OHNS HOPKINS WHITING SCHOOL of ENGINEERING

Department of Computer Science

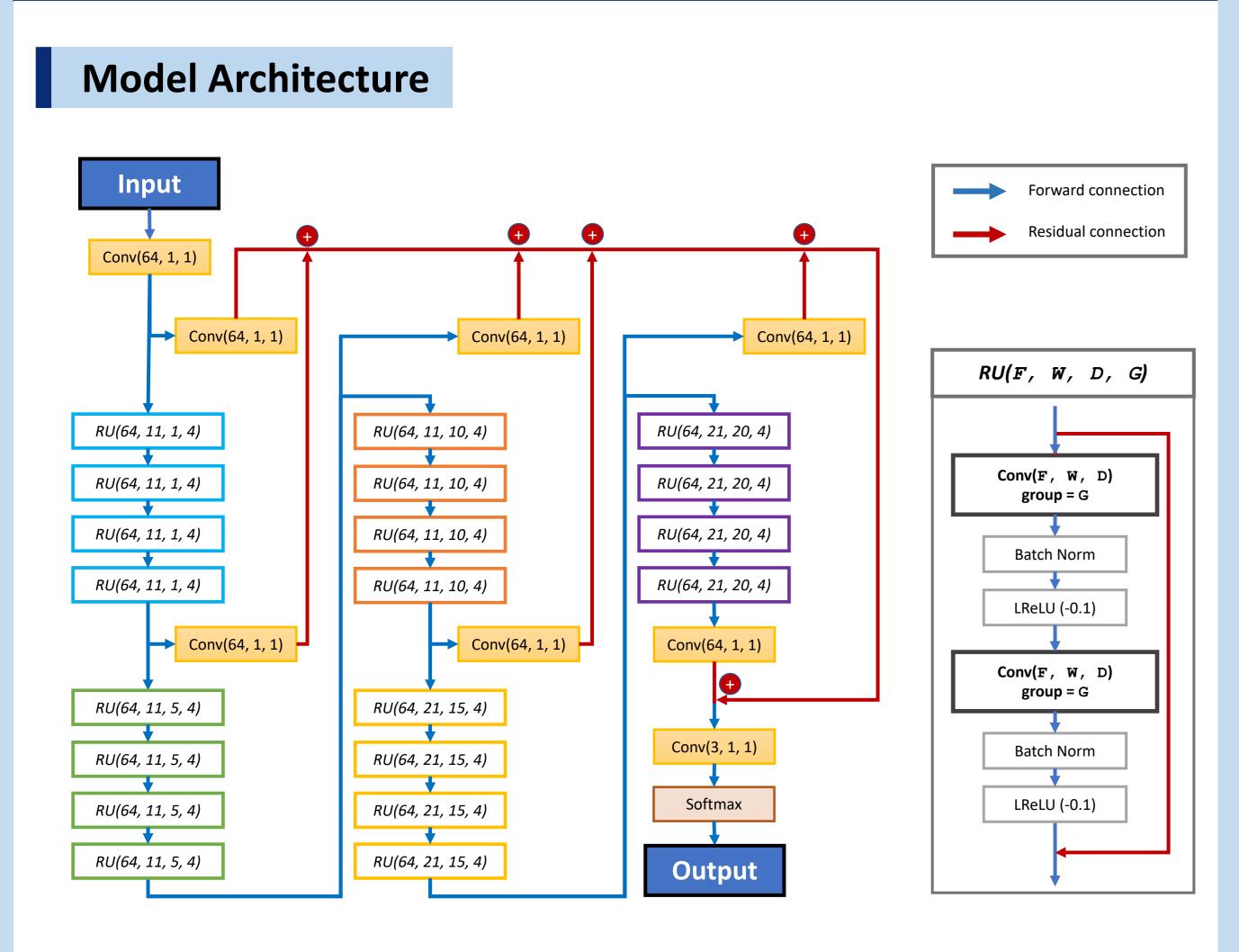
¹Department of Computer Science,

bioRχiv doi.org/10.1101/2023.07.27.550754

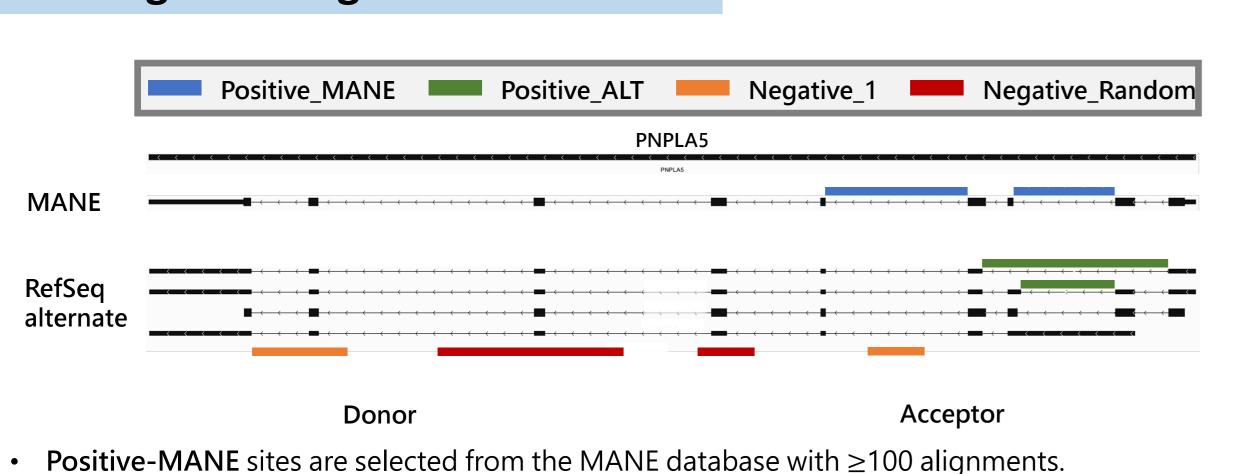


- Splam introduces the idea of training a neural network on donor and acceptor pairs together, inspired by the splicing machinery itself, which recognizes both ends of each intron at the same time.
- 2. Splam uses a limited window of 400 bp flanking each splice site, again motivated by the biological process of splicing, which relies primarily on signals within this window
- 3. Splam recognizes splice sites from genomic sequence alone more accurately than existing methods.
- Splam can improve the accuracy of transcript assemblies by removing spurious alignments produced by spliced aligners.

Methods



Training & testing dataset creation



- **Positive-Alt** sites are in RefSeq but not in MANE, also with \geq 100 alignments.
- Negative-1 sites occur on the opposite gene strand, supported by only 1 alignment. • **Negative-Random** sites are random GT-AG pairs on the opposite strand, not overlapping with known sites and lacking alignment support.

Splam: a deep-learning-based splice site predictor that improves spliced alignments

Kuan-Hao Chao^{1,2,*}, Alan Mao^{1,2,3}, Steven Salzberg^{1,2,3,4,*}, Mihaela Pertea^{1,2,3,*}

