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## The salivary microbiota in health and disease

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### ABSTRACT

The salivary microbiota (SM), comprising bacteria shed from oral surfaces, has been shown to be individualized, temporally stable and influenced by diet and lifestyle. SM reflects local bacterial alterations of the supragingival and subgingival microbiota, and periodontitis and dental-caries associated characteristics of SM have been reported. Also, data suggest an impact of systemic diseases on SM as demonstrated in patients with a wide variety of systemic diseases including diabetes, cancer, HIV and rheumatoid arthritis. The presence of systemic diseases seems to influence salivary levels of specific bacterial species, as well as  $\alpha$ - and  $\beta$ -diversity of SM. The composition of SM might thereby potentially mirror oral and general health status. The contentious development of advanced molecular techniques such as metagenomics, metatranscriptomics and metabolomics has enabled the possibility to address bacterial functions rather than presence in microbial samples. However, at present only a few studies have employed such techniques on SM to reveal functional and metabolic characteristics in oral health and disease. Future studies are therefore warranted to illuminate the possible impact of metabolic functions of SM on oral and general health status. Ultimately, such an approach has the possibility to reveal novel and personalized therapeutic avenues in oral and general medicine.

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### Introduction

Saliva is the fluid covering the surfaces of the oral cavity. Being instrumental in physiological processes such as mastication, swallowing and speech [1], saliva also harbors essential biological constituents including proteins and enzymes, which are essential for maintenance of oral homeostasis [2]. For example, salivary mucins and glycoproteins are the sole nutritional source in early plaque development [3], and salivary antimicrobials are critically involved in maintaining a symbiotic relationship between host and its resident microbiota [4]. This relationship is constantly stressed by internal and external ecological perturbations [5], and while the oral microbiota is resilient to minor ecological changes [6], prolonged perturbation can induce dysbiosis of the resident microbiota, which may lead to the two major oral diseases, i.e. periodontitis and dental caries [7].

By tradition microbiological analysis of patients with periodontitis has been performed on the subgingival microbiota, whereas in patients with caries, research has focused primarily on the supragingival plaque. In general medicine samples of stools are usually subject to analysis of the gastrointestinal microbiota. However, saliva is an easy and non-invasive alternative to such sampling strategies [8], and the salivary microbiota (SM) has been shown to reflect local bacterial alterations in supragingival and subgingival microbiotas [9,10]. Furthermore, oral



bacterial species are reported in gut microbial samples [11,12]. Therefore, saliva might be a feasible alternative to local samples in studies of the microbiota in oral and general health and disease [13].

The purpose of the present review is to report recent research on the bacterial part of SM in oral and general health and disease, and to discuss future perspectives for this line of research. The main focuses of the present review are on the two most prevalent oral diseases i.e. periodontitis and dental caries, as well as on diabetes and cancer. The literature included is limited to studies of the bacterial part of SM using molecular techniques.

### The salivary microbiota in oral health

Saliva is sterile when being secreted into the oral cavity [14]. However, when sampled a diverse microbiota is present in saliva [15]. Accordingly, SM has been shown to be a conglomerate of bacteria shed from oral surfaces with the throat, the tongue and the tonsils as the main sites of origin [16]. The SM has been demonstrated to be individualized [17,18] and temporarily stable [19,20] in orally healthy individuals. On the other hand, circadian oscillations of SM have also been documented [21].

The composition of SM in health is to some extent shaped by environmental factors [22]. Cross-sectional data suggest that the composition differentiates in individuals living under different climate conditions

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[23], and a recent longitudinal study showed alterations of SM in members during an Antarctic expedition [24]. Likewise, several studies have documented an impact of diet on SM [25–27].

The SM mirrors dentate status. Accordingly, the composition of SM has been reported to differentiate between dentate and edentulous individuals [28], and full-mouth extraction impacts on SM [29]. Furthermore, SM is affected by dental developmental stages [30,31], and early life development of SM is a coordinated process, influenced by ecological perturbations such as mode of delivery, breastfeeding length and antibiotic treatment [32].

## The salivary microbiota in oral disease

### Periodontitis

The SM has been widely compared in patients with periodontitis and orally healthy controls. A substantial part of the studies has employed polymerase chain reaction (PCR)-based techniques with special emphasis on the applicability of salivary levels of specific bacterial species as a biomarker of periodontitis.

