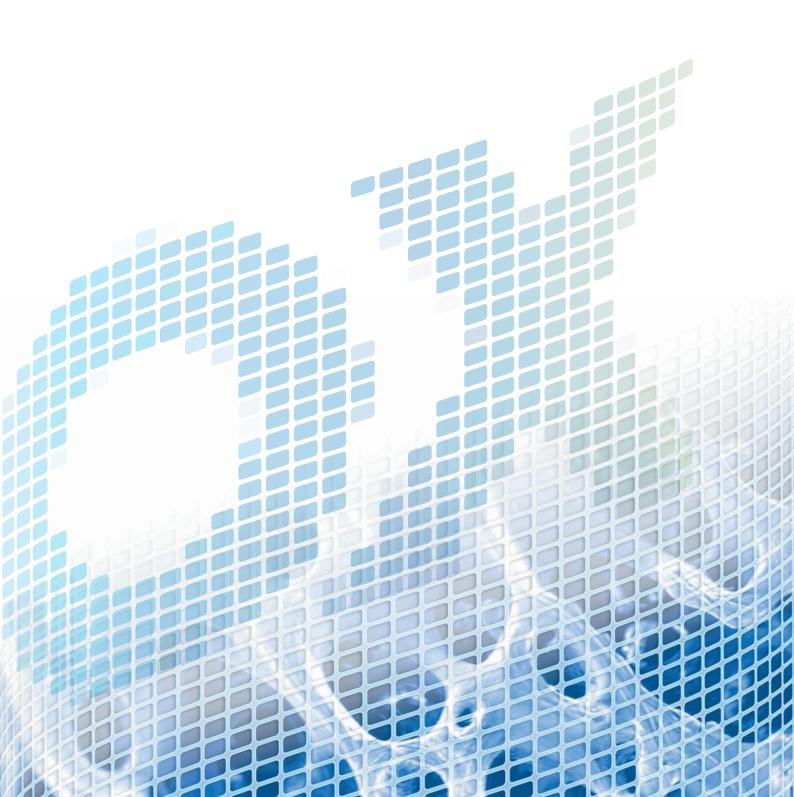


The new GBR standard



Ethics. Safety. Osteoconductivity.

A 15 years experience for the benefit of the Oral Surgeon

What is it? OSTEOXENON[®] is an advanced line of bone substitutes for regenerating bone in dental surgeries. OSTEOXENON[®] comes from a 15 years experience in Orthopedics, where this same material is grafted for huge bone reconstructions. This same biotechnological know-how and the same manufacturing process are now being applied to create bone substitutes for Oral and Maxillo-facial Surgery.

OSTEOXENON[®] is conceived and manufactured totally in Italy.

Why Equine? **OSTEOXENON**[®] is an heterologous material. Its origin is equine. This choice is not a chance.

Ethics **OSTEOXENON® is accepted by the patient:** In a multi-ethnical population comprising people belonging to different religions, patients would not accept other bone grafts (porcine or bovine).

Safety OSTEOXENON® is safe: the European Directive 2003/32/CE¹ defines equine-derived materials as safer, since no diseases, transmittable from horses to men, are currently known.

Osteoconductivity OSTEOXENON[®] is osteoconductive: Mammals share a very similar trabecular structure. Equine bones can be cut in order to achieve sections showing the same trabecular structure of human bone.

Human Bone



The two bone sections are identical. (Source: Bioteck Research Lab)

OSTEOXENON®







Enzymatic deantigenation: Biotechnology serving the Oral Surgeon

To deantigenate means eliminating all those elements that the immune system will recognize as antigens, inducing an unwanted reaction.

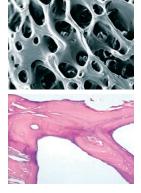
Manufacturing process

OSTEOXENON[®] is achieved through an enzymatic deantigenation process, devised by Bioteck – a leader Company in the field of Bone Substitutes manufacturing.

The **enzymatic process** is an extremely advanced method. It is based on the application of lastgeneration biotechnological processes. Mixtures of lytic enzymes clean up animal bone from any antigenic component, making it totally biocompatible.