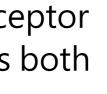
²Center for Computational Biology,

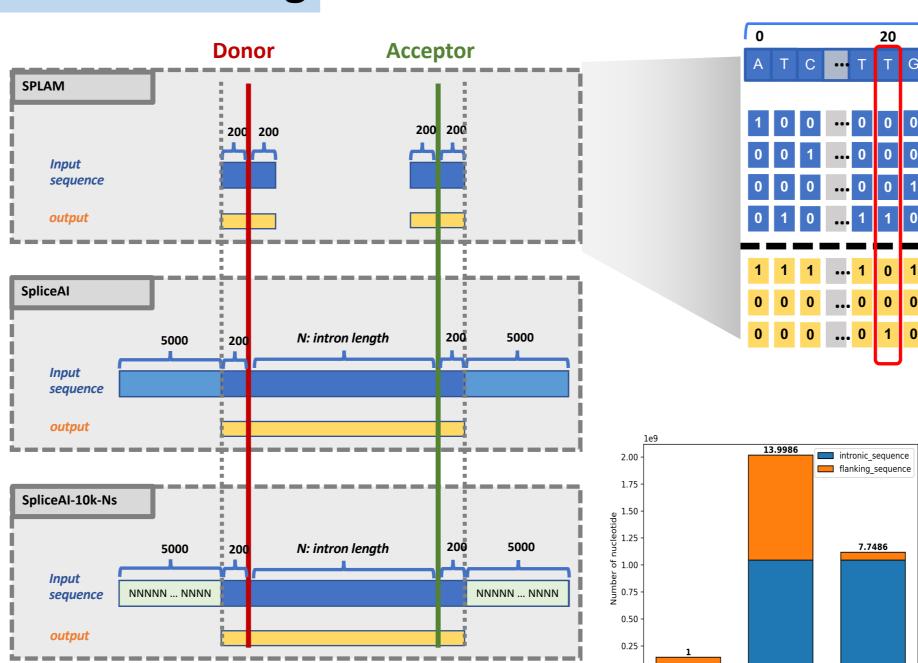
³Department of Biomedical Engineering,

Ccb.jhu.edu/splam

github.com/Kuanhao-Chao/splam

Data encoding

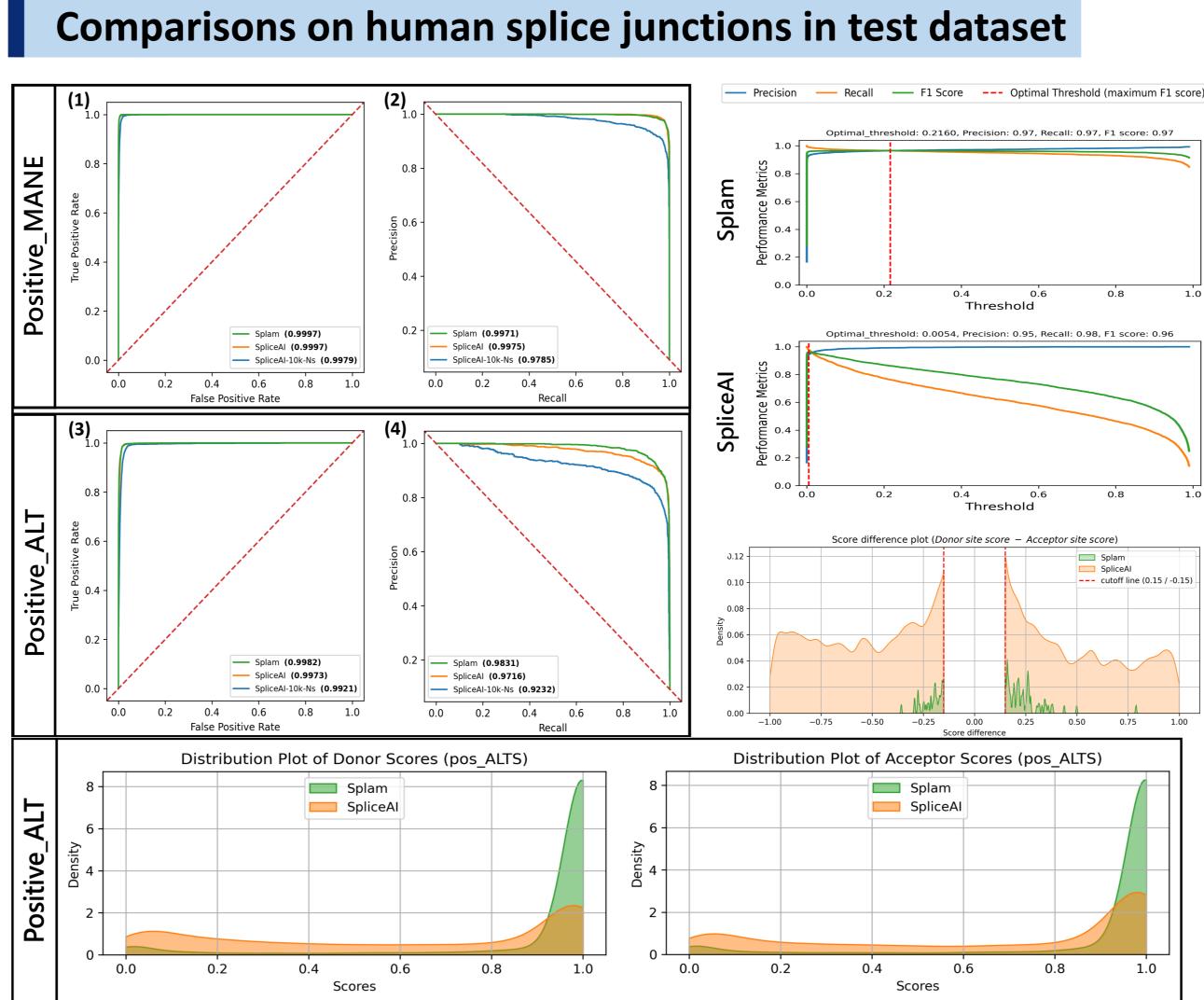




- Splam's input (top row) uses 400nt flanking the donor site and another 400nt flanking the acceptor site. The output is labels for the 800nt region (shown in yellow).
- SpliceAI's input (second row) follows its standard configuration, using 200nt upstream and downstream of the donor and acceptor sites, the entire intron, and 10Kb of flanking sequences.

