Available online www.unicrossjournals.com



UNICROSS JOURNAL OF SCIENCE AND TECHNOLOGY, UJOST

 RESEARCH ARTICLE
 VOL. 3(1)
 MARCH 31, 2024 ISSN:2814-2233

 Date Approved March 31, 2024
 Pages 126 - 137

EPIDEMIOLOGICAL ASSESSMENT OF BODY WEIGHT LOSSES IN POULTRY CAUSED BY ECTOPARASITES: A CASE STUDY OF OLD ORLU, IMO STATE

Uttah, Emmanuel Chukwunenye¹, Eze, Chinedu Nobert², Uttah, Faustina Onyinye¹

¹Department of Animal and Environmental Biology, University of Cross River State, Calabar. ²Department of Biology, University of Agriculture and Environmental Sciences, Umuagwo, Imo State.

The study was aimed at assessing body weight losses in poultry caused by ectoparasites in Old Orlu, Imo State. Confined and free-range chickens were collected from four popular markets in Old Orlu, namely, the Orlu Main market, Eke Mgbidi market, Afor Umuaka market, and Orie Nnempi market. A total of 1000 birds were examined; comprised of 250 from each of the four markets, and sampled equally for confined and free range birds. The highest prevalence of infestation (92.2%) was recorded among the 1.1 - 1.5 kg weight category. The least prevalence rate (65.8%) was among the heaviest weight category (>3.0 kg). Infestation was significantly higher among those in > 2.0 kg weight category than those in the \leq 2.0 kg weight category (×²test= 12.342; p < 0.001). Chicken of the low weight (1.1 – 1.5 kg) category contributed the highest percentage (29.3 %) of all parasites counted in both the males (31.7 %) and females (26.3 %). whereas chicken of the heaviest weight (>3.0 kg) category contributed the least percentage (5.0 %) of all parasites counted; both the male (4.7%) and female (5.3%) chicken. The ectoparasite intensity of those that were ≤ 2.0 kg in weight were significantly higher than the intensity of those that were > 2.0 kg in weight (\times 2-test = 26.222; p < 0.001). In conclusion, high prevalence and high intensity of ectoparasites resulted in significant body weight losses in chickens in Old Orlu leading to economic losses. Routine actionable protocols for the prevention and control of ectoparasites in Old Orlu is recommended.

Keywords: Ectoparasites, prevalence, poultry, body weight loss, Orlu

1.0 Introduction

Poultry is one of the most important sources of protein for man, and this has resulted into a blossoming Poultry population industry. in Nigeria is about 160 million; with chickens comprising about 72.4 million (Akintunde et al., 2015). Backyard poultry constitutes the most important form of poultry production (Mohammed and Sunday, 2015). The poultry industry plays a vital role in national economies as a revenue provider (Odenu et al., 2016). Its role includes improving income of small farmers with small land holdings (Poulsen et al., 2000; Yoriyo et al.,2008); contributing up to 15% of the country's gross domestic product (GDP) and accounting for 36% of total protein intake of the country (Odenuet al., 2016; Akintunde et al., 2015). The trado-cultural and socioreligious of acceptability the production. marketing and consumption of the poultry products could be the principal reason for this success story (Beyene et al., 2014; Mohammed and Sunday, 2015).

The main constraints in poultry production in Nigeria are the common poultry diseases caused by poor housing, poor management, and arthropod-borne infections

Adeoti, (Akintunde 2014: and Mohammed and Sunday, 2015). Arthropod ectoparasites especially have major impact on husbandry, productivity and welfare of domestic chickens (Balaet al., 2011; Desoky et al., 2015). Ectoparasitic taxonomic groups such as lice, mites and fleas, live on domestic chickens (Tamiru et al., 2015; Angyiereyiri et al., 2015); and cause severe dermatitis and allergies (Bala et al., 2011), anaemia (Zeryehun and Yohannes, 2015), anorexia or death (Permin et al., 2002), and may act as vectors for pathogenic agents, such as Rickettsia disease (murine typhus), bacterial disease (plague) and viral disease (myxomatosis) resulting in serious diseases not only in domestic chickens. but also in human population (Bala et al., 2011; Asresie and Eshetu, 2015; George et al., 2015). Ectoparasites may also cause diseases such as scaly leg and depluming mange. These eventually lead to losses in egg and meat production (Zeryehun and Yohannes, 2015).

Notwithstanding the status of ectoparasites, as one of the major causes of decrease in chicken productivity, they are rarely studied (Dinka *et al.*, 2010), and often neglected (Dube and Aisien, 2005).