Scanning Electron Microscopy

Hematoxylin-eosin staining



OSTEOXENON® The enzymatic process eliminates all cells. (SEM Service, Biology Dept, Padova University, Italy and Prof. N. Pennelli Histological Lab, Padova, Italy)

The enzymatic process has two main features: the temperature applied is 37°C and the process is selective. These features give **OSTEOXENON**[®] **unique-in-the-world-properties, as far as both biological response and clinical outcome are concerned.**

	Total remodeling	Collagen effects
Enzymatic deantigenation	Enzymes work in a water solution at 37°C (physiologic conditions).	By adapting the composition of the enzymatic mixture, the process can be made selective (some molecular families can be preserved).
OSTEOXENON®	The mineral component undergoes no modification, either chemical or physical.	The collagen component (Type I Bone Collagen) is totally preserved.
Biological benefit	The material is not only biocompatible. The mineral component is recognized by osteoclasts as endogenous. After 6-12 months all the grafted material is remodeled and replaced by the bone of the patient.	Type I bone collagen stimulates a great number of cellular and sub-cellular processes which are at the basis of bone regeneration.
Clinical benefit	A real bone regeneration is achieved. Not only grafting a scaffold, but a true restitutio ad integrum of the lost tissue. If osseointegrated implants are going to be placed, they will be inserted into the patient's bone, without the presence of any exogenous material.	The probability of success of regenerative surgery will be greater, since the biological conditions are optimal.
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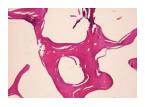
37°C: is it really important?

Nature says it is...

Preserving bone structure

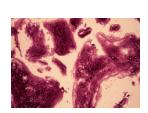
Some manufacturers apply a **thermal deantigenation process**, heating the material at a very high temperature (greater than 600°C!). The organic component sublimates, and can be easily withdrawn. Unfortunately such method causes some chemical and physical modifications to the mineral bone component, altering both its morphology and mechanical properties. Biological properties are compromised: thermal processed bone biomaterials are not only **fragile**, but also **very slowly resorbable**³, not permitting to achieve a real bone regeneration.

OSTEOXENON®



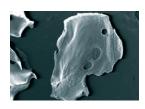
The material is identical to human bone (all cells are eliminated by the enzymatic deantigenation). Hematoxylin-eosin staining (20x). Dr. Tshering Dorji, Milano, Italy.

Thermally deproteinized bovine bone



The surface of granules is deeply altered. Their appearance is totally unnatural. Hematoxylin-eosin staining (20x).Dr. Tshering Dorji, Milano, Italy.

OSTE<mark>OX</mark>ENON®



The surface of granules is homogeneous, showing no fracture lines. *SEM Service , Biology Dept, Padova University, Italy .*

Thermally deproteinized bovine bone



The surface appears somewhat "dusty". The granule is clearly fragile. *SEM Service , Biology Dept, Padova University, Italy.*





Type I bone collagen. Which effects?

OSTEOXENON® contains, inside its structure, type I bone collagen unaltered

The importance of collagen physical process called **epitaxy**⁵. type I bone collagen, in fact: The benefits of adhesion of the cells to the grafted material collagen stimulating action of the endogenous growth factors modulating bone mineralization controlling the remodeling process proliferation and differentiation in osteoblasts action cells more sensitive to the regenerating signals **Osteoblasts** A. osteoblast portion C. mineralized collagen fibers OX[®] grafts Epitaxy of the OX[®] series. native collagen in the grafts.

Grafting bone collagen into the defect creates a precise biological condition: osteoblasts themselves, in fact, produce a collagen fiber which is then mineralized by Calcium salts⁵.

The same tridimensional structure of the collagen fiber allows the crystal formation through a

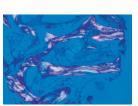
Beyond this physical effect, collagen exerts also many important biological actions⁶⁻¹⁵:

- > interacts with the beta 1 subunit of the integrins of the cellular surface of the osteoblasts to foster
- > acts as a coactivator necessary for the action of the morphogenetic proteins (BMPs) to foster the
- > binds the soluble growth factors, turning them into insoluble factors: it thus protects them from proteolysis and increases their half-life, lengthening the duration of regenerative stimulation
- > controls access of the extracellular factors to the bone crystal being formed, physiologically
- > modulates transduction of the proliferation and differentiation signal in the osteoblastic cells,
- > interacts with the mesenchymal cells coming from the bone marrow, inducing their adhesion,
- > promotes bone regeneration when grafted in bone defects, wielding a direct pro-regenerative
- > it can even stimulate the expression of the coding genes for receptor II of the BMPs, making the

Osteoblasts produce a great amount of collagen matrix that becomes a substrate for the deposition of Calcium salts. B. collagen fibers (still not mineralized)

When grafted in rat's muscles **OX**[®] bone substitutes induce the formation of Calcium salts crystals. Probably this is catalyzed by the same presence of Dept. of Biomedical Experimental Sciences, Padova University, Italy.

Bone collagen



Bone collagen presence in the **OX**[®] bone grafts **can be shown** also through polarized light: collagen fibers, having a regular structure, show a typical refringence that makes them appear brighter. Prof. N. Pennelli Histological Lab, Padova, Italy.

