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FIG. 1 *Leptodeira polysticta* holding a *Lepidophyma tuxtlae* sideways. IBH-RF-491 (Colección Nacional de Anfibios y Reptiles, Photographic Reference).

México, Mexico City. 189 pp.). Because *L. tuxtlae* is mainly a leaf litter dwelling species, the *L. polysticta* probably began chasing its prey in the leaf litter and pursued it to the highest part of a small stone fence. The *L. polysticta* bit the *L. tuxtlae* at the pelvis and immediately coiled around the middle part of the body before beginning to ingest the prey from the side (Fig. 1). Around 6 min after the capture, the snake uncoiled, perhaps because of the awkward position in which the *L. tuxtlae* was captured and having given the time necessary for venom to kill or immobilize the *L. tuxtlae*. After this time the *L. polysticta* regurgitated the lizard and began swallowing it again starting from the head, but without coiling. The *L. polysticta* used the anterior part of its body to wrap the prey, without an initial twist, so that the snake's belly faced away from its head (Greene and Burghardt 1978. Science 200:74–77). The total time of ingestion was ca. 12 min.

Duellman (1958, *op. cit.*) observed that "[*Leptodeira*] would grab hold of [a small frog], usually quickly work to the head, and swallow the frog…With larger frogs the snake strikes, grabs hold, and rapidly chews, opening its mouth widely until the enlarged, grooved fangs are engaged. Then it holds the prey until movement stops." All reports of *Leptodeira* feeding behavior to date involve frogs (except two on fish; Céspedes and Abarca 2014. Mesoamer. Herpetol. 1:288–289; Solís and Guerrero 2016. Herpetol. Rev. 47:313), and none mention constriction (De Carvalho et al. 2007. Herpetol. Rev. 38:89; Hagman and Schulte 2007. Herpetol. Rev. 38:90; Cabrera-Guzmán et al. 2009, *op. cit.*; Dehling 2009. Herpetol. Rev. 40:356; Cruz-Sáenz et al. 2010. Herpetol. Rev. 41:366; Hernández-Ríos et al. 2011. Herpetol. Rev. 42:100; Mata-Silva et al. 2012. Herpetol. Rev. 43:660; Santos-Silva et al. 2014. Herpetol. Notes 7:123–126; Neta et al. 2015. Herpetol. Rev. 46:452; Bello-Sánchez et al. 2018. Herpetol. Rev. 49:756), even when attempts are made to swallow very large anurans, which may take hours and are sometimes unsuccessful (Rodríguez et al. 2011. Herpetol. Rev. 42:616; Sales et al. 2013. Herpetol. Rev. 44:524; Vargas-Salinas and Aponte-Gutiérrez 2013. Herpetol. Notes 6:189–191; Santos-Silva et al. 2014, *op. cit.*; Engeman and Engeman 2015. Herpetol. Rev. 46:104–105; García-Mata et al. 2017. Bull. Chicago Herpetol. Soc. 52:139–145; Céspedes et al. 2018. Herpetol. Notes 11:959–960). Although venom activity in *Leptodeira* has been relatively well-studied (Lemoine et al. 2004. Wilderness Environ. Med. 15:82–89; Weinstein et al. 2014. Clin. Toxicol. 52:277–282; Torres-Bonilla et al. 2016. Toxicon 119:345–351) and despite numerous reports of feeding in the wild, this is the first documentation of constriction in *Leptodeira*, suggesting that perhaps this behavior is used only when consuming lizards. Despite this unique behavior, we cannot assure that constricting behavior plays a key role in killing their prey.

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***MICRURUS CORALLINUS* (Coral Snake). REPRODUCTION.** *Micrurus corallinus* is a medium-sized, semifossorial elapid snake (up to 950 mm SVL) found mostly in Atlantic forest areas in southeastern South America (Marques 1996. Amphibia-Reptilia 17:277–285). Female reproduction is highly seasonal, with oviposition occurring in mid wet season (December–January; Marques 1996, *op. cit.*). Apart from clutch size and offspring size, information on many fecundity parameters remains largely unknown. Here, I provide information on egg size, egg mass, relative clutch mass (RCM), and embryonic stage at oviposition of *M. corallinus*.

Information was obtained from seven gravid females collected by laypersons at various locations in the Atlantic forest domain (southeastern Brazil) and donated to the Instituto Butantan (São Paulo, Brazil) between 2006 and 2013. Oviposition occurred from late November to mid-January (1–8 days post capture). Clutch size averaged 5.7 ± 3.4 (SD) eggs (range: 3–13 eggs) and was positively correlated with maternal body mass ($r^2$ = 0.916; $P$ = 0.0007). Egg mass averaged 4.0 ± 0.8 g (range: 3.2–5.3 g), and total clutch mass averaged 23.0 ± 13.6 g (range: 10.0–50.5 g). RCM (total clutch mass/body mass of mother after oviposition; Shine 1980. Oecologia 46:92–100) averaged 0.54 ± 0.12 (range: 0.39–0.78). Egg size was measured in four clutches. Eggs were quite elongated and averaged 37.7 ± 4.2 mm in length (range: 33.1–43.2 mm) and 12.8 ± 0.9 mm in width (range: 12.1–13.9 mm). The fecundity parameters of *M. corallinus* (*i.e.*, low clutch size, with elongated eggs and high RCM) are recurrent among several unrelated fossorial or semifossorial snakes (Braz et al. 2014. Herpetol. J. 24:49–57). Dissection of two eggs from two clutches revealed developing embryos at early growth phase (stages 26 and 27 according to Zehr 1962. Copeia 1962:322–329), which corresponds to the upper limit of the stages at oviposition in most snakes (Blackburn 1995. J. Theor. Biol. 174:199–216).

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***MICRURUS NIGROCINCTUS* (Central American Coral Snake) and *LEPTODEIRA RHOMBIFERA* (Central American Banded Cat-eyed Snake). DIET and PREDATION.** *Micrurus nigrocinctus* (Elapidae) is distributed widely from southeastern Mexico to northern Colombia, whereas *Leptodeira rhombifera* (Dipsadidae) ranges from Guatemala south to Central Panama (Savage 2002. The Amphibians and Reptiles of Costa Rica: A Herpetofauna between Two Continents, between Two Seas. University of Chicago Press, Chicago, Illinois. 934 pp.; McCranie