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A simplified approach for nicotine quantification in electronic cigarette liquids using GC-Orbitrap mass spectrometry

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Keywords

Accurate mass, electronic cigarettes, e-liquids, gas chromatography, GC-MS, quantification, GC, high-resolution, Orbitrap mass spectrometry, nicotine

Goal

To demonstrate the performance of Thermo Scientific[™] Exactive[™] GC Orbitrap[™] GC-MS mass spectrometer for quantitative analysis of e-liquids components.

Introduction

Since the introduction of electronic cigarettes in 2007, their use has increased worldwide as an alternative to conventional tobacco cigarettes. Electronic cigarette liquids (e-liquids) can contain nicotine, a highly addictive drug that is toxic in high doses, may increase heart attack risk, and can affect reproductive health.¹ Studies have shown that nicotine can also interfere with memory and attention processing especially in adolescents.²

To protect human health, and to meet the obligations of the European Union under the WHO Framework Convention on Tobacco Control,³ the Tobacco Products Directive 2014/14/EU (TPD)⁴ in Article 20, contains rules for nicotine containing electronic cigarettes and refill containers. The Medicines and Healthcare products Regulatory Agency (MHRA),⁵ is responsible for implementing the majority of the provisions under Article 20, including the restrictions on the nicotine strength of no more than 20 mg/mL.

Quantitative assessments of current analytical technologies for e-liquids include LC and GC coupled to both analog detectors (GC-FID, LC-UV) as well as MS.⁶⁻¹¹ But many of these methods can lack the sensitivity and selectivity required. High mass resolution performance and exceptional mass accuracy make GC Orbitrap a powerful solution for both qualitative and quantitative analysis of e-liquids.



The aim of this work was to demonstrate the quantitative performance of the Exactive GC Orbitrap system for the analysis of nicotine in e-liquids samples.

Experimental

Sample and standard preparation

Flavored and flavorless e-liquid samples with specified nicotine levels of 0, 6, or 12 mg/mL were purchased locally and analyzed. Shortfill samples were also analyzed, which are supplied at 0 mg/mL specified nicotine level. Shortfills are e-liquids that can be purchased in bottles larger than the regulated limit of 10 mL, into which the user can add a nicotine shot prior to use. They are not regulated under TPD within the UK as they contain 0% nicotine upon purchase.

For target quantitative analysis of nicotine in e-liquid samples, liquid split/splitless sample injection was used. Each e-liquid sample was first diluted 50 μ L to 5 mL with acetonitrile (LC/MS grade), mixed, then further diluted taking 100 μ L of the diluted sample and 50 μ L of the internal standard (0.1 mg/mL, 8-hydroxyquinoline), then made up to 1 mL with acetonitrile in a GC vial ready for analysis.

Calibration standards (ranging from 46 to 13,792 ng/mL nicotine) were prepared in acetonitrile, diluting from a certified e-liquid standard acquired from LGC (Teddington, UK). In addition, the calibration standards contained an internal standard (8-hydroxyquinoline) to a final concentration of 5000 ng/mL.

Instrument and method setup

An Exactive GC Orbitrap mass spectrometer coupled with a Thermo Scientific[™] TRACE[™] 1310 gas chromatograph, configured with a Thermo Scientific[™] TriPlus[™] RSH[™] autosampler and a Thermo Scientific[™] Instant Connect split/splitless (SSL) injector, was used in all experiments.

Compound separation was achieved on a Thermo Scientific[™] TG-WaxMS 30 m x 0.25 mm i.d. x 0.25 µm film capillary column (P/N 26088-1420). The mass spectrometer was tuned and calibrated in <1.5 min using FC43 (CAS 311-89-7) to achieve mass accuracy of <0.5 ppm. The system was operated using electron ionization (El) mode, using full-scan and 60,000 mass resolution (full width at half maxima, measured at m/z 200). Additional details of the instrument parameters are shown in Tables 1 and 2.

