

# A Look at *Column Choices*

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Columns and Supplies  
Technical Support

# Overview

## What to consider when choosing a column?

- Column chemistry
  - Silica surface
  - Bonded phase
  - End-capping
- Particle size options
  - Totally porous
  - Superficially porous
- Method conditions
- Column lifetime



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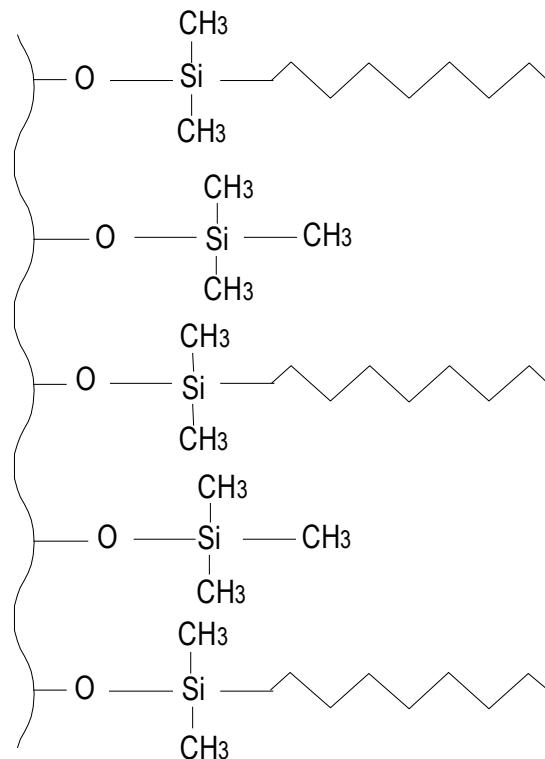
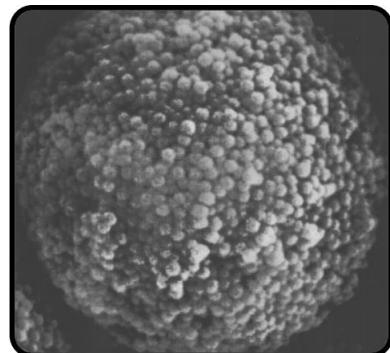
# Silica Column Characteristics

## Surface area

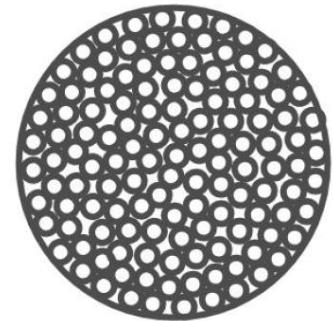
- Pore size
- Particle size

## Surface chemistry

- Bonding
- Silanols



# Pore Size



## Small molecules

- 80 – 120 Å
- Maximizes loading and retention

## Peptides, proteins, other large biomolecules

- 120 Å (Peptides)
- 300 Å to 450 Å (or higher)
- Maintain high efficiency

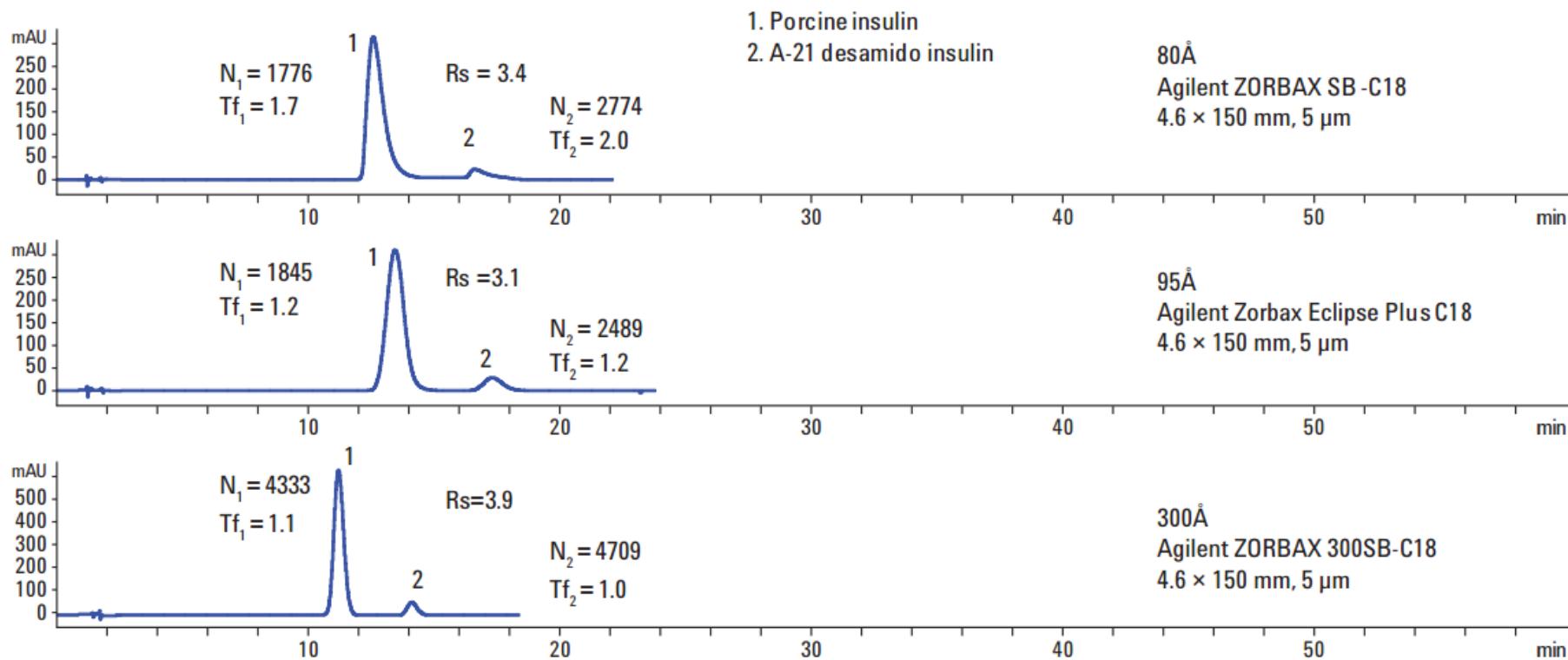


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# Pore Size



Chromatograms on 4.6 × 150 mm, 5 µm columns with different pore size.

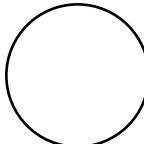
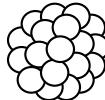


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# Particle Technologies

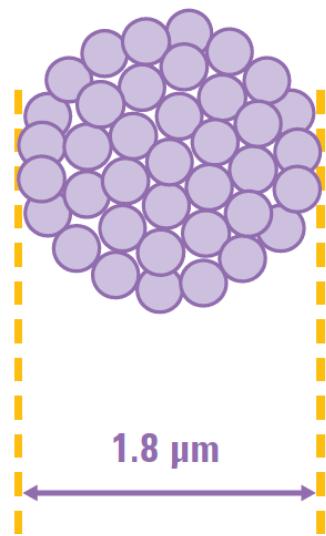
Year(s) of Acceptance	Particle Size	Most Popular Nominal Size	Plates / 15cm
1950's		100µm	200
1967		57µm (pellicular)	1,000
1972		10µm	6,000
1985		5µm	12,000
1992		3.5µm	22,000
2003		$\leq 2\mu\text{m}$	>30,000
2007/2008		2.7 µm	32,000



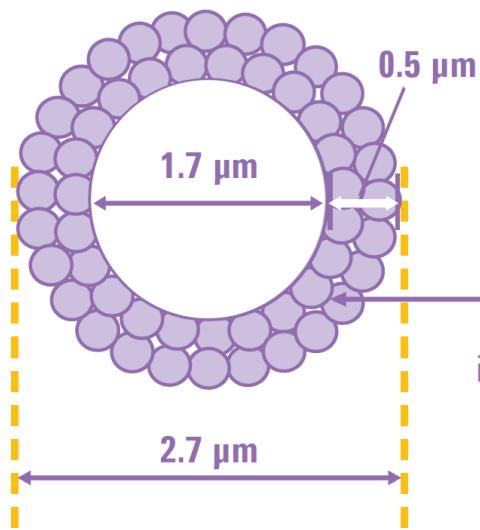
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# Particle Technologies

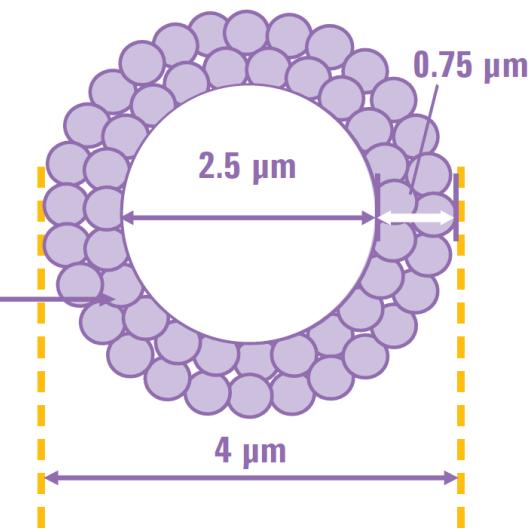
1.8  $\mu\text{m}$  totally porous



2.7 and 4  $\mu\text{m}$  superficially porous



Pore size  
120 $\text{\AA}$ ,  
ideal for small  
molecules

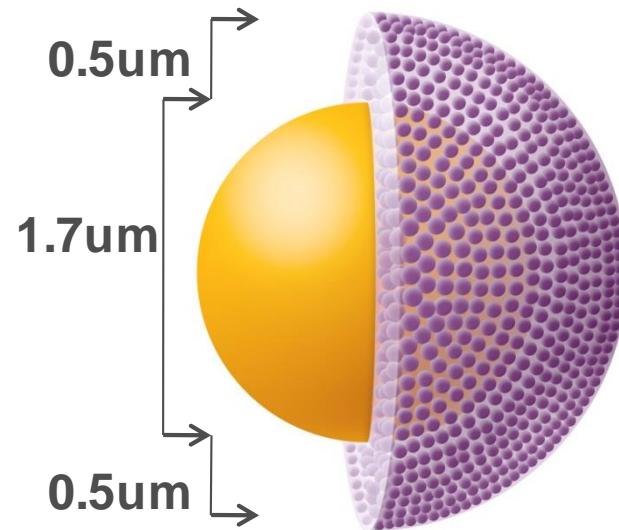


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# Superficially Porous Column Technologies

## Poroshell 120 columns:

- Efficiency  $\approx$  90% of sub-2  $\mu\text{m}$
- Pressure  $\approx$  40-50% of sub-2  $\mu\text{m}$
- N  $\approx$  2X 3.5  $\mu\text{m}$  (totally porous)
- $d_p = 2.7 \mu\text{m}$
- 2  $\mu\text{m}$  frit to reduce clogging
- $P_{\text{limit}} = 600$  bar for HPLC or UHPLC
- Particles
  - 1.7  $\mu\text{m}$  solid core
  - 0.5  $\mu\text{m}$  diffusion path
  - 2.7  $\mu\text{m}$  total diameter



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# Comparing Efficiency and Pressure with Different Types of Columns

Particle Size/Type	Pressure	Efficiency	LC Compatibility
5 µm Totally Porous	80 bar	5,000	All 400 bar instruments
3.5 µm Totally Porous	123 bar	7,800	All 400 bar instruments
2.7 µm Poroshell 120	180 bar	12,000	All LCs/UHPLCs (up to 600 bar)
1.8 µm Totally Porous	285 bar	12,500	All LCs/UHPLCs (up to 1200 bar)

Columns: 4.6 x 50mm, Mobile Phase: 60% ACN:40% Water Flow Rate: 2 mL/min

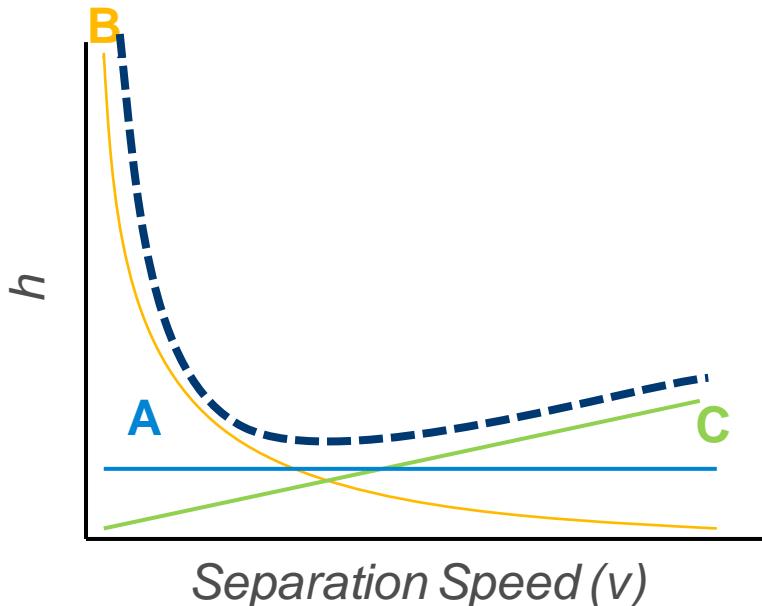


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# Efficiency Improvement with Superficially Porous Particles

van Deemter equation:

$$h = A + B/\nu + C \cdot \nu$$



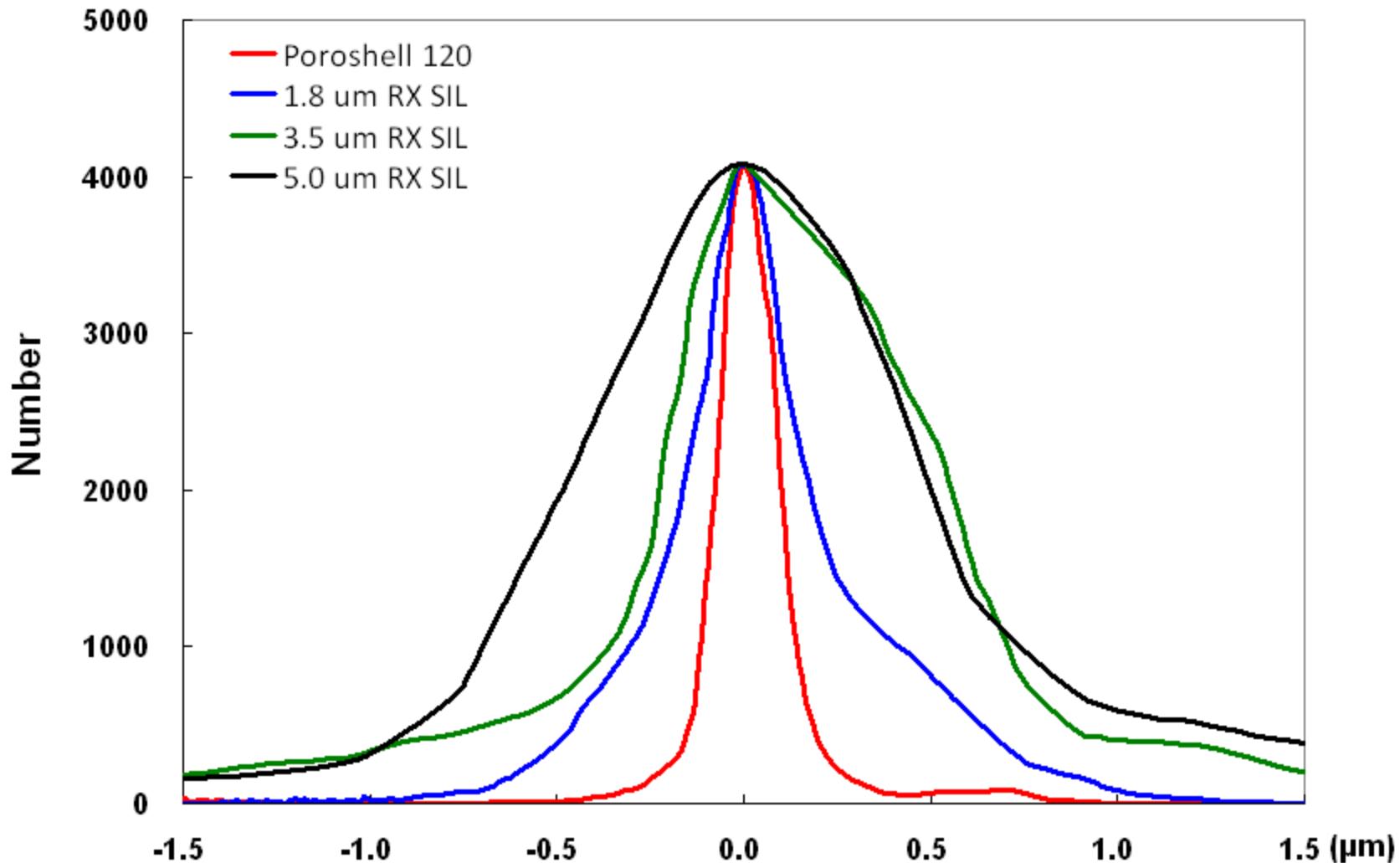
Lower  $h$  = higher efficiency!