SpliceAI : Jaganathan, Kishore, et al. "Predicting splicing from primary sequence with deep learning." Cell 176.3 (2019): 535-548.

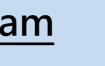
Results



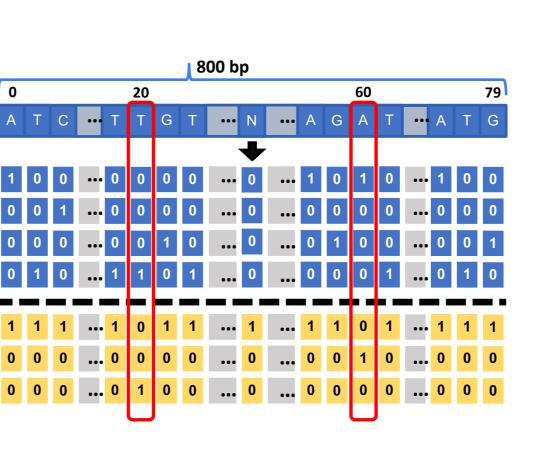
- ROC / PR curves results are shown at the junction level, where the junction score is determined by the minimum of its donor and acceptor scores.
- Discrimination threshold plots show the precision (blue curve), recall (orange curve), and F1 score (green curve) calculated at different thresholds. The optimal threshold (maximum F1 score) is indicated by a red dashed line.
- Kernel density plot shows the differences between donor and acceptor scores (donor score acceptor score) • Distribution of Splam and SpliceAI scores on alternative splice junctions.

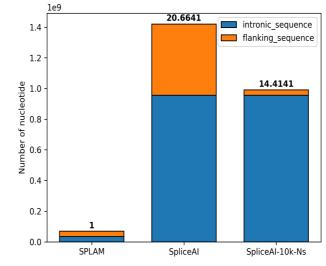


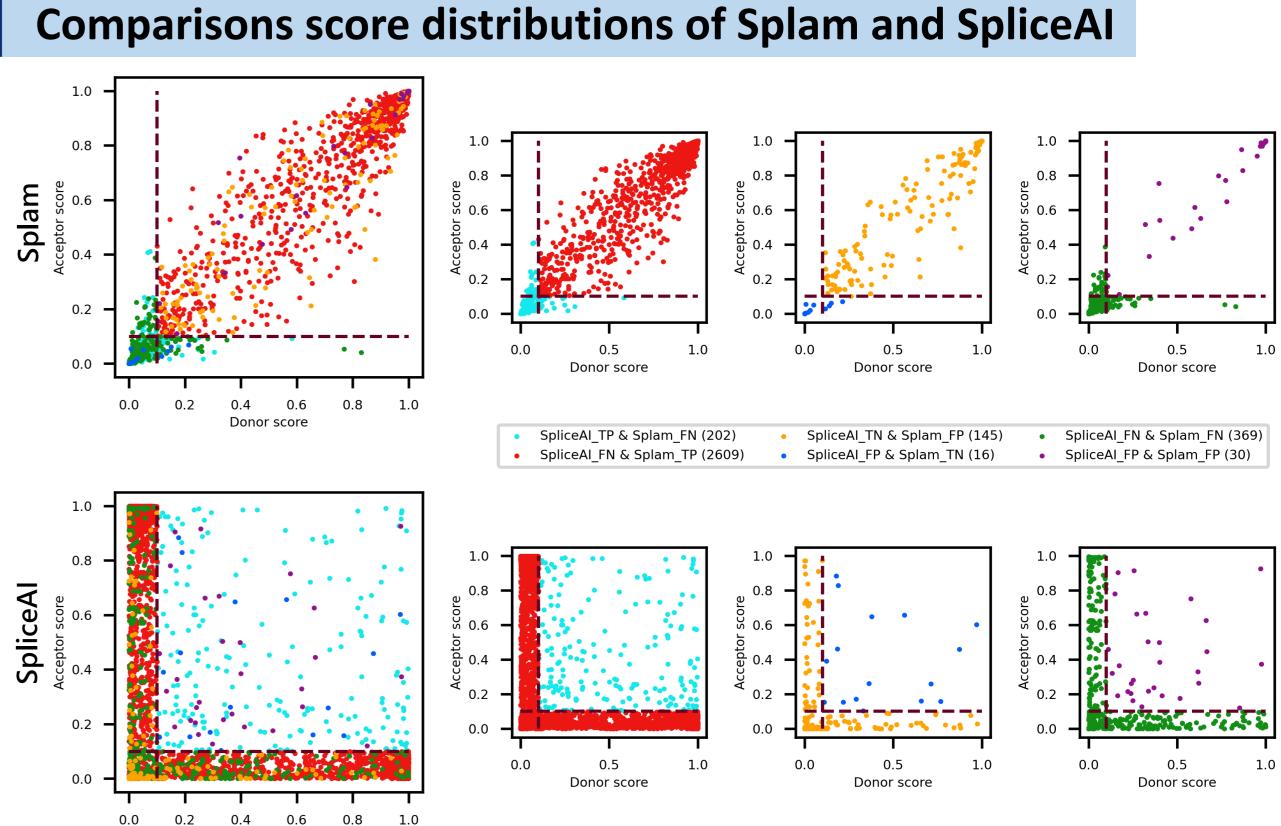
⁴Department of Biostatistics, Johns Hopkins University



