

PERSPECTIVE

Immunotherapy in the Precision Medicine Era: Melanoma and Beyond

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Abbreviations: APC, antigen-presenting cell; BCG, bacillus Calmette-Guérin; CD28, cluster of differentiation 28; CTLA-4, cytotoxic T-lymphocyte-associated protein 4; EGFR, epidermal growth factor receptor; FDA, Food and Drug Administration; MAPK, mitogen-activated protein kinase; PARP, poly(ADP-ribose) polymerase; PD-1, programmed cell death protein 1; PD-L1, programmed cell death-ligand 1; TCR, T cell

Innovations in cancer immunotherapy over the past decade have reinvigorated the field, which currently stands poised to transform treatment of certain important cancer types. While recent discoveries in immunotherapy have occurred in an era of intense focus on precision medicine—spurred in large part by advances in sequencing technologies allowing for unprecedented insights into individual and tumor genomes—the field of cancer immunotherapy is rooted in a history that is anything but precise. The idea that the immune system could be used to target cancer has existed for over a century. In the 1890s, William Coley provided a spark by noting a connection between spontaneous tumor regressions and concurrent bacterial infections [1]. Suspecting an immune mechanism, Coley and many others in the decades that followed experimented with live bacteria, toxins, vaccines, and countless other agents to coax the immune system to attack cancer, yielding only a tiny number of positive responses and little understanding of the underlying immune mechanisms. As recently as 2010, immunotherapy remained a strikingly imprecise endeavor, with the only United States Food and Drug Administration (FDA)-approved cancer immunotherapies being a pair of systemic cytokines—interferon- α and interleukin-2—and intravesical bacillus Calmette-Guérin (BCG). However, as recent successes reshape the field, efforts to comprehend and harness precise mechanisms in immunotherapy will be required.

Breakthroughs in Melanoma

Perhaps the most profound impacts of both immunotherapy and precision medicine, in the form of targeted therapy, can be appreciated in the transformation of treatment for metastatic melanoma during the past decade. The discovery of the oncogenic V600E driver mutation in BRAF, which is present in ~50% of melanomas, led to the development of vemurafenib and dabrafenib, which are inhibitors of the mutant BRAF kinase [2]. Although targeting mutant BRAF kinase and inhibiting the downstream mitogen-activated protein kinase (MAPK) pathway produces high response rates in melanoma patients, the duration of responses is brief [3], highlighting the need for additional therapies. Meanwhile, preclinical evidence suggested that blocking negative regulators, or checkpoints, of T cell responses could improve antitumor immune responses (Fig 1). The first such negative regulator tested was cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), a T cell surface receptor that binds with high affinity to the costimulatory ligands B7-1 and B7-2 found on the surface of antigen-presenting cells. The interaction with CTLA-4 prevents binding of the costimulatory ligands to cluster of differentiation 28 (CD28), a major costimulatory receptor on the T cell necessary for a robust T cell response. Despite concerns about uncontrollable autoimmunity resulting from systemic

receptor; VEGF, vascular endothelial growth factor; VEGFR, vascular endothelial growth factor receptor.

