

## Expected complete data log-likelihood and EM

In our EM algorithm, the expected complete data log-likelihood (“ $Q$ ”) is a function of a set of model parameters  $\tau$ , i.e.

$$Q(\tau) = \sum_{m=1}^M \left( \sum_{z_m, l_m} \log(f(b_m, r_m, g_m | z_m, l_m, \tau)) p_m^*(z_m, l_m) \right),$$

where  $M$  is the total marker number,  $m$  is the SNP marker index,  $b_m$  is the observed BAF,  $r_m$  is the observed LRR,  $g_m$  is the error-free genotype,  $z_m = (z_{m1}, z_{m2})$  is ordered haplotype cluster memberships,  $l_m$  is the aberration type,  $\tau$  is the model parameters set,  $p_m^*(z_m, l_m) \equiv p(z_m, l_m | \tau^*, b, r, g)$  is the conditional marginal distribution, given parameter estimates  $\tau^*$ . We further assume that conditioned on  $(z_m, l_m)$ ,  $r_m$  and  $(g_m, b_m)$  are independent (see Materials and Methods). Thus

$$Q(\tau) = \sum_{m=1}^M \left( \sum_{z_m, l_m} (\log(f(r_m | l_m, \tau)) + \log(f(b_m, g_m | z_m, l_m, \tau))) p_m^*(z_m, l_m) \right).$$

We maximize  $Q$  at each EM cycle by solving the equation that sets to zero its partial derivative w.r.t. each parameter. For some parameters, a closed-form solution is available; for others, a numerical method must be applied.

In our experience, when the tumor mixture is high (e.g. above 10%), we can approximate the M-step by maximizing  $Q$  w.r.t. each individual parameter in  $\tau$  marginally, rather than maximizing in a multivariate manner. However, for extreme low tumor purity (e.g. about 3%), to avoid convergence problems, we must take the approach of expected conditional maximization (ECM), meaning we have to re-compute the posterior probability of latent states with the updated estimates after maximizing each parameter. The computation is more expensive with ECM.

## Estimation of the mixture proportion

The derivative of  $Q$  w.r.t. tumor DNA mixture proportion ( $w$ ) is composed of the following two summations involving derivatives of BAF and LRR densities respectively:

$$\begin{aligned} \frac{\partial}{\partial w} Q(w) = & \sum_{m=1}^M \left( \sum_{z_m, l_m} \left( \frac{\partial}{\partial w} \log(f(r_m | l_m, \tau)) \right) p_m^*(z_m, l_m) \right) + \\ & \sum_{m \in \{i \text{ st } g_i = 1\}} \left( \sum_{z_m, l_m} \left( \frac{\partial}{\partial w} \log(f(b_m, g_m | z_m, l_m, \tau)) \right) p_m^*(z_m, l_m) \right), \end{aligned} \quad (1)$$

where  $M$  is the total number of SNP makers, and the inner sum is over all combinations of  $z$  and  $l$ . Since BAFs are informative at heterozygous sites only (germline homozygous sites

have the derivative of zero w.r.t.  $w$ ), the second summation in equation (1) is limited to germline heterozygous sites.

We assume LRRs follow the same normal distribution as defined in GPHMM except for the addition of a sample-specific scale factor, i.e.

$$f(r|l, w, o_r, \sigma_r^2, q) = \frac{1}{\sigma_r} \phi \left( \frac{r - \mu^{(r)}(l, w, q) - o_r}{\sigma_r} \right),$$

where

$$\mu^{(r)}(l, w, q) \equiv q \cdot \log_2 \frac{(1-w)2 + w(\alpha(l) + \beta(l))}{2}, \text{ and}$$

$\phi$  is pdf of the standard normal distribution,  $l$  the latent aberration type,  $\sigma_r^2$  the variance,  $o_r$  the global baseline shift, and  $q$  the LRR scale. The functions  $\alpha(l_m)$  and  $\beta(l_m)$  have domains on the state space of  $l$  and give parent-specific allele copy numbers. The derivative in the first summation of equation [1] is

$$\begin{aligned} \frac{\partial}{\partial w} \log(f(r_m|l_m, \tau)) = \\ \frac{(r_m - o_r - q \log_2 \frac{(1-w)2 + w(\alpha(l_m) + \beta(l_m))}{2})}{\sigma_r^2} \cdot \frac{q(-1 + 0.5(\alpha(l_m) + \beta(l_m)))}{\log_e(2)}. \end{aligned}$$