Periodic examination of the flock can help to detect an early infestation and can help to prevent a larger flock outbreak (Maina, 2005). This is extremely necessary to forestall damages to local poultry Moreki et al., 2003), such as injuries caused during blood-feeding, weight loss, blemishes. and lowered egg production (Kaufman, 2019). This study has become necessary to epidemiological conduct an assessment of weight losses in poultry due to ectoparasitic infestations. The limited scope will be to chickens. Gallus gallus domestica. in communities in the Old Orlu area of Imo State.

2.0 Materials and methods2.1 Description of the study area

Orlu town is the headquarters of the old Orlu Region in the present Imo State, Nigeria, located between 58358° N and 7.01968° E, having an area of 129.8 km² and a population of 420,000 in 2006 census (NPC 2006). It is presently referred to as the Imo West Senatorial zone.

Orlu is characterized by two distinct tropical seasons, the rainy and dry seasons. The rainy season (April to September) with annual rainfall between 200mm and 450mm, while the dry season (November to March) also comes with harmattan weather from December to February during which night temperature is relatively very low.

Majority of inhabitants in Orlu are predominantly agrarian and depend on poultry products either consumed as food (meat and egg) or traded as a source of income.

2.2 Sampling and field examination of the birds

The study consisted mainly of confined and free-range chickens, collected from four popular markets in different areas of the Old Orlu. These markets were Orlu Main market, Eke Mgbidi market, Afor Umuaka market, and OrieNnempi market. In all, a total of 1000 birds were examined. This comprised of 250 chickens examined from each of the four markets and their environs, and sampled equally for confined and free range birds.

2.3 Procedure for examination of birds

The objectives of this study was comprehensively explained to chicken owners prior to the start of the bird examinations, to allay suspicion and to encourage active and honest participation.

Ectoparasites were collected from the birds by displaying the feathers horizontally against their anatomical

EPIDEMIOLOGICAL ASSESSMENT OF BODY WEIGHT LOSSES IN POULTRY CAUSED BY ECTOPARASITES: A CASE STUDY OF OLD ORLU, IMO STATE Uttah, et al.

direction of alignment so as to expose them. Lice were collected from hosts by parting the hairs or feathers, gently brushing the base of the feathers with a fine soft brush so as to prevent injuries on the chickens. Mites were collected by scarping the skin surface with the edge of a slide. All the parasites collected were counted and placed in sampling bottles containing 70% alcohol.

Each chicken examined was assigned a serial number on the sampling bottle for easy identification. Information was obtained regarding each of the chicken sampled including sex, feather patterns, and age of the chickens.

2.4 Identification of parasites

The ectoparasites were placed in 10 % KOH (clearing agent) two to three days before identification. Species determination was based on microscopic examination using dissecting and binocular microscopes morphological study to their characteristics.

2.5 Laboratory analysis

Laboratory analysis involved a thorough examination of the parasites for identification. Samples of the observed parasites were removed with a thumb forceps or camel hair brush and transferred to a petri dish containing 10% alcohol. They were cleared with lactophenol and fixed on a microscopic slide using a little quantity of polyvinyl, alcohol and lactophenol solution before detailed morphological examination and identification using a compound microscope.

2.6 Data analysis

Chi square (χ^2 -test) was used to test the significance of differences between the parameters tested while student's t-test was used to analyze difference in parasites intensity. Values of P<0.05 will be considered as statistical significant.

The Geometric Mean Intensity (GMI) of infestation was calculated as antilog ($\sum \log (x+1)/n$), with x being the number of ectoparasites counted, and *n* being the number of positive individuals examined.

3.0 Results and discussion

3.1 Prevalence of ectoparasitic infestations in relation to weight

Prevalence of ectoparasitic infestations in relation to weight is presented in Table 1. The highest prevalence (92.2%) was among the 1.1 - 1.5 kg weight category followed by 90.2% among 0.6 - 1.0 kg weight category. The least prevalence rates (65.8% and 71.3%) were among the heaviest weight categories >3.0 kg and 2.5 - 3.0 kg weight categories respectively.

A comparison of prevalence among relatively less weighty (2kg or less) and more weighty (2.2 kg and above) chicken (see Figure 1) shows that prevalence among the former was significantly higher than among the latter (\times^2 -test= 12.342; p < 0.001).