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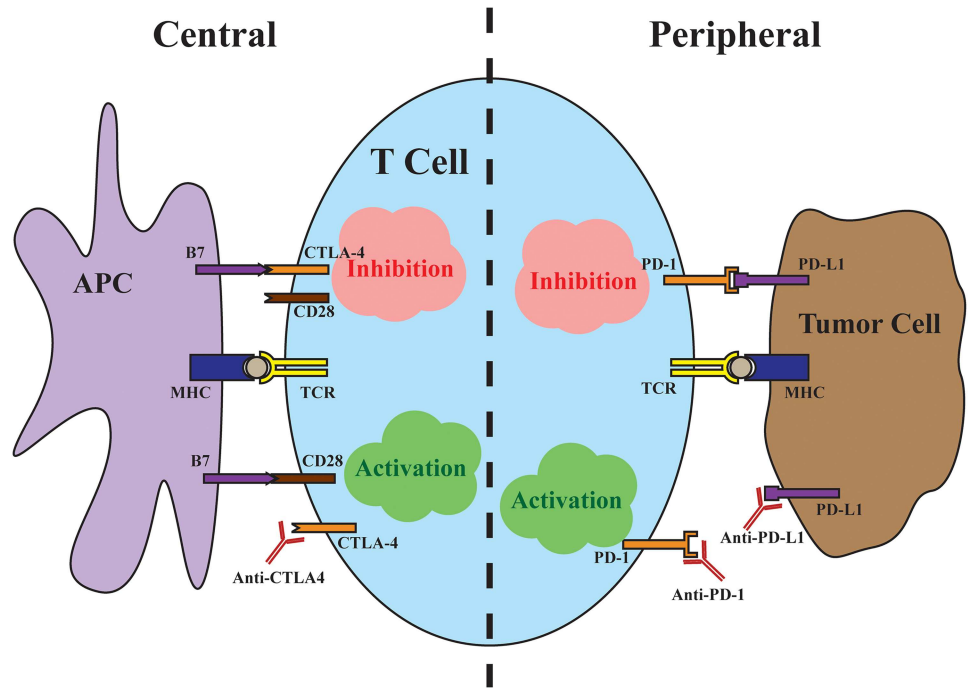


Fig 1. Central and peripheral immune checkpoints. Central immune inhibition occurs as a result of CTLA-4 binding to the costimulatory B7 ligands found on antigen-presenting cells (APCs) with high affinity, preventing signaling through CD28. Antibodies targeting CTLA-4 release this checkpoint to allow T cell activation. Peripheral immune inhibition occurs through binding of the PD-L1 found on tumor cells to the PD-1 receptor on T cells. Antibodies targeting either the ligand or the receptor release this checkpoint.

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CTLA-4 blockade, ipilimumab, a monoclonal antibody against CTLA-4, improved overall survival in patients with metastatic melanoma [4].

The successes of ipilimumab were quickly followed by trials targeting the immune inhibitory interaction between programmed cell death protein 1 (PD-1), found on T cells, and its ligand, PD-L1, found on tumor cells. In contrast to the central immune inhibitory effects of CTLA-4, inhibition through PD-1 predominantly occurs at the periphery, with tumor cells up-regulating PD-L1 in response to local immune signals such as interferon- γ . In a major head-to-head clinical trial, pembrolizumab, a monoclonal antibody against PD-1, demonstrated improved progression-free and overall survival with fewer high-grade adverse events compared to ipilimumab [5]. Importantly, in a minority of cases, checkpoint inhibition produces strikingly durable responses, with up to a third of advanced melanoma patients alive at five years of follow-up [6]. The majority, unfortunately, fail to respond to immunotherapy, with objective response rates to checkpoint inhibitor monotherapy ranging from 10%–40%, with PD-1/PD-L1 targeted antibodies benefiting larger proportions of patients [4,5,7]. Because the mechanisms of CTLA-4 and PD-1 blockade are distinct and potentially complementary, dual checkpoint blockade increases response rates to around 60%, albeit with significantly more adverse events [8,9]. In a matter of a few years, immune checkpoint blockade has become first-line therapy for advanced melanoma and transformed the outlook for patients. Looking ahead, uncovering predictive markers of response to immunotherapy and ultimately understanding the mechanisms of response will be necessary to extend the promise of immunotherapy to broader groups of cancer patients.

Efforts to uncover the precise mechanisms of immune checkpoint inhibition and identify patients likely to benefit from treatment are well under way. Pathologists have long recognized the prognostic significance of tumor-infiltrating lymphocytes [10], and, unsurprisingly, pre-treatment CD8+ T cell density is associated with favorable response to immune checkpoint blockade [11]. However, preclinical data suggest that CD8+ T cells in the tumor microenvironment can promote immune inhibition as a result of interferon- γ -mediated induction of PD-L1 on tumor cells [12]. Response to anti-PD1 therapy in melanoma as well as other cancers has been suggested to positively correlate with tumor expression of PD-L1 [13,14], highlighting the central role of this ligand in tumor immune evasion, although PD-L1 expression thus far appears to be an imperfect predictive biomarker of response. A comprehensive understanding of immunotherapy requires a deeper analysis of the interaction between immune system and tumor.