We focus on low purity samples, where the perturbed BAF will remain relatively close to one-half and the truncation of BAFs at 0 or 1 (for heterozygotes) is of minimal concern. Thus, at germline heterozygous sites, we assume the potentially mixed BAF is distributed as

$$f(b|h, l, w, o_b, \sigma_b^2) = \frac{1}{\sigma_b} \phi \left( \frac{b - \mu^{(b)}(h, l, w) - o_b}{\sigma_b} \right),$$

where  $\phi$  is the pdf of the standard normal distribution,  $\sigma_b^2$  is the variance of BAF,  $o_b$  is a global baseline shift,  $h$  is the inherited allele configuration (either "AB" or "BA") and

$$\mu^{(b)}(h, l, w) \equiv \frac{0.5w(\beta(l) - \alpha(l))(-1)^{\mathbb{1}(h="AB")}}{(1-w)2 + w(\alpha(l) + \beta(l))} + 0.5.$$

For simplicity, we subtract 0.5 from observed BAFs, then we can drop 0.5 from  $\mu^{(b)}(h, l, w)$  expression and it has opposite signs for allele configurations "AB" and "BA". The derivative in the second summation of equation (1) is

$$\frac{\partial}{\partial w} \log f(b_m, g_m = 1 | z_m = (j, k), l_m, w) = \frac{1}{\sigma_b^2} \left( (b_m - o_b) \frac{1 - \Omega_m}{1 + \Omega_m} - \mu_m^{AB} \right) \frac{\partial}{\partial w} \mu_m^{AB},$$

where

$$\Omega_m \equiv \exp\left(\frac{-2b_m\mu_m^{AB}}{\sigma_b^2}\right) \frac{p(h_m = \text{"BA"}|z_m = (j, k))}{p(h_m = \text{"AB"}|z_m = (j, k))} = \exp\left(\frac{-2b_m\mu_m^{AB}}{\sigma_b^2}\right) \frac{\theta_{jm}(1 - \theta_{km})}{\theta_{km}(1 - \theta_{jm})},$$

$$\mu_m^{AB} \equiv \mu^{(b)}(h_m = \text{"AB"}, l_m, w) = \frac{-0.5(\alpha(l_m) - \beta(l_m))w}{(1 - w)2 + (\alpha(l_m) + \beta(l_m))w},$$

$$\frac{\partial}{\partial w}\mu_m^{AB} = \frac{-\alpha(l_m) + \beta(l_m)}{((\alpha(l_m) + \beta(l_m) - 2)w + 2)^2}, \text{ and}$$

$\theta_{im}$  is the probability that allele is "B" given haplotype cluster membership is  $i$  at maker  $m$ , as defined in fastPHASE model [1].

After substituting the two derivatives in equation (1), we do not have a closed-form solution. Therefore we rely on numerical root-finding methods. In practice, we use the secant method with previous  $w$  estimates as initial values.

## Estimation of BAF global baseline shift ( $o_b$ )

The derivative of  $Q$  w.r.t.  $o_b$  is

$$\frac{\partial}{\partial o_b}Q(o_b) = \sum_{m \in \{i \text{ st } g_i=1\}} \left( \sum_{z_m, l_m} \frac{\partial}{\partial o_b} \log(f(b_m, g_m|z_m, l_m, \tau)) p_m^*(z_m, l_m) \right).$$

Therefore, the new estimate of  $o_b$  is

$$\hat{o}_b = \frac{1}{M^{het}} \sum_{m \in \{i \text{ st } g_i=1\}} \sum_{z_m, l_m} \left( b_m - \mu_m^{AB} \frac{1 - \Omega_m}{1 + \Omega_m} \right) p_m^*(z_m, l_m),$$

where  $M^{het}$  is the number of germline heterozygous SNP markers.

## Estimation of BAF variance ( $\sigma_b^2$ )

The derivative of  $Q$  w.r.t.  $\sigma_b^2$  is

$$\frac{\partial}{\partial \sigma_b^2}Q(\sigma_b^2) = \sum_{m \in \{i \text{ st } g_i=1\}} \left( \sum_{z_m, l_m} \frac{\partial}{\partial \sigma_b^2} \log(f(b_m, g_m|z_m, l_m, \tau)) p_m^*(z_m, l_m) \right).$$

And using the normality assumption for BAF distribution,

$$\frac{\partial}{\partial \sigma_b^2} \log(f(b_m, g_m = 1|z_m = (j, k), l_m, w)) =$$

$$\frac{1}{2\sigma_b^4} \left( -\sigma_b^2 + (b_m - o_b)^2 + (\mu_m^{AB})^2 - 2(b_m - o_b)(\mu_m^{AB}) \frac{1 - \Omega_m}{1 + \Omega_m} \right).$$

We apply numerical root-finding method to obtain the new estimate.