Accordingly, a number of cross-sectional PCR-based studies have compared salivary levels of putative periodopathogens, especially *Porphyromonas gingivalis*, *Tannerella forsythia*, *Treponema denticola*, *Prevotella intermedia* and *Aggregatibacter actinomycetemmitans* in saliva from patients with periodontitis to those of orally healthy controls. A recent study from 2019 reported salivary levels of the JP2 clone of *A. actinomycetemmitans* to associate with clinical attachment loss in Moroccan adolescents [34], whereas a large-scale study comprising 977 Japanese individuals showed salivary levels of *P. gingivalis* to correlate with percentage of sites with probing pocket depth  $\geq 4$  mm [35]. In addition, a cross-sectional study of a Finnish population ( $n = 462$ ) documented that combined salivary levels of *P. gingivalis*, *P. intermedia* and *T. forsythia* were associated with periodontitis [36]. In addition, a recent study from 2019 demonstrated higher salivary levels of two hitherto uncultured bacterial species (*Fretibacterium* sp. human oral taxon 360 and *Fretibacterium* sp. human oral taxon 356) in patients with periodontitis as compared to orally healthy controls [37]. However, no comparison of subgingival and salivary levels of the selected bacteria was performed in the above-mentioned studies, which is why the origin of periodopathogens in saliva was unknown. Nevertheless, other studies have used PCR and next-generation sequencing (NGS) of the 16 rDNA gene to compare subgingival and salivary levels of putative periodopathogens. Taken together, these studies have demonstrated a strong correlation of subgingival and salivary levels of putative periodopathogens [38–41].

Moreover, several studies have aimed to differentiate patients with periodontitis from orally healthy controls by means of salivary levels of putative periodopathogens. For example, a recently published NGS-based study showed that relative abundance of *P. gingivalis* could discriminate patients with periodontitis from orally healthy controls with an AUC (area under curve) of 0.80 [42], and a PCR-based study of 9 selected periodopathogens reported that it was possible to discriminate the severity of periodontitis based on salivary levels of the bacteria tested [43]. Furthermore, salivary levels of periodopathogens have been used in periodontal risk assessment. For example, in a longitudinal study of 24 months duration, the combination of salivary levels of *P. gingivalis* and serum levels of *P. gingivalis*' IgG antibodies was associated with periodontal disease progression [44].

A few studies have used NGS to characterize the salivary microbiota in patients with periodontitis, and compare data with that of orally healthy controls. Accordingly, a recently published study in a Swedish cohort showed a significant periodontitis associated-microbiota with increased levels of *T. forsythia*, *Filifactor alocis* and *Parvimonas micra* [45]. Furthermore, several interventional studies using NGS have demonstrated an impact of non-surgical periodontal treatment on the composition of SM [9,46–48]. Interestingly, two of these studies showed a positive correlation of subgingival and salivary levels of putative periodopathogens before and after periodontal treatment [9,47]. Thus, data suggest that even though periodopathogens are occasionally found off the tongue [49], spillover of bacteria from the subgingival area are probably the primary site of origin of periodopathogens identified in saliva. This is why salivary levels of periodopathogens might be used as a biomarker of periodontitis.

### Dental caries

The SM has been characterized in patients with severe early childhood caries (SECC), as well as in adolescent and adult populations with dental caries. In all cases, data from patients with caries have been compared to that of age-matched orally healthy controls.

Recently, two NGS-based studies performed a cross-sectional comparison of SM in patients with SECC and children  $< 5$  yrs. Without caries, and both studies reported caries-associated characteristics of SM [50,51]. Notably, co-analysis of *Candida albicans* demonstrated that carriage of *C. albicans* in children with SECC attenuated the differences observed [51]. In 2018 three longitudinal studies on SM in children with dental caries were published [52–54]. One of these studies compared their findings in patients with recurrent caries ( $n = 7$ ) with those of patients with a history of caries ( $n = 6$ ) and caries-free

controls (n = 15). The main finding was that salivary levels of *Fusobacterium*, *Prevotella*, *Leptotrichia* and *Capnocytophaga* species could predict recurrent caries with an AUC = 0.95 [54]. Likewise, another study reported that the composition of SM in combination with information on salivary levels of host defense peptides could predict caries progression [53].

The SM in adolescents with dental caries has recently been compared to that of orally healthy adolescents [55,56]. Accordingly, an NGS-based retrospective cross-sectional study of a Swedish cohort (n = 62) showed significant caries-associated differences of SM. Specifically, higher salivary abundance of bacterial species, such as *Scardovia wiggsiae* and *Streptococcus mutans*, were identified in patients with caries. Furthermore, patients with salivary presence of *Bifidobacterium longum* had an increased caries risk [56]. In addition, a cross-sectional study of 154 adolescents confirmed distinct differences of SM in caries patients vs. healthy controls, which were partly driven by the co-occurrence of *S. wiggsiae* and *S. mutans* [55].

Increased levels of *Veillonella* and *Bifidobacterium* species were found in SM of adult patients with caries compared to that of healthy controls [57]. Likewise, a recently published study demonstrated caries-associated characteristics of SM in elderly patients with caries as compared to healthy controls [58].

