Table 2. Total amounts of ammonia loss and soil pH after 12 days of incubation.

| Treatment | NH₃ loss (% of urea-N) | рН |
|-----------|------------------------|--------------------|
| Т0 | 0.00 ^c | 3.88 ^e |
| T1 | 37.48 ^a | 7.18 ^{cd} |
| T2 | 32.15 ^b | 7.12 ^d |
| Т3 | 33.65 ^b | 7.22 ^c |
| T4 | 34.66 ^b | 7.30 ^b |
| T5 | 33.20 ^b | 7.40 ^a |

Different alphabets indicate significant difference between means using Duncan' test at $p \le 0.05$.

 Table 3. Effect of treatments on soil exchangeable ammonium and nitrate accumulation after 12 days of incubation.

| Treatment | Exchangeable NH₄ ⁺ -N (ppm) | Available NO₃ ⁻ -N (ppm) |
|-----------|---|--|
| Т0 | 67.72 ^d | 30.36 ^b |
| T1 | 905.98 ^c | 42.03 ^a |
| T2 | 997.05 ^b | 46.70 ^a |
| Т3 | 1032.07 ^b | 39.70 ^a |
| T4 | 1048.42 ^b | 39.70 ^a |
| T5 | 1144.15 ^a | 42.03 ^a |

Different alphabets indicate significant difference between means using Duncan' test at $p \le 0.05$.

loss increased by rain or re-wetting of the soil after several days of application because of an increase in the rate of urea hydrolysis. After the maximum NH₃ loss as previously discussed, NH₃ loss declined until day 12 of incubation where the loss was 1% of nitrogen added as urea. Treatments T2, T3, T4 and T5 significantly reduced NH₃ loss when compared with T1 (Table 2). When applied with water, urea penetrates into the soil layer and held by the soil colloids consequently lowering the air velocity when compared with surface application. During urea infiltration, hydrolysis occurs and converts urea into NH_4^+ after application (Nkrumah et al., 1989). Hence, hydrolysis of urea not only occurs at the surface but in the soil profile and directly reduces NH₃ loss. This observation was consistent with the result reported by Sigunga et al. (2002) and Nyord et al. (2008) where reduction of NH₃ loss in the soil was affected by the depth of soil injected with fertilizer.

After 12 days of incubation, soil pH (Table 2) increased with increasing level of zeolite (T3 to T5) when compared with liquid urea alone (T2). Application of zeolite created more soil alkalinity and this was possible because of the high pH of the zeolite added. Increase in soil pH upon application of zeolite has been previously reported (Lin et al., 1998). Higher accumulation of NH₄⁺ after 12 days of urea application was found for treatment T5, while the lowest was recorded for T1 (Table 3). There was no

significant difference in NH4⁺ content for liquid urea only (T2) and addition of zeolites (T3 and T4) except for T5 where 12 g of zeolite was added. Application of zeolites to the soil increased the amount of NH4⁺ and reduced NH₃ loss when compared with urea without additive due to the exchange sites of zeolite which can retain NH4⁺ produced from urea during hydrolysis (Ahmed et al., 2008; Latifah et al., 2010). High NH_4^+ and low NH_3 loss for T5 was because the CEC of the zeolite favoured formation of NH₄⁺. In related studies, Ahmed et al. (2008) and Latifah et al. (2010) reported that mixing triple superphosphate (TSP) or Sago waste water with zeolite increased accumulation of NH_4^+ over NH_3 in the soil. The authors showed that application of liquid urea combined with zeolite had no significant effect on NO3 when compared with granular urea. This was because high NH₃ loss due to soil pH exceeding soil buffer capacity for T1 and zeolite has good selectivity for NH4⁺ (Pabalan and Bertetti, 2001) than formation of NO₃ in the soil.

Conclusion

Ammonia volatilization from urea can be reduced by mixing urea with zeolite. The exchanger site from zeolite was able to retain ammonium ion from urea as compared to the formation of ammonia as well as increasing the accumulation of ammonium ion.

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REFERENCES

- Ahmed OH, Aminuddin H, Husni MHA (2006). Reducing ammonia loss from urea and improving soil-exchangeable ammonium retention through mixing triple superphosphate, humic acid and zeolite. Soil Use Manage, 22: 315-319.
- Ahmed OH, Aminuddin H, Husni MHA (2008). Ammonia volatilization and ammonium accumulation from urea mixed with zeolite and triple superphosphate. Acta Agric. Scand. Sect. B Soil Plant Sci., 58: 182-186.
- Ahmed OH, Braine CH, Nik Muhamad AM (2010). Minimizing ammonia loss from urea through mixing with zeolite and acid sulphate soil. Intl. J. Phy. Sci., 5(14): 2198-2202.
- Bremner JM (1965). Total nitrogen. In, Black et al. (eds) Method of soil analysis, Part 2, American Society of Agronomy, Madison, Wisconcin, pp. 1149-1178.
- Cabrera ML, Kissel DE, Vaio N, Craig JR, Rema JA, Morris LA (2005). Loblolly pine needles retain urea fertilizer that can be lost as ammonia. Soil Sci. Soc. Am. J., 69: 1525-1531.
- Cottenie A (1980). Soil testing and plant testing as a basis of fertilizer recommendation. FAO Soils Bull. 38: 70-73.
- Fan MX, Mackenzie AF (1993). Urea and phosphate interactions in fertilizer microsites: Ammonia volatilization and pH changes. Soil Sci. Soc. Am. J., 57: 839-845.
- Ferguson GA, Pepper G (1987). Ammonium retention in sand amended with clinoptillolite. Soil Sci. Soc. Am. J., 51: 231-234.