- **A term – eddy diffusion and flow distribution**
  - particle size & packing quality important
  - narrow particle size distribution
- **B term – longitudinal diffusion**
  - Slightly lower due to longitudinal diffusion reduction
- **C term – mass transfer**
  - shorter diffusion paths
  - better with superficially porous particles
  - more effect on large molecules



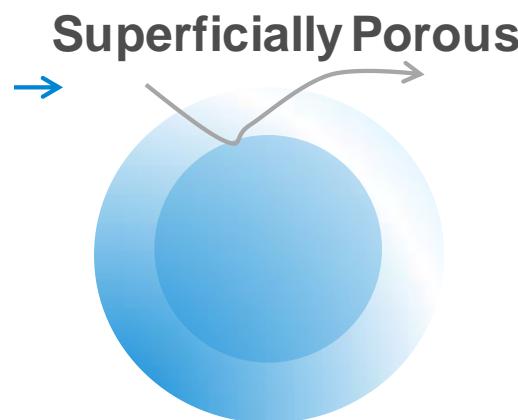
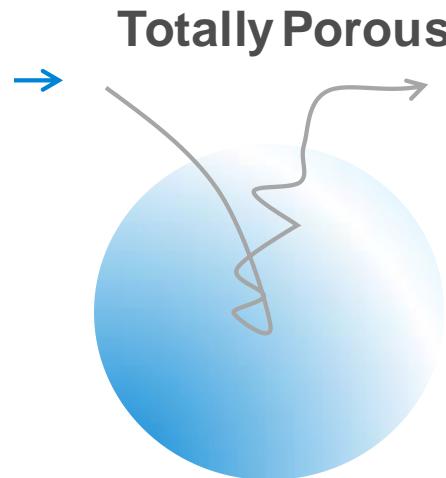
# Comparison of Particle Size Distributions

## Totally Porous and Poroshell 120 Particles



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# Analyte Mass Transfer Improvements through Lower Diffusion



- **Totally porous particles**
  - diffusion throughout particle
- **Poroshell 120**
  - diffusion limited to outer shell

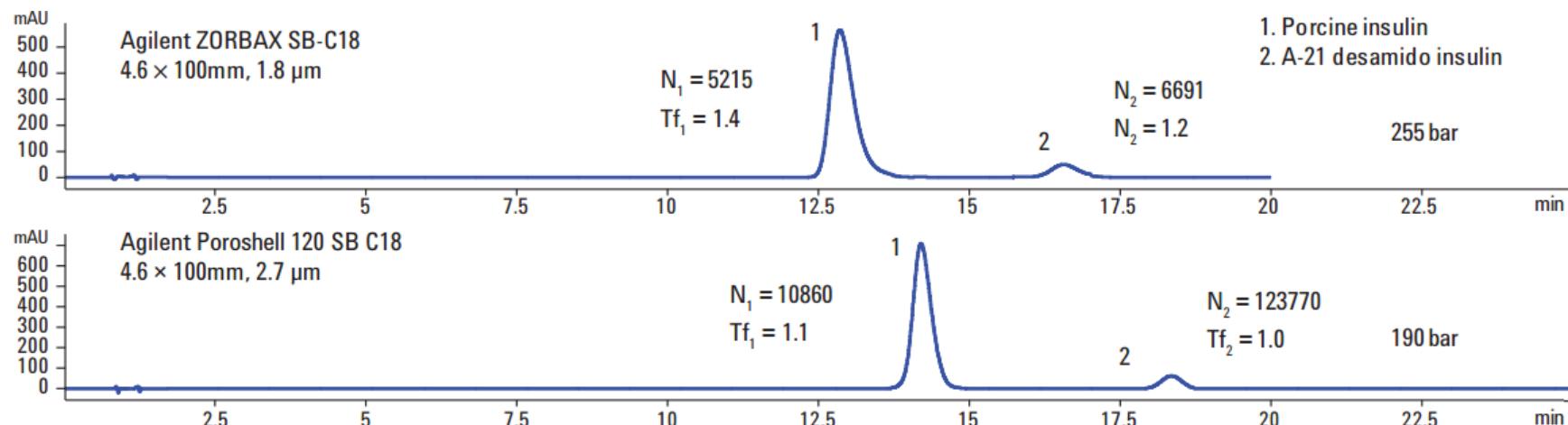
van Deemter equation:  
$$h = A + B/\nu + C \cdot \nu$$

- **Results:**
  - Lower C term
  - Higher efficiency
- **And**
  - Higher flow rate with
  - Minimal impact on efficiency



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# Pore Size and Efficiency

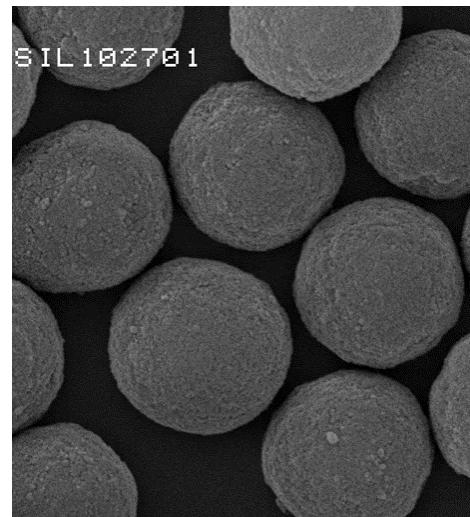
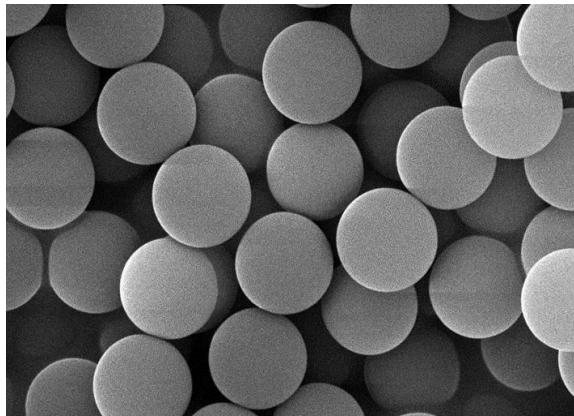


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# Making a Poroshell Particle



## Make the solid core

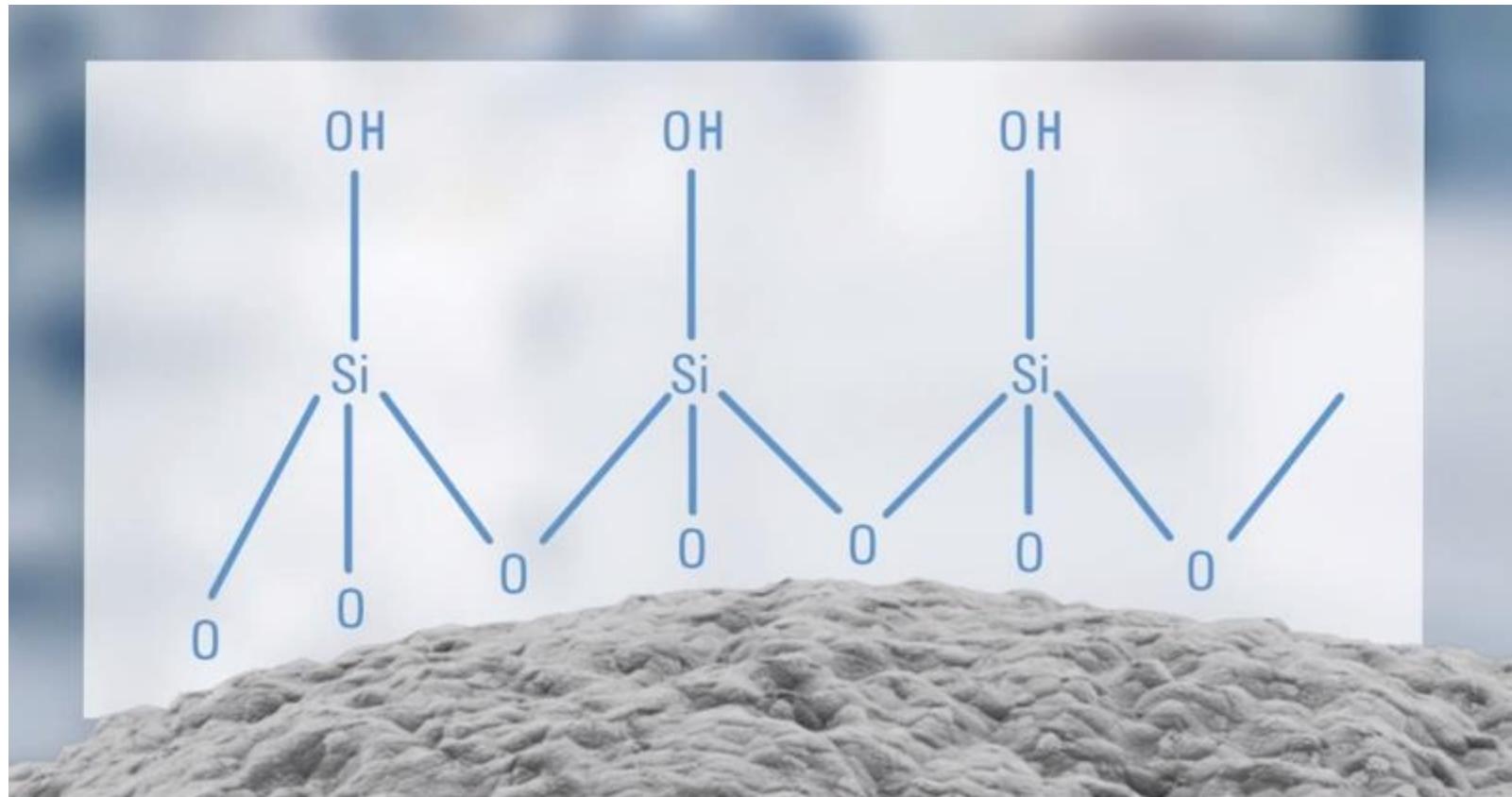
- Smooth surface
- Tight particle size distribution
- Tightly packed column bed
- Higher efficiency

## Apply the porous shell

- Single coacervation step
- High yields
- Better reproducibility

## Apply the bonded phase

# Silica Particle Surface

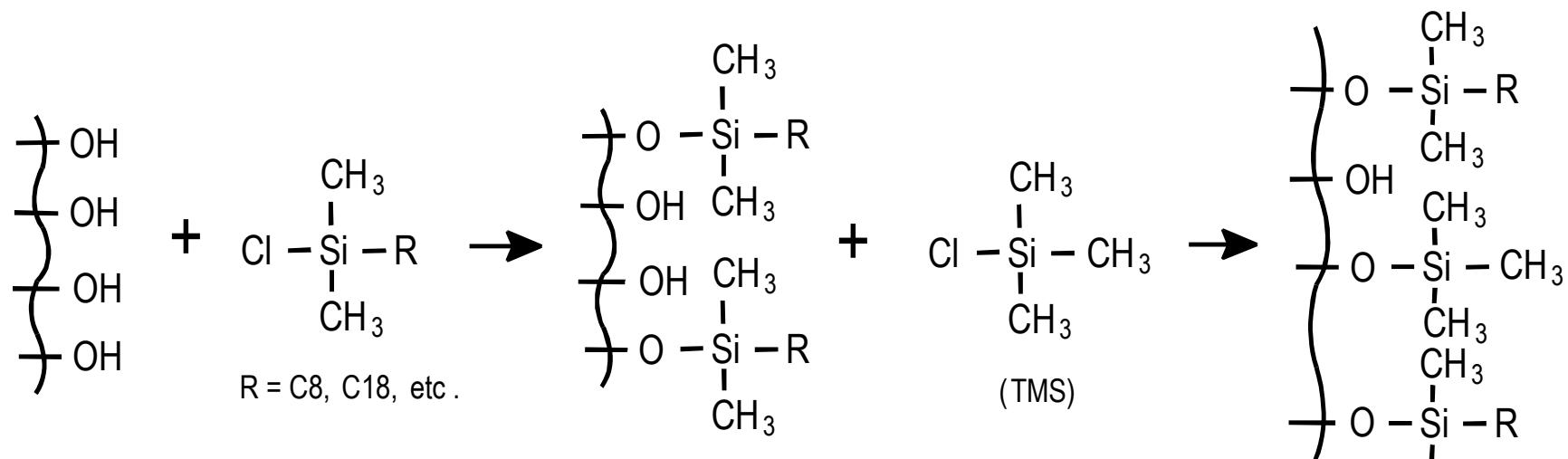


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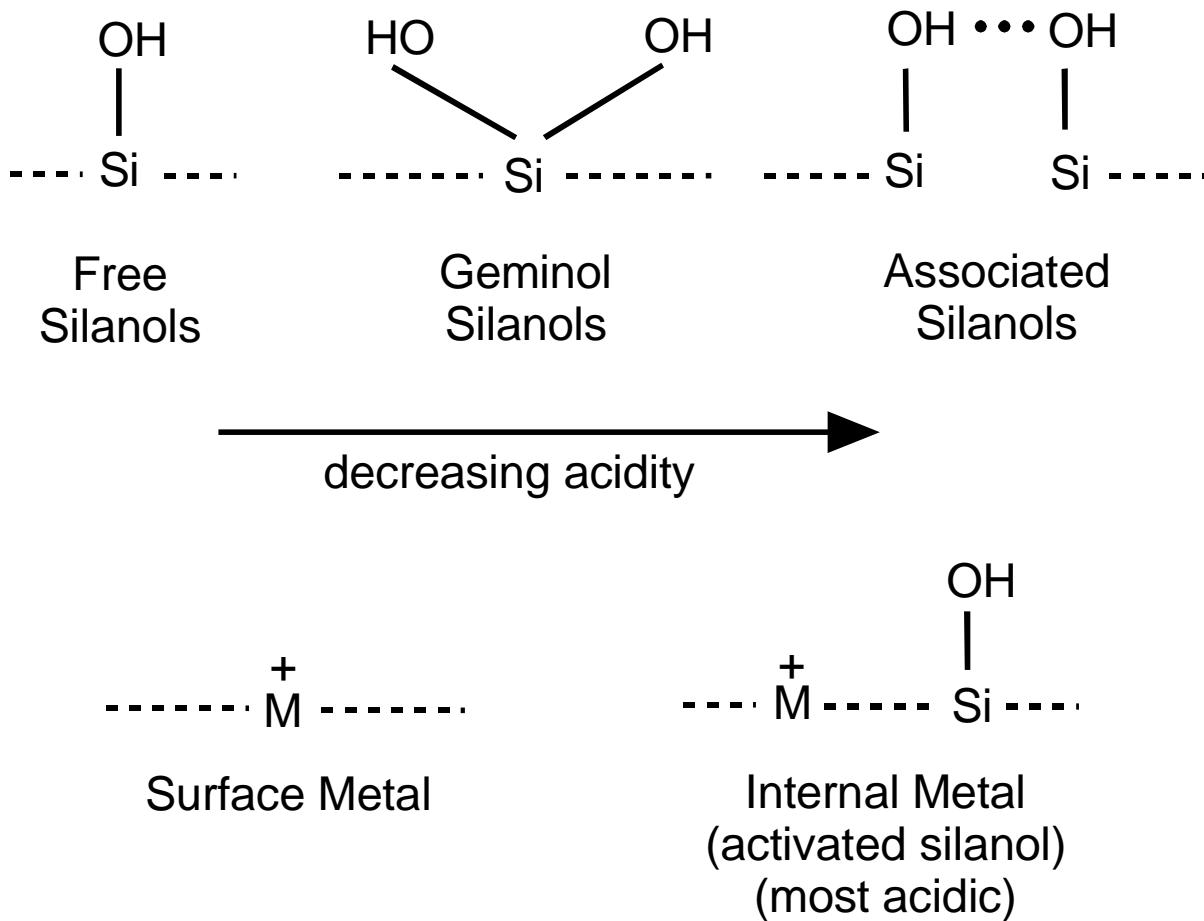
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# Typical Stationary Phase Bonding and Endcapping Reaction

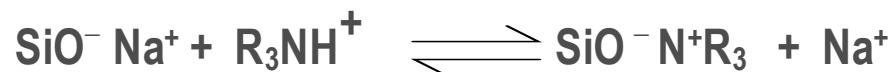


# The Surface of Silica Supports



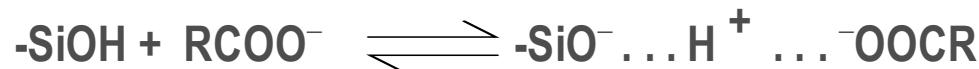
# Potential Secondary Interactions

## Ion-exchange



1. Ionized silanols ( $\text{SiO}^-$ ) will ion-exchange with protonated bases ( $\text{R}_3\text{NH}^+$ ) which can cause tailing and method variability. This occurs most often at mid pH where silanols are ionized.