Collectively, data from various NGS-based studies of the 16S rRNA gene have reported SM to differentiate patients with periodontitis and dental caries from that of orally healthy controls. Furthermore, SM differs in patients with periodontitis vs. patients with caries [59,60]. Thus, it seems that the presence of treatment requiring oral disease associates with characteristics of SM. The breakthrough of more advanced molecular techniques such as metagenomics, metatranscriptomics and metabolomics has enabled the possibility to add bacterial functions such as carbohydrate metabolism and proteolytic activity to their presence in microbial samples [61]. Accordingly, a few studies have described functional and metabolic characteristics of SM in oral health and disease [62,63]. Thus, future studies will have the possibility to focus on the possible impact of metabolic functions of the salivary microbiota as an etiological agent in periodontitis and dental caries. Such an approach may be important for the development of novel personalized therapeutic avenues.

## The salivary microbiota in systemic disease

### Diabetes and obesity

Poor oral health status associates with increased risk of systemic disease, and especially the bidirectional relationship of periodontitis and type 2 diabetes

(T2DM) have been described in detail [64]. Much focus has been paid on the role of low-grade inflammation as the link between oral and systemic diseases [65]. However, the potential impact of systemic disease on SM has also been addressed.

Several studies have compared SM in patients with diabetes to that of healthy controls by means of NGS, and in general, data show that diabetes associates with a decrease in bacterial diversity of SM [66–68]. In addition, higher salivary levels of *P. gingivalis*, *T. forsythia* and *F. alocis* were reported in patients with gestational diabetes [69], whereas only minor differences were identified in children with T2DM, when compared to obese and healthy controls, respectively [70].

Also, SM in obese individuals has been compared to that of lean controls. A metagenomic study from 2018 showed decreased bacterial diversity and richness in saliva from obese individuals. Furthermore, functional analysis documented higher bacterial expression of immune disease-related genes in obese individuals [71]. Likewise, obesity was reported to modify salivary bacterial diversity in patients with T2DM [72]. In addition, a PCR-based analysis demonstrated higher salivary levels of *P. gingivalis*, *T. forsythia* and *Fusobacterium nuclatum* in obese patients with and without T2DM, as compared to lean controls [73].

Finally, a potential impact of salivary glucose concentration on SM has been investigated. Specifically, DNA-DNA hybridization of >2500 saliva samples collected from Kuwaiti children showed high salivary glucose concentration to associate with a decrease in bacterial diversity [74], and NGS of SM revealed increased abundance of *Leptotrichia*, *Staphylococcus*, *Catonella* and *Bulleidia* species in individuals with impaired fasting glucose [75].

Thus, data suggest that T2DM, obesity and poorly controlled clearance of glucose i.e. impaired fasting glucose is associated with comparable impacts on SM.

### Cancer

The SM has been studied in patients with oral squamous cell carcinoma (OSCC). One large-scale report characterized SM in 138 patients with OSCC by means of NGS and compared data with that of saliva from 151 matched healthy controls. The main finding was higher salivary abundance of periodontitis-associated species, such as *Prevotella tanneriae*, *F. nucleatum*, and *P. intermedia* in patients with OSCC [76]. Likewise, two studies have reported SM to differentiate in patients with oral leukoplakia [77] and patients with other epithelial precursor lesions to OSCC [78], when compared to that of orally healthy controls. It is, however, more interesting that specific alterations of SM were present in samples from OSCC patients versus patients

with leukoplakia and epithelial precursor lesions. Specifically, higher salivary levels of *Solobacterium* species and lower levels of *Streptotoccus* species were recorded in OSCC patients compared to leukoplakia patients [77], whereas salivary abundance of *Bacillus*, *Enterococcus*, *Parvimonas*, *Peptostreptococcus* and *Slackia* species discriminated patients with OSCC from patients with epithelial precursor lesions [78]. Thus, cross-sectional data suggest that SM might be used in screening of OSCC. However, one study has shown that the microbiota identified in OSCC lesions is different from the concomitant SM in patients with OSCC [79]. Thus, prospective data are warranted to evaluate if the specific strains of SM are causally involved in the development of OSCC.

The effect of OSCC treatment i.e. radiotherapy and chemotherapy on SM has been longitudinally evaluated. One study showed that radiotherapy was associated with an increase in salivary levels of *Lactobacillus* species, which was reversed to baseline levels 1 year after radiotherapy. Interestingly, strong correlation was observed in salivary levels of *Lactobacillus* species and fluctuations of saliva flow rates and salivary pH levels during radiotherapy [80]. Another report demonstrated that chemotherapy-induced oral mucositis was associated with a decrease in salivary levels of health-associated bacterial genera, including *Streptococcus*, *Actinomyces* and *Veillonella* in combination with an increase of the periodontitis-associated genera *Fusobacterium* and *Prevotella*, and increased transcription of genes related to innate immunity and apoptosis in oral epithelial cells from patients with oral mucositis [81].