3.2 Parasite Geometric Mean Intensity (GMI) in relation to weight of chicken

The GMI of ectoparasitic infestation in relation to weight of chicken is presented in Table 2. The highest GMI were 7.91 and 7.57 for chicken belonging to the 1.1 - 1.5 kg and 0.6 - 1.0 kg weight categories respectively. The least GMI were 6.14 and 6.37 for chicken belonging to the heaviest weight categories, that is, >3.0 kg and the 2.6 - 3.0 kg respectively. Relative contributions to parasite intensity by chicken of various weight categories is presented in Figure 2. Chicken of the 1.1 - 1.5weight category contributed the highest percentage (29.3 %) of all parasites counted in both the males (31.7 %) and females (26.3 %). Furthermore, chicken of the heaviest weight category (>3.0)kg) contributed the least percentage (5.0 %) of all parasites counted in both males (4.7%) and females (5.3%).

For further comparative analysis, the chicken were divided into two broad categories namely, those that were \leq 2.0 kg and those that were > 2.0 kg, and the parasite intensity of these two groups were compared and presented in Figure 3. The overall parasite intensity of those \leq 2.0 kg in weight were significantly higher than the parasite intensity of those that were > 2.0 kg in weight (×²-test = 26.222; p < 0.001).

Weight (kg)	Numbe	er Examine	d	Number Positive (%)			
	Males	Females	Total	Males	Females	Total	
00 - 0.5	68	52	120	58 (85.3)	44 (84.6)	102 (85.0)	
0.6 - 1.0	93	81	174	84 (90.3)	73 (90.1)	157 (90.2)	
1.1 - 1.5	171	97	268	156 (91.2)	91 (93.8)	247 (92.2)	
1.6 - 2.0	86	77	163	69 (80.2)	63 (81.8)	132 (81.0)	
2.1 - 2.5	58	51	109	47 (81.0)	37 (72.5)	84 (77.1)	
2.5 - 3.0	46	41	87	33 (71.7)	29 (70.7)	62 (71.3)	
>3.0	41	38	79	27 (65.8)	25 (65.8)	52 (65.8)	
Total	563	437	1000	474 (84.2)	362 (82.8)	836 (83.6)	

Table 1.Prevalence of ectoparasitic infestations in relation to weight (kg)



Figure 1. Comparing the prevalence between relatively less weighty (2kg or less) and more weighty (2.2 kg and above) chicken

Weight	Males			Females			Total		
(kg)	No.	No.	GMI	No.	No.	GMI	No.	No.	GMI
	counted	infected		counted	infected		counted	infected	
0-0.5	642	58	6.46	456	44	6.12	1098	102	7.00
0.6-1.0	1003	84	6.91	946	73	6.85	1949	157	7.57
1.1-1.5	1631	156	7.40	1106	91	7.01	2737	247	7.91
1.6-2.0	871	69	6.77	774	63	6.65	1645	132	7.41
2.1-2.5	427	47	6.06	444	37	6.10	871	84	6.77
2.6-3.0	331	33	5.80	253	29	5.53	584	62	6.37
>3.0	244	27	5.50	221	25	5.40	465	52	6.14
Total	5149	474	8.55	4200	362	8.34	9349	836	9.14

Table 2.GMI of the ectoparasitic infestation in relation to weight (kg)



Figure 2. Relative percentage contributions to parasite intensity by chicken of various weight categories



Figure 3. Comparing the percentage contributions to parasite intensity between chicken ≤ 2.0 kg and those >2.0 kg.

Two relationships were established in this study between ectoparasitic infestation and body weight loss in chicken. Firstly, prevalence of infestation was significantly higher among birds in the lower weight (\leq 2.0 kg) than those in the higher weight

EPIDEMIOLOGICAL ASSESSMENT OF BODY WEIGHT LOSSES IN POULTRY CAUSED BY ECTOPARASITES:A CASE STUDY OF OLD ORLU, IMO STATEUttah, et al.