Table 1. GC and injector conditions

TRACE 1310 GC system parameters								
Injection volume (µL):	1.0							
Liner:	Thermo Scientific [™] LinerGOLD [™] 4 mm i.d. 78.5 mm length (P/N 453A1255-UI)							
Inlet (°C):	260							
Carrier gas (mL/min):	He, 1.2							
Inlet module and mode:	SSL, split mode							
Split ratio:	10:1							
Purge flow (mL/min):	5.0							
Column:	TG-WaxMS 30 m x 0.25 mm i.d. x 0.25 µm film capillary column (Thermo Scientific [™] TraceGOLD [™] GC Column) (P/N 26088-1420							
Oven temperature program:	RT (min)	Rate (°C/min)	Target temperature (°C)	Hold time (min)				
Initial	0	-	40	3.00				
Final	3.00	13	250	6.00				
Run time	25	-	-	-				

Table 2. Mass spectrometer conditions

Exactive GC Orbitrap mass spectrometer parameters					
Transfer line (°C):	250				
lonization type:	EI				
lon source (°C):	230				
Electron energy (eV):	70				
Acquisition mode:	Full-scan				
Mass range (Da):	75–500				
Mass resolution:	60,000 FWHM at <i>m/z</i> 200				

Data processing

Data were acquired and processed using

Thermo Scientific[™] TraceFinder[™] software. TraceFinder single platform software integrates instrument control, method development functionality, and qualitative and quantitation data processing.

Results and discussion

The objective of this study was to evaluate the utility of Orbitrap-based GC-MS technology for quantitative analysis of e-liquids for nicotine, using direct liquid injection, accessing various analytical parameters including chromatographic resolution, instrument sensitivity, and linearity.

Chromatography

Good chromatographic separation was obtained using the GC conditions described in Table 1. The total ion chromatogram (TIC) for nicotine and 8-hydroxyquinoline (internal standard) are shown in Figure 1. In order to develop a quantitative workflow for nicotine analysis using TraceFinder software, a compound database (CDB) to store compound information was produced (see Figure 2). The overlaid extracted ion chromatograms (EICs) achieved for target and confirming ions for nicotine over the calibration range are shown in Figure 3.



Figure 1. Total ion chromatogram (TIC) for [1] nicotine and [2] 8-hydroxyquinoline (internal standard) in a standard at 13,792 ng/mL for nicotine and 5000 ng/mL for 8-hydroxyquinoline. For peaks at retentions times 12.12, 11.81, and 17.30 minutes, additional compound names were identified, based on the top NIST library search results achieved for the background subtracted mass spectra for each peak.

Compound Data	mpound Database - e-cig SSL*													
Tree View Pane	▼ ₽×	Peak	View Pane											
Compound Name			Compound	Peak Label II	Peak Workflow	п	Associated	п	m/7	Re	tention Time	RT Window (sec)	Target Ratio	п
Expand All	Collapse All	8-	Name	T can caber +	T car from to		Target Peak	Ŧ	10/2	- (m	in)		larger natio	T
All Results		1	8-hydroxy quinoline	T1: 145.05219	TargetPeak	•		•	145.05219	16	42	30.00	15	
4 8-hydroxy	quinoline 05219	2	8-hydroxy quinoline	T1C1: 117.05731	Confirming	•	T1: 145.05219	•	117.05731	16	42	30.00	15	
▲ T1C	1: 117.05731	3	8-hydroxy quinoline	T1C2: 89.03863	Confirming	•	T1: 145.05219	•	89.03863	16	42	30.00	15	
▲ Nicotine	2: 89.03863	4	Nicotine	T1: 84.08083	TargetPeak	•		•	84.08083	13	80	30.00	15	
▲ ▲T1: 84.0	8083	5	Nicotine	T1C1: 133.07614	Confirming	-	T1: 84.08083	-	133.07614	13	80	30.00	15	
T1C	1: 133.07614 2: 161.10740	6	Nicotine	T1C2: 161.10740	Confirming	•	T1: 84.08083	•	161.10740	13	80	30.00	15	

Figure 2. CDB for nicotine and 8-hydroxyquinolone, detailing the target and confirming ions (*m/z*), retention time (RT), RT window, and the target ion ratio for each compound



Figure 3. Overlaid EICs for target and confirming ions for [A] the lowest (46 ng/mL) and [B] the highest (13,792 ng/mL) calibration standards in the developed quantification method

Quantification and sensitivity

The quantitative performance of the Exactive GC Orbitrap GC-MS system was tested for nicotine. System sensitivity, linearity, and peak area repeatability were evaluated. Additionally, mass accuracy of the target compounds was assessed across the concentration ranges. Linearity was assessed using 11 calibration levels ranging from 46 to 13,792 ng/mL (equivalent to 0.046 to 13.79 mg/mL in the prepared e-liquid samples). Data was acquired using full scan, with compound detection based on retention time (±0.5 min window), accurate mass (±5 ppm window), and ion ratio of quantification vs. confirming ion (±15% window). Collecting full scan data enables the retrospective qualitative targeted or non-targeted screening of the collected data if required at a later date.¹² Nicotine was easily detected in the lowest calibration standard, 46 ng/mL (equivalent to 0.046 mg/mL in the prepared e-liquid samples).