Cross-sectional studies have evaluated SM in patients with cancers outside the oral cavity. Accordingly, high salivary levels of *T. forsythia* and *A. actinomycescomitans* and low bacterial diversity were reported in patients with precancerous gastric lesions [82]. On the other hand, gastrointestinal cancers of various origins were associated with an increased salivary bacterial diversity and high levels of *P. gingivalis* as compared to matched healthy controls [83]. In line, a study from 2019 used NGS of SM to discriminate patients with throat cancer from healthy controls and patients with vocal cord polyps with an AUC = 0.87 [84]. Finally, two cross-sectional studies have compared SM in patients with lung cancer to healthy controls. The main findings were higher levels of *Streptococcus* and *Veillonella* species in saliva from patients with non-small cell lung cancer [85], and lower salivary levels of *Streptococcus* species in combination with low bacterial diversity in female non-smokers with lung cancer as compared to matched healthy controls [86].

Therefore, data suggest that various non-oral cancers are associated with different alterations of SM. Data is, however, based solely on cross-sectional

studies, which hampers the possibility to draw any conclusions on causality at present.

### Other systemic diseases

Periodontitis has several important comorbidities such as rheumatoid arthritis and atherosclerosis [65]. Interestingly, SM has been reported to differ in patients with such comorbidities as compared to healthy controls. For example, rheumatoid arthritis associated with dysbiosis of SM with depletion of *Haemophilus* species in saliva, dental plaque and fecal samples, which was partly normalized by treatment of rheumatoid arthritis [87]. In addition, salivary levels of four periopathogens, i.e. *P. gingivalis*, *T. denticola*, *T. forsythia* and *P. intermedia*, have been suggested to be independently involved in lowering serum levels of high-density lipoproteins, which may be associated with an increased risk of atherosclerosis [88].

Immune defects have an impact on SM. Accordingly, immune deficiency as expressed by the manifest human immunodeficiency virus (HIV) has been reported to influence  $\alpha$ - and  $\beta$ -diversity of SM [89]. Furthermore, increased salivary levels of periopathogenic species including *Prevotella melanogenica* and *Rothia mucilaginosa* were shown to correlate with the extent of CD4 + T cell depletion in patients with HIV [90].

In several of the above-mentioned studies, correlation changes to the salivary and fecal microbiotas were evident [87,89], which highlight the possibility to use saliva-based screening as a substitute to fecal samples in microbiologic studies of systemic diseases. An example of such an approach was published in a recent longitudinal study, which linked dysbiosis of SM during the first 7 years of life with development of allergy [32].

### Future perspectives

The accumulated evidence suggests that SM is individualized and relatively stable over time as long as oral and general health is maintained. In addition, local bacterial changes associated with periodontitis and dental caries are reflected by alterations of SM. Furthermore, presence of systemic disease appears to have an impact on SM. Thus, SM seems to reflect oral and general health status. However, future studies are needed to reveal if changes of SM precede clinical signs of disease, which would enable the possibility to use SM in the prediction of future disease risk. Ideally, this could be performed targeting subtypes or strains of specific bacterial species in SM such as *P. gingivalis* or *A. actinomycescomitans* [91] in periodontitis or *S. mutans* in caries. An elegant example of such an approach was recently published in a study showing that analysis of *S. mutans* in saliva based on adhesion subtypes could be used for future caries risk

prediction [92]. It is also interesting that in a recent cross-sectional study from 2017 data showed that orally healthy individuals can be divided into five ecotypes based on characteristics of SM, the salivary metabolome and host-related biochemical salivary parameters [93]. In addition, a recent longitudinal study in hospitalized cancer patients demonstrated that increased variability of SM was associated with adverse clinical outcomes [94]. Notably, these studies demonstrate the possibility to use SM in risk assessment and treatment evaluation of oral and systemic disease. Accordingly, prospective longitudinal studies are urgently needed to reveal the full potential of using SM in the field of precision medicine.

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No potential conflict of interest was reported by the author.

