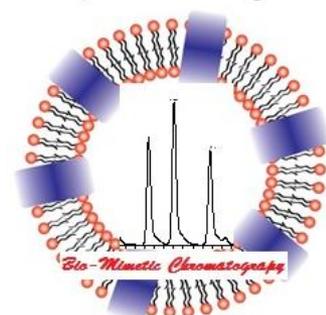


# Comparison of log P/D with bio-mimetic properties

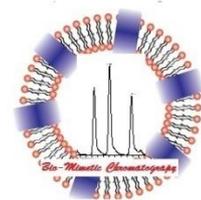


Klara Valko

*Bio-Mimetic Chromatography*

Consultancy for Successful Drug Discovery

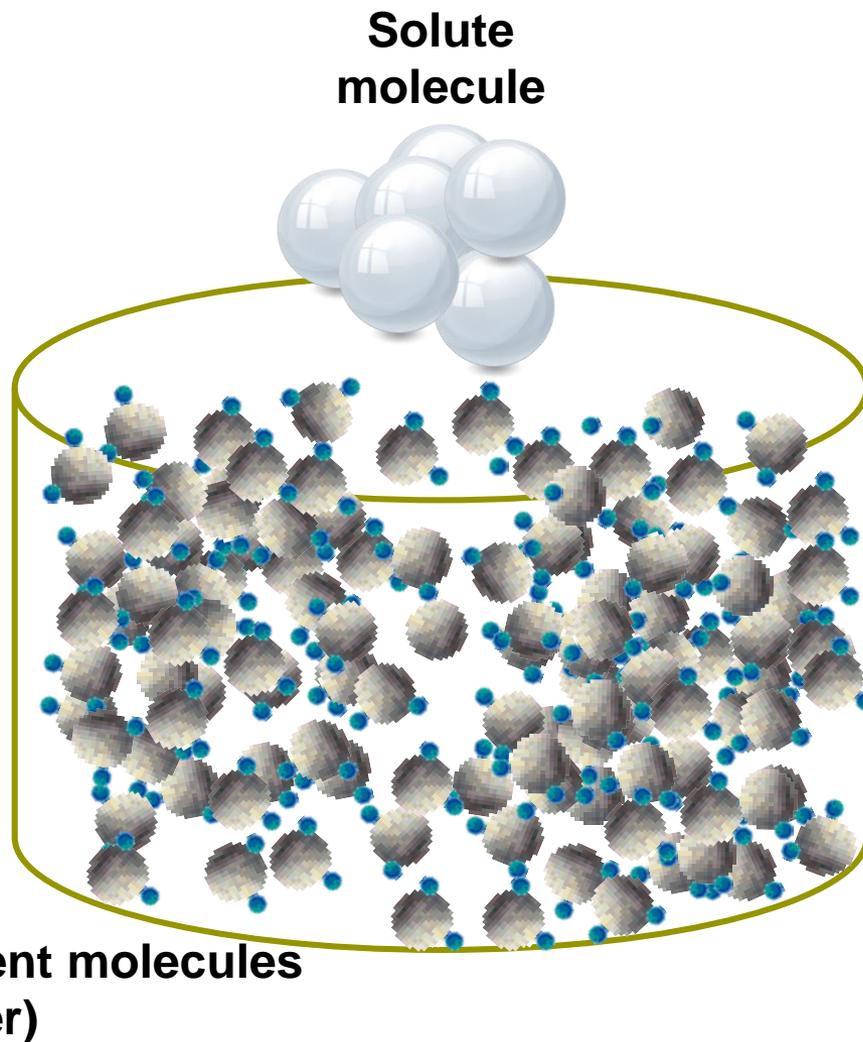
# Solvation process



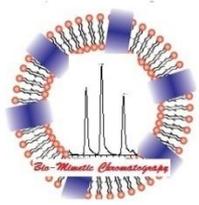
First we need  
to create a  
cavity

Requires energy that  
depends on the size  
of the solute

Size descriptor: **V**



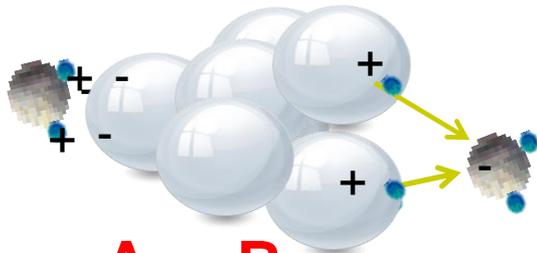
# Solvation process



Second step to build  
new interactions  
between the solute and  
solvent molecules

We gain back  
energy.

