

GC/MS Application Note





Determination of MCPD and glycidyl esters in foodstuff

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Short summary

- Highly efficient automation due to the PAL DHR Dual Head instrument under the CHRONOS control software. The system analyzes a sample in 45 minutes.
- AOCS Cd 29c-13 (or DGF C-VI 18(10)) is the most efficient method.
- The Clean-Technology provides excellent robustness of the system.

Introduction

3-Monochloropropane-1,2-diol (3-MCPD), 2-monochloropropane-1,3-diol (2-MCPD) and glycidol are known as contaminants in foodstuffs. MCPD fatty acids may occur during the refinement process of oils and fats at high temperature or in the presence of chloride containing salts. However, refinement is an essential art of the manufacturing process. It removes undesired odorants and flavoring substances as well as possible traces of toxic compounds (pesticides, heavy metals or mycotoxins). There is an increasing importance of the analysis of these contaminants due to their suspected carcinogenicity. Animal experiments revealed that uptake of free 3-MCPD is increasing the risk of cancer. In March 2016 the European Food Safety Authority (EFSA) declared a reduced value for the tolerable daily uptake of 0.8 µg/kg body weight. There are numerous methods for the determination of MCPD-esters (see references). They can be divided into two groups: the direct determination using LC-MS/MS or the indirect analytics by GC-MS. The direct analysis is more timeconsuming because every single ester must be determined by LC/MS. Therefore the indirect method is more frequently used for routine applications. In both cases the manual sample preparation is a time-consuming and error-prone process. Axel Semrau® therefore has developed methods for the automated sampe preparation following current methods.

Automation workflow

AOCS Cd 29c-13 or DGF C-VI 18(10) were selected as the most efficient methods. An evaporation step was left out, since it is not required for an accurate measurement. An additional step has been integrated with the in-house developed Clean-

Technology. In the process, the analytes which are bound to fatty acid esters are first converted into their free forms by transesterification.

According to the DGF method two parts are required for the determination of the amount of glycidol. The transesterification in part A is quenched by adding sodium chloride. Here, MCPD fatty acid esters and glycidyl esters are both converted into free MCPD. In part B the transesterification is quenched by the addition of a sodium bromide solution. In this case, only MCPD fatty acid esters are converted into free MCPD. In further steps, free MCPD is extracted and then derivatized. Subsequently, a cleaning step and measurement via GC-MS/MS is performed. The difference of the values determined in part A and B is then multiplied by an experimentally determined transformation coefficient (t). The result is the amount of glycidol in the foodstuff sample (figure 1).



Figure 1: Workflow for the sample preparation for the indirect analytics of MCPDs with the classical DGF method and DGF Fast & Clean

With this method one sample is analyzed in only 45 minutes (part A & B). As a consequence, the new system provides highest precision in combination with a significant time saving. The modular PAL3 system (www.palsystem.com) allows for the automation of other methods and therefore is perfectly suited for the automation of sample preparation. For example, the 3-in-1 method by SGS (Kuhlmann, AOCS Cd 29b-13) may be partially automated, but a short manual interaction still is required. For the Unilever method (AOCS Cd 29a-13) a centrifuge can be integrated on the PAL System. Also the Zwagerman-Overman method has been automated on a PAL System. Likewise the earlier Weisshaar method has been implemented.

Instrument setup

- PAL DHR dual head system with 160 cm x-rail with RTC/RSI head (see details in figure 2)
- CHRONOS Software
- Bruker EVOQTM GC-TQ with 456-GC (instrument parameters see table 1)

The PAL DHR with 160 cm x-rail offers the space required for all modules. Since PAL System is a platform concept, the instrument can be adapted to other methods or applications. The Dilutor Module delivers three solvents (n-hexane, extracting agent and iso-octane).

To avoid condensation of the derivatization reagent (phenylboronic acid) on the analytical column or ion source of MS, the GC system is equipped with a backflush option. This increases the uptime of the GC-MS to at least 200 injections and delivers robust data (see figure 3). Alternatively, the installation is possible with instruments of other manufacturers or on existing GC-MS devices.