### References

- [1] Pedersen A, Sorensen CE, Proctor GB, et al. Salivary functions in mastication, taste and textural perception, swallowing and initial digestion. *Oral Dis.* 2018;24(8):1399–1416.
- [2] Lynge Pedersen AM, Belstrom D. The role of natural salivary defences in maintaining a healthy oral microbiota. *J Dent.* 2019;80(Suppl 1):S3–S12.
- [3] Jakubovics NS. Saliva as the sole nutritional source in the development of multispecies communities in dental plaque. *Microbiol Spectr.* 2015;3:3.
- [4] Marsh PD, Do T, Beighton D, et al. Influence of saliva on the oral microbiota. *Periodontol 2000.* 2016;70(1):80–92.
- [5] Marsh PD. In sickness and in health-what does the oral microbiome mean to us? An ecological perspective. *Adv Dent Res.* 2018;29(1):60–65.
- [6] Rosier BT, Marsh PD, Mira A. Resilience of the oral microbiota in health: mechanisms that prevent dysbiosis. *J Dent Res.* 2018;97(4):371–380.
- [7] Sanz M, Beighton D, Curtis MA, et al. Role of microbial biofilms in the maintenance of oral health and in the development of dental caries and periodontal diseases. Consensus report of group 1 of the Joint EFP/ORCA workshop on the boundaries between caries and periodontal disease. *J Clin Periodontol.* 2017;44(Suppl 18):S5–S11.
- [8] Bhattarai KR, Kim HR, Chae HJ. Compliance with saliva collection protocol in healthy volunteers: strategies for managing risk and errors. *Int J Med Sci.* 2018;15(8):823–831.
- [9] Belstrom D, Grande MA, Sembler-Moller ML, et al. Influence of periodontal treatment on subgingival and salivary microbiotas. *J Periodontol.* 2018a;89(5):531–539.
- [10] Belstrom D, Sembler-Moller ML, Grande MA, et al. Impact of oral hygiene discontinuation on supragingival and salivary microbiomes. *JDR Clin Trans Res.* 2018b;3(1):57–64.
- [11] Franzosa EA, Morgan XC, Segata N, et al. Relating the metatranscriptome and metagenome of the human gut. *Proc Natl Acad Sci U S A.* 2014;111(22):E2329–E2338.
- [12] Schmidt TS, Hayward MR, Coelho LP, et al. Extensive transmission of microbes along the gastrointestinal tract. *Elife.* 2019;8:e42693.
- [13] Belibasakis GN, Bostanci N, Marsh PD, et al. Applications of the oral microbiome in personalized dentistry. *Arch Oral Biol.* 2019;104:7–12.
- [14] Schroder SA, Bardow A, Eickhardt-Dalboe S, et al. Is parotid saliva sterile on entry to the oral cavity? *Acta Otolaryngol.* 2017;137(7):762–764.
- [15] Hasan NA, Young BA, Minard-Smith AT, et al. Microbial community profiling of human saliva using shotgun metagenomic sequencing. *PLoS One.* 2014;9(5):e97699.
- [16] Segata N, Haake SK, Mannon P, et al. Composition of the adult digestive tract bacterial microbiome based on seven mouth surfaces, tonsils, throat and stool samples. *Genome Biol.* 2012;13(6):R42.
- [17] Hall MW, Singh N, Ng KF, et al. Inter-personal diversity and temporal dynamics of dental, tongue, and salivary microbiota in the healthy oral cavity. *NPJ Biofilms Microbiomes.* 2017;3:2.
- [18] Leake SL, Pagni M, Falquet L, et al. The salivary microbiome for differentiating individuals: proof of principle. *Microbes Infect.* 2016;18(6):399–405.
- [19] Cameron SJ, Huws SA, Hegarty MJ, et al. The human salivary microbiome exhibits temporal stability in bacterial diversity. *FEMS Microbiol Ecol.* 2015;91(9):fiv091.
- [20] Van GS, Rohrig N, Schattenberg F, et al. A cytometric approach to follow variation and dynamics of the salivary microbiota. *Methods.* 2018;134–135:67–79.
- [21] Takayasu L, Suda W, Takanashi K, et al. Circadian oscillations of microbial and functional composition in the human salivary microbiome. *DNA Res.* 2017;24(3):261–270.
- [22] Shaw L, Ribeiro ALR, Levine AP, et al. The human salivary microbiome is shaped by shared environment rather than genetics: evidence from a large family of closely related individuals. *MBio.* 2017;8:5.
- [23] Li J, Quinque D, Horz HP, et al. Comparative analysis of the human saliva microbiome from different climate zones: Alaska, Germany, and Africa. *BMC Microbiol.* 2014;14:316.
- [24] Bhushan B, Yadav AP, Singh SB, et al. Diversity and functional analysis of salivary microflora of Indian Antarctic expeditionaries. *J Oral Microbiol.* 2019;11(1):1581513.
- [25] De FF, Vannini L, La SA, et al. The same microbiota and a potentially discriminant metabolome in the saliva of omnivore, ovo-lacto-vegetarian and Vegan individuals. *PLoS One.* 2014;9(11):e112373.
- [26] Hansen TH, Kern T, Bak EG, et al. Impact of a vegan diet on the human salivary microbiota. *Sci Rep.* 2018;8(1):5847.
- [27] Lassalle F, Spagnoletti M, Fumagalli M, et al. Oral microbiomes from hunter-gatherers and traditional farmers reveal shifts in commensal balance and pathogen load linked to diet. *Mol Ecol.* 2018;27(1):182–195.
- [28] Gazdeck RK, Fruscione SR, Adami GR, et al. Diversity of the oral microbiome between dentate and edentulous individuals. *Oral Dis.* 2019;25(3):911–918.