Solute –solvent  
interactions

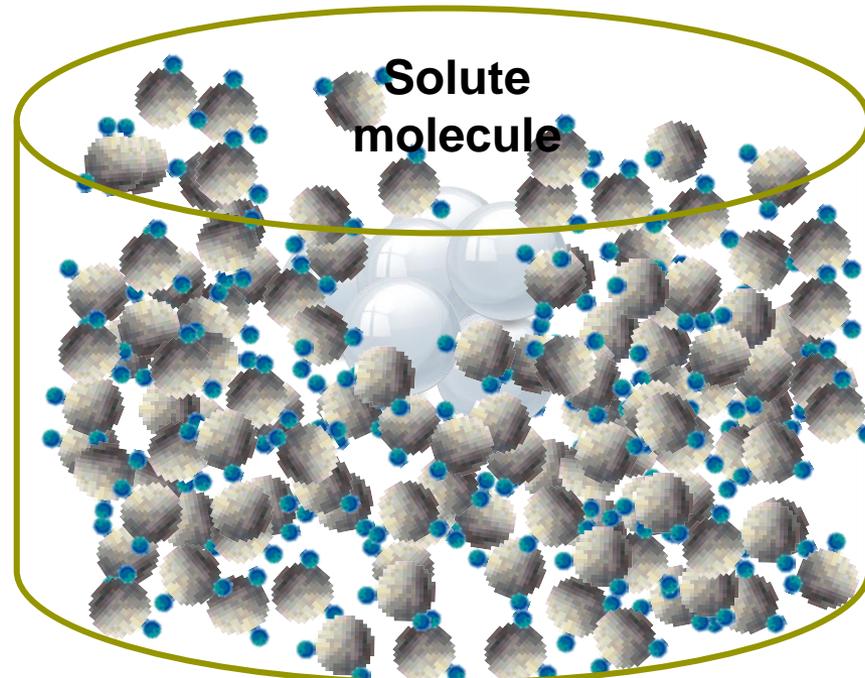


**A**    **B**



Dipole –dipole **S**

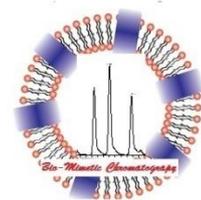
Excess molar refraction **E**



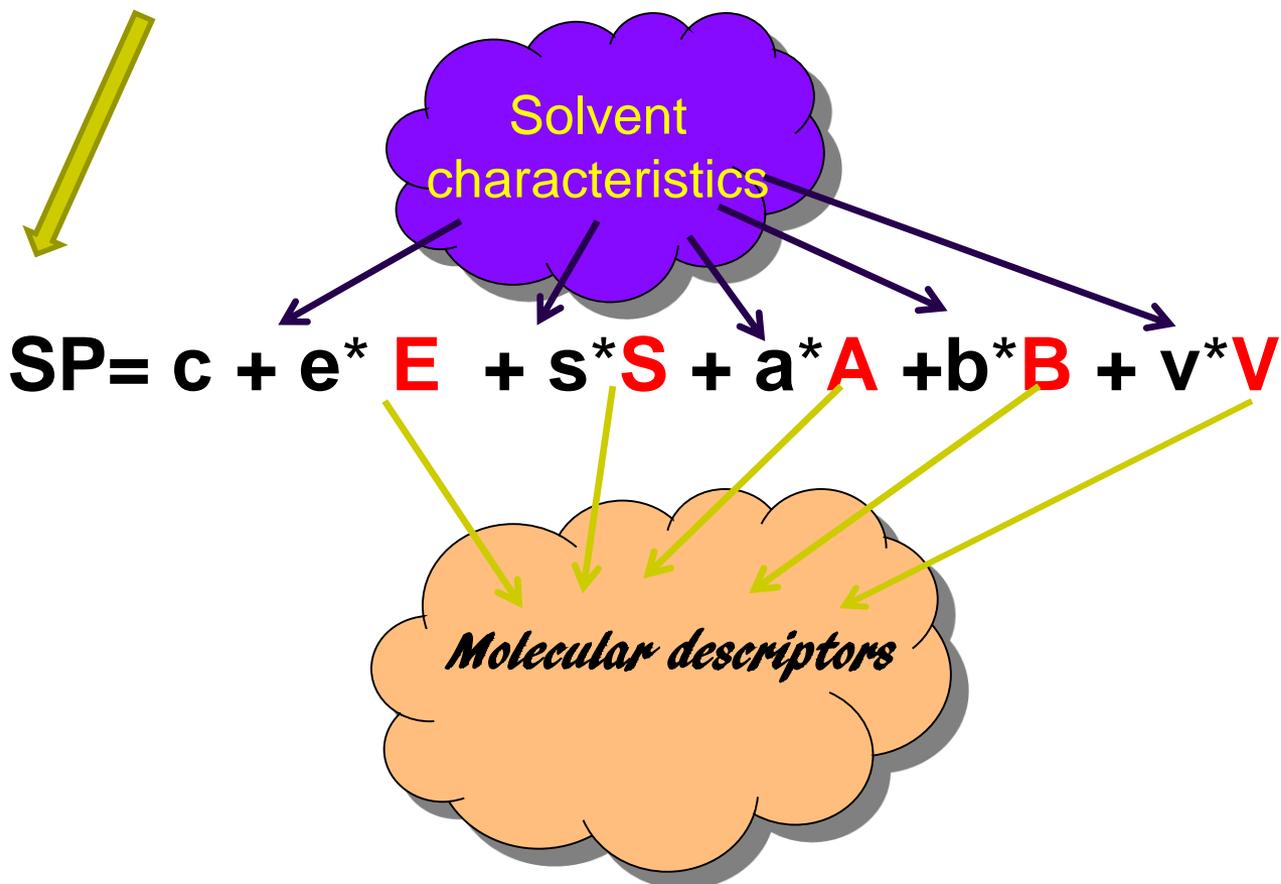
Solute molecule

Solvent molecules  
(water)

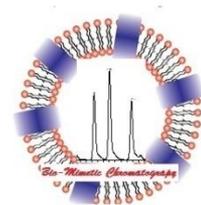
# Molecular descriptors describe various solvation and partition processes



SP is the solute property in a given system