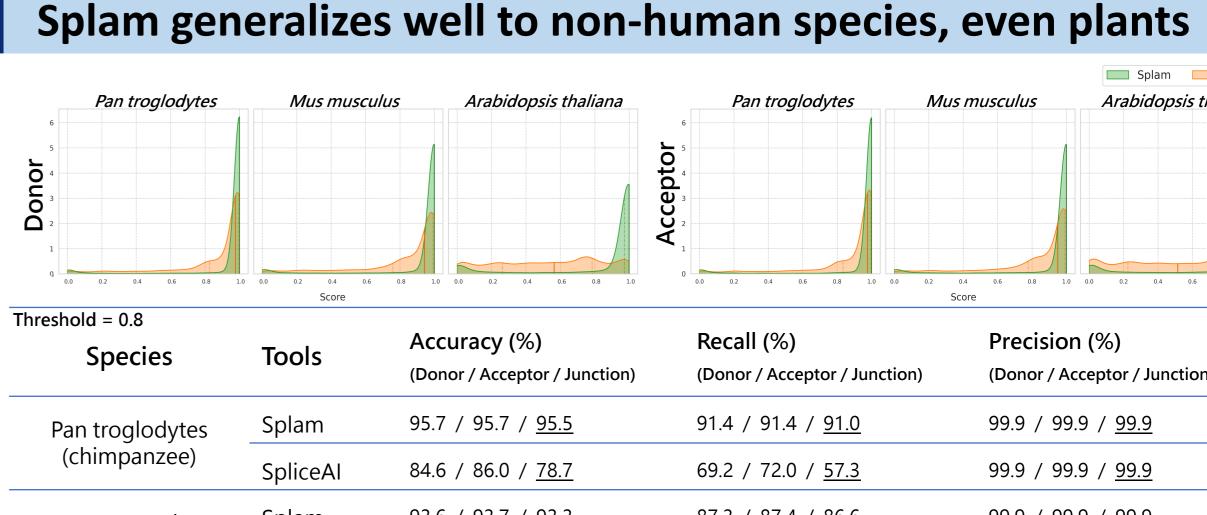
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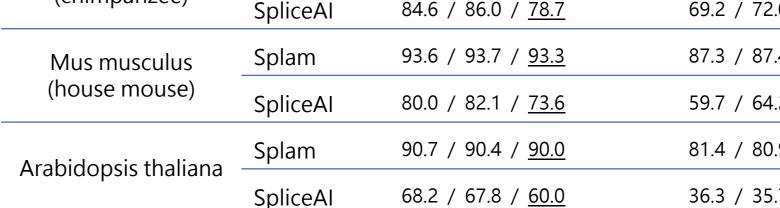


