

**S1 Table. Strains, plasmids and primers used in this study.**

	<b>Genotype/description</b>	<b>Source</b>
<b>Strains</b>		
BY4741 ( <i>S.cerevisiae</i> )	<i>MATa his3Δ0 leu2Δ0 met15Δ0 ura3Δ0</i>	Open Biosystems
DH5α ( <i>E. coli</i> )	<i>F- gyrA96 (Nal<sup>r</sup>) recA1 endA1 thi-1 hsdR17 (rk-mk-) gluV44 deoR Φ80d Δ(lacZ)M15</i>	[1]
C43(DE3) ( <i>E. coli</i> )	<i>F- ompT hsdSB (rB- mB-) gal dcm (DE3)</i>	[2]
<b>Plasmids</b>		
pET17b-P450	Vector with <i>Gs</i> P450 gene (cDNA)	This study
pYeDP60U-CPR1	Vector with <i>Gs</i> CPR1 gene (cDNA)	This study
pYeDP60U-CPR2	Vector with <i>Gs</i> CPR1 gene (cDNA)	This study
<b>Primers</b> 5' → 3' orientation		
CYP630B18- fw_ <i>Nde</i> I	<u>GACATCCATATGTATGCCCTTCTGTGCCAGC</u>	
CYP630B18- rev_ <i>Eco</i> RI	GGAGGTGGAGTTGAAGGAAGAGCTCTCATCAC TGAGAATT <u>CGATGTC</u>	
CPR1- pYeD_ <i>Bam</i> HI	CTAAATTACCGGATCATGGCGAACTGG ACACGCTGGAC	
CPR1-pYeD_ <i>Eco</i> RI	CCAATACCAGGAGGACGTTTGGT <u>GAAATTCGC</u> GGGGGATC	
CPR2- pYeD_ <i>Bam</i> HI	CTAAATTACCGGATCATGTTTGTCC TGCCTTTGGCTCTGC	
CPR2-pYeD_ <i>Eco</i> RI	GCTATATCGAGGATGTGTGGGGTT <u>GAAATTCG</u> CGGGGGATC	
<b>PCR conditions</b>		
CYP6330B18	95°C/3' 15x(95°C/30'' (55°C/65°C)/45'' 72°C/60'') 25x(95°C/30'' 55°C/45'' 72°C/60'') 72°C/5'	
CPR1	95°C/3' 35x(95°C/30'' 55°C/30'' 72°C/45'') 72°C/5'	
CPR2	95°C/3' 15x(95°C/30'' (55°C/65°C)/45'' 72°C/60'') 25x(95°C/30'' 55°C/45'' 72°C/60'') 72°C/5' 95°C/3' 14x(95°C/10'' (62°C/72°C)/45'' 72°C/60'') 29x(95°C/10'' 62°C/55'' 72°C/60'') 72°C/5'	

Restriction enzyme cut sites are underlined.