# Solvation equations for biomimetic distributions



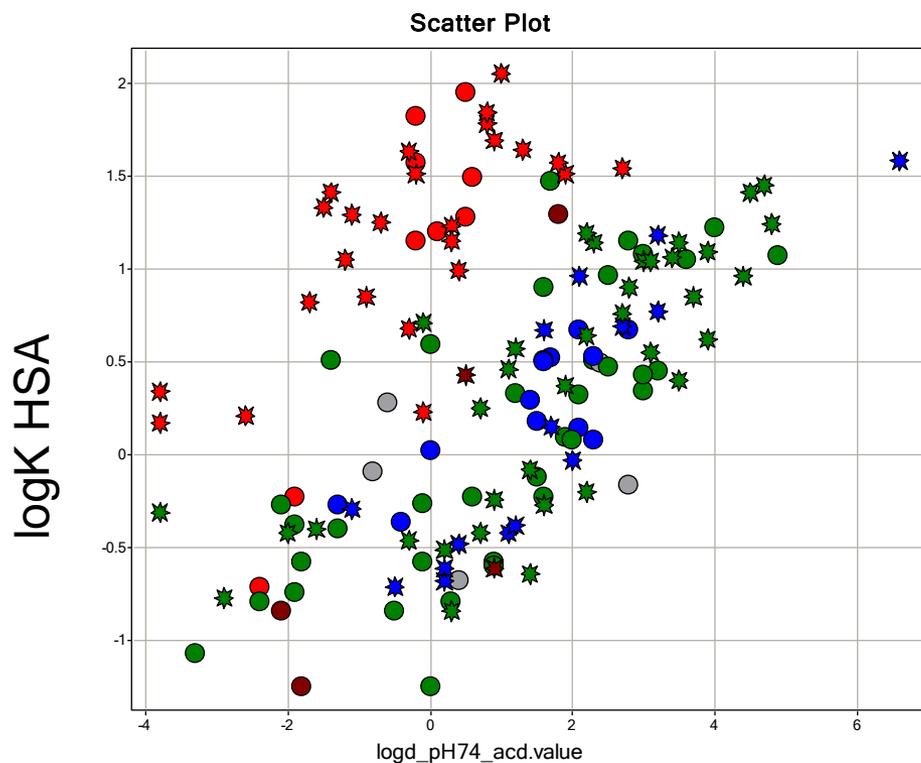
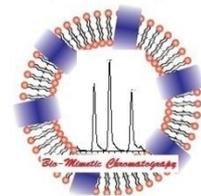
	e/v	s/v	a/v	b/v
• Log K (HSA)	0.02	-0.07	0.16	-1.21
• Log K (AGP)	0.46	-0.38	-0.33	-0.85
• $CHI_{RP,AcN}$	0.09	-0.24	-0.30	-0.98
• $\log P_{octanol}$	0.15	-0.28	0.01	-0.91
• Log K (IAM)	0.11	-0.03	0.01	-1.05
• Blood/brain	0.59	-1.03	-0.84	-0.78
• water/skin	0.00	-0.33	-0.35	-1.95

•The octanol/water partition system is an excellent model for compounds binding to HSA and IAM.