From biological benefits to clinical success

OSTEOXENON® gives clinical success a biological rationale

OSTEOXENON[®] bone grafts provide the oral surgeon with the real answers a bone substitute should give:

- > total replacement with the own patient's bone (total remodeling)
- > total volume preservation
- > regenerative stimulation

Total replacement

OSTEOXENON[®] is remodeled and resorbed through the action of osteoclasts.

This occurs following a totally physiologic kinetic: as the patient's bone remodels in 6-12 months, the same happens to **OSTEOXENON**[®]: after this period of time it is totally replaced by the bone of the patient.

This is possible since **OX**[®], unlike other materials, is recognized as an optimal substrate by osteoclasts, which remodel it in a physiological way¹⁶. Only in this case, in fact, the process can end with the complete substitution of the graft.



Adjacent post-extractive sockets. **OX**[®] (position 46) and deproteinized bovine bone (position 47). X-rays and 6-months second surgery. Bovine bone did not undergo remodeling, and discrete granules can be still observed. **OX**[®] instead underwent total remodeling, being replaced by

the bone of the patient. (Dr. M. Ludovichetti, Padova, Italy)

Volume preservation

If the material remodels physiologically, no volume loss can be observed. If resorption is too fast (for example, as it happens with Calcium Sulphate), or too slow (like it happens with hydroxyapatite), the endogenous bone volume is never equal to the volume grafted.

OSTEOXENON[®], instead, undergoing osteoclastic remodeling, allows to preserve the volume being grafted¹⁷.

Regenerative stimulation

OSTEOXENON[®], since it contains native type I bone collagen, creates the best condition for bone regeneration to occur.

But it provides also the oral surgeon with the possibility of stimulating the regenerative process with osteopromoting DBMs (Demineralized Bone Matrixes) that prompt the osteogenic process.

In vitro studies showed, in fact, that their action is based on the **stimulation** of blood vessels endothelial cells to migrate into the graft, and of bone marrow cells to express pro-regenerative growth factors.

There is a first evidence of their capability of accelerating bone regeneration¹⁸.

Surely this allows to increase the probability of success of bone regeneration surgeries.

Results



Bone regeneration, osteopromoting DBMs added. Results after 6 months. The quality of the regenerated tissue is easily appreciable from the hematoxylin-eosin staining. (*Prof. Danilo Alessio Di Stefano, Milan, Italy*)



References



- 1. 2003/32/CE Directive of European Commission. http://eur-lex.europa.eu/LexUriServ/LexUriServ. do?uri=CELEX:32003L0032:IT:HTML
- 2. Steele, D. Gentry; Claud A. Bramblett (1988). The Anatomy and Biology of the Human Skeleton. Texas A&M University Press.
- 3. Perrotti V, Nicholls BM, Horton MA, Piattelli A. Human osteoclast formation and activity on a xenogenous bone mineral.J Biomed Mater Res A. 2008 May 21. [Epub ahead of print]
- 4. Pagnutti S, Maggi S, Di Stefano DA, Ludovichetti M. An enzymatic deantigenation method allows achieving physiological remodeling and even osteopromoting bone grafting materials. Biotechnol. & Biotechnol. Eq. 2007. 21 (4): 491-495
- 5. John P. Bilezikian, Lawrence G. Raisz. Principles of Bone Biology. Academic Press, 2008.
- Baslé MF, Lesourd M, Grizon F, Pascaretti C, Chappard D. Type I collagen in xenogenic bone material regulates attachment and spreading of osteoblasts over the beta1 integrin subunit. Orthopade. 1998 Feb;27(2):136-42
- 7. Sampath TK, Reddi AH. Dissociative extraction and reconstitution of extracellular matrix components involved in local bone differentiation. PNAS 1981 Dec;78(12):7599-603
- 8. Paralkar VM, Nandedkar AK, Pointer RH, Kleinman HK, Reddi AH. Interaction of osteogenin, a heparin binding bone morphogenetic protein, with type IV collagen. J Biol Chem. 1990 Oct 5;265(28):17281-4.
- 9. Toroian D, Lim JE, Price PA. The size exclusion characteristics of type I collagen: implications for the role of noncollagenous bone constituents in mineralization. J Biol Chem. 2007 Aug 3;282(31):22437-47.
- Green J, Schotland S, Stauber DJ, Kleeman CR, Clemens TL. Cell-matrix interaction in bone: type I collagen modulates signal transduction in osteoblast-like cells. Am J Physiol. 1995 May;268(5 Pt 1): C1090-103.
- 11. Liu G, Hu YY, Zhao JN, Wu SJ, Xiong Z, Lu R.Effect of type I collagen on the adhesion, proliferation, and osteoblastic gene expression of bone marrow-derived mesenchymal stem cells. Chin J Traumatol. 2004 Dec;7(6):358-62.
- 12. Mizuno M, Fujisawa R, Kuboki Y. Type I collagen-induced osteoblastic differentiation of bone-marrow cells mediated by collagen-alpha2beta1 integrin interaction. J Cell Physiol. 2000 Aug;184(2):207-13.
- 13. Gungormus M. The effect on osteogenesis of type I collagen applied to experimental bone defects. Dent Traumatol. 2004 Dec;20(6):334-7.
- 14. Gungormus M, Kaya O. Evaluation of the effect of heterologous type I collagen on healing of bone defects. J Oral Maxillofac Surg. 2002 May;60(5):541-5.
- 15. Regazzoni C, Winterhalter KH, Rohrer L. Type I collagen induces expression of bone morphogenetic protein receptor type II. Biochem Biophys Res Commun. 2001 May 4;283(2):316-22.
- 16. Perrotti V, Nicholls BM, Piattelli A. Human osteoclast formation and activity on an equine spongy bone substitute. Clin Oral Implants Res. 2009 Jan;20(1):17-23
- Di Stefano DA, Artese L, Iezzi G, Piattelli A, Pagnutti S, Piccirilli M, Perrotti V.Alveolar Ridge Regeneration with Equine Spongy Bone: A Clinical, Histological, and Immunohistochemical Case Series. Clin Implant Dent Relat Res. 2008 Sep 9.
- 18. Di Stefano DA, Cazzaniga A. Chirurgia Ossea Ricostruttiva Pre e Perimplantare. Elsevier 2008.