- [29] De Waal YC, Winkel EG, Raangs GC, et al. Changes in oral microflora after full-mouth tooth extraction: a prospective cohort study. *J Clin Periodontol.* 2014;41(10):981–989.
- [30] Crielaard W, Zaura E, Schuller AA, et al. Exploring the oral microbiota of children at various developmental stages of their dentition in the relation to their oral health. *BMC Med Genomics.* 2011;4:22.
- [31] Mason MR, Chambers S, Dabdoub SM, et al. Characterizing oral microbial communities across dentition states and colonization niches. *Microbiome.* 2018;6(1):67.
- [32] Dzidic M, Collado MC, Abrahamsson T, et al. Oral microbiome development during childhood: an ecological succession influenced by postnatal factors and associated with tooth decay. *Isme J.* 2018;12(9):2292–2306.
- [33] Dzidic M, Abrahamsson TR, Artacho A, et al. Oral microbiota maturation during the first 7 years of life in relation to allergy development. *Allergy.* 2018;73(10):2000–2011.
- [34] Ennibi OK, Claesson R, Akkaoui S, et al. High salivary levels of JP2 genotype of *Aggregatibacter actinomycetemcomitans* is associated with clinical attachment loss in Moroccan adolescents. *Clin Exp Dent Res.* 2019;5(1):44–51.
- [35] Chigasaki O, Takeuchi Y, Aoki A, et al. A cross-sectional study on the periodontal status and prevalence of red complex periodontal pathogens in a Japanese population. *J Oral Sci.* 2018;60(2):293–303.
- [36] Salminen A, Kopra KA, Hyvarinen K, et al. Quantitative PCR analysis of salivary pathogen burden in periodontitis. *Front Cell Infect Microbiol.* 2015;5:69.
- [37] Khemwong T, Kobayashi H, Ikeda Y, et al. *Fretibacterium* sp. human oral taxon 360 is a novel biomarker for periodontitis screening in the Japanese population. *PLoS One.* 2019;14(6):e0218266.
- [38] Belstrom D, Sembler-Moller ML, Grande MA, et al. Microbial profile comparisons of saliva, pooled and site-specific subgingival samples in periodontitis patients. *PLoS One.* 2017b;12(8):e0182992.
- [39] Haririan H, Andrukhov O, Bertl K, et al. Microbial analysis of subgingival plaque samples compared to that of whole saliva in patients with periodontitis. *J Periodontol.* 2014;85(6):819–828.
- [40] He J, Huang W, Pan Z, et al. Quantitative analysis of microbiota in saliva, supragingival, and subgingival plaque of Chinese adults with chronic periodontitis. *Clin Oral Investig.* 2012;16(6):1579–1588.
- [41] Nickles K, Scharf S, Rollke L, et al. Comparison of two different sampling methods for subgingival plaque: subgingival paper points or mouthrinse sample? *J Periodontol.* 2017;88(4):399–406.
- [42] Damgaard C, Danielsen AK, Enevold C, et al. *Porphyromonas gingivialis* in saliva associates with chronic and aggressive periodontitis. *J Oral Microbiol.* 2019;11(1):1653123.
- [43] Kim EH, Joo JY, Lee YJ, et al. Grading system for periodontitis by analyzing levels of periodontal pathogens in saliva. *PLoS One.* 2018b;13(11):e0200900.
- [44] Morozumi T, Nakagawa T, Nomura Y, et al. Salivary pathogen and serum antibody to assess the progression of chronic periodontitis: a 24-mo prospective multicenter cohort study. *J Periodontal Res.* 2016;51(6):768–778.