## Hydrogen bonding

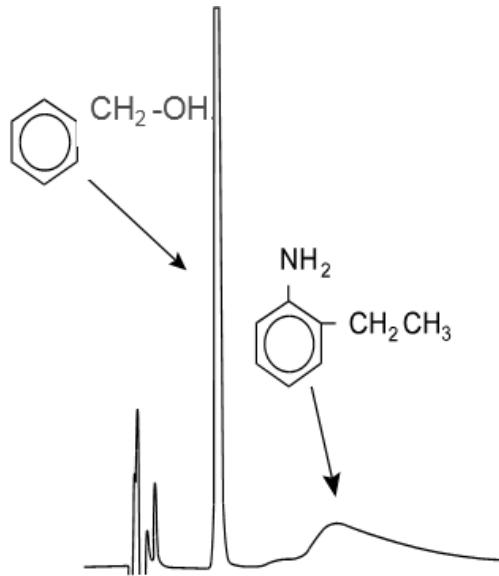


2. Unprotonated acids can compete for  $\text{H}^+$  with protonated silanols. This can occur at low pH.



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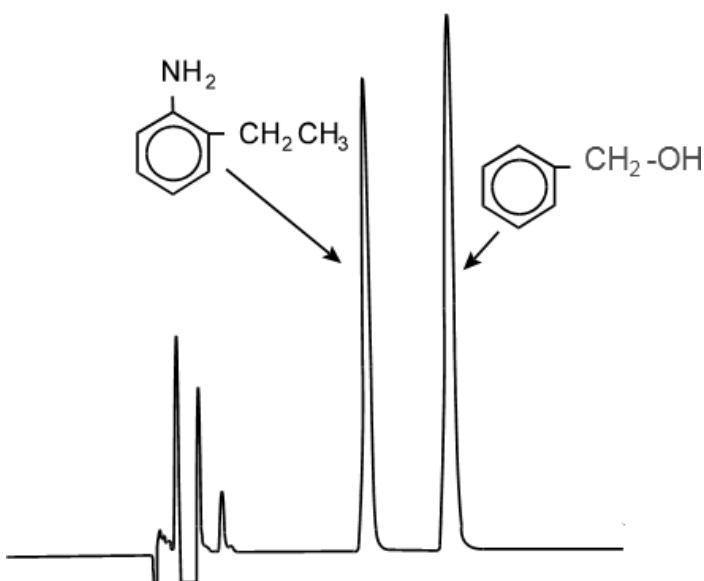
# Highly Purified Zorbax Rx-Sil



Original ZORBAX, 1973 and other type A silicas

- basic compounds can tail

Conditions: Flow Rate: 2.0 mL / min.  
Mobile Phase: 5% 2-Propanol in Heptane



ZORBAX Rx-Sil, 1987 and other Type B silicas

- basic compounds have less tailing
- lower effective silanol pKa



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# Poroshell 120 Column Chemistries

*Multiple bonded phases for flexibility in method development*



## Poroshell 120 EC-C18 and C8

- Robust end-capped C18/C8 for best peak shape at pH 2-9

## Poroshell 120 Stablebond C18 and C8

- Robust chemistries for pH<2
- Non-endcapped for alternate selectivity

## Poroshell HPH-C18 and HPH-C8

- Long lifetime at high pH

## Poroshell 120 Phenyl-Hexyl

- Excellent choice for pi-pi interactions
- Selectivity similar to phenyl, diphenyl, or other phenyl-hexyl columns

## Poroshell 120 SB-Ag

- Proprietary bonded phase is an excellent choice for polar analytes

## Poroshell 120 Bonus-RP

- Embedded polar group provides unique selectivity for polar compounds

## Poroshell 120 EC-CN

- Flexible end-capped CN chemistry for normal and reversed-phase separations

## Poroshell 120 HILIC

- Bare silica HILIC for use in hydrophilic interaction chromatography of polar molecules

## Poroshell 120 PFP

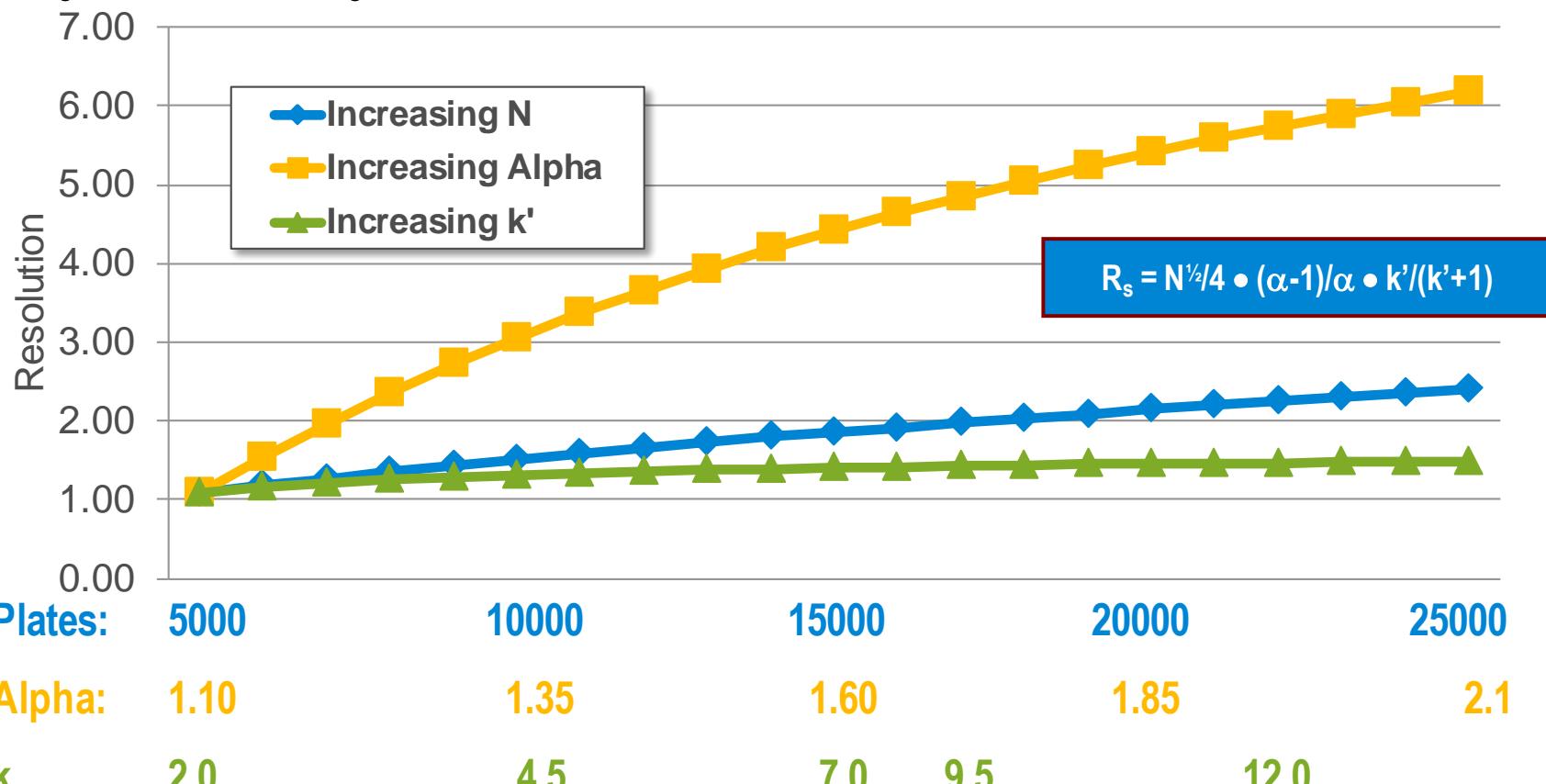
- Pentafluorophenyl chemistry for orthogonal selectivity relative to C18



# Porshell 120 Column Chemistries

Bonded Phase	Pore Size (Å)	Temp. Limit (°C)	pH Range	Endcapped	Carbon Load (%)	Surface Area (m <sup>2</sup> /g)
EC-C18	120	60	2 - 8	Double	10	130
EC-C8	120	60	2 - 8	Double	5	130
SB-C18	120	90	1 - 8	No	8	130
SB-C8	120	80	1 - 8	No	5.5	130
HPH-C18	100	60	3 - 11	Double	Proprietary	95
HPH-C8	100	60	3 - 11	Double	Proprietary	95
Phenyl-Hexyl	120	60	2 - 8	Double	9	130
SB-Aq	120	80	1 - 8	No	Proprietary	130
Bonus-RP	120	60	2 - 9	Triple	9.5	130
HILIC	120	60	0 - 8	No	N/A	130
EC-CN	120	60	2 - 8	Double	3.5	130
PFP	120	60	2 - 8	Yes	5.1	130

# Why So Many Phase Chemistries?



Selectivity Impacts Resolution Most

- Change bonded phase } Typical Method Development Parameters
- Change mobile phase
- Plates are easiest to increase

# Method Development Scheme

## Changing selectivity to improve resolution:

### Mobile phase (1<sup>st</sup> choice to change because it's easy)

- Mobile phase – organic modifier (ACN, MeOH etc.)
- Mobile phase – pH – over a wide pH range – pH 1-12 if needed

### Bonded phase (optimization for robust methods)

- Phases other than C18/C8
- Phenyl-Hexyl, Polar-embedded, CN, PFP



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# Why Is Changing the Bonded Phase Effective?

- Differences in interactions between polar and non-polar compounds.
- Other types of interactions with a bonded phase can be exploited (pi-pi interactions etc.)
- These all change with bonded phase!
- Changing the bonded phase can improve selectivity/resolution, reduce analysis time

**When you use Poroshell 120 columns the comparison of bonded phases can be done quickly!!**

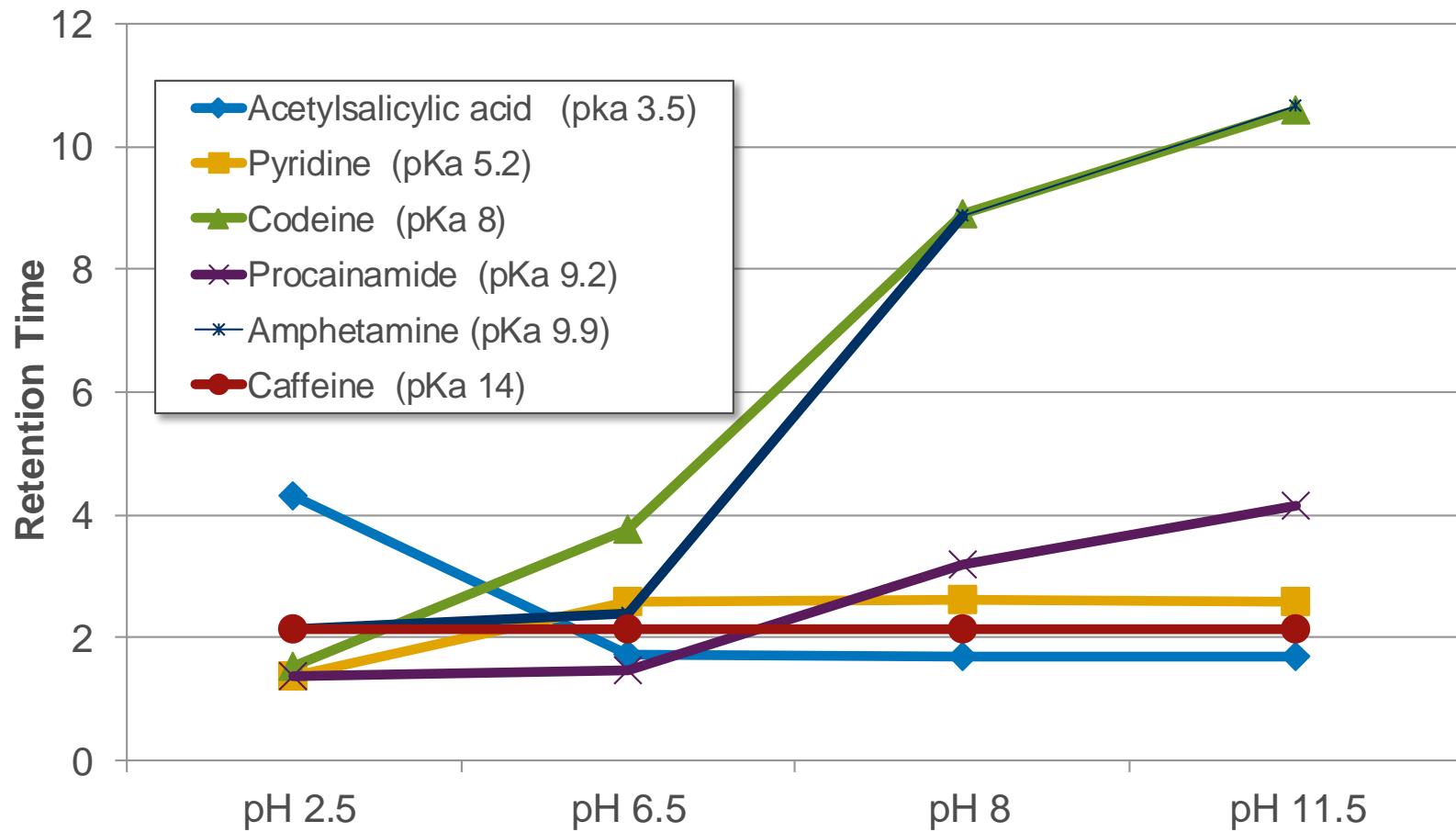
- Multiple column choices available with different high speed technologies make this easy



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# Change in Retention with pH for Ionizable Compounds is Compound-Dependent

More retention for non-charged analytes (i.e. acids at low pH and bases at high pH)



Mobile Phase: 45% MeOH, 55% 20 mM Phosphate Buffer



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# Change in Retention with pH for Ionizable Compounds is Key to Method Development

- Non-charged analytes have better retention (i.e. acids at low pH and bases at high pH)
- Silanols on silica ionize at mid-pH, increasing retention of basic analytes (i.e possible ion-exchange interactions)
- Choose mobile phase pH to optimize retention and selectivity during method development
- Poroshell 120 EC-C18 and C8 can be used over a wide pH range
- Other choices exist for high pH



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# First Steps in a Method Development Scheme

- **Poroshell 120 EC-C18 or Eclipse Plus C18**

- Low pH
- Adjust %ACN/MeOH for  $0.5 < k < 20$

More resolution needed

## Change % organic

More resolution needed

- **Change organic modifier**

- Adjust % organic for  $0.5 < k < 20$

More resolution needed

## • Change bonded phase

- Phenyl-Hexyl, Bonus-RP, CN, SB-C18, SB-C8, PFP, HILIC

➤ Select a high quality (Poroshell 120 EC or Eclipse Plus) C18 bonded phase first for good retention and resolution with typical acidic, basic and neutral samples.