The products

Once grafted, OX[®] bone substitutes behave according to the physiologic kinetic of patient's bone remodeling, and are completely replaced by newly-formed bone in a natural time.



> **OX37** Cancellous granules 1 bottle - 0.25 g \approx 0.5 cc granules 0.5/1 mm

> OX30 Cancellous granules 1 bottle - 0.5 g ≈ 1 cc granules 0.5/1 mm

> OX33 Cancellous granules 1 bottle - 1 g ≈ 2 cc granules 2/3 mm

> OX34 Cancellous granules 1 bottle - 1 g ≈ 2 cc granules 2/4 mm

> OX38 Cancellous granules 1 bottle - 2 g ≈ 4 cc granules 0.5/1 mm

> OX39 Cancellous granules 1 bottle - 2 g ≈ 4 cc granules 2/3 mm

> OX40 Cortical granules 1 bottle - 0.5 g ≈ 1 cc granules 0.5/1 mm

> OX35 Cortical-cancellous Mix 1 bottle - 0.25 g ≈ 0.5 cc granules 0.5/1 mm

> **OX31** Cortical-cancellous Mix 1 bottle - 0.5 g ≈ 1 cc granules 0.5/1 mm

> OX32 Cortical-cancellous Mix 1 bottle - 1 g ≈ 2 cc granules 0.5/1 mm

> OX41 Cortical-cancellous Mix 1 bottle - 2 g ≈ 4 cc granules 0.5/1 mm

> OMC-030 Calcitos 6 bottles - 0.5 g ~ 1 cc granules 0.5/1 mm



> OX01 Cancellous
 1 pc 25 x 25 x 3 mm
 > OX02 Cortical
 1 pc 25 x 25 x 2-2.5 mm



> OX21
 2 syringes, 0.25 ml each
 > OX22
 2 syringes, 0.50 ml each

OX® Collagen Gel



> OX06
 2 syringes, 0.25 ml each
 > OX07
 2 syringes, 0.50 ml each



> OX51 1 pc 10 x 10 x 10 mm > OX52 1 pc 10 x 10 x 20 mm > OX54 2 pcs 10 x 20 x 3 mm > OX55 2 pcs 10 x 20 x 5 mm > OX55R 1 pc 15 x 30 x 5 mm

OX® Membrane

> BCG-XC30 Collagen
 1 membrane 30 x 25 x 0.2 mm
 > HRT-001 Pericardium

1 membrane 30 x 25 x 0.2 mm

> HRT-002 Pericardium 1 membrane 50 x 30 x 0.2 mm

> HRT-003 Pericardium
2 membranes 15 x 20 x 0.2 mm

> OX03 Cortical 1 membrane 25 x 25 x 0.2 mm

- > OX04 Cortical
- 1 membrane 50 x 25 x 0.2 mm

OX® Angiostad DBM

> OX11
 Osteopromoting gel
 2 syringes, 0.50 ml each



> OX14 Osteopromoting granules 1 bottle, 0.5 cc



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