The entire system is controlled by the user-friendly software CHRONOS, making even complex procedures easy to handle. The CHRONECT application solutions by Axel Semrau® are pre-installed in application laboratories and tested (Factory Acceptance Test). The systems are then delivered ready-to-use to the customer and put into operation at the customer's site (Site Acceptance Test).

Results

3-MCPD and 2-MPCD and the corresponding deuterated variants were detected using the triple quadrupole mass spectrometer. For each compound one ion was selected for qualification and one for quantification. The respective collision energies for the fragmentation of the parent ions were determined experimentally. Important parameters for the GC-MS device are shown in table 1.



3 Vortex Mixer

- 4 Standard Wash Station
- 5 Fast Wash Station

- 8 Park Stations (2 x)
- Not shown: Dilutor Tool/Module and Liquid Tools



Figure 3: Full scan TICs showing an injection with (green) and without Clean Technology (red). The removal of PBA greatly increases uptime of the GC-MS.

The DGF "classical" method was validated first. Samples were prepared according to the DGF guidelines and then analyzed. For validation, rac-1,2-Bis-palmitoyl-3-chloropropanediol und rac-1,3-Distearoyl-2-chloropropanediol were used. Defined amounts of the two compounds were added to a native olive oil and subsequently prepared using the PAL DHR system. Native olive oil is suitable as a blank matrix since it is pressed at low temperatures and therefore should not contain any MCPD esters. Control measurements confirmed this assumption with a blank value of ~ 0.02 mg/kg sample for 3-MCPD and 2-MCPD. The results of the validation experiments are given in Table 2.

Both, part A and B of the DGF method were validated. The measurements lead to a detection limit of 0.026 mg/kg sample (3 x S/N) with a limit of determination of 0.041 mg/kg sample (10 x S/N) for 3-MCPD. The same approach was used for the method DGF Fast and Clean. Here a reference oil (FAPAS) was analyzed additionally. The oil's 3-MCPD and 2-MCPD content was known from a interlaboratory test.

Injector:	SSL, 250°C; 1 μL injection volume, splitle	ss (split	1:30 afte	er 1 minute)		
Temperature GC (°C)	Heating rate (°C/min)			time (min)	Total (min)	
85.0					0.10	
200.0	200.0				1.68	
300.0	200.0				12.18	
Pressure control:	1.5 mL/min constant flow, backflush after 2 min					
GC column:	Rxi-5 MS , 0.25 mm inner diameter, 0.25 μm film;					
Oven program:						
Temperature (°C)	Heating rate (°C/min)	Holding time (min)		Total (min)		
80.0			1.00		1.00	
150.0	10.0	0.00		8.00		
320.0	30.0				23.67	
GC-MS:	Transfer line 280 °C, CID-gas argon, MRM-mode					
Compound:	Retention time (min)	Precur	sor ion	Product ion	Mode	
2-MCPD	7.71	198.00		104.00	Quantifier	
		196.00		104.00	Qualifier	
2-MCPD-d5	7.66	203.00		107.00	Quantifier	
		201.00		93.00	Qualifier	
3-MCPD	7.36	196.00		147.00	Quantifier	
		196.00		91.00	Qualifier	
3-MCPD-d5	7.32	201.00		150.00	Quantifier	
		201.00)	93.00	Qualifier	

Table 1: GC-MS parameters for the detection of 3-MCPD and 2-MCPD

	DGF "classical"		DGF Fast & Clean		
	Recovery	% SD	Recovery	% SD	
3-MCPD part A	102.6	3.9	91.6	7.7	
3-MCPD part B	94.3	3.9	101.9	8.8	
2-MCPD part B	133.2	6.0	116.2	8.9	

Table 2: Recovery (RC in %) and reproducibility (RP in % SD) values obtained from the methods DGF "classical" and DGF "fast" for part A and part B on four consecutive days.