➤ Optimize the organic component of the mobile phase to change selectivity

➤ Choose alternate bonded phases to completely optimize method if needed

➤ Evaluate other bonded phases and conditions for most robust method

➤ **With superficially porous or sub-2um column choices, steps can be done quickly**

• **Makes it possible to find a robust method**



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# Method Development Scheme – Evaluating Mid pH

From low pH



- Poroshell 120 EC-C18 or Eclipse Plus C18
  - pH 7 (6-9) acetate or other buffer,
  - Adjust % ACN for  $0.5 < k < 20$

More resolution needed



Change % organic

More resolution needed



- Change organic modifier (MeOH)
- Adjust % organic for  $0.5 < k < 20$

More resolution needed



- Try Phenyl-Hexyl, Bonus-RP, etc

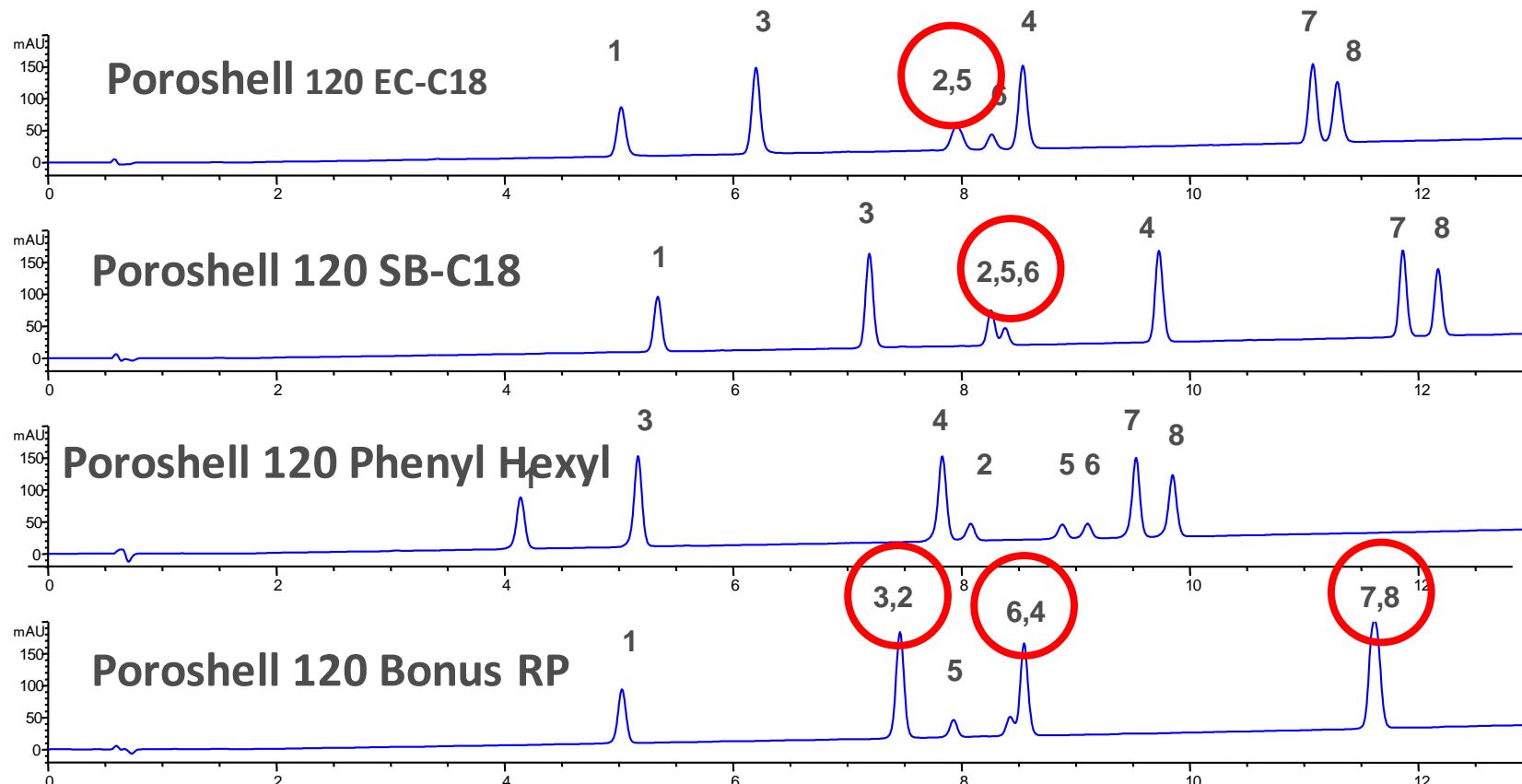
- Mid pH can provide better selectivity
- It may be more compatible with your sample
- The process for investigating mid pH is the same as for low pH
- Poroshell 120 EC- and Eclipse Plus deliver outstanding peak shape and lifetime performance at mid pH
- Alternate bonded phases should also be considered if improved selectivity is desired



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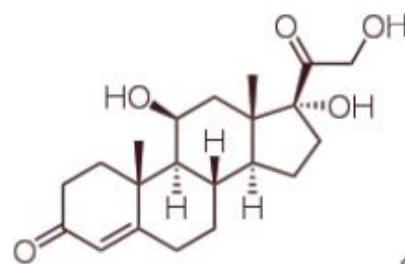
# Separation of 8 Steroids with Methanol Gradient

Best Resolution of all analytes with Poroshell 120 Phenyl-Hexyl

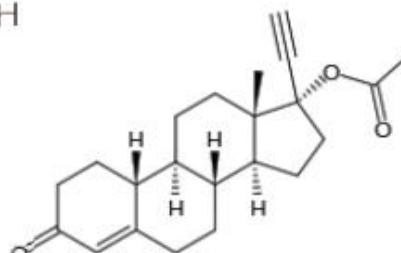


1. Hydrocortisone, 2.  $\beta$ -Estradiol, 3. Androstadiene 3,17 dione, 4. Testosterone,
5. Ethinylestradiol, 6. Estrone, 7. Norethindrone acetate, 8. Progesterone

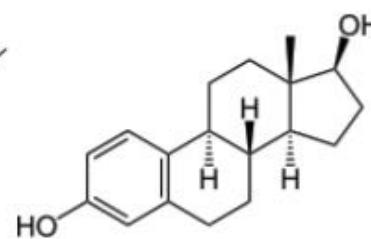
# Separation of 8 Steroids with Methanol Gradient



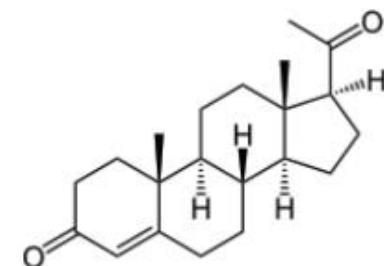
Hydrocortisone



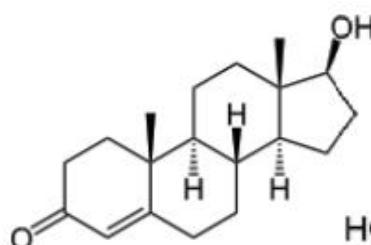
Norethindrone  
acetate



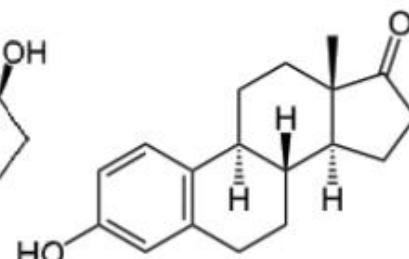
$\beta$  Estradiol



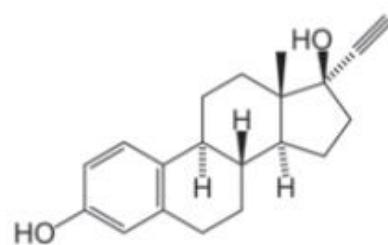
Progesterone



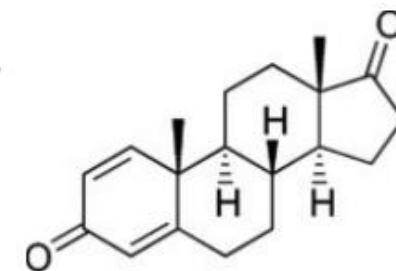
Testosterone



Estrone



Ethinylestradiol

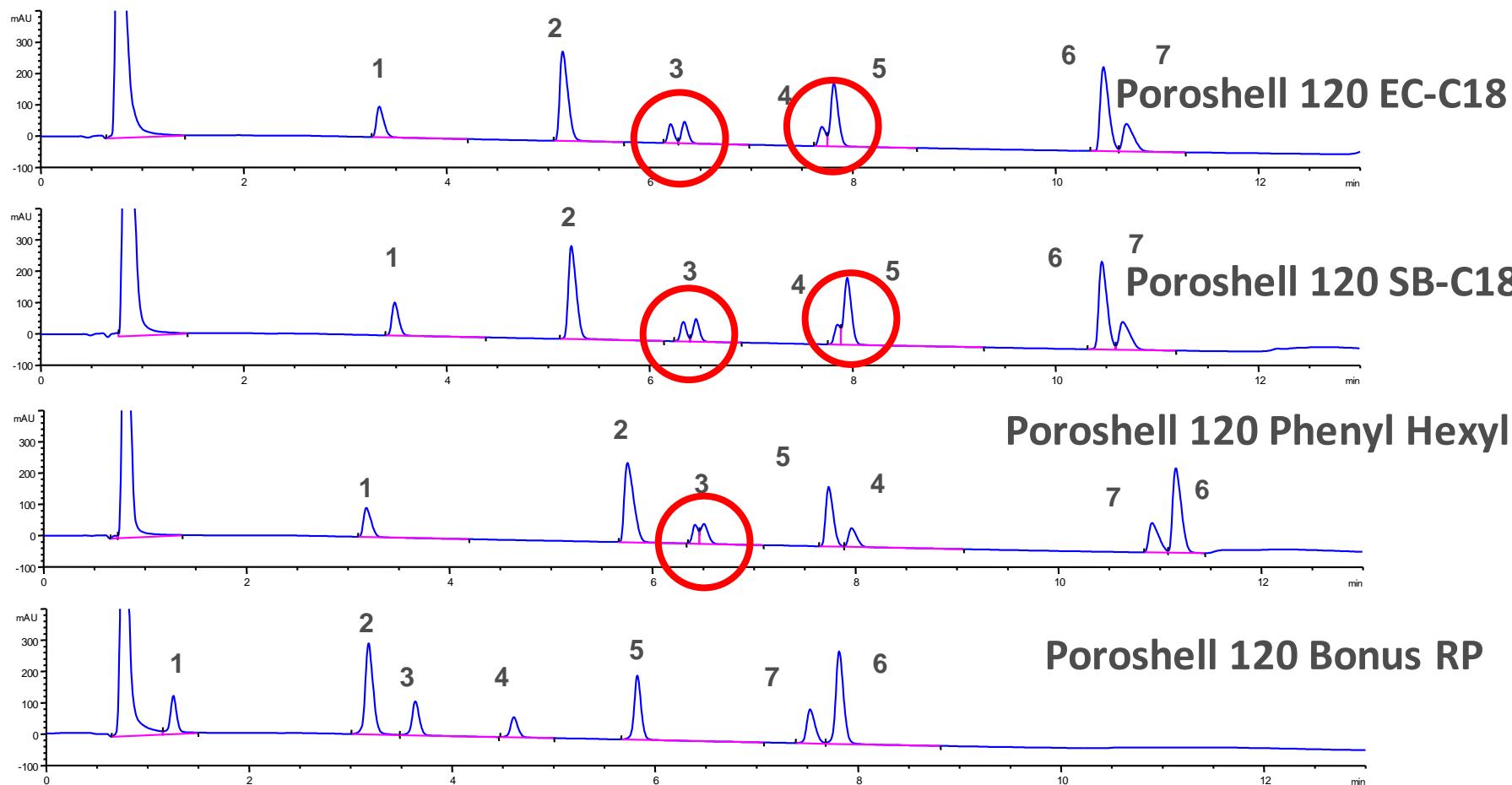


Androstadiene 3,17 dione



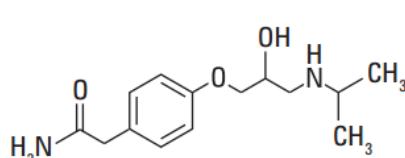
# Beta Blockers with Methanol Gradient

Best Resolution of all analytes with Poroshell Bonus-RP

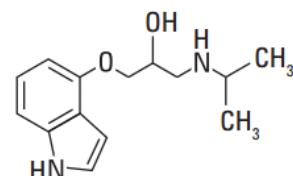


1. Atenolol, 2. Pindolol, 3. Naldolol, 4. Metoprolol, 5. Acebutolol, 6. Propranolol, 7. Alprenolol

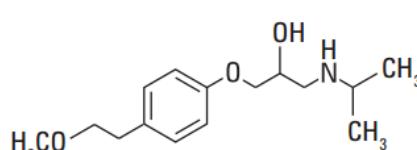
# Beta Blockers with Methanol Gradient



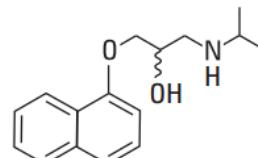
Atenolol



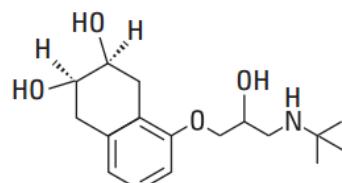
Pindolol



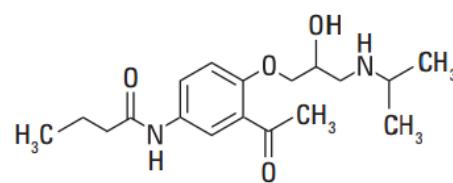
Metoprolol



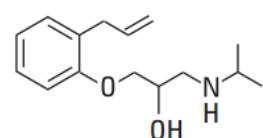
Propranolol



Nadolol



Acebutolol

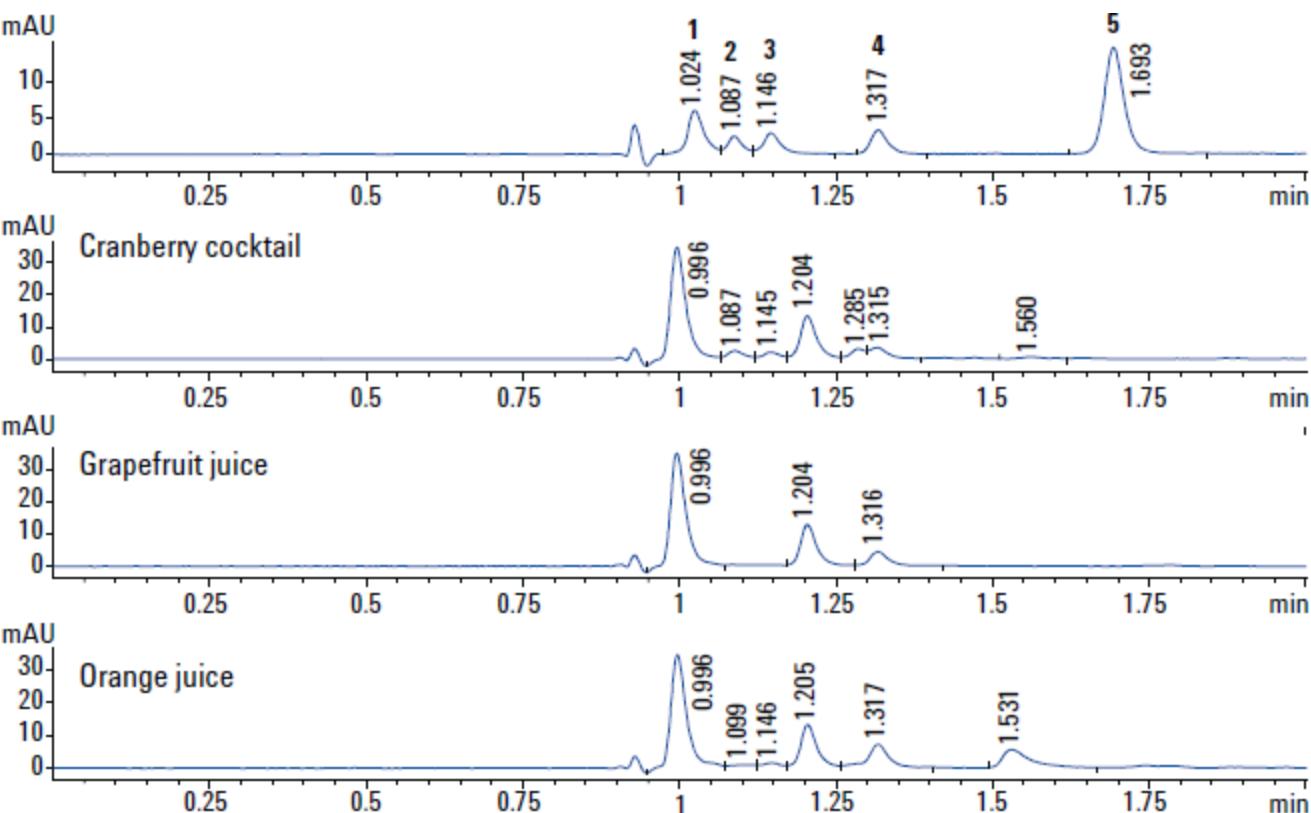


Alprenolol

Samples prepared at 10 mg/mL in DMSO  
diluted in water to a final concentration of  
0.1 mg/mL

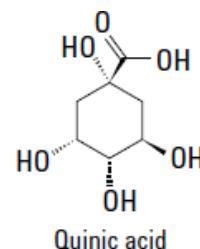


# ZORBAX SB-Aq Phase



## Peak ID

1. Tartaric acid
2. Quinic acid
3. Malic acid
4. Citric acid
5. Fumaric acid



Column: Agilent Poroshell 120 SB-Aq, 3 × 100 mm, 2.7 µm  
(p/n 685975-314)

Eluent: 100 mM Potassium phosphate buffer, pH 2.5

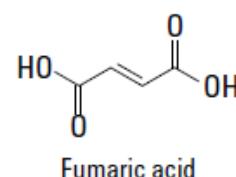
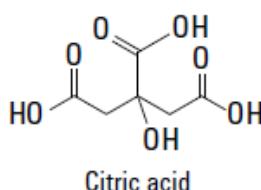
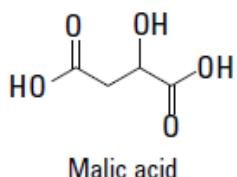
Injection volume: 5 µL