Mass accuracy

Maintaining mass accuracy and spectral fidelity is critical for correct compound identification in potentially complex e-liquid samples. Figure 4 illustrates the mass accuracy achieved over the calibration range for nicotine; a mass accuracy of <1 ppm was achieved for each ion in the spectra.



Figure 4. Mass spectra for the lowest (46 ng/mL) and highest calibration (13,792 ng/mL) standards for nicotine. Consistent <1 ppm mass accuracy was obtained for each ion in the spectra. Annotated are the measured mass, the elemental composition, and the theoretical mass as well as the mass accuracy (ppm).

Peak area repeatability in matrix

In order to have confidence in the routine nicotine quantitation results achieved, the stability of responses in the matrix is critical. Repeatability of nicotine responses in matrix were assessed by carrying out n=3 repeat injections of two e-liquids with stated nicotine concentrations of 6 mg/mL. Excellent repeatability was obtained as shown in Table 3, with %RSD for quantification ions peak area counts between 3.7% and 3.9%.

1.6

1.2-

1.0-0.8-

0.6

0.4

0.2

0.0

Area 1.4

월 120000000

100000000

80000000

60000000

40000000

20000000

0-

Linearity of response

To assess compound linearity, 11 calibration levels (46 to 13,792 ng/mL) were quantified using an internal standard calibration, using a 1/x weighting factor. Excellent peak area repeatability for the internal standard (8-hydroquiniline) was acheived, as shown in Figure 5A, with a %RSD of 5.3. Excellent linearity was demonstrated for nicotine, with an R² value of 0.9991, an average residual %RSD of 4.4, and an amount deviation tolerance at each point of <10%. An example calibration curve for nicotine is shown in Figure 5B where both the coefficient of determination (R²) and the residual %RSD are annotated.

Table 3. Nicotine results summary for replicate injections (n=3) for two e-liquid samples with stated nicotine concentration of 6 mg/mL, f = branded flavored, and g = flavorless. The concentration results (mg/mL) are internal standard corrected.



4000 Figure 5. (A) Average peak area for 8-hydroxyquinline (internal standard), and (B) example calibration curve for nicotine, illustrating the linearity obtained, over 11 calibration levels ranging from 46 to 13,792 ng/mL (two injections per calibration level)

6000

8000

10000

12000

14000

2000

Quantitation of nicotine in e-liquid samples

Ten e-liquid samples were prepared and analyzed as detailed; concentrations of the nicotine identified are illustrated in Figure 6A. The samples analyzed were quantified using an internal standard calibration, with the internal standard added to all samples and standards to the same concentration. For the three e-liquids with specified nicotine concentrations, the results versus the specified nicotine concentrations are shown in Figure 6B. Many studies have shown that nicotine compared to the label claims,^{13,14} which is illustrated here with the deviations of the results from the claimed concentrations between -8% and +21%.

Α	Sample	Description	Declared nicotine concentration (mg/mL)	Nicotine concentration (mg/mL)
	а	Flavorless	0	<0.046
	b	Flavored (branded)	0	<0.046
	С	Flavored (branded)	0	<0.046
	d	Flavored (vanilla)	0	<0.046
	е	Flavored (mint)	0	<0.046
	f	Flavored (branded)	6	5.5
	g	Flavorless	6	7.3
	h	Flavored (lemon)	12	12.2
	i	Flavored (strawberry)	0	<0.046
	j	Flavored (lemon)	0	<0.046

В



Figure 6. (A) Calculated concentration of nicotine detected in the analyzed e-liquid samples, and (B) detected nicotine results versus declared value, for the three e-liquids with specified nicotine concentration

Conclusions

- The results of this study demonstrate that using Orbitrap-based GC-MS technology, with high mass resolution performance, and exceptional mass accuracy, provides excellent solutions for the quantitative analysis of e-liquids.
- In the targeted analysis of nicotine using liquid injection: linearity was demonstrated with R² = 0.999 and residual values RSD% = 4.4%, over 11 calibration levels ranging from 46 to 13,792 ng/mL (equivalent to 0.046 to 13.79 mg/mL in the prepared e-liquid samples); mass accuracy of <1 ppm was obtained for all ions in the nicotine spectra (from the lowest to the highest standard).
- Quantitative targeted analysis for nicotine in ten e-liquid samples, including flavored and flavorless, with declared nicotine levels of 0, 6, or 12 mg/mL, and two shortfill flavored e-liquid samples was performed. Replicate measurements for nicotine containing samples indicated excellent precision with %RSD < 3 achieved.