Genomics-Guided Advances

The rapid progress in cancer genomics in recent years has enabled a close examination of the genetic makeup of thousands of individual cancers across the entire spectrum of major cancer types. Amidst the search for mutations responsible for driving carcinogenesis, it has become evident that overall mutation rates vary dramatically both within and between cancer types [15]. While the vast majority of mutations contribute little, if at all, to intrinsic biological function in tumor development and survival, they represent novel, “non-self” epitopes that, if processed and presented, carry the potential to elicit a tumor-specific immune response [16]. By mining exome sequencing data to predict neoantigens present in a tumor, several groups have demonstrated a strong correlation between total neoantigen burden and positive response to immune checkpoint blockade [17,18]. These insights provide a mechanistic rationale to expand checkpoint inhibition to mismatch repair-deficient cancers of any type, because the dramatically elevated mutation rates in these cancers produce significantly more neoantigens and the potential for a more robust immune response [19].

Although the importance of neoantigens is now firmly established, substantial overlap in neoantigen burden exists between nonresponders and responders, highlighting the need to more precisely identify the tumor-specific antigens required for a successful immune response. Emerging evidence suggests that clonal neoantigens—that is, neoantigens present in all cancer cells—rather than total neoantigen load are critical for T cell recognition and clinical benefit from checkpoint blockade [20]. The critical neoantigens also need not be shared between patients. One targeted approach with the potential to improve and increase responses to checkpoint inhibition is to produce personalized vaccines specific to the neoantigens present in a patient’s tumor [16,21].

However, selecting and manufacturing the optimal neoantigen vaccines within a clinically meaningful time frame remains challenging, and successful targeting of tumor neoantigens requires antigen-specific T cells, which may not be present. Indeed, the efficacy of CTLA-4 blockade likely stems in part from an expansion of the peripheral T cell receptor (TCR) repertoire [22], which presumably includes neoantigen-specific TCRs. Nascent experimental and computational methods may help to resolve the critical TCR–antigen interactions in each tumor [23,24], providing the critical blueprints for delivering precision immunotherapy.

As the precise mechanisms of immune checkpoint inhibition continue to be unraveled, great interest has emerged in combining checkpoint blockade with current targeted therapies. Substantial research on BRAF mutant melanoma indicates that BRAF inhibition synergizes with immune checkpoint blockade by facilitating T cell recognition of melanoma antigens in a more favorable tumor microenvironment [25,26]. Although one early trial combining

immune checkpoint blockade with BRAF inhibition stalled because of hepatotoxicity [27], many additional trials are ongoing to optimize dosing and sequencing of the combination. An attractive feature of combining immune checkpoint blockade with targeted therapy is the potential to leverage current advances in precision medicine to impact many cancer types. For example, in BRCA1-deficient ovarian cancer, inhibition of poly(ADP-ribose) polymerases (PARPs), a family of enzymes involved in DNA repair, markedly increases mutational load in tumor cells and has demonstrated synergistic potential with CTLA-4 blockade [28]. Targeting driver mutations such as those in the genes encoding *c-KIT* or epidermal growth factor receptor (*EGFR*) with specific inhibitors appears to foster antitumor immunity [29,30]. In many cancers, disrupting tumor angiogenesis by inhibiting vascular endothelial growth factor (VEGF)/vascular endothelial growth factor receptor (VEGFR) promotes immune cell recruitment and infiltration, which may derive additional benefit from checkpoint inhibition [31]. In all of these cases and others, clinical trials are ongoing to determine the ability of targeted therapies to potentiate response to immune checkpoint inhibition.

Future of Precision Immunotherapy

Propelled by recent successes, enthusiasm for immunotherapy has surged beyond targeting CTLA-4 and PD-1 pathways. Many additional immune regulators, both stimulatory and inhibitory, are being explored as potential targets for cancer immunotherapy, each with a unique set of advantages and potential risks. Novel insights into neoantigens and combination with immune checkpoint inhibition have breathed new life into cancer vaccines, which had been tested for decades with little success. In addition, cell-based therapies may be required in some patients to elicit a robust antitumor response. Adoptive cell transfer of autologous T cells selected for tumor reactivity [32] and genetically engineered T cells with chimeric antigen receptors targeting tumor antigens [33] have demonstrated promise in melanoma and other cancer types. As new immunotherapies expand the arsenal of cancer therapies, deeper mechanistic insight will be required to inform clinical decisions. The exciting advances toward understanding and delivering precision immunotherapy are poised to change the way cancer is treated.