- Gould WD, Hagedorn C, McCready RGL (1986). Urea transformations and fertilizer efficiency in soil. Adv. Agron., 40: 209-238.
- Harper LA, Catchpoole VR, Davis R, Weir KL (1983). Ammonia volatilization: Soils, plant, and microclimate effects on diurnal and seasonal fluctuations. Agron. J., 75: 212-218.
- Keeney DR, Nelson DW (1982). Nitrogen-inorganic Forms. In, Page et al. (eds) Methods of Soil Analysis, Part 2, (2nd ed), Agron. Monograph, 9. ASA and SSSA, Madison, Wisconsin.
- Latifah O, Ahmed OH, Nik Muhamad AM (2010). Minimizing ammonia volatilization in waterlogged soils through mixing of urea with zeolite and sago waste water. Intl. J. Phy. Sci., 5(14): 2193-2197.
- Lewis MD, Moore FDI, Goldsberry KL (1984). Ammonium-exchanged clinoptilolite and granulated clinoptilolite with urea as nitrogen fertilizers. In, Pond WG, Mumpton FA (eds) Zeo-Agriculture: Use Natural Zeolites in Agriculture and Aquaculture, Westview Press, Boulder, CO.
- Lin CF, Lo SS, Lin HY, Lee Y (1998). Stabilization of cadminum contaminated soils using synthesized zeolite. J. Hazard. Mater., 60: 217-226.
- Mengel K, Kirkby EA (1987). Principles of Plant Nutrition (4th ed). International Potash Institute, Bern, Switzerland, p. 642.
- Ming DW, Dixon JB (1986). Clinoptilolite in South Texas soils. Soil Sci. Soc. Am. J., 50: 1618-1622.
- Nkrumah M, Griffith SM, Ahmad N (1989). Lysimeter and field studies on ¹⁵N in a tropical soil. II. Transformation of (NH₂)₂CO-¹⁵N in a tropical loam in lysimeter and field plots. Plant Soil. 114: 13-18.
- Nyord T, Sogaard HT, Hansen MN, Jensen LS (2008). Injection methods to reduced ammonia emission from volatile liquid fertilisers applied to growing crops. Biosyst. Eng., 100: 235-244.
- Pabalan RT, Bertetti FP (2001). Cation-exchange properties of natural zeolites. In, Bish D, Ming DW (eds) Natural Zeolites: Occurrence, Properties, Applications. Reviews in Mineralogy Geochem. Mineralogical Soci. Am., pp. 453-518.
- Papadopoulos I (2000). Fertigation: Present and future prospects. In, Ryan J (eds) Nutrient Management under Pressurized Irrigation Systems in the Mediterranean Region: Proceedings of the IMPHOS International Fertigation Workshop organized by the World Phosphate Institute, Amman, Jordan. ICARDA, Aleppo, Syria, pp. 232-245.
- Paramananthan S (2000). Soils of Malaysia: Their characteristics and identification (Volume 1). Acad. Sci. Malaysia, Kuala Lumpur, Malaysia, p. 616.

- Peech HM (1965). Hydrogen-ion activity. In, Black et al. (eds) Methods of soil analysis part 2, American Society of Agronomy, Madison, Wisconsin, pp. 914-926.
- Raison RJ, McGarity JW (1978). Effect of plant ash on nitrogen fertilizer transformations and ammonia volatilization. Soil Sci. Soc. Am. J., 42: 140-143.
- Sadanandan AK (2000). Agronomy and Nutrition of Black Pepper. In, Ravindran PN (eds) Black Pepper, Hardwood Academic Publishers, pp. 163-223.
- Sharpley AN, Meisinger JJ, Breeuwsma A, Sims JT, Daniel TC, Schepers JS (1998). Impacts of animal manure management on ground and surface water quality. In, Hatfield JL (eds) Animal waste utilization: Effective use of manure as a soil resource, Ann Arbor Press, Chelsea, MI, pp. 173-242.
- Sigunga DO, Janssen BH, Oenema O (2002). Ammonia volatilization from vertisols. Eur. J. Soil Sci., 53(2): 195-202.
- Siva KB, Aminuddin H, Husni MHA, Manas AR (1999). Ammonia volatilization from urea as affected by tropical-based palm oil palm effluent (pome) and peat. Comm. Soil Sci. Plant Anal., 30: 785-804.
- Sommer SG, Schjoerring JK, Denmead OT (2004). Ammonia emission from mineral fertilizers and fertilized crops. Adv. Agron., 82: 557-622.
- Tan KH (2005). Soil sampling, preparation and analysis (2nd ed). Boca Raton, Florida, USA, p. 623.
- Voss RD (1984). Potential for used of urease inhibitors. In, Hauck RD (eds) Nitrogen in Crop Production, American Society of Agronomy, Madison, pp. 571-577.
- Zwingmann N, Balbir S, Ian DRM, Robert JG (2009). Zeolite from alkali modified kaolin increases NH₄⁺ retention by sandy soil: Column experiments. Appl. Clay Sci., 46: 7-12.