(>2.0 kg) category. Secondly, intensity of ectoparasites was significantly higher among birds in the lower weight (< 2.0 kg) than those in the higher weight (>2.0)kg) category. These findings are corroborated by the observations of Riwidiharsoet al. (2020) that the higher the ectoparasites attack, the lower the chicken's body weight. In another study, strong relationship was established between ectoparasite prevalence and chicken weight loss (Nik-Hasan et al., 2015; Riwidiharso et al., 2020). The highly infested birds with high parasite intensity lost weight due to effects of parasitization such as anaemia, increased stress, feather pecking, and restlessness (Koutinas et al., 2019). According to Ikpeze et al. (2008), adverse effects of ectoparasitic infestation included irritation, reduced mating potentials in cocks, reduced egg laying in hens, and loss of weight in broilers, pullets and cockerels. Ectoparasites generated a decrease in weight gain among broiler chicken (Sáenz, 2021). All things being equal, chronically infested chickens were found to be underweight when compared with their non-infested counterparts (Akinwunmi et al., 1978). Interference with feed consumption by ectoparasites would lead to emaciation, and this together with anaemia (also caused by ectoparasites) brought about weight loss in chicken (Mishra et al., 2017). Elucidating further on this, Okechukwu and Ikpeze (2020) stated that infestations by biting, chewing, and suckinglice caused featherperking, feather-loss, and restlessness in infested chickens, which led to reduced feed in-take, general debility, weight loss and low egg production.

Ectoparasitic infestations could result in mortality (Sáenz, 2021), and disease transmission (Kouam *et al.*, 2022). Furthermore, skin-damage by biting and sucking activities of lice might reduce market value of chickens (Ikpeze 2008).

Indeed. ectoparasites in chicken are prevalent and a very important factor in poultry (Riwidiharso et al., 2020). They have been shown in many studies in both the tropical and temperate regions to cause economic damage (Kaufman, 2019), and to adversely affect economical productivity (Mungube et al., 2006 Nyoni et al., 2012; Onyekachi, 2021). In the village chicken production systems particularly, ectoparasitic infestation is of great economic importance (Firaol et al., 2014). Globally, the annual loss in poultry production, caused by external parasites, has been estimated at one billion US dollars (Akinwunmi et al., 1978).

Chicken mortality caused by ectoparasitic infestations is higher than those attributed to some chicken viral infectious diseases such as Newcastle disease and fowl pox disease (Nnadi and George, 2010; Opara *et al.*, 2014). High mortality of village poultry pose a serious threat not only to food security and livelihood of many rural families (Musa *et al.*, 2008), but to efforts of Government to alleviate poverty and discourage urban migration for white-collar jobs (Lawal *et al.*, 2016).

4.0 Conclusion/ Recommendation

In conclusion, high prevalence and high intensity of ectoparasites result in significant body weight losses among chickens leading to serious economic losses. This calls for planned action on the part of Government for mitigation. In this regard, routine actionable protocols for the prevention and control of ectoparasites should be put in place in Old Orlu, and Imo State in general. Furthermore, mounting robust educative campaigns to create awareness, among poultry farmers, on the economic importance of ectoparasites on the quantity and quality of poultry production has become extremely necessary in Nigeria.

Acknowledgement

This work was supported by an Institutionbased Research Grant(IBR) from the Tertiary Education Trust Fund (TETFund) toEze, C.N. and Uttah, E.C. (Epidemiological analysis of losses in poultry caused by ectoparasites: a case study of Orlu, Imo state) through the University of Agriculture & Environmental Sciences, Umuagwo, Imo State.

References

- Angyiereyiri E.D., Sackey I, Bonu-Ire MST (2015).Survey on Arthropod Ectoparasites on Goats and Domestic Fowls in Vunania, Navrongo, Ghana. *Canadian Journal of Pure and Applied Science***9**(2): 3371-3377.
- Akintunde, O,K, and Adeoti, A.I. (2014). Assessment of factors affecting the level of poultry disease management in southwest, Nigeria. Trends in Agric. Econs, 7(2): 41-56.
- Akintunde, O,K,, Adeoti, A.I., Okoruwa, V.O., Omonona, B.T. and Abu, A.O. (2015). Effect of disease management on profitability of chicken egg production in Southwest Nigeria. Asian J. of Poul. Sci., 9(1): 1-18.
 Akinwunmi, J. A., Adegeye, A. T., Ikpi, A. E. and Olayide, S. O. (1978). Economic analysis of Nigerian poultry

industry. Federal Livestock Department (FLD), Lagos.