Flow rate: 0.5 mL/min

Temperature: 50 °C

Detector: DAD, at 226 nm

5991-1992EN



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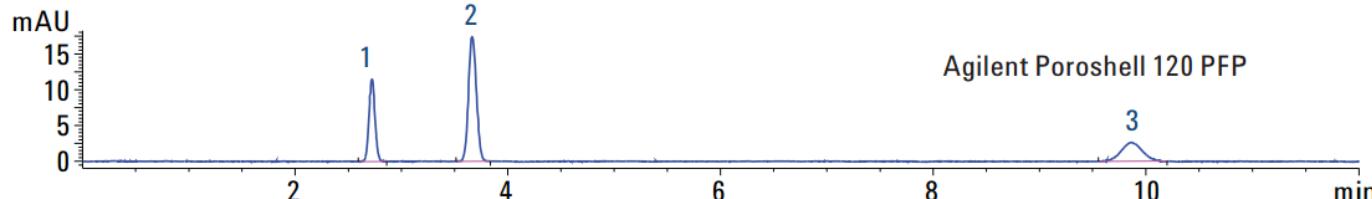
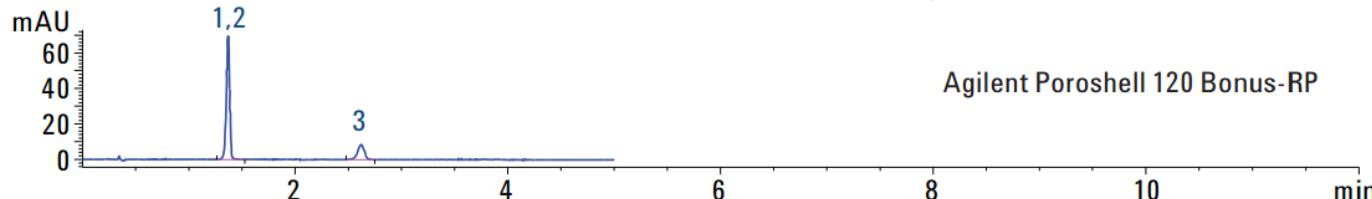
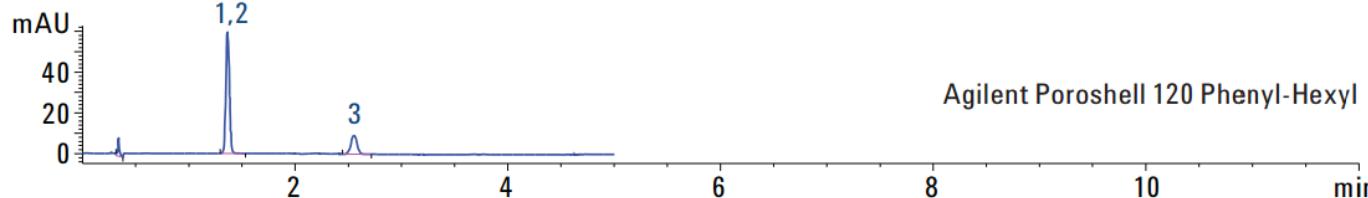
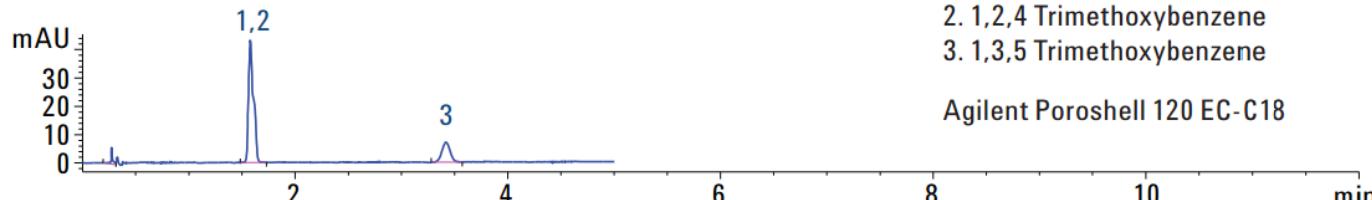
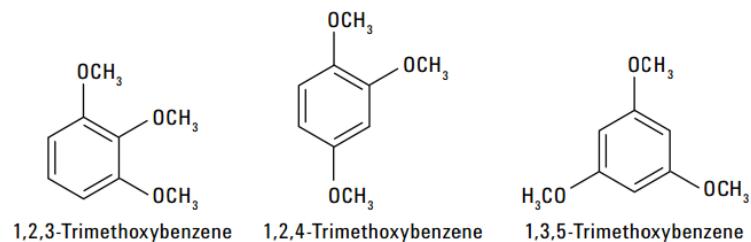
# New Phase on Poroshell 120

## Poroshell 120 PFP

- USP L43
- pentafluorophenyl bonded phase
- Excellent choice for polar analytes
- Unique pi-pi interactions
- The ring system electron deficient, making it a Lewis acid
- Allows for electronic interactions with electron-donating Lewis bases
- Alternative phase chemistry orthogonal to C18 chemistries
- **Recommended operating range**
  - pH 2-8
  - Maximum temp: 60°C



# Positional isomers on Poroshell 120 phases



Poroshell 120,  
4.6 × 50 mm,  
70:30 water:MeOH,  
1.5 mL/min, 40 °C,  
254 nm



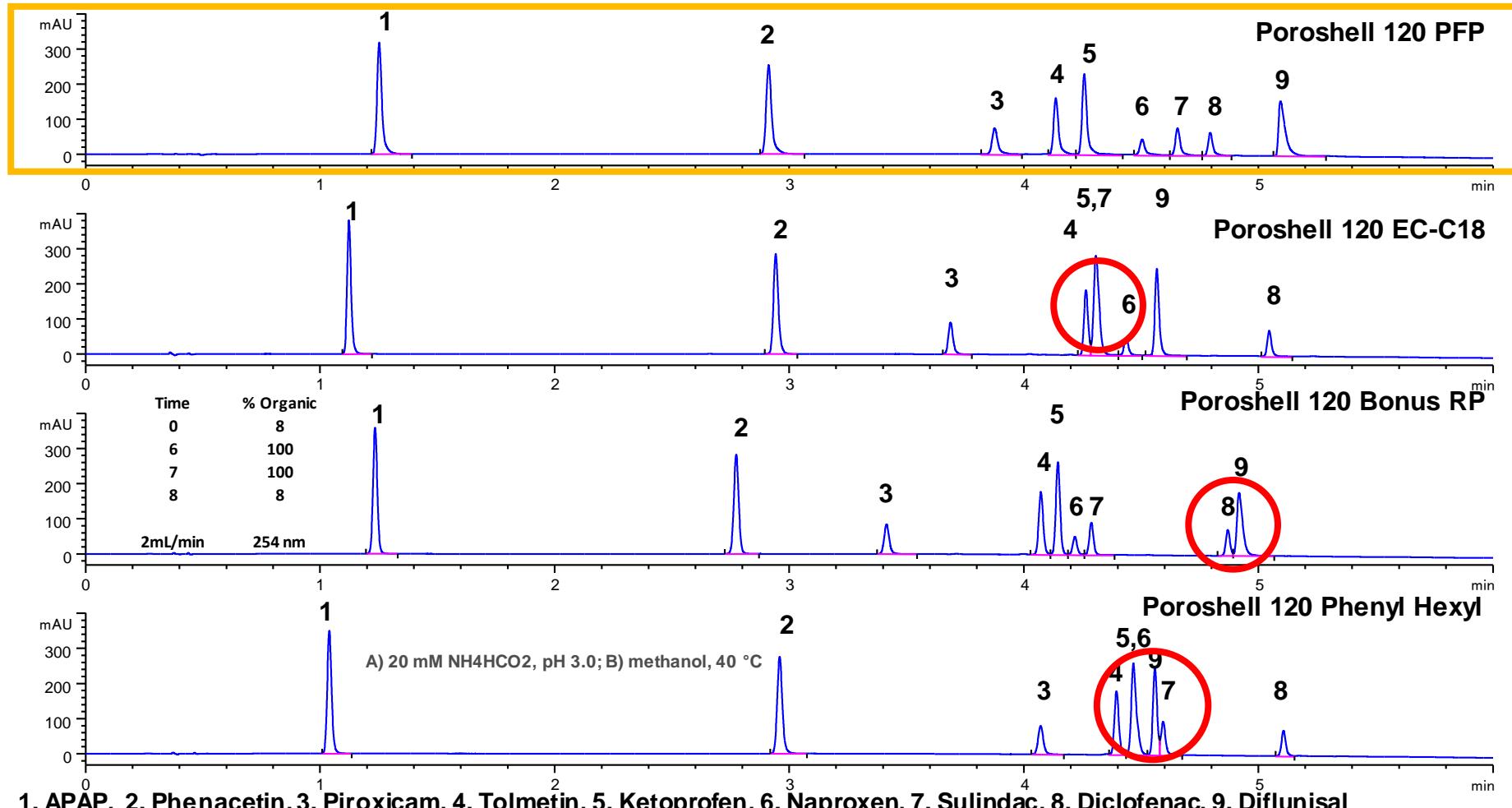
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# NSAID Separation Poroshell 120 with a Methanol Gradient

Best Resolution of all analytes with Poroshell 120 PFP

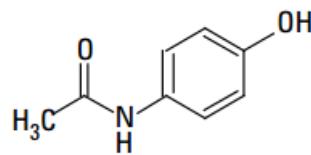


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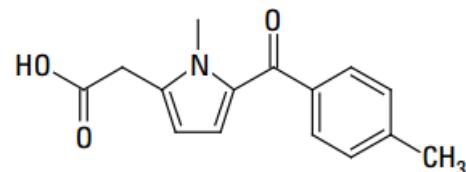
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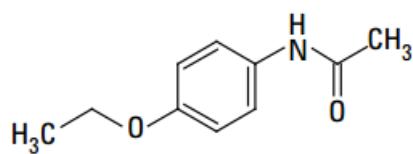
# NSAID Separation Poroshell 120 with a Methanol Gradient



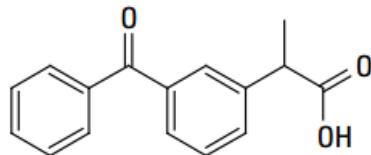
Acetaminophen (APAP)



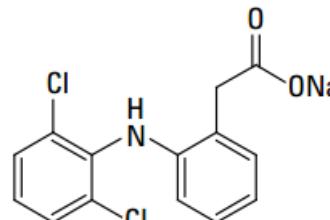
Tolmetin



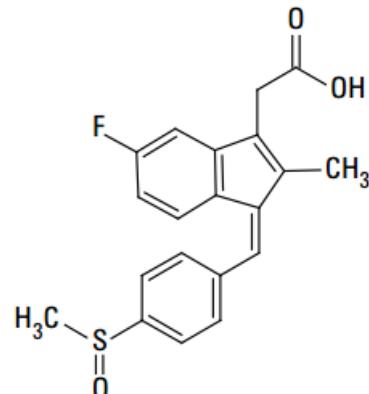
Phenacetin



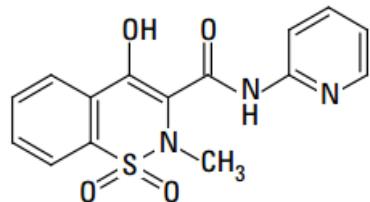
Ketoprophen



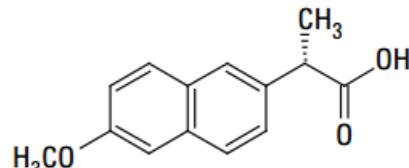
Diclofenac



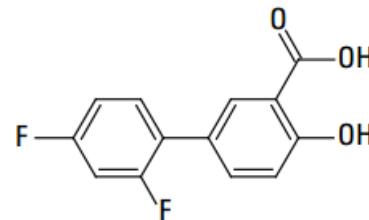
Sulindac



Piroxicam



Naproxen



Diflunisal



# Method Development at High pH

From Mid pH



- **Poroshell 120 HPH-C8 or C18**
  - pH 10.5 (9-12) 5 mM ammonia, or TEA, or 10 – 50 mM organic or borate buffers
  - T = 25°C (ambient – 40°C)
  - Adjust MeOH for  $0.5 < k < 20$

More resolution needed



- Change organic modifier
- Adjust for  $0.5 < k < 20$

Try different HPLC mode - HILIC

## Reasons to Consider High pH

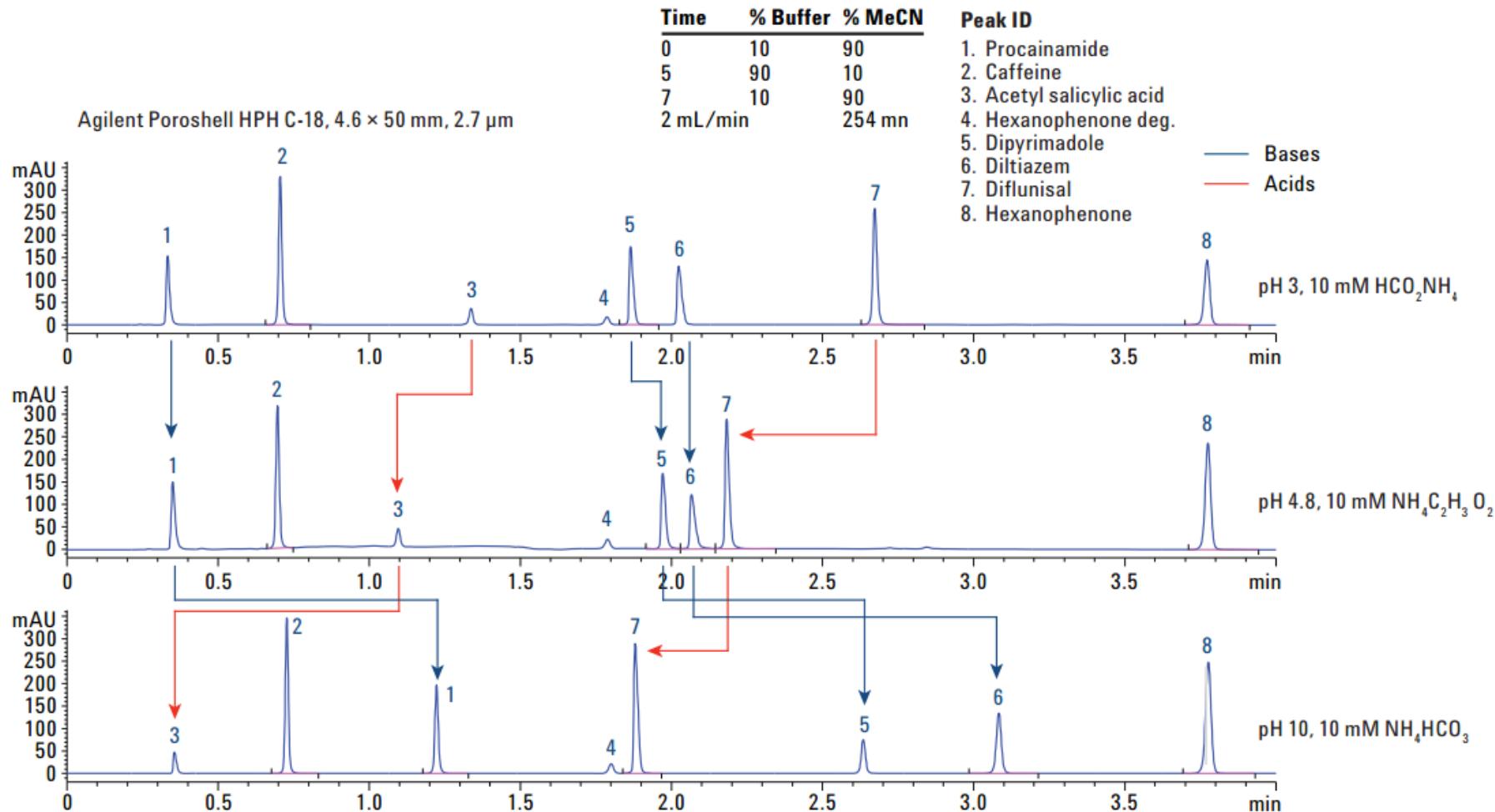
- Increase retention of basic compounds by analyzing them in non-charged form
- Improve selectivity



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# Poroshell HPH-C18 Low, Mid, & High pH

Why use multiple pH's?

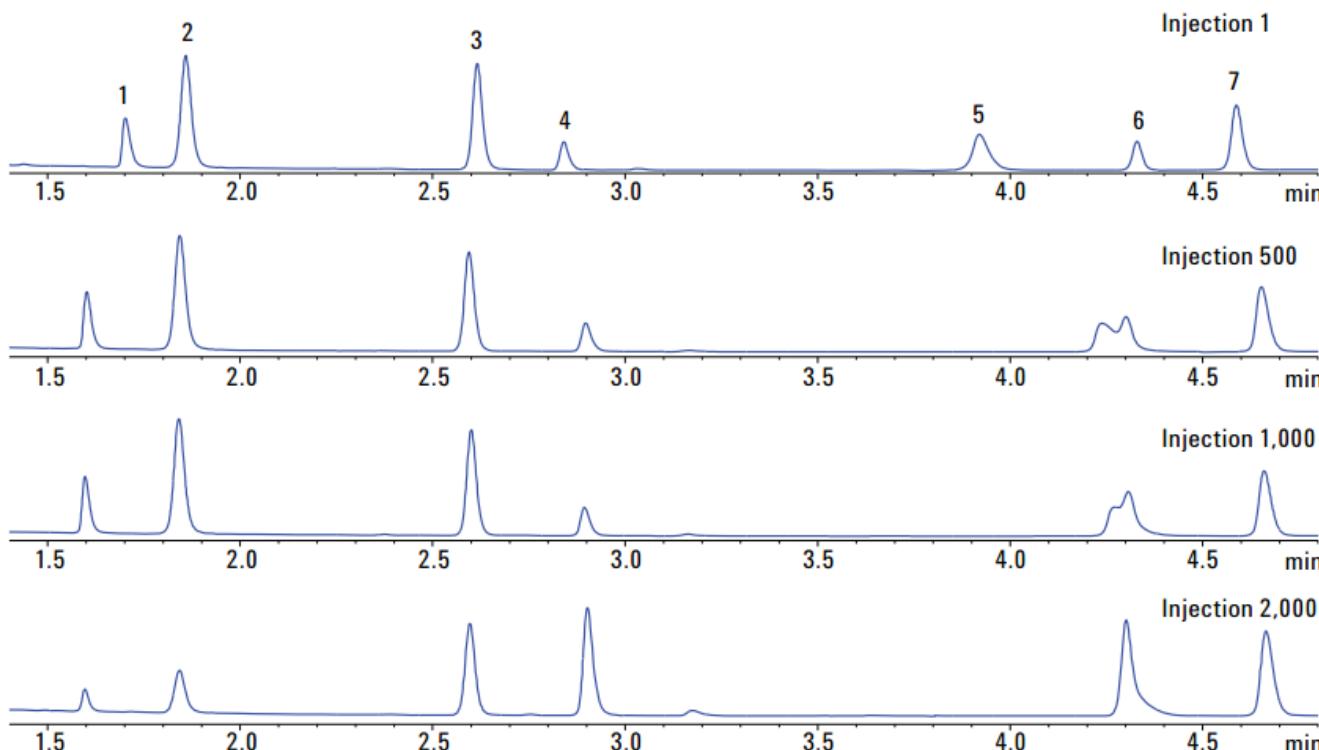


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# Column Lifetime with High pH Methods



Other brand column on an ammonium bicarbonate gradient at pH 10.