- [45] Lundmark A, Hu YOO, Huss M, et al. Identification of salivary microbiota and its association with host inflammatory mediators in periodontitis. *Front Cell Infect Microbiol.* 2019;9:216.
- [46] Chen C, Hemme C, Beleno J, et al. Oral microbiota of periodontal health and disease and their changes after nonsurgical periodontal therapy. *Isme J.* 2018;12(5):1210–1224.
- [47] Kageyama S, Takeshita T, Asakawa M, et al. Relative abundance of total subgingival plaque-specific bacteria in salivary microbiota reflects the overall periodontal condition in patients with periodontitis. *PLoS One.* 2017;12(4):e0174782.
- [48] Yamanaka W, Takeshita T, Shibata Y, et al. Compositional stability of a salivary bacterial population against supragingival microbiota shift following periodontal therapy. *PLoS One.* 2012;7(8):e42806.
- [49] Tanner AC, Paster BJ, Lu SC, et al. Subgingival and tongue microbiota during early periodontitis. *J Dent Res.* 2006;85(4):318–323.
- [50] Hurley E, Barrett MPJ, Kinirons M, et al. Comparison of the salivary and dental microbiome of children with severe-early childhood caries to the salivary microbiome of caries-free children. *BMC Oral Health.* 2019;19(1):13.
- [51] Xiao J, Grier A, Faustoferra RC, et al. Association between oral candida and bacteriome in children with severe ECC. *J Dent Res.* 2018;97(13):1468–1476.
- [52] Kim BS, Han DH, Lee H, et al. Association of salivary microbiota with dental caries incidence with dentine involvement after 4 years. *J Microbiol Biotechnol.* 2018a;28(3):454–464.
- [53] Simon-Soro A, Sherriff A, Sadique S, et al. Combined analysis of the salivary microbiome and host defence peptides predicts dental disease. *Sci Rep.* 2018;8(1):1484.
- [54] Zhu C, Yuan C, Ao S, et al. The predictive potentiality of salivary microbiome for the recurrence of early childhood caries. *Front Cell Infect Microbiol.* 2018;8:423.
- [55] Eriksson L, Lif HP, Esberg A, et al. Microbial Complexes and Caries in 17-Year-Olds with and without streptococcus mutans. *J Dent Res.* 2018;97(3):275–282.
- [56] Eriksson L, Lif HP, Johansson I. Saliva and tooth biofilm bacterial microbiota in adolescents in a low caries community. *Sci Rep.* 2017;7(1):5861.
- [57] Zhou J, Jiang N, Wang S, et al. Exploration of human salivary microbiomes—insights into the novel characteristics of microbial community structure in caries and caries-free subjects. *PLoS One.* 2016;11(1):e0147039.
- [58] Jiang Q, Liu J, Chen L, et al. The oral microbiome in the elderly with dental caries and health. *Front Cell Infect Microbiol.* 2018;8:442.
- [59] Belstrom D, Paster BJ, Fiehn NE, et al. Salivary bacterial fingerprints of established oral disease revealed by the human oral microbe identification using next generation sequencing (HOMINGS) technique. *J Oral Microbiol.* 2016;8:30170.
- [60] Takeshita T, Kageyama S, Furuta M, et al. Bacterial diversity in saliva and oral health-related conditions: the Hisayama Study. *Sci Rep.* 2016;6:22164.
- [61] Takahashi N. Oral microbiome metabolism: from “Who are they?” to “What are they doing?”. *J Dent Res.* 2015;94(12):1628–1637.