## Estimation of variance and global baseline shift for LRR ( $\sigma_r^2$ , $o_r$ )

It is easy to show that the solutions that maximize  $Q$  w.r.t.  $\sigma_r^2$  and  $o_r$  are the following expressions:

$$\hat{o}_r = \frac{1}{M} \sum_{m=1}^M \sum_{l_m} \left( r_m - \mu^{(r)}(l_m, w) \right) p_m^*(l_m)$$

and

$$\hat{\sigma}_r^2 = \frac{1}{M} \sum_{m=1}^M \sum_{l_m} \left( r_m - \mu^{(r)}(l_m, w) - o_r \right)^2 p_m^*(l_m),$$

where  $p_m^*(l_m) = \sum_{z_m} p_m^*(z_m, l_m)$ .

## Estimation of LRR scale coefficient ( $q$ )

It has been pointed out that amplitude of LRR varies from sample to sample and that the observed amplitude is usually smaller than the standard value  $\log_2\left(\frac{\text{tumor copy number}}{2}\right)$  [2]. In GAP, this is modeled with a simple coefficient of contraction that is specific to the sample. GPHMM models the expected LRR as

$$\mu^{(r)}(l, w) \equiv 2\log_{10}(2) \cdot \log_2 \left( \frac{\text{average allele copy number in mixture}}{2} \right).$$

In our model, extra flexibility is achieved by replacing the constant  $2\log_{10}(2)$  in GPHMM with a LRR scale parameter ( $q$ ) and the new estimate for updating  $q$  is

$$\hat{q} = \frac{\sum_{m=1}^M \sum_{z_m, l_m} p_m^*(z_m, l_m) (r_m - o_r) \log_2 \frac{(1-w)2 + w(\alpha(l_m) + \beta(l_m))}{2}}{\sum_{m=1}^M \sum_{z_m, l_m} p_m^*(z_m, l_m) \left( \log_2 \frac{(1-w)2 + w(\alpha(l_m) + \beta(l_m))}{2} \right)^2}.$$

## Estimation of a GC content coefficient

Local GC content may induce a “wave” effect in the LRR data [3]. Therefore adjusting for GC content can reduce the noise in LRR signal, as demonstrated in GPHMM [4]. Similar to GPHMM, we use average GC-percentage in a 1Mb window around each SNP maker.

Let  $x_m$ , ( $m = 1 \cdots M$ ) denote the average GC content at marker  $m$  and  $t$  a global coefficient for GC content. Then we can re-write the density for LRR data as

$$f(r_m | x_m, l_m, w, o_r, \sigma_r^2, q, t) = \frac{1}{\sigma_r} \phi \left( \frac{r - \mu^{(r)}(l_m, w, q) - o_r - t \cdot x_m}{\sigma_r} \right).$$

It is easy to show the estimate for  $t$  is

$$\hat{t} = \frac{\sum_{m=1}^M \sum_{z_m, l_m} p_m^*(z_m, l_m)(r_m - o_r - \mu^{(r)}(l_m, w, q))x_m}{\sum_{m=1}^M \sum_{z_m, l_m} p_m^*(z_m, l_m)x_m^2}.$$

The above estimations for rest of the parameters remain valid if we replace  $r_m$  with  $r_m - t \cdot x_m$ .

## Identification of over-represented allele in tumor DNA

After the EM algorithm converges, the latent aberration state and haplotype cluster membership at marker  $m$  has joint posterior probability  $p^c(z_m, l_m) = p(z_m, l_m | g, r, b, \nu, \hat{t})$ . We then compute the probability that the allele ‘‘B’’ is over-represented at a germline heterozygous marker  $m$  as follows:

$$\begin{aligned} \sum_{z_m, l_m} p(\text{‘‘B’’ is over-represented} | z_m, l_m) p_m^c(z_m, l_m) = \\ \sum_{z_m, l_m} \sum_{h_m \in \{(A,B), (B,A)\}} \mathbb{1}\{\text{‘‘B’’ is over-presented} | h_m, l_m\} p(h_m | z_m) p_m^c(z_m, l_m), \end{aligned}$$

where  $\mathbb{1}\{\cdot\}$  is an indicator function. The probability for the allele ‘‘A’’ can be similarly obtained.

## Mean copy of haplotype cluster in tumor DNA

It is possible that a causal factor is correlated with a particular haplotype background, either due to an untyped ‘‘causal’’ germline allele well tagged by a haplotype or to a ‘‘haplotype effect’’ itself. Therefore it may be helpful to test the association of phenotypes with the mean copy number of a haplotype cluster. Suppose we obtain the posterior probability  $p_m^c(z_m, l_m)$  as defined above, the mean copy of haplotype cluster  $k$  at marker  $m$  is

$$\sum_{z_m, l_m} (\mathbb{1}\{z_{m1} = k\} \alpha(l_m) + \mathbb{1}\{z_{m2} = k\} \beta(l_m)) p_m^c(z_m, l_m),$$

where  $z_m = (z_{m1}, z_{m2})$ .

## References

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