## Peak ID

1. Methyl salicylate
2. 4-Chlorocinnamic acid
3. Acetophenone
4. Quinine
5. Nortriptyline
6. Heptanophenone
7. Amitriptyline

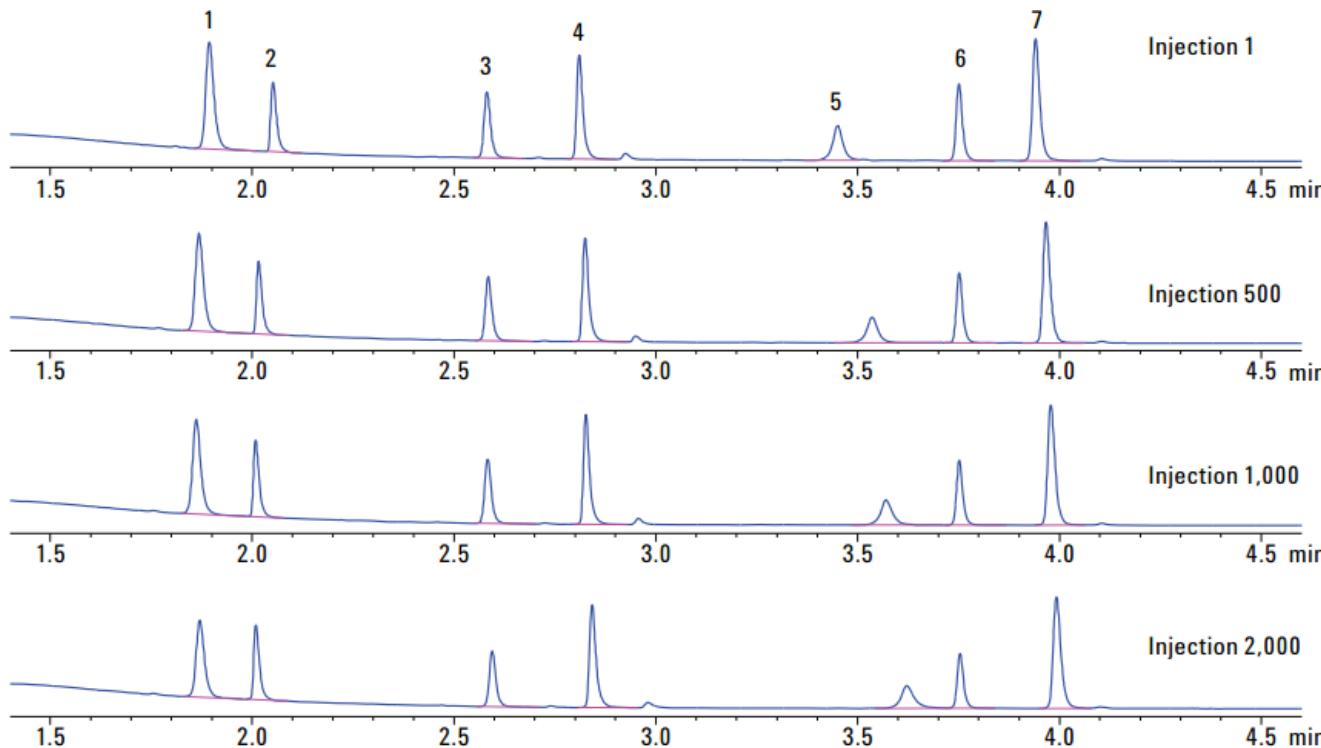
## Conditions

Column:	Other brand, 2.1 x 50 mm, 3 $\mu$ m
Eluent:	A) 10 mM ammonium bicarbonate adjusted to pH 10.0 in water
Flow rate:	B) acetonitrile
Gradient:	0.4 mL/min
Time (min)	%B
0	5
5	95
5.1	5
Total run time:	7 min



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# Column Lifetime with High pH Methods



## Peak ID

1. Methyl salicylate
2. 4-Chlorocinnamic acid
3. Acetophenone
4. Quinine
5. Nortriptyline
6. Heptanophenone
7. Amitriptyline

## Conditions

Column: Agilent Poroshell 120 HPH-C18, 2.1 × 50 mm, 2.7 µm  
Eluent: A) 10 mM ammonium bicarbonate adjusted to pH 10.0 in water  
Flow rate: 0.4 mL/min  
Gradient:

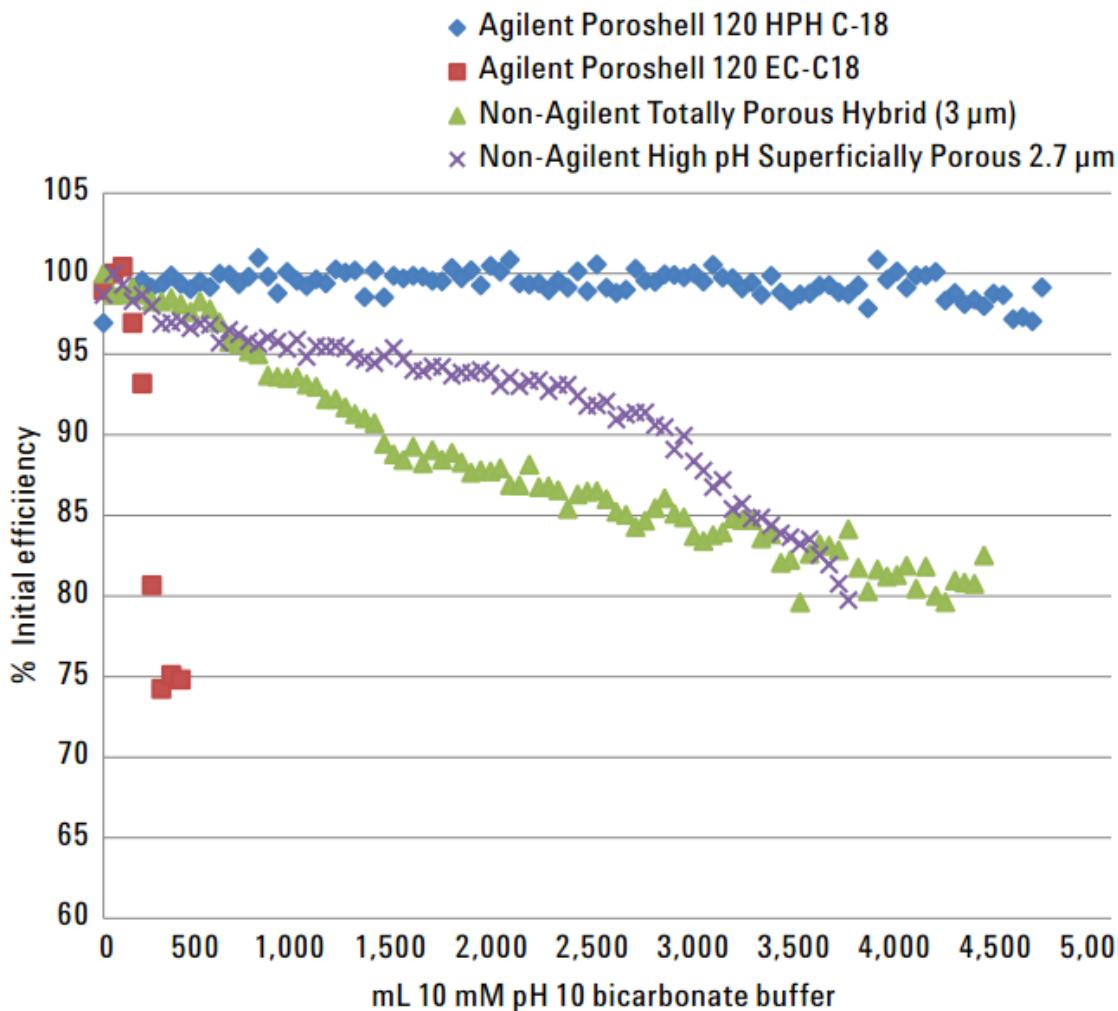
Time (min)	%B
0	5
5	95
5.1	5

  
Total run time: 7 min



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# Column Lifetime with High pH Methods



## Conditions

Columns: 2.1 × 50 mm, 2.7 µm  
Eluent: A) 0.1% ammonium hydroxide in water  
B) acetonitrile  
Flow rate: 0.4 mL/min  
Gradient:  

Time (min)	% B
0	5
3	95
3.5	5

  
Total run time: 4 min

Excellent column life  
with HPH-C18 and C8



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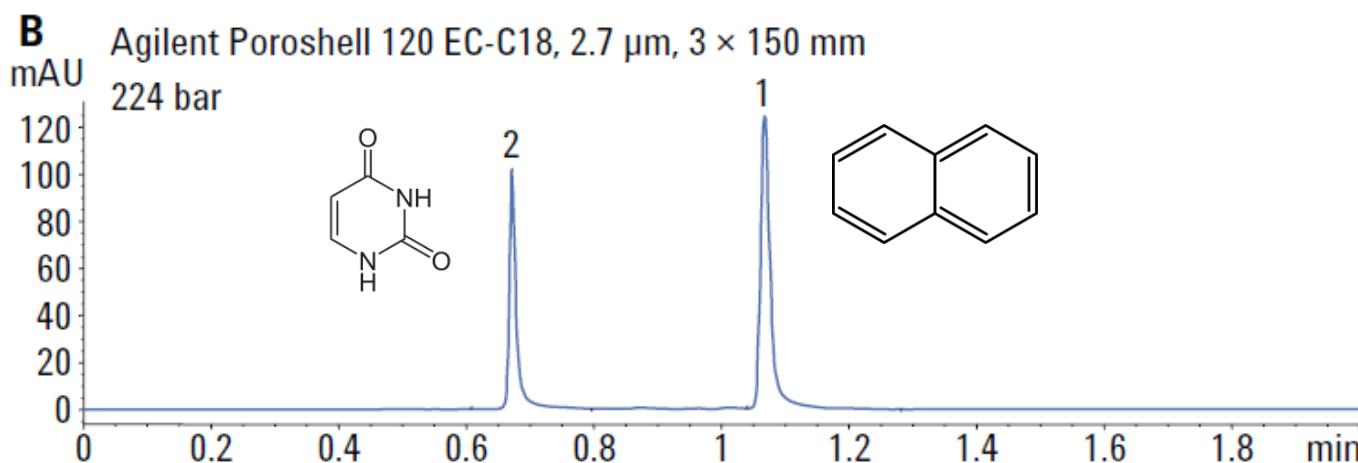
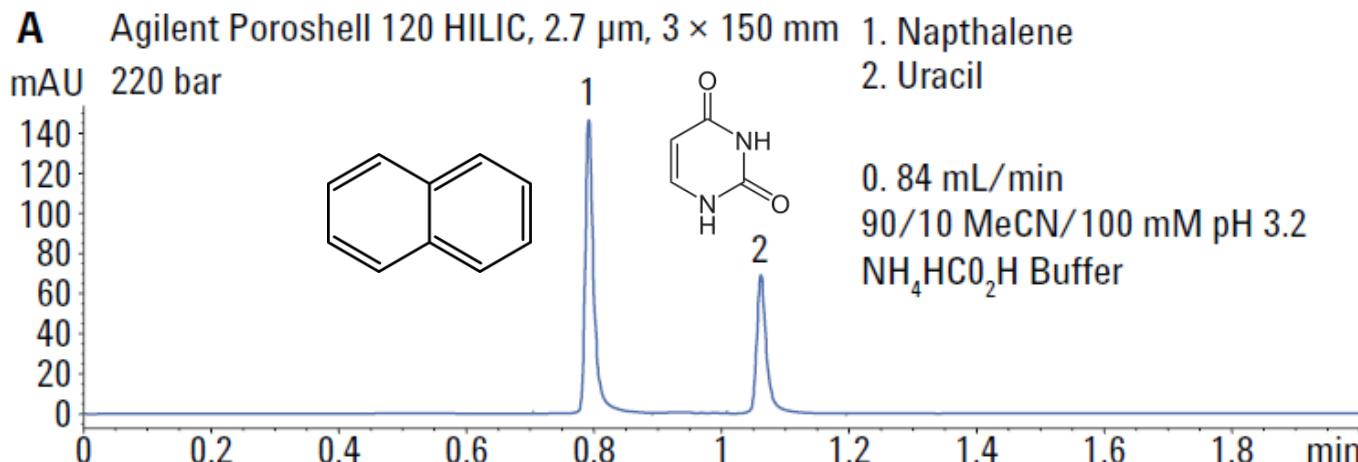
# HILIC

## Hydrophilic Interaction Chromatography

- HILIC offers more retention than reversed-phase for very polar bases
- Polar stationary phase:
  - Silica
  - Amine
  - Amide
- Polar mobile phase:
  - Water is the strong solvent
  - Typically ACN/water
  - Buffer controls ionization of analyte and stationary phase
  - Typically ammonium acetate or ammonium formate
- Retention/elution is from least to most polar

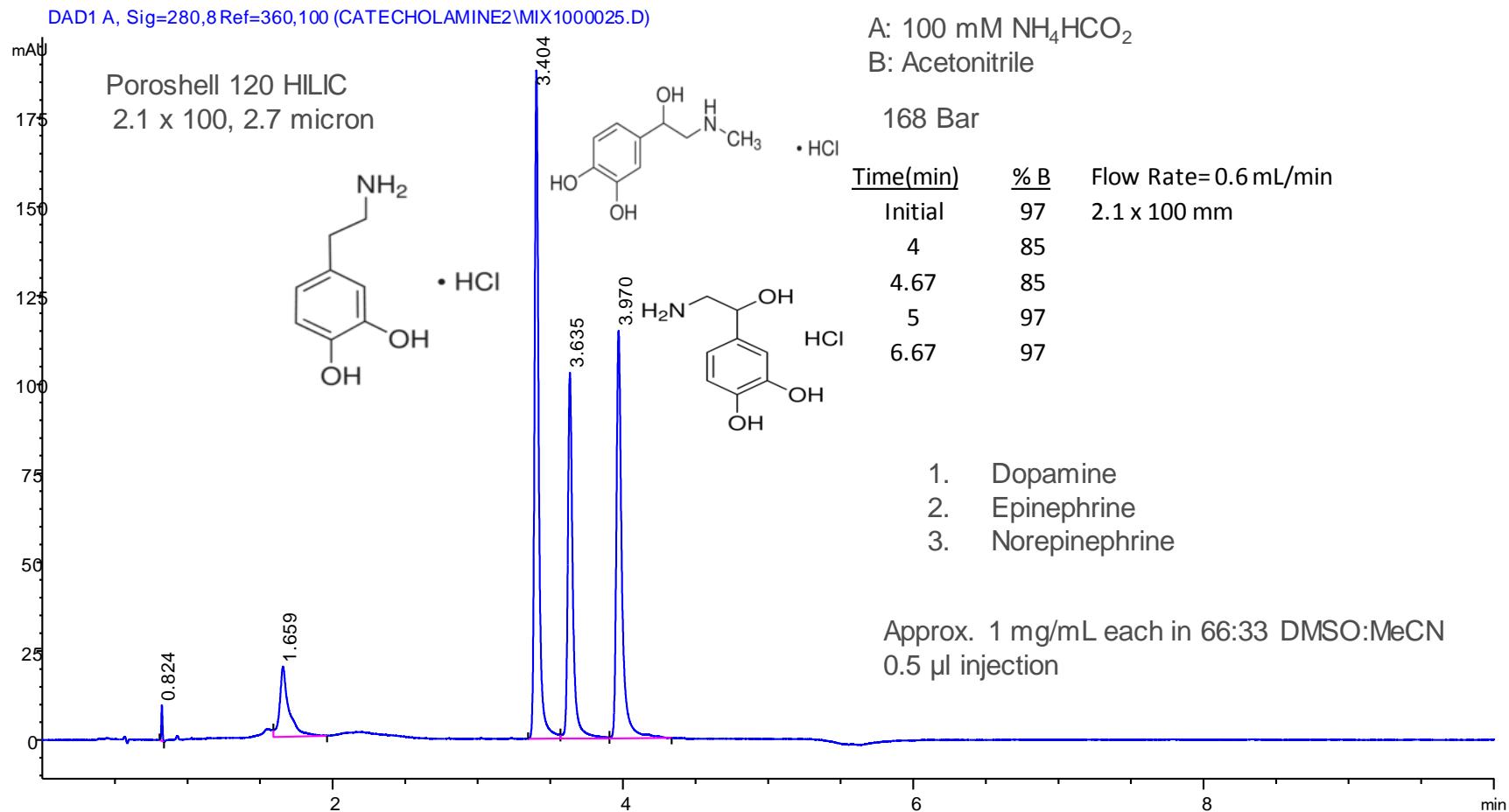


# HILIC – comparison with C18



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# HILIC Separation of Catecholamines Poroshell 120 2.1 x 100, 2.7 micron