- Asresie, A., and Eshetu, M. (2015). Traditional Chicken Production System and Marketing in Ethiopia: *A review. J.* of Mar. and Cons. Res., **8**: 27-34.
- Bala AY, Anka SA, Waziri A, Shehu H (2011). Preliminary Survey of ectoparasites Infesting Chickens (Gallus domesticus) in Four Areas of Sokoto Metropolis. *Nigerian Journal of Basic and Applied Science* **19**: 173-180.
- Beyene, K., Bogale, B. and Chanie, M. (2014). Study on effects and occurrence of nematodes in local and exotic chickens in and around Bahir Dar, Northwest Ethiopia. American-*Eurasian J. of Sci. Res.*, **9**(3): 62-66. Desoky, A. S.S., Abdel-Gwad, K. H., Maher Ali, A., and Nafady, A. A. (2015). Comparison between spraying and washing method of reduction ratios on animal ectoparasites by using Diazinon 60% EC under field conditions in farm animals. African J. of Agric. Sci. and *Tech.*, **3**(6):294-298
- Dinka, H., Chala, R., Dawo, F., Bekana, E., and Leta, S. (2010). Major constraints and health management of village poultry production in Rift Valley of Oromia,
- Ethiopia. American-Eurasian J. Agric and EnvironSci, **9**:529-533.
- Dube, S. and Aisien, M.S.O. (2005). Studies on the family gastrothylacidae stiles et Goldberger, 1910 occurring in Nigerian fulani cattle. *ActaZoolTaiwanica***15**: 1-10.

EPIDEMIOLOGICAL ASSESSMENT OF BODY WEIGHT LOSSES IN POULTRY CAUSED BY ECTOPARASITES: A CASE STUDY OF OLD ORLU, IMO STATE Uttah, et al.

- Firaol T, Dagmawit A, Askale G, Solomon S, Morka D, Waktole T(2014).
 Prevalence of Ectoparasite Infestation in Chicken in andAround Ambo Town, *Ethiopia. J. Vet. Sci. Technolo.* 5(4):1 -5.
- George, D. R., Finn, R. D., Graham, K. M., Mul, M. F., Maurer, V., Moro, C. V., and Sparagano, O. A. (2015). Should the poultry red mite *Dermanyssusgallinae* be of wider concern for veterinary and medical science?.*Parasites & Vectors*8(1), 178.
- Ikpeze, O. O., Amagba, I. C. and Eneanya, C. I. (2008). Preliminary Survey of Ectoparasites of Chicken in Awka, South-Eastern Nigeria. Animal Research International5(2): 848 – 851.
- Kaufman, P. E. (2019). External parasites of poultry. IFAS Extension, University of Florida, Publication #ENY-290 <u>https://edis.ifas.ufl.edu/publication/IG1</u> <u>40</u>. Accessed 21st Dec., 2023.
- Kouam, M. K., Armand N. F., Herman F. B., Arnaud B. H. T., Thomas T. T. (2022). Prevelance and clinical signs of chewing lice local chickens in (Gallus gallusdomesticus) in Menoua Division, Western highlands of Cameroon. Veterinary *Parasitology:* Regional and Report **Studies** Vol. **34**. https://doi.org/10.1016/j.vprsr.2022.100 772
- Lawal, J.R. Bello, A.M., Balami, Y., Waki, Z. B., Yusuf, J., and Dauda, E. S. (2016). Prevalence and Economic Significance

of Ectoparasites Infestation in Village Chickens (Gallus gallusdomesticus) in Gombe, Northeastern Nigeria Direct Research Journal of Agriculture and Food Science **4**(5): 94-103.

- Maina A N.(2005) Prevalence, intensity and lesion associated with gastrointestinal parasites of indigenous chicken in Kenya. MSc thesis. University of Nairobi.
- Mishra S, Pednekar R, Mohanty BS, Gatne M. (2017). Prevalence, economic loss and control of lice infestation in poultry. *Intl J Sci Environ Tech***6** (3): 1745-1757.
- Mohammed, B.R. and Sunday, O. S. (2015).
 An Overview of the Prevalence of Avian Coccidiosis in Poultry Production and Its Economic Importance in Nigeria. *Vet. Res. Int.*, 3(3): 35-45.
- Moreki, J.C. (2003). Village Chicken and Poverty Alleviation, Animal Production and Health Division, UNDP, New York, NY, USA; South African Centre for Cooperation in Agricultural Research and Training, Gaborone, Botswana.
- Mungube, E. O., Bauni, S. M., Tenhagen, B. A., Nzioka, S. M., Muhammed, L, and Nginyi, J. M.(2008).Prevalence of parasite of local scavenging chicken in a selected semi-arid zone in Eastern Kenya, Africa. Trop/ca/ Animal Health Production Bulletin 40: 101 -109.
- Musa U, Abdu PA, Dafwang II, Edache JA, Ahmed MS, Bawa GS, Karsin PD, Emannaa PE (2008). A survey of causes of mortality in some Local chicken flocks in Plateau state: In: Proceedings of the 33rd Annual Conference of the

Nigeria Society of Animal Production (NSAP), pp.551 – 554.