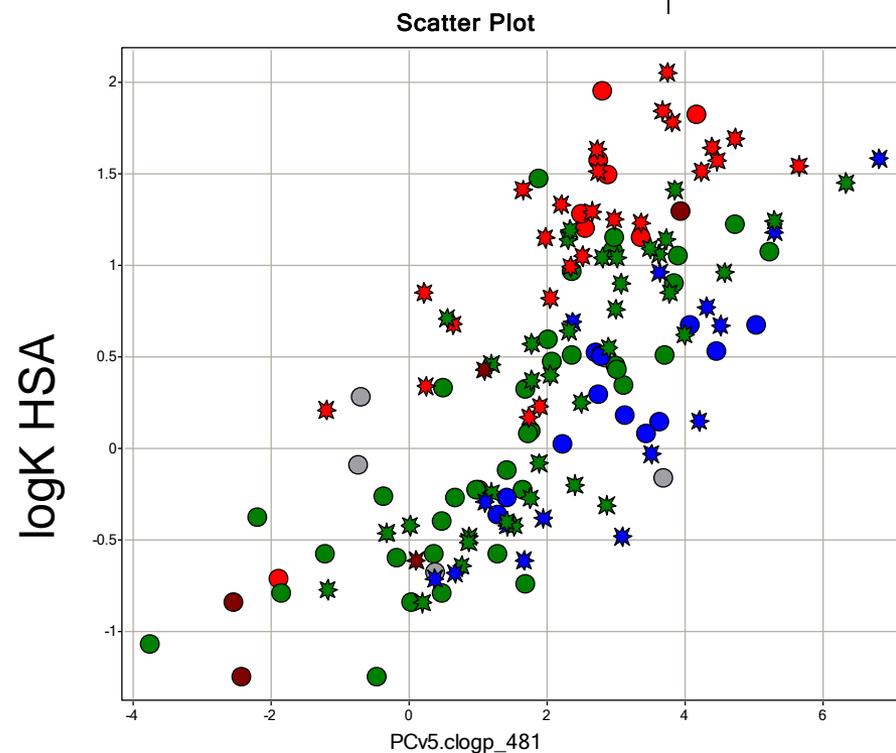
•AGP binding , CHI (reversed phase with acetonitrile) and blood/brain barrier partition are different from octanol/water.

•Equations are derived from the data of non-ionized compounds.