Sample	Measurement	3-MCPD (mg/kg)	2-MCPD (mg/kg)	Glycidol (mg/kg)
Reference oil (FAPAS T2646QC)	manual	0.59	0.31	0.26
	DGF "classical"	0.49	0.30	0.23
	DGF Fast & Clean	0.50	0.38	0.36
Unkown	manual	0.78	0.39	0.64
cooking oil	DGF Fast & Clean	0.80	0.58	0.73
Democra de la	manual	0.14	< 0.10	0.10
Rapeseed on	DGF Fast & Clean	0.11	0.08	0.13
Cumfleringen eil III	manual	0.84	0.39	0.15
Sumower on-HL	DGF Fast & Clean	0.73	0.60	0.29
Sunflower oil-HO	manual	0.31	0.15	0.49
	DGF Fast & Clean	0.25	0.19	0.58

Table 3: Comparison between results obtained with manual, DGF "classical" and DGF Fast&Clean.



Figure 4: The chromatogram shows 0.05 mg/kg 3-MCPD in olive oil.

Figure 5: Overlay of chromatograms for 10 consecutive runs demonstrating excellent reproducibility of the automated measurements of 2- and 3-MCPD in different concentrations.

Discussion of results

Summary

The validation of the methods was performed according to standard procedures. Recovery and reproducibility were determined.

Recoveries between 91 and 133 % were found using DGF "classical" (Table 2).

For the method DGF Fast & Clean recoveries of 91 - 116 % were observed. Here, the values for 2-MCPD are above 100 % as well.

The data for this study were acquired on four consecutive days. Table 4 highlights the excellent comparability of DGF "Classical", DGF Fast & Clean and SGS 3-in-1.

DGF Fast & Clean provides, especially in comparison to the classical DGF method, a significant time saving. Overlapped processing enables the analysis of 36 samples in 24 hours. Applying this method, results are available after 45 minutes only, as shown in figure 7.

- The automation of the sample preparation for the analysis of MCPD and glycidol improves the sample throughput and reproducibility.
- The process presented can prepare and analyze a sample in a short period of time (45 minutes for a sample with part A and B). Axel Semrau® therefore recommends the AOCS Cd 29c-13 (or DGF C-VI 18(10)) as the most efficient method.
- The PAL DHR Dual Head System greatly increases the efficiency of the process. The two heads perform tasks in parallel and independently. CHRONOS optimizes throughput by calculating the most efficient allocation of instrument resources.
- The Clean-Technology for the GC-MS provides excellent robustness of the system.
- The fully automated application is perfectly suited for the routine analytics of 2- and 3-MCPD as well as glycidol in food control labs.

	Fat mixture 1		Fat mixture 2		Table 4: Results of a mix- ture of sunflower oil and
	3-MCPD-Ester (mg/kg)	Glycidyl-Ester (mg/kg)	3-MCPD-Ester (mg/kg)	Glycidyl-Ester (mg/kg)	rapeseed oil with DGF Fast & Clean in comparison with the manual DGF method
"DGF Fast &Clean" DGF C-VI 18 AOCS Cd 29c-13	0.14	0.05	0.11	<0.05	as well as the SGS 3-in-1 method
"DGF" manual DGF C-VI 18 AOCS Cd 29c-13	0.15	0.08	0.13	0.05	
"SGS 3- in 1- method"; manual AOCS Cd 29b-13	0.14	<0.05	0.10	<0.05	

1 Part A (automated) Part B (manual) 0,9 Part B (automated) 0,8 Part B (manual) 07 Glycidyl ester (automated) ≡ Glycidyl ester (manual) 0,6 [mg/kg] 0.5 0,4 0,3 0.2 0.1 0 Butter biscuits Shortbread biscuits Chocolate Chocolate eggs Milk powder Condensed milk Hummus

Figure 6: Results for 2- and 3-MCPD and glycidol content of selected foodstuffs.



Figure 7: Overlapping in CHRONOS boosts productivity : Interlaced analysis of Part A and Part B of one sample in 45 minutes.

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