- National Population Commission (NPC) (2006) Nigerian Population Census Report. National Population Commission, Abuja.
- Nik-Hassan NRN, Awang A, Rahman AMD. (2015). Parasitic burden and its relation with the bodyweight of free-range chicken in oil palm dominated Sandakan District of Malaysian Borneo. Intl J Livestock Res. DOI: 10.5455/ijlr.20150909073638.
- Nnadi PA, and George SO (2010). A crosssectional survey on parasites of chickens in selected villages in the subhumidzones of southeastern *Nigeria*. *J. Parasitol. Res.* Pp.1–6.
- Nyoni NMB, Masika PJ. (2012). Village chicken production practices in the Amatola Basin of the Eastern Cape Province, South Africa. African Journal of Agriculture Resources **17**:2647-2652.
- Odenu R.A., Mohammed B.R., Simon M.K., Agbede R.I.S. (2016). Ecto-parasites of Domestic Chickens (Gallus gallusdomesticus) in Gwagwalada Area Council, *Alexandria Journal of Veterinary Sciences* **51**(1): 140-146

Abuja, Nigeria-West Africa

- Okechukwu, P.C. and Ikpeze, O.O. (2020). Ectoparasites found on intensivelyreared chickens at semi-urban Emene in south-Eastern Nigeria. *The Biomedical Diagnostics*, **4**(2): 92-101,
- Onyekachi, O. (2021). Prevalence of Ectoparasites Infestation of Chickenin Three Poultry Farms in Awka. *Asian Basic and Applied Research*

*Journal***3**(1): 41-53, 2021; Article no.ABAARJ.386.

- Opara MN, Osowa DK, Maxwell JA (2014). Blood and Gastrointestinal Parasites of Chickens and Turkeys Reared in the Tropical Rainforest Zone of Southeastern Nigeria. *Open J. Vet. Med.* **4**:308-313.
- Permin, A., Esmann, J. B., Hoj, C. H., Hove, T., and Mukaratiwa, S. (2002). Ecto, Endo and Haemoparasites in free range chickens in the Goromonzi district in Zambabwe. *Preventive Veterinary Medicine* 54: 213-224.
- Poulsen, J., Permin, O., Hindsbo, I., Yelifari, P., Nansen, and Bloch, P. (2000).Prevalence and distribution of gastrointestinal helminthes and haemoparasites in young scavenging chicken in upper Eastern region of Ghana, West Africa preventing Veterinary Medicine 45 (3-4): 237-245.
- Riwidiharso, E., Darsono, I. and WidhionoImam, W. (2020). Prevalence and diversity of ectoparasites in scavenging chickens (*Gallus domesticus*) and their association to body weight. *Biodiversitas Journal of Biological Diversity***21**(7):3163-3169.
- Sáenz, J.A.C. (2021). Parasites in poultry. What are and how to avoid them? <u>https://www.veterinariadigital.com/en/a</u> <u>rticulos/parasites-in-poultry-what-are-</u> <u>and-how-to-avoid-them/</u>. Accessed 21st Dec., 2023.
- Tamiru, F., Dagmawit, A., Askale, G., Solomon, S., Morka, D., and Waktole, T. (2014). Prevalence of Ectoparasite Infestation in Chicken in and Around Ambo Town, Ethiopia. J. Ve.t Sci Technol., 5:189.

EPIDEMIOLOGICAL ASSESSMENT OF BODY WEIGHT LOSSES IN POULTRY CAUSED BY ECTOPARASITES: A CASE STUDY OF OLD ORLU, IMO STATE Uttah, et al.

- Yoriyo, K.P., Adarg, K.L., Adamu, S. U., and Panda, S. M. (2008). Prevalence of gastrointestinal helminthes of free-range chickens and guinea fowls in bauchi and its environment, Nigeria. *Bulletin of Pure and Applied Science* 27 (1):1-6.
- Koutinas, A.F., Papazahariadou, M.G., Rallis, T.S., Tzivara, N.H. Himonas, C.A. (2015). Flea species from dogs and cats in northern Greece: environmental and clinical implications. *Journal of Veterinary Parasitology*58: 109-115.

- Kaufman, P.E. (2019). External parasites of poultry. <u>https://edis.ifas.ufl.edu</u>
- Zeryehun T, Yohannes Υ. (2015). Ectoparasite infestation of free chickens reared under scavenging traditional backyard production system in Wolayita Zone, Southern Ethiopia. Ethiopian Veterinary Journal19(2):55-66.