# Are we just measuring another lipophilicity by HSA binding?

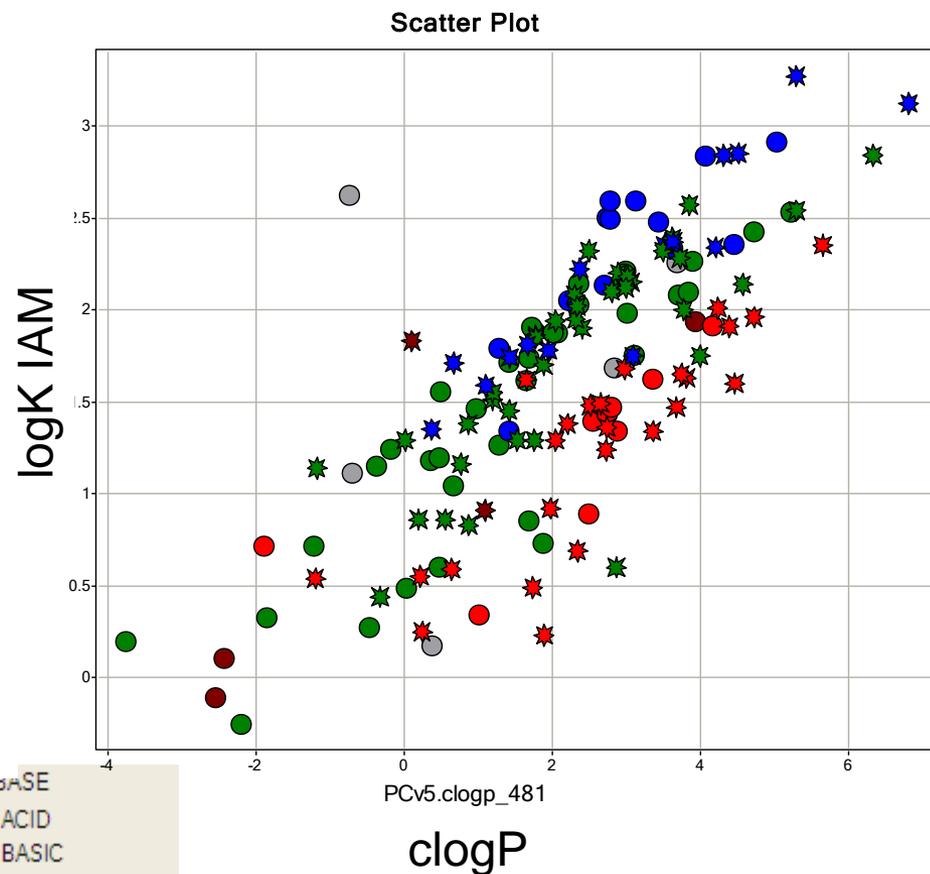
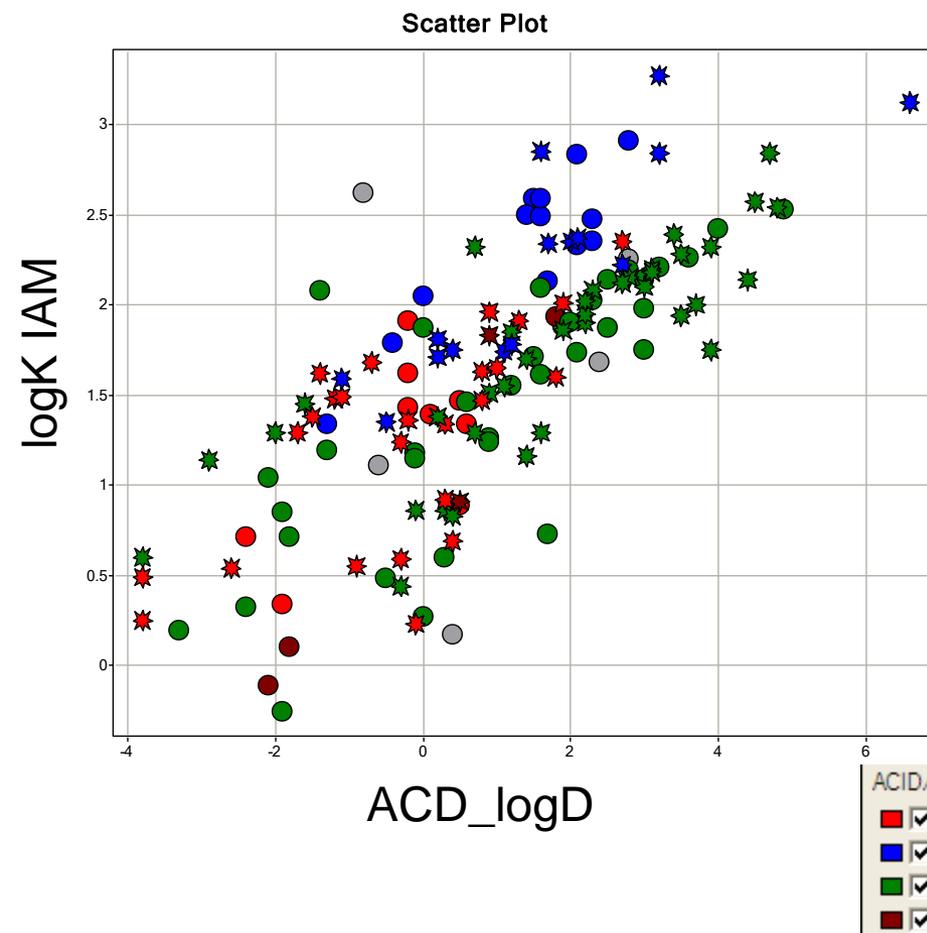
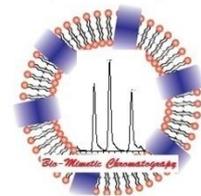