- [62] Belstrom D, Constancias F, Liu Y, et al. Metagenomic and metatranscriptomic analysis of saliva reveals disease-associated microbiota in patients with periodontitis and dental caries. *NPJ Biofilms Microbiomes*. 2017a;3:23.
- [63] Liebsch C, Pitchika V, Pink C, et al. The saliva metabolome in association to oral health status. *J Dent Res*. 2019;98(6):642–651.
- [64] Borgnakke WS, Ylostalo PV, Taylor GW, et al. Effect of periodontal disease on diabetes: systematic review of epidemiologic observational evidence. *J Clin Periodontol*. 2013;40(Suppl 14):S135–S152.
- [65] Holmstrup P, Damgaard C, Olsen I, et al. Comorbidity of periodontal disease: two sides of the same coin? An introduction for the clinician. *J Oral Microbiol*. 2017;9(1):1332710.
- [66] Ogawa T, Honda-Ogawa M, Ikebe K, et al. Characterizations of oral microbiota in elderly nursing home residents with diabetes. *J Oral Sci*. 2017;59(4):549–555.
- [67] Sabharwal A, Ganley K, Miecznikowski JC, et al. The salivary microbiome of diabetic and non-diabetic adults with periodontal disease. *J Periodontol*. 2019;90(1):26–34.
- [68] Saeb ATM, Al-Rubeaan KA, Aldosary K, et al. Relative reduction of biological and phylogenetic diversity of the oral microbiota of diabetes and pre-diabetes patients. *Microb Pathog*. 2019;128:215–229.
- [69] Gogeneni H, Buduneli N, Ceyhan-Ozturk B, et al. Increased infection with key periodontal pathogens during gestational diabetes mellitus. *J Clin Periodontol*. 2015;42(6):506–512.
- [70] Janem WF, Scannapieco FA, Sabharwal A, et al. Salivary inflammatory markers and microbiome in normoglycemic lean and obese children compared to obese children with type 2 diabetes. *PLoS One*. 2017;12(3):e0172647.
- [71] Wu Y, Chi X, Zhang Q, et al. Characterization of the salivary microbiome in people with obesity. *PeerJ*. 2018;6:e4458.
- [72] Tam J, Hoffmann T, Fischer S, et al. Obesity alters composition and diversity of the oral microbiota in patients with type 2 diabetes mellitus independently of glycemic control. *PLoS One*. 2018;13(10):e0204724.
- [73] Al-Rawi N, Al-Marzooq F. The relation between periodontopathogenic bacterial levels and resistin in the saliva of obese type 2 diabetic patients. *J Diabetes Res*. 2017;2017:2643079.
- [74] Goodson JM, Hartman ML, Shi P, et al. The salivary microbiome is altered in the presence of a high salivary glucose concentration. *PLoS One*. 2017;12(3):e0170437.
- [75] Wang RR, Xu YS, Ji MM, et al. Association of the oral microbiome with the progression of impaired fasting glucose in a Chinese elderly population. *J Oral Microbiol*. 2019b;11(1):1605789.
- [76] Hsiao JR, Chang CC, Lee WT, et al. The interplay between oral microbiome, lifestyle factors and genetic polymorphisms in the risk of oral squamous cell carcinoma. *Carcinogenesis*. 2018;39(6):778–787.
- [77] Hashimoto K, Shimizu D, Hirabayashi S, et al. Changes in oral microbial profiles associated with oral squamous cell carcinoma vs leukoplakia. *J Investig Clin Dent*. 2019;10(4): e12445.
- [78] Lee WH, Chen HM, Yang SF, et al. Bacterial alterations in salivary microbiota and their association in oral cancer. *Sci Rep*. 2017;7(1):16540.
- [79] Zhang Z, Yang J, Feng Q, et al. Compositional and functional analysis of the microbiome in tissue and saliva of oral squamous cell carcinoma. *Front Microbiol*. 2019b;10:1439.
- [80] Muller VJ, Belibasakis GN, Bosshard PP, et al. Change of saliva composition with radiotherapy. *Arch Oral Biol*. 2019;106:104480.
- [81] Hong BY, Sobue T, Choquette L, et al. Chemotherapy-induced oral mucositis is associated with detrimental bacterial dysbiosis. *Microbiome*. 2019;7(1):66.
- [82] Sun J, Zhou M, Salazar CR, et al. Chronic periodontal disease, periodontal pathogen colonization, and increased risk of precancerous gastric lesions. *J Periodontol*. 2017;88(11):1124–1134.
- [83] Kageyama S, Takeshita T, Takeuchi K, et al. Characteristics of the salivary microbiota in patients with various digestive tract cancers. *Front Microbiol*. 2019;10:1780.
- [84] Wang L, Yin G, Guo Y, et al. Variations in oral microbiota composition are associated with a risk of throat cancer. *Front Cell Infect Microbiol*. 2019a;9:205.
- [85] Zhang W, Luo J, Dong X, et al. Salivary microbial dysbiosis is associated with systemic inflammatory markers and predicted oral metabolites in non-small cell lung cancer patients. *J Cancer*. 2019a;10(7):1651–1662.
- [86] Yang J, Mu X, Wang Y, et al. Dysbiosis of the salivary microbiome is associated with non-smoking female lung cancer and correlated with immunocytochemistry markers. *Front Oncol*. 2018;8:520.
- [87] Zhang X, Zhang D, Jia H, et al. The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment. *Nat Med*. 2015;21(8):895–905.
- [88] Choi Y-H, Kosaka T, Ojima M, et al. Relationship between the burden of major periodontal bacteria and serum lipid profile in a cross-sectional Japanese study. *BMC Oral Health*. 2018;18(1):77.
- [89] Jimenez-Hernandez N, Serrano-Villar S, Domingo A, et al. Modulation of saliva microbiota through prebiotic intervention in HIV-infected individuals. *Nutrients*. 2019;11:6.
- [90] Lewi T, Hong B-Y, Weiser B, et al. Oral microbiome in HIV-infected women: shifts in abundance of pathogenic and beneficial bacteria are associated with aging, HIV load, CD4 count, and antiviral therapy. *Aids Res Hum Retroviruses*. 2019;35(3):276–286.
- [91] Johansson A, Claesson R, Höglund Åberg C, et al. Genetic profiling of *Aggregatibacter actinomycetemcomitans* Serotype B isolated from periodontitis patients living in Sweden. *Pathogens*. 2019;8(3):ii: E153.
- [92] Esberg A, Sheng N, Mårell L, et al. *Streptococcus Mutans* adhesin biotypes that match and predict individual caries development. *EBio Med*. 2017;24:205–215.
- [93] Zaura E, Brandt BW, Prodan A, et al. On the ecosystemic network of saliva in healthy young adults. *Isme J*. 2017;11(5):1218–1231.
- [94] Galloway-Pena JR, Smith DP, Sahasrabhojane P, et al. Characterization of oral and gut microbiome temporal variability in hospitalized cancer patients. *Genome Med*. 2017;9(1):21.