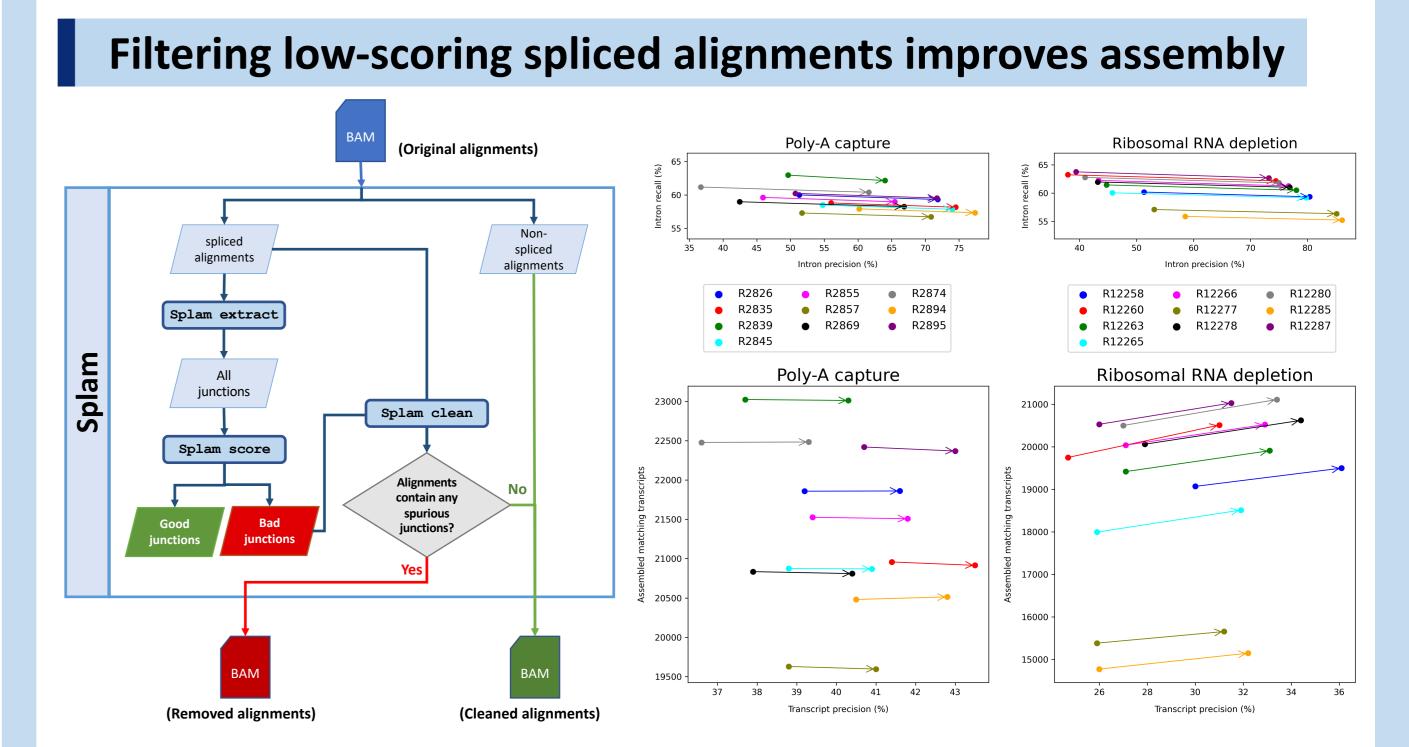




- Each dot represents a splice junction. The red dashed lines are the 0.1 cutoff threshold for labeling splice sites as true positives (TPS), true negatives (TNS), false positives (FPS), or false negatives (FNS).
- Subplots in the second & third columns show cases where one program was correct while the other was incorrect (TP and FN, or FP and TN); the fourth column shows cases where both programs made incorrect predictions (FP and FN).







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Splam SpliceAl Arabidopsis thaliana Pan troalodyte Mus musculus Score

%) Acceptor / Junction)	Precision (%) (Donor / Acceptor / Junction)
1.4 / <u>91.0</u>	99.9 / 99.9 / <u>99.9</u>
2.0 / <u>57.3</u>	99.9 / 99.9 / <u>99.9</u>
7.4 / <u>86.6</u>	99.9 / 99.9 / <u>99.9</u>
4.2 / <u>47.3</u>	99.9 / 99.9 / <u>99.9</u>
0.9 / <u>80.2</u>	99.9 / 99.9 / <u>99.9</u>
5.7 / <u>20.0</u>	99.8 / 99.9 / <u>99.9</u>