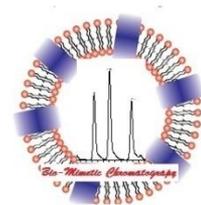
ACD\_logD



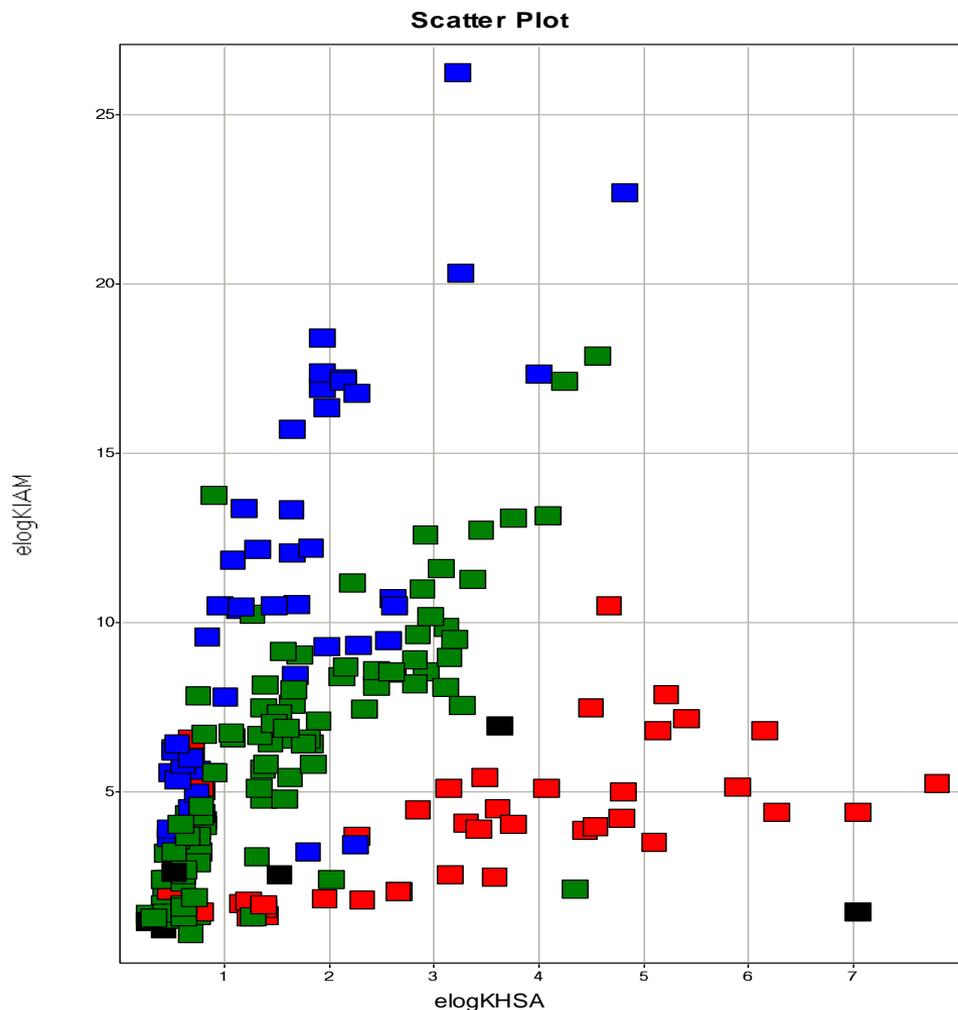
clogP

# Are we just measuring another lipophilicity by IAM binding?





# IAM binds positively charged compounds and HSA binds negatively charged compounds



Our mechanistic model shows the importance of HSA and IAM binding in modelling volume of distribution.