References

- United States Department of Health and Human Services. How Tobacco Smoke Causes Disease: The Biology and Behavioral Basis for Smoking-Attributable Disease: a report of the Surgeon General. Centers for Disease, Centers for Disease Control and Prevention, Atlanta, GA, USA, 2010.
- Goriounova, N.A.; Mansvelder, H.D. Short-and Long-Term Consequences of Nicotine Exposure during Adolescence for Prefrontal Cortex Neuronal Network Function. *Cold Spring Harbor Perspectives in Medicine*, v.2(12), 2012.
- World Health Organization Framework Convention on Tobacco Control signed in Geneva on 21 May 2003. [Online] http://apps.who.int/iris/bitstream/ handle/10665/42811/9241591013.pdf;sequence=1 (accessed Nov 15, 2018).
- The Tobacco Products Directive (2014/14/EU). [Online] https://ec.europa.eu/ health//sites/health/files/tobacco/docs/dir_201440_en.pdf (accessed Nov 15, 2018).
- E-cigarettes: regulations for consumer products. Medicines and Healthcare products Regulatory Agency. [Online] https://www.gov.uk/guidance/e-cigarettesregulations-for-consumer-products (accessed Nov 15, 2018).
- Flora, J. W.; Wilkinson, C. T.; Sink, K. M.; McKinney, D. L.; Miller, J. H. Nicotinerelated impurities in e-cigarette cartridges and refill e-liquids. *Journal of Liquid Chromatography & Related Technologies*, 2016, *39*(17-18), 821–829.
- Liu, X.; Joza, P.; Rickert, B. Analysis of nicotine and nicotine-related compounds in electronic cigarette liquids and aerosols by liquid chromatography-tandem mass spectrometry. *Beiträge zur Tabakforschung International/Contributions to Tobacco Research* 2017, *27*(7), 154–167.
- Aszyk, J.; Kubica P., Kot-Wasik, A.; Namiésnik, J.; Wasik, A. Comprehensive determination of flavouring additives and nicotine in e-cigarette refill solutions. Part I: liquid chromatography-tandem mass spectrometry analysis. *Journal of Chromatography A* 2017, *1519*, 45–54.
- Herrington, B. J. S.; Myers, C.; Rigdon, A. Analysis of Nicotine and Impurities in Electronic Cigarette Solutions and Vapor. Restek, State College, PA, USA, 2015
- CORESTA Recommended Method No. 62). Determination of Nicotine in Tobacco and Tobacco Products by Gas Chromatographic Analysis. 2005. [Online] https:// www.coresta.org/determination-nicotine-tobacco-and-tobacco-products-gaschromatographic-analysis-29185.html (accessed Dec 19, 2018).
- Ghalop, V.V.; Kosmider, L.; Halquist, M.S. A Standardized Approach to Quantitative Analysis of Nicotine in e-Liquids Based on Peak Purity Criteria using High-Performance Liquid Chromatography. *Journal of Analytical Methods in Chemistry* 2018, Article ID 1720375.
- Cooper, J.; Allen, C.; Cojocariu, C. Comprehensive chemical characterization of e-cigarette liquids using high resolution Orbitrap GC-MS. Thermo Scientific Application Note 10708, March 2019.
- Davis, B.; Dang, M.; Kim, J.; Talbot P. Nicotine concentrations in electronic cigarette refill and do-it-yourself fluids. *Nicotine & Tobacco Research* 2015, *17*(2), 134–141.
- Trehy, M. L.; Ye, W.; Hadwiger, M. E. et al. Analysis of electronic cigarette cartridges, refill solutions, and smoke for nicotine and nicotine related impurities. *Journal of Liquid Chromatography & Related Technologies* **2011**, *34*(14), 1442–1458.

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