References

1. Coley W. The treatment of malignant tumors by repeated inoculations of erysipelas: with a report of ten original cases. *Am J Med Sci.* 1893; 105:487–511.
2. Holderfield M, Deuker MM, McCormick F, McMahon M. Targeting RAF kinases for cancer therapy: BRAF-mutated melanoma and beyond. *Nat Rev Cancer.* 2014; 14(7):455–67. doi: [10.1038/nrc3760](https://doi.org/10.1038/nrc3760) PMID: [24957944](https://pubmed.ncbi.nlm.nih.gov/24957944/)
3. Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med.* 2011; 364(26):2507–16. doi: [10.1056/NEJMoa1103782](https://doi.org/10.1056/NEJMoa1103782) PMID: [21639808](https://pubmed.ncbi.nlm.nih.gov/21639808/)
4. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med.* 2010; 363(8):711–23. doi: [10.1056/NEJMoa1003466](https://doi.org/10.1056/NEJMoa1003466) PMID: [20525992](https://pubmed.ncbi.nlm.nih.gov/20525992/)
5. Robert C, Schachter J, Long GV, Arance A, Grob JJ, Mortier L, et al. Pembrolizumab versus Ipilimumab in Advanced Melanoma. *N Engl J Med.* 2015; 372(26):2521–32. doi: [10.1056/NEJMoa1503093](https://doi.org/10.1056/NEJMoa1503093) PMID: [25891173](https://pubmed.ncbi.nlm.nih.gov/25891173/)
6. Hodi FS, Kluger H, Sznol M, Carvajal R, Lawrence D, Atkins M, et al. Abstract CT001: Durable, long-term survival in previously treated patients with advanced melanoma (MEL) who received nivolumab (NIVO) monotherapy in a phase I trial. *Cancer Research.* 2016; 76(14 Supplement):CT001–CT.
7. Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, et al. Nivolumab in previously untreated melanoma without BRAF mutation. *N Engl J Med.* 2015; 372(4):320–30. doi: [10.1056/NEJMoa1412082](https://doi.org/10.1056/NEJMoa1412082) PMID: [25399552](https://pubmed.ncbi.nlm.nih.gov/25399552/)

8. Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N Engl J Med*. 2015; 372(21):2006–17. doi: [10.1056/NEJMoa1414428](https://doi.org/10.1056/NEJMoa1414428) PMID: [25891304](https://pubmed.ncbi.nlm.nih.gov/25891304/)
9. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, et al. Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. *N Engl J Med*. 2015; 373(1):23–34. doi: [10.1056/NEJMoa1504030](https://doi.org/10.1056/NEJMoa1504030) PMID: [26027431](https://pubmed.ncbi.nlm.nih.gov/26027431/)
10. Larsen TE, Grude TH. A retrospective histological study of 669 cases of primary cutaneous malignant melanoma in clinical stage I. 3. The relation between the tumour-associated lymphocyte infiltration and age and sex, tumour cell type, pigmentation, cellular atypia, mitotic count, depth of invasion, ulceration, tumour type and prognosis. *Acta Pathol Microbiol Scand A*. 1978; 86A(6):523–30. PMID: [716913](https://pubmed.ncbi.nlm.nih.gov/716913/)
11. Tumeah PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature*. 2014; 515(7528):568–71. doi: [10.1038/nature13954](https://doi.org/10.1038/nature13954) PMID: [25428505](https://pubmed.ncbi.nlm.nih.gov/25428505/)
12. Spranger S, Spaepen RM, Zha Y, Williams J, Meng Y, Ha TT, et al. Up-regulation of PD-L1, IDO, and T (regs) in the melanoma tumor microenvironment is driven by CD8(+) T cells. *Sci Transl Med*. 2013; 5(200):200ra116. doi: [10.1126/scitranslmed.3006504](https://doi.org/10.1126/scitranslmed.3006504) PMID: [23986400](https://pubmed.ncbi.nlm.nih.gov/23986400/)
13. Taube JM, Klein A, Brahmer JR, Xu H, Pan X, Kim JH, et al. Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. *Clin Cancer Res*. 2014; 20(19):5064–74. doi: [10.1158/1078-0432.CCR-13-3271](https://doi.org/10.1158/1078-0432.CCR-13-3271) PMID: [24714771](https://pubmed.ncbi.nlm.nih.gov/24714771/)
14. Garon EB, Rizvi NA, Hui R, Leigh N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. *N Engl J Med*. 2015; 372(21):2018–28. doi: [10.1056/NEJMoa1501824](https://doi.org/10.1056/NEJMoa1501824) PMID: [25891174](https://pubmed.ncbi.nlm.nih.gov/25891174/)
15. Lawrence MS, Stojanov P, Polak P, Kryukov GV, Cibulskis K, Sivachenko A, et al. Mutational heterogeneity in cancer and the search for new cancer-associated genes. *Nature*. 2013; 499(7457):214–8. doi: [10.1038/nature12213](https://doi.org/10.1038/nature12213) PMID: [23770567](https://pubmed.ncbi.nlm.nih.gov/23770567/)
16. Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, et al. Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature*. 2014; 515(7528):577–81. doi: [10.1038/nature13988](https://doi.org/10.1038/nature13988) PMID: [25428507](https://pubmed.ncbi.nlm.nih.gov/25428507/)
17. Van Allen EM, Miao D, Schilling B, Shukla SA, Blank C, Zimmer L, et al. Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. *Science*. 2015; 350(6257):207–11. doi: [10.1126/science.aad0095](https://doi.org/10.1126/science.aad0095) PMID: [26359337](https://pubmed.ncbi.nlm.nih.gov/26359337/)
18. Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. *N Engl J Med*. 2014; 371(23):2189–99. doi: [10.1056/NEJMoa1406498](https://doi.org/10.1056/NEJMoa1406498) PMID: [25409260](https://pubmed.ncbi.nlm.nih.gov/25409260/)
19. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. *N Engl J Med*. 2015; 372(26):2509–20. doi: [10.1056/NEJMoa1500596](https://doi.org/10.1056/NEJMoa1500596) PMID: [26028255](https://pubmed.ncbi.nlm.nih.gov/26028255/)
20. McGranahan N, Furness AJ, Rosenthal R, Ramskov S, Lyngaa R, Saini SK, et al. Clonal neoantigens elicit T cell immunoreactivity and sensitivity to immune checkpoint blockade. *Science*. 2016; 351(6280):1463–9. doi: [10.1126/science.aaf1490](https://doi.org/10.1126/science.aaf1490) PMID: [26940869](https://pubmed.ncbi.nlm.nih.gov/26940869/)
21. Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, et al. Cancer immunotherapy. A dendritic cell vaccine increases the breadth and diversity of melanoma neoantigen-specific T cells. *Science*. 2015; 348(6236):803–8. doi: [10.1126/science.aaa3828](https://doi.org/10.1126/science.aaa3828) PMID: [25837513](https://pubmed.ncbi.nlm.nih.gov/25837513/)
22. Robert L, Tsoi J, Wang X, Emerson R, Homet B, Chodon T, et al. CTLA4 blockade broadens the peripheral T-cell receptor repertoire. *Clin Cancer Res*. 2014; 20(9):2424–32. doi: [10.1158/1078-0432.CCR-13-2648](https://doi.org/10.1158/1078-0432.CCR-13-2648) PMID: [24583799](https://pubmed.ncbi.nlm.nih.gov/24583799/)
23. Li B, Li T, Pignon JC, Wang B, Wang J, Shukla SA, et al. Landscape of tumor-infiltrating T cell repertoire of human cancers. *Nat Genet*. 2016; 48(7):725–32. doi: [10.1038/ng.3581](https://doi.org/10.1038/ng.3581) PMID: [27240091](https://pubmed.ncbi.nlm.nih.gov/27240091/)
24. Yadav M, Jhunjunwala S, Phung QT, Lupardus P, Tanguay J, Bumbaca S, et al. Predicting immunogenic tumour mutations by combining mass spectrometry and exome sequencing. *Nature*. 2014; 515(7528):572–6. doi: [10.1038/nature14001](https://doi.org/10.1038/nature14001) PMID: [25428506](https://pubmed.ncbi.nlm.nih.gov/25428506/)
25. Frederick DT, Piris A, Cogdill AP, Cooper ZA, Lezcano C, Ferrone CR, et al. BRAF inhibition is associated with enhanced melanoma antigen expression and a more favorable tumor microenvironment in patients with metastatic melanoma. *Clin Cancer Res*. 2013; 19(5):1225–31. doi: [10.1158/1078-0432.CCR-12-1630](https://doi.org/10.1158/1078-0432.CCR-12-1630) PMID: [23307859](https://pubmed.ncbi.nlm.nih.gov/23307859/)
26. Cooper ZA, Juneja VR, Sage PT, Frederick DT, Piris A, Mitra D, et al. Response to BRAF inhibition in melanoma is enhanced when combined with immune checkpoint blockade. *Cancer Immunol Res*. 2014; 2(7):643–54. doi: [10.1158/2326-6066.CIR-13-0215](https://doi.org/10.1158/2326-6066.CIR-13-0215) PMID: [24903021](https://pubmed.ncbi.nlm.nih.gov/24903021/)