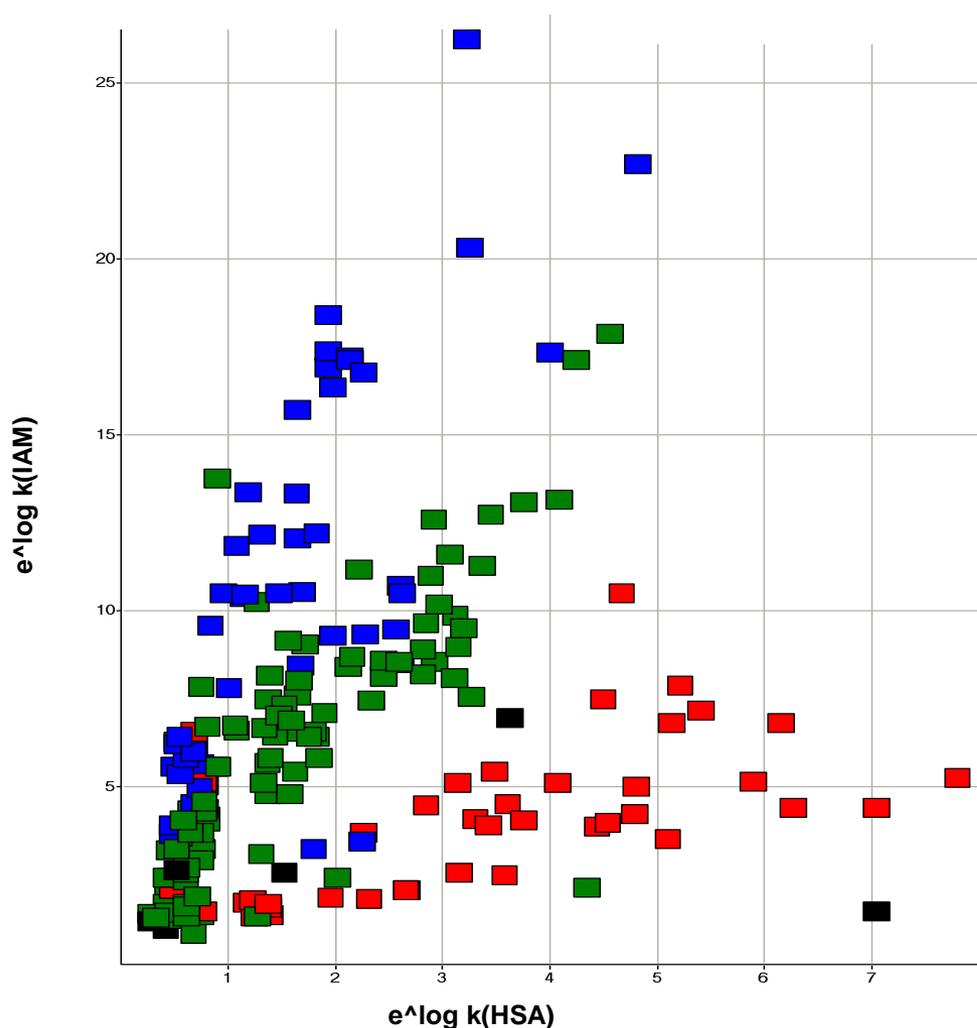
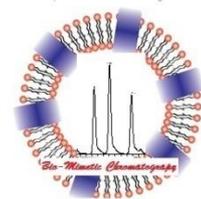
Both HSA binding and membrane partition can be related to lipophilicity of the compounds.

However the presence of positive or negative charge makes a significant difference between the two types of binding.

■ Positively charged compound   ■ Negatively charged compound   ■ Neutral compound

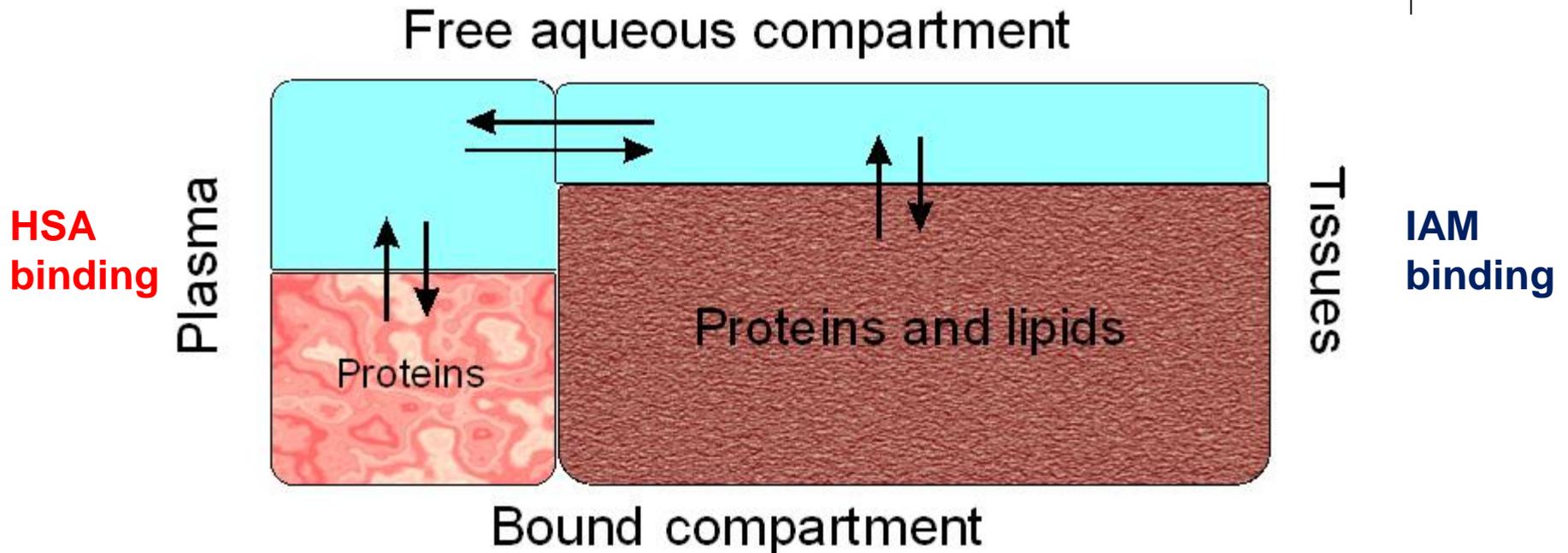
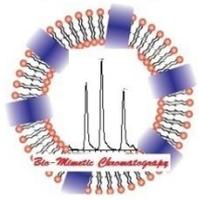
# IAM binds positively charged compounds