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# Poroshell 120 Options

## Poroshell 120 (2.7 µm)

Size (mm)	EC-C18	EC-C8	SB-C18	SB-C8	NEW HPH-C18	NEW HPH-C8
4.6 x 150	693975-902	693975-906	683975-902	683975-906	693975-702	693975-706
4.6 x 100	695975-902	695975-906	685975-902	685975-906	695975-702	695975-706
4.6 x 75	697975-902	697975-906	687975-902			
4.6 x 50	699975-902	699975-906	689975-902	689975-906	699975-702	699975-706
4.6 x 30	691975-902	691975-906	681975-902			
3.0 x 150	693975-302	693975-306	683975-302	683975-306	693975-502	693975-506
3.0 x 100	695975-302	695975-306	685975-302	685975-306	695975-502	695975-506
3.0 x 75	697975-302	697975-306	687975-302			
3.0 x 50	699975-302	699975-306	689975-302	689975-306	699975-502	699975-506
3.0 x 30	691975-302	691975-306	681975-302			
2.1 x 150	693775-902	693775-906	683775-902	683775-906	693775-702	693775-706
2.1 x 100	695775-902	695775-906	685775-902	685775-906	695775-702	695775-706
2.1 x 75	697775-902	697775-906	687775-902			
2.1 x 50	699775-902	699775-906	689775-902	689775-906	699775-702	699775-706
2.1 x 30	691775-902	691775-906	681775-902			
Size (mm)	Phenyl-Hexyl	SB-Aq	Bonus-RP	HILIC	EC-CN	NEW PFP
4.6 x 150	693975-912	683975-914	693968-901	693975-901	693975-905	693975-408
4.6 x 100	695975-912	685975-914	695968-901	695975-901	695975-905	695975-408
4.6 x 50	699975-912	689975-914	699968-901	699975-901	699975-905	699975-408
3.0 x 150	693975-312	683975-314	693968-301	693975-301	693975-305	693975-308
3.0 x 100	695975-312	685975-314	695968-301	695975-301	695975-305	695975-308
3.0 x 50	699975-312	689975-314	699968-301	699975-301	699975-305	699975-308
2.1 x 150	693775-912	683775-914	693768-901	693775-901	693775-905	693775-408
2.1 x 100	695775-912	685775-914	695768-901	695775-901	695775-905	695775-408
2.1 x 50	699775-912	689775-914	699768-901	699775-901	699775-905	699775-408

Note: Poroshell 120 columns have a 600 bar/9000 psi pressure limit.

## Poroshell 120 Fast Guards for UHPLC

Size (mm)	EC-C18	EC-C8	SB-C18	Phenyl-Hexyl	NEW PFP
4.6 x 5	820750-911	820750-913	820750-912	820750-914	
3.0 x 5	823750-911	823750-913	823750-912	823750-914	
2.1 x 5	821725-911	821725-913	821725-912	821725-914	821725-915



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# Method Development Kits



Method Development Kits	Description (One of each)	Dimension	Part No.
Poroshell 120 Selectivity	EC-C18, Phenyl-Hexyl, Bonus-RP	2.1 x 50 mm	5190-6155
Poroshell 120 Selectivity	EC-C18, Phenyl-Hexyl, Bonus-RP	4.6 x 50 mm	5190-6156
Poroshell 120 Aqueous	SB-Aq, Phenyl-Hexyl, Bonus-RP	2.1 x 50 mm	5190-6157
Poroshell 120 Aqueous	SB-Aq, Phenyl-Hexyl, Bonus-RP	4.6 x 50 mm	5190-6158
Poroshell 120 USP L1, L7, and L10	EC-C18, EC-C8, EC-CN	4.6 x 100 mm	5190-6159
Poroshell 120 USP L1, L7, and L10	EC-C18, EC-C8, EC-CN	3.0 x 100 mm	5190-6160
ZORBAX RRHD pH	SB-C18, Eclipse Plus C18, and Extend-C18	2.1 x 50 mm	5190-6152
ZORBAX Eclipse Plus	C18, C8, Phenyl-Hexyl	2.1 x 50 mm	5190-6153
ZORBAX RRHD Aqueous	SB-Aq, Bonus-RP, Eclipse Plus Phenyl-Hexyl	2.1 x 50 mm	5190-6154



# Poroshell 120 Options

Agilent Poroshell 120 4 µm

NEW

Size (mm)	EC-C18	EC-C8	PFP	Phenyl-Hexyl	HILIC
4.6 x 250	690970-902	690970-906	690970-408	690970-912	690970-901
4.6 x 150	693970-902	693970-906	693970-408	693970-912	693970-901
4.6 x 100	695970-902	695970-906	695970-408	695970-912	695970-901
4.6 x 50	699970-902	699970-906	699970-408	699970-912	699970-901
3.0 x 250	690970-302	690970-306	690970-308	690970-312	690970-301
3.0 x 150	693970-302	693970-306	693970-308	693970-312	693970-301
3.0 x 100	695970-302	695970-306	695970-308	695970-312	695970-301
3.0 x 50	699970-302	699970-306	699970-308	699970-312	699970-301
2.1 x 250	650750-902	650750-906	650750-408	650750-912	650750-901
2.1 x 150	693770-902	693770-906	693770-408	693770-912	693770-901
2.1 x 100	695770-902	695770-906	695770-408	695770-912	695770-901
2.1 x 50	699770-902	699770-906	699770-408	699770-912	699770-901

Guard columns for 4 µm

NEW

Size (mm)	EC-C18
4.6 x 5	820750-916
3.0 x 5	823750-916
2.1 x 5	821725-916



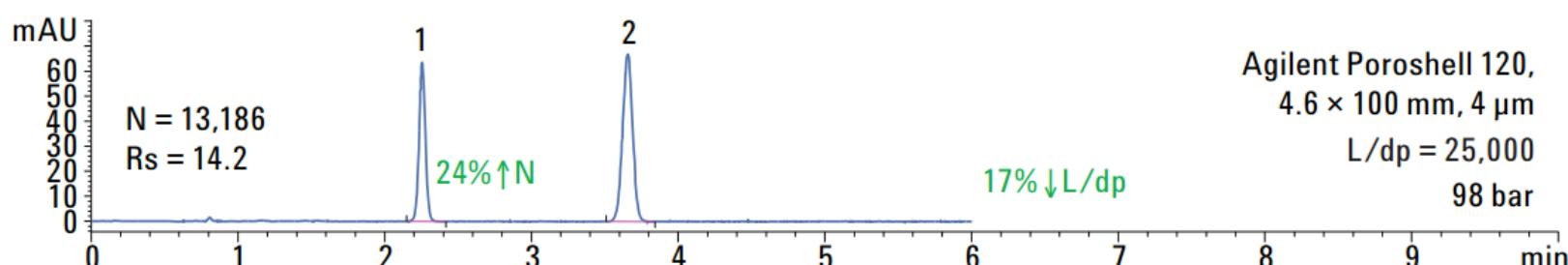
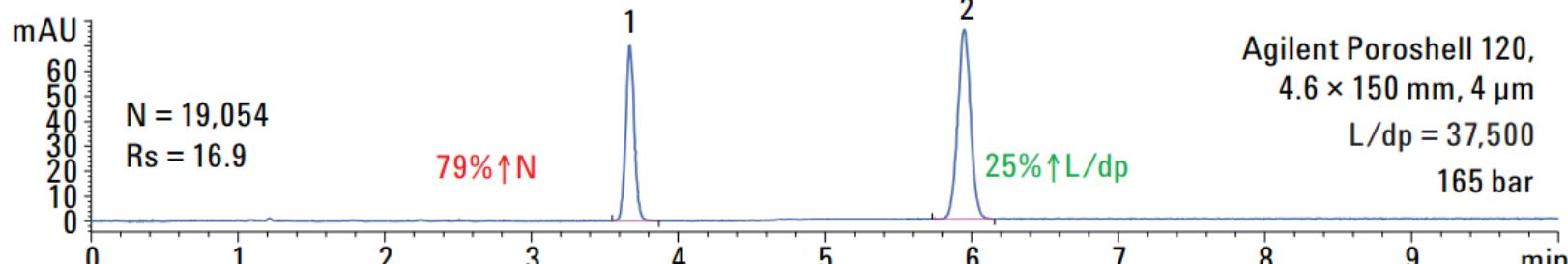
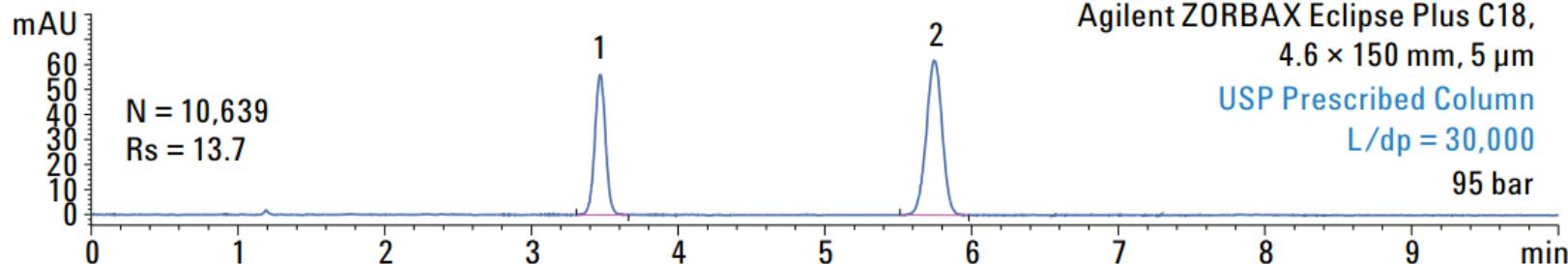
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# Benefits of 4 $\mu$ m Poroshell 120

System Suitability Method Requirement: N > 4000, Rs > 11.5



1. Naproxen

2. Butyrophenone

50:49:1 MeCN:water:acetic acid

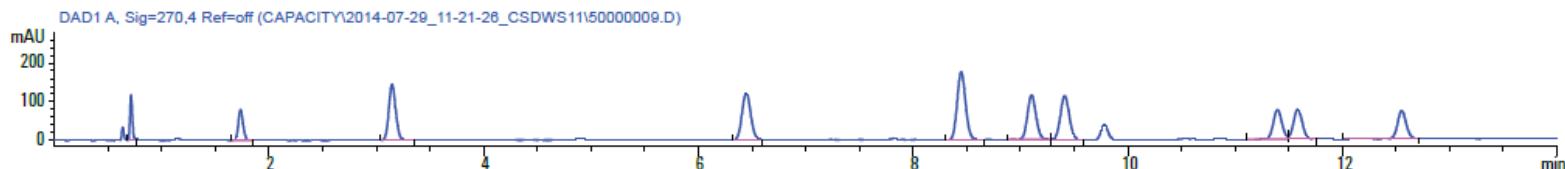
Flow rate: 1.2 mL/min

5991-5408EN

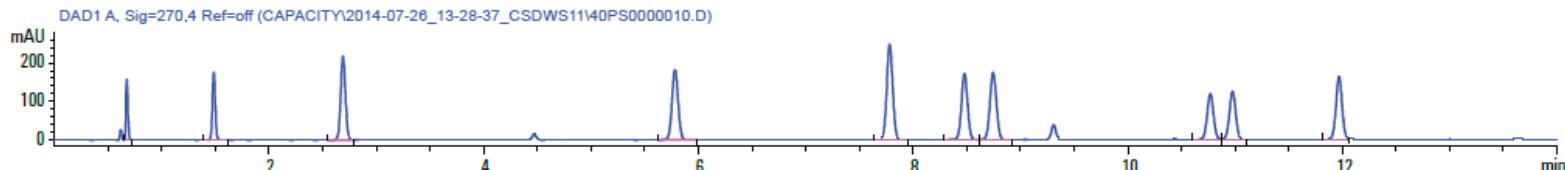
# Consistent Selectivity Across Particle Sizes