27. Ribas A, Hodi FS, Callahan M, Konto C, Wolchok J. Hepatotoxicity with combination of vemurafenib and ipilimumab. *N Engl J Med*. 2013; 368(14):1365–6. doi: [10.1056/NEJMc1302338](https://doi.org/10.1056/NEJMc1302338) PMID: [23550685](https://pubmed.ncbi.nlm.nih.gov/23550685/)
28. Higuchi T, Flies DB, Marjon NA, Mantia-Smaldone G, Ronner L, Gimotty PA, et al. CTLA-4 Blockade Synergizes Therapeutically with PARP Inhibition in BRCA1-Deficient Ovarian Cancer. *Cancer Immunol Res*. 2015; 3(11):1257–68. doi: [10.1158/2326-6066.CIR-15-0044](https://doi.org/10.1158/2326-6066.CIR-15-0044) PMID: [26138335](https://pubmed.ncbi.nlm.nih.gov/26138335/)
29. Akbay EA, Koyama S, Carretero J, Altabef A, Tchaicha JH, Christensen CL, et al. Activation of the PD-1 pathway contributes to immune escape in EGFR-driven lung tumors. *Cancer Discov*. 2013; 3(12):1355–63. doi: [10.1158/2159-8290.CD-13-0310](https://doi.org/10.1158/2159-8290.CD-13-0310) PMID: [24078774](https://pubmed.ncbi.nlm.nih.gov/24078774/)
30. Balachandran VP, Cavnar MJ, Zeng S, Bamboat ZM, Ocuin LM, Obaid H, et al. Imatinib potentiates antitumor T cell responses in gastrointestinal stromal tumor through the inhibition of Ido. *Nat Med*. 2011; 17(9):1094–100. doi: [10.1038/nm.2438](https://doi.org/10.1038/nm.2438) PMID: [21873989](https://pubmed.ncbi.nlm.nih.gov/21873989/)
31. Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, et al. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc Natl Acad Sci U S A*. 2012; 109(43):17561–6. doi: [10.1073/pnas.1215397109](https://doi.org/10.1073/pnas.1215397109) PMID: [23045683](https://pubmed.ncbi.nlm.nih.gov/23045683/)
32. Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, et al. Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy. *Clin Cancer Res*. 2011; 17(13):4550–7. doi: [10.1158/1078-0432.CCR-11-0116](https://doi.org/10.1158/1078-0432.CCR-11-0116) PMID: [21498393](https://pubmed.ncbi.nlm.nih.gov/21498393/)
33. Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N Engl J Med*. 2014; 371(16):1507–17. doi: [10.1056/NEJMoa1407222](https://doi.org/10.1056/NEJMoa1407222) PMID: [25317870](https://pubmed.ncbi.nlm.nih.gov/25317870/)