## HSA binds negatively charged compounds



- Positively charged compound
- Negatively charged compound
- Neutral compound

# In vivo distribution of drugs

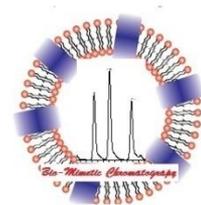


Human Clinical Volume of distribution showed good correlation to the binding **difference** of compounds between phospholipids (IAM) and albumin (HSA)

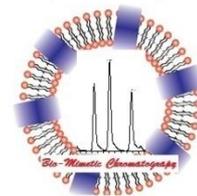
Drug efficiency showed good correlation with the **sum** of the two types of binding (IAM + HSA) which is the reciprocal value of the unbound volume of distribution  $V_{du}$

# Conclusion

- The biomimetic protein and phospholipid binding data obtained by HPLC can be used to model compounds *in vivo* distribution.
- Both protein and phospholipid bindings are governed by lipophilicity.
- There are major differences in the effect of charge on the octanol/water lipophilicity, protein binding and phospholipid binding.
- Positively charged compounds bind strongly to phospholipids and AGP, while negatively charged compounds bind strongly to albumin.
- The *in vivo* volume of distribution can be modelled by the difference of compounds binding to phospholipids (IAM) and albumin (HSA).
- The overall non-specific binding of the compounds *in vivo*, the drug efficiency and the unbound volume of distribution can be modelled by the sum of IAM and HSA binding.
- Measurement of the physico-chemical binding behaviour of compounds using biomimetic HPLC are helpful in drug design and discovery and can predict *in vivo* distribution of compounds.



# References



- Benet et al. Changes in plasma protein binding have little clinical relevance (Clin. Pharm. Therap. 71 (2002) 115.)
- Trainor The importance of plasma protein binding in drug discovery (Expert Opinion in Drug Discovery, 2, (2007) 52.
- Lombardo, F., Obach, R. S., Shalaeva, M. Y., Gao, F., Prediction of volume of distribution values in humans for neutral and basic drugs using physico-chemical measurements and plasma protein binding data. J. Med. Chem. 45 (2002) 2867.
- Hollosy, F., Valko, K., Hersey, A., Nunhuck, S., Keri, Gy., Bevan, C., Estimation of volume of distribution in humans from high throughput HPLC-based measurements of human serum albumin and immobilized artificial membrane partitioning. J. Med. Chem. 49 (2006) 6958.
- Valko, K. L., Nunhuck, S. B., Hill, A. P., Estimating unbound volume of distribution by *In vitro* HPLC-based human serum albumin and immobilized artificial membrane-binding measurements. J. Pharm. Sci., 100 (2011) 849.
- S. Braggio, D. Montanari, et al. Expert Opin Drug Discovery, Drug efficiency: a new concept to guide lead optimization programs towards the selection of better clinical candidates. 5 (7) (2010) 609
- D. Montanari, E. Chiarparin, M. P. Gleeson, S. Braggio, R. Longhi, K. Valko, T. Rossi, Application of drug efficiency index in drug discovery: a strategy towards low therapeutic dose. Exp. Opin Drug Discovery 6 (9) (2011) 913-920
- Testa et al. Perspectives in Drug Discovery and Design: 19 (2000) 179-211