Phenol Mix gradient at 1.5 mL/min

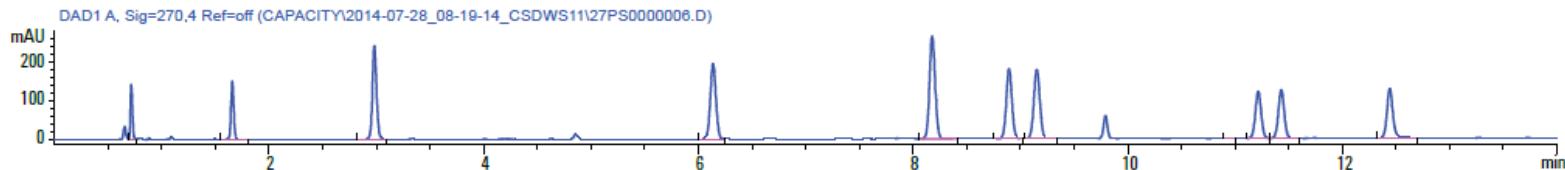
ZORBAX Eclipse Plus C18 5  $\mu$ m



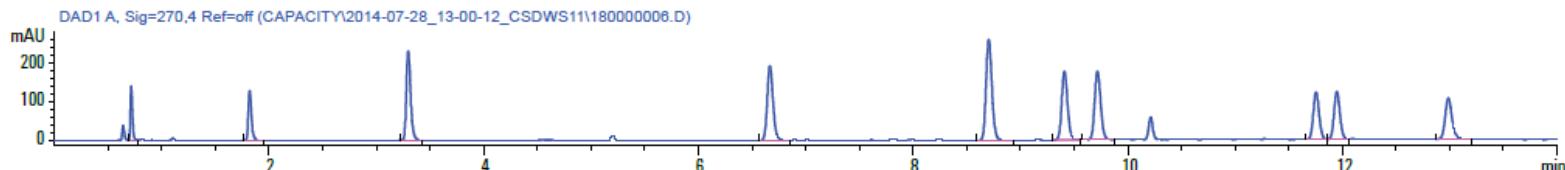
Poroshell 120 EC-C18 4  $\mu$ m



Poroshell 120 EC-C18 2.7  $\mu$ m



ZORBAX Eclipse Plus C18 1.8  $\mu$ m

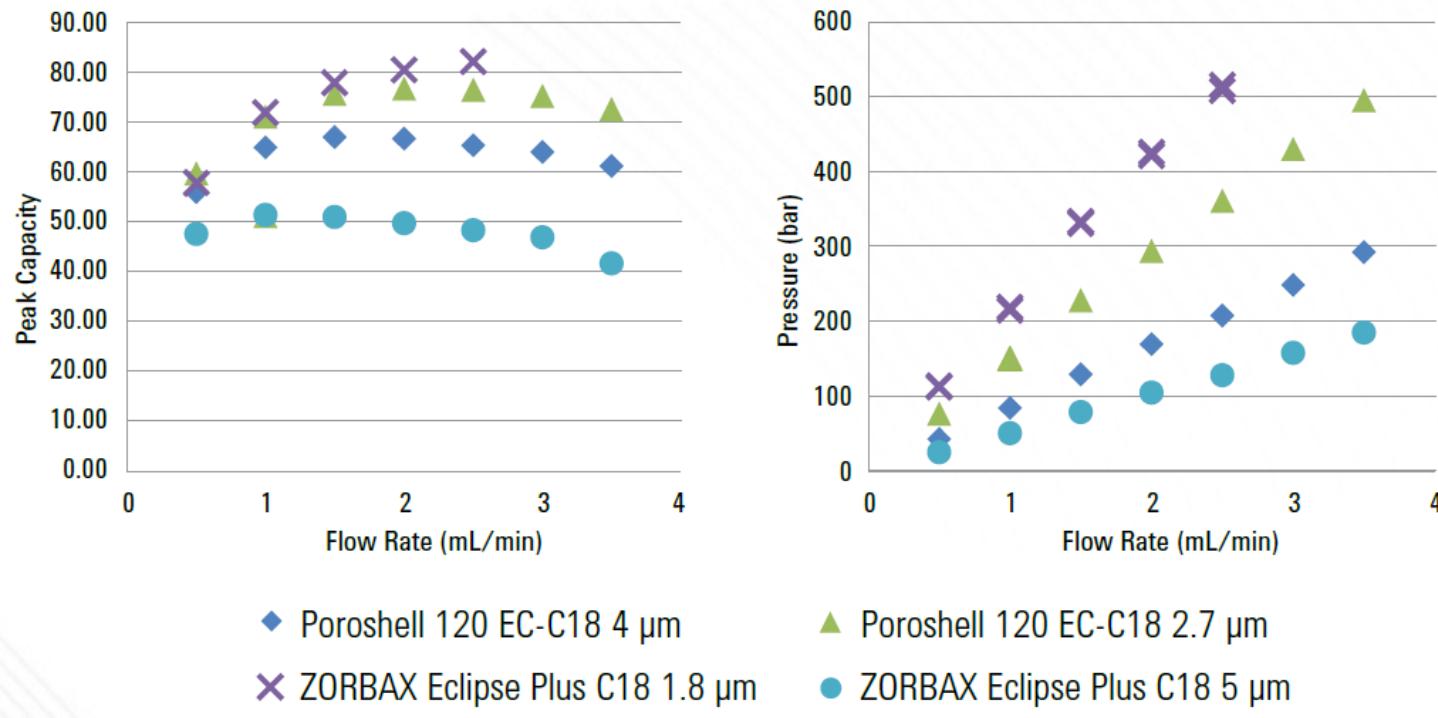


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April 8, 2015

# Consistent Selectivity Across Particle Sizes



Columns: All columns 4.6 x 100 mm

Instrument: Agilent 1260, pulse damper and mixing column bypassed

Mobile phase: A: 0.1% Formic acid  
B: MeOH + 0.1% formic acid

Flow rate: 0.4 mL/min

Temperature: 25 °C

Detection: 260 nm

Sample: 8 steroids

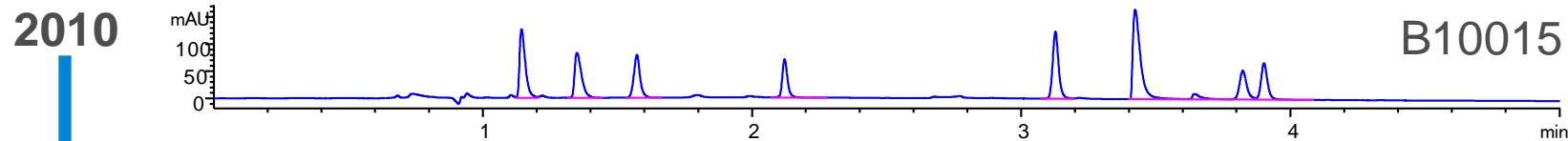
Gradient: 40-80% MeOH/14 min



# Lot Reproducibility

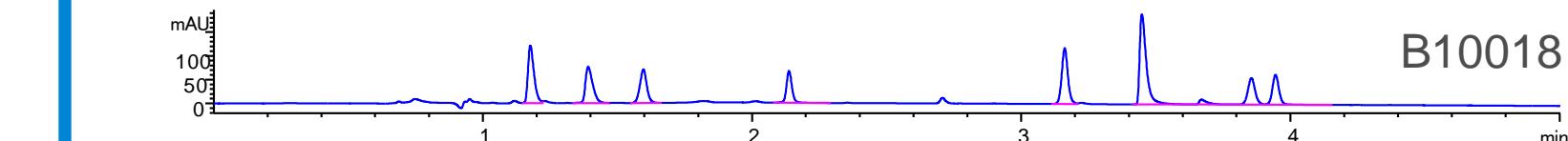
Batch-to-batch reproducibility of Poroshell 120 columns

2010



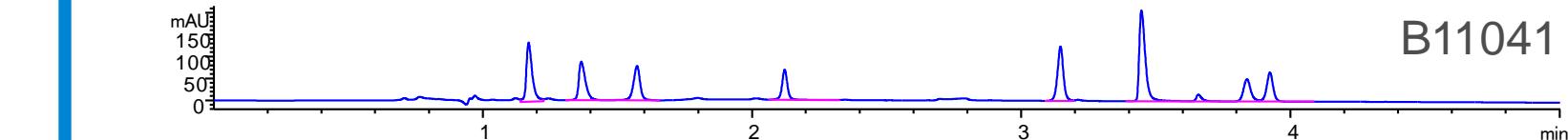
B10015

2012



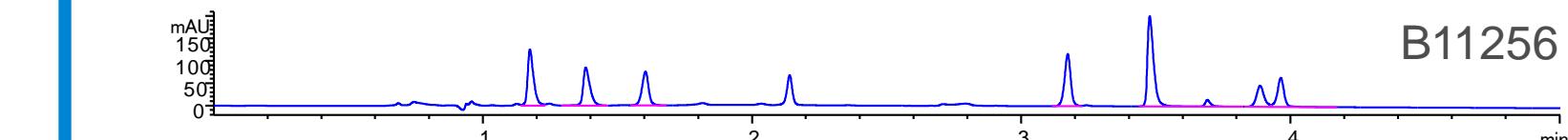
B10018

2010



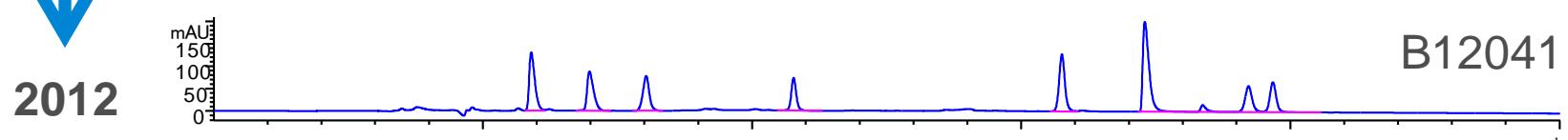
B11041

2012



B11256

2012



B12041

Beverage Additives



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# Method Validation Kits

## Agilent ZORBAX Rapid Resolution High Definition (RRHD) Method Validation Kits

Size (mm)	Particle Size ( $\mu\text{m}$ )	Eclipse Plus C18	Eclipse Plus C8	Eclipse XDB-C18	Extend-C18	Eclipse Plus Phenyl-Hexyl	Bonus-RP	SB-C18	SB-C8	SB-Phenyl	SB-Aq
3.0 x 150	1.8	959759-302K	959759-306K	981759-302K				859700-302K	859700-306K		
3.0 x 100	1.8	959758-302K	959758-306K	981758-302K	758700-302K	959758-312K		858700-302K	858700-306K	858700-312K	858700-314K
3.0 x 50	1.8	959757-302K	959757-306K	981757-302K	757700-302K	959757-312K		857700-302K	857700-306K	857700-312K	857700-314K
2.1 x 150	1.8	959759-902K	959759-906K	981759-902K	759700-902K	959759-912K	859768-901K	859700-902K	859700-906K	859700-912K	859700-914K
2.1 x 100	1.8	959758-902K	959758-906K	981758-902K	758700-902K	959758-912K	858768-901K	858700-902K	858700-906K	858700-912K	858700-914K
2.1 x 50	1.8	959757-902K	959757-906K	981757-902K	757700-902K	959757-912K	857768-901K	857700-902K	857700-906K	857700-912K	857700-914K

## Agilent ZORBAX Method Validation Kits

Size (mm)	Particle Size ( $\mu\text{m}$ )	Eclipse Plus C18	Eclipse Plus C8	Eclipse XDB-C18	Eclipse XDB-C8	Extend-C18	Eclipse Plus Phenyl-Hexyl	Bonus-RP	SB-Aq	SB-C18	SB-C8	SB-Phenyl
4.6 x 250	5	959990-902K	959990-906K	990967-902K	990967-906K	770450-902K	959990-912K	880668-901K	880975-914K	880975-902K	880975-906K	880975-912K
4.6 x 150	5	959993-902K	959993-906K	993967-902K	993967-906K	773450-902K		883668-901K	883975-914K	883975-902K	883975-906K	883975-912K
3.0 x 150	5	959993-302K										
4.6 x 250	3.5									884950-567K		
4.6 x 150	3.5	959963-902K	959963-906K	963967-902K	963967-906K	763953-902K	959963-912K	863668-901K	863953-914K	863953-902K	863953-906K	863953-912K
4.6 x 100	3.5	959961-902K	959961-906K	961967-902K	961967-906K	764953-902K	959961-912K	864668-901K	861953-914K	861953-902K	861953-906K	861953-912K
4.6 x 50	3.5	959943-902K	959943-906K	935967-902K	935967-906K	735953-902K	959943-912K	835668-901K	835975-914K	835975-902K	835975-906K	835975-912K
4.6 x 150	1.8	959994-902K							829975-914K	829975-902K	829975-906K	829975-912K
4.6 x 100	1.8	959964-902K	959964-906K	928975-902K	928975-906K	728975-902K	959964-912K	828668-901K	828975-914K	828975-902K	828975-906K	828975-912K
4.6 x 50	1.8	959941-902K	959941-906K	927975-902K	927975-906K	727975-902K	959941-912K	827668-901K	827975-914K	827975-902K	827975-906K	827975-912K
3.0 x 100	1.8				928975-306K			828668-301K				
3.0 x 50	1.8				927975-306K			827668-301K				
2.1 x 100	1.8				928700-906K							
2.1 x 50	1.8				927700-906K							

## Agilent Poroshell 120 Method Validation Kits

Size (mm)	Particle Size ( $\mu\text{m}$ )	EC-C18	EC-C8	Phenyl-Hexyl	SB-C18	SB-C8	SB-Aq	Bonus-RP
4.6 x 150	2.7	693975-902K	693975-906K	693975-912K	683975-902K	683975-906K	683975-914K	693968-901K
4.6 x 100	2.7	695975-902K	695975-906K	695975-912K	685975-902K	685975-906K	685975-914K	695968-901K
4.6 x 50	2.7	699975-902K	699975-906K	699975-912K	689975-902K	689975-906K	689975-914K	699968-901K
3.0 x 150	2.7	693975-302K	693975-306K	693975-312K	683975-302K	683975-306K	683975-314K	693968-301K
3.0 x 100	2.7	695975-302K	695975-306K	695975-312K	685975-302K	685975-306K	685975-314K	695968-301K
3.0 x 50	2.7	699975-302K	699975-306K	699975-312K	689975-302K	689975-306K	689975-314K	699968-301K
2.1 x 150	2.7	693775-902K	693775-906K	693775-912K	683775-902K	683775-906K	683775-914K	693768-901K
2.1 x 100	2.7	695775-902K	695775-906K	695775-912K	685775-902K	685775-906K	685775-914K	695768-901K
2.1 x 50	2.7	699775-902K	699775-906K	699775-912K	689775-902K	689775-906K	689775-914K	699768-901K

# Summary

## What to consider when choosing a column?

- Make sure you choose the best pore size
- Consider newer particle technologies
  - Poroshell 120 2.7 µm
  - Poroshell 120 4.0 µm
- Take advantage of selectivity changes
  - Mobile phase
  - pH
  - Bonded phase

**Thank you!**

[LC-column-support@agilent.com](mailto:LC-column-support@agilent.com)



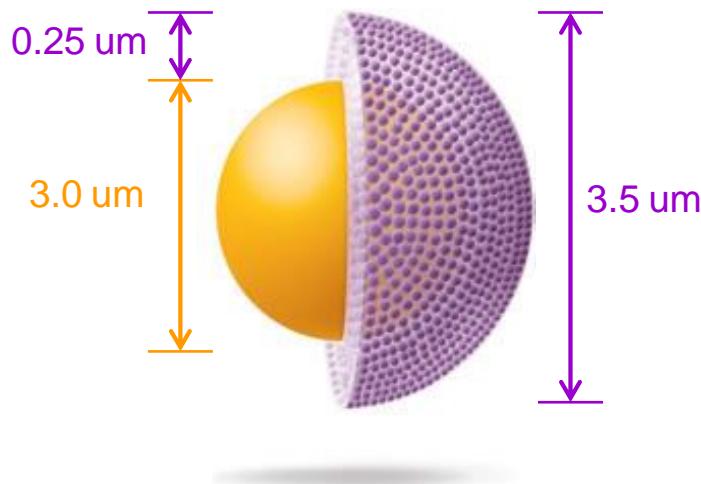
# AdvanceBio RP-mAb



## Particle

- 3.5 µm SP particle
- 0.25 µm porous layer depth
- 450Å pore diameter

The optimum large molecule resolution for use with both HPLC and UHPLC systems



## Phases

- C4
- SB-C8
- Diphenyl

The most popular phases for proteins, plus a choice for unique selectivity

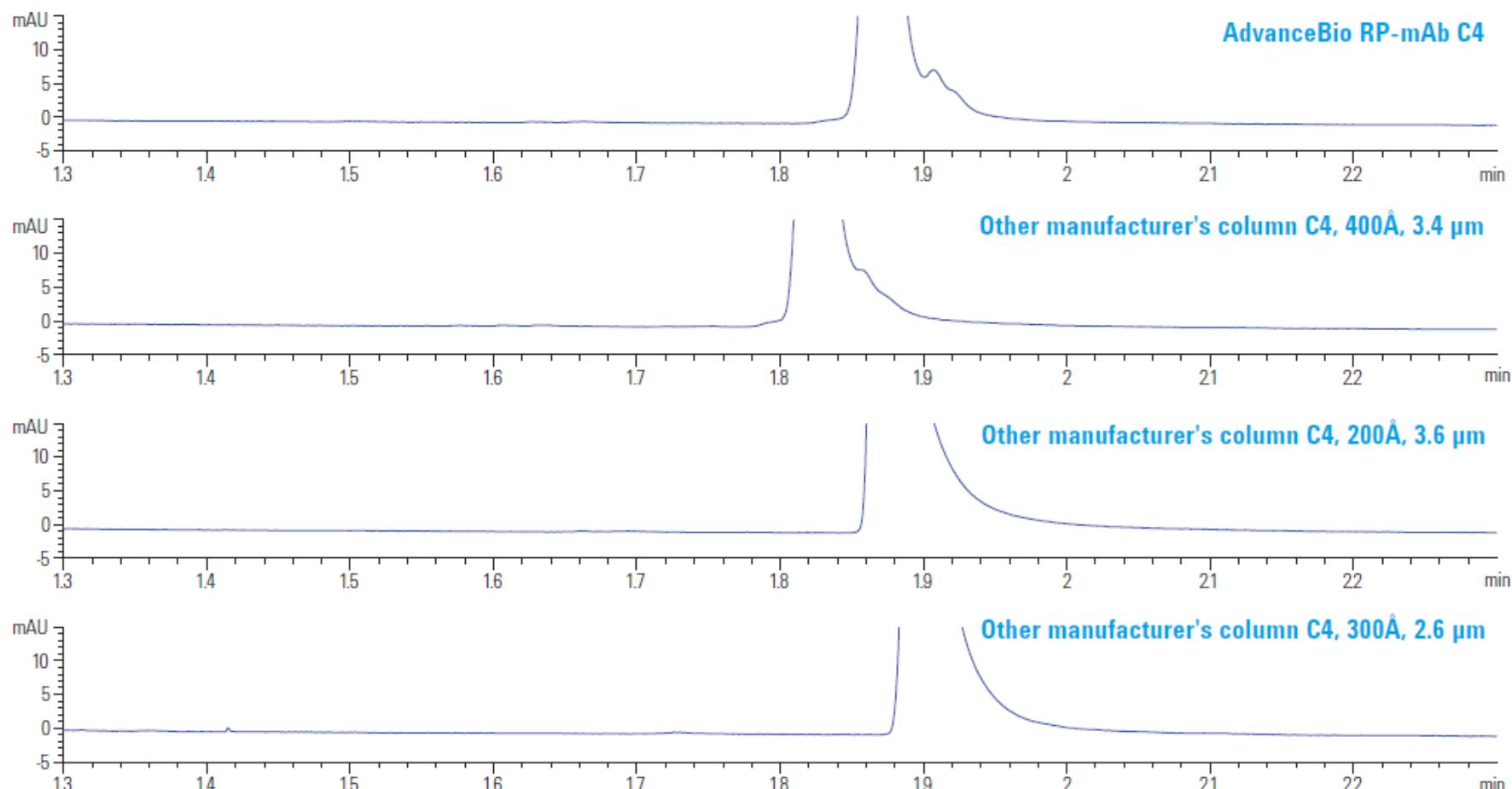


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April 8, 2015

# AdvanceBio RP-mAb



Column dimensions: 2.1 x 100 mm, 3.5 µm

Mobile phase A: 0.1% TFA in water:IPA (98:2)

Flow rate: 1.0 mL/min

Temperature: 80 °C

Sample: 5 µL injection of humanized recombinant Herceptin IgG1 intact from Creative Biolabs (1 mg/mL)

Mobile phase B: IPA:acetonitrile:MPA (70:20:10)

Gradient: 10-58% B in 4 min, 1 min wash at 95% B, 1 min re-equilibration at 10% B

Detection: UV @ 254 nm



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