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A 9000-year carbon isotopic record of acid-soluble organic matter in a stalagmite from Heshang Cave, central China: Paleoclimate implications

Xiuli Li^a, Chaoyong Hu^{a,*}, Junhua Huang^b, Shucheng Xie^a, Andy Baker^c

^a State Key Laboratory of Biogeology and Environmental Geology, China University of Geosciences, Wuhan, Hubei 430074, China
^b State Key Laboratory of Geological Processes and Mineral Resources, China University of Geosciences, Wuhan 430074, China

^c State Key Laboratory of Geological Processes and Mineral Resources, China University of Geosciences, Wunan 430 ^c Connected Waters Initiative Research Centre, UNSW Australia, NSW 2052, Australia

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ABSTRACT

Organic matter preserved in speleothems has the potential to reflect the changes of the overlying vegetation and soil ecosystem. Here we report the first near 50-yr-resolution acid-soluble organic matter (ASOM) carbon isotope $(\delta^{13}C_{ASOM})$ sequence derived from a laminated stalagmite (HS-4) spanning the last 9 ky in Heshang Cave, central China. The $\delta^{13}C_{ASOM}$ values vary between -25.8% and -22.0%, with lower values from 9 to 4 ka BP and less negative values from 4 to 0 ka BP. We postulate that the $\delta^{13}C_{ASOM}$ sequence is mainly controlled by temperature and water balance. Temperature could affect both vegetation physiology and microbial degradation in soil horizons. The influence of temperature on the $\delta^{13}C_{ASOM}$ is supported by the negative correlation (r = -0.48, p < 0.001) between the $\delta^{13}C_{ASOM}$ record and the paleotemperature record in the nearby Dajiuhu peatland. The water balance can affect the retention time of organic matter in soils. Under drier conditions, the soil organic matter will be retained longer and is more likely to be biologically degraded, resulting in more negative $\delta^{13}C_{ASOM}$ values. Our results reveal that $\delta^{13}C_{ASOM}$ in speleothems has the potential to reflect the response of vegetation and/or soil processes to paleoclimate changes.

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1. Introduction

Stalagmites are one of the most important archives for highresolution paleoclimate reconstructions in the Quaternary, with many climatic proxies developed (McDermott, 2004; Blyth et al., 2008; Fairchild and Treble, 2009). Among these proxies, the carbon isotope composition of calcite ($\delta^{13}C_{carb}$) has been widely investigated and is normally interpreted as an indicator of overlying vegetation changes (e.g. Genty et al., 2001; Frappier et al., 2002; McDermott, 2004; Cosford et al., 2009). However, the carbon isotopic signal of dissolved CO₂ can be affected by other processes, especially fractionation during carbonate precipitation and degassing (Baker et al., 2011; Dreybrodt and Scholz, 2011; Frisia et al., 2011), leaving an uncertainty in the interpretation of $\delta^{13}C_{carb}$ as a record of vegetation (e.g. Genty et al., 2003; Blyth et al., 2007).

In comparison to inorganic carbon, organic matter acts as a minor carbon pool in speleothems. Plenty of researchers have exploited the paleoecological and paleoenvironmental potential of lipid biomarkers in speleothems (e.g. Xie et al., 2003; Blyth et al., 2008, 2014; Huang et al., 2008a,2008b; Rushdi et al., 2011; Yang et al., 2011; Blyth and Schouten, 2013). Other than lipid biomarkers, humic substances, especially fulvic acids have been widely investigated due to their

E-mail address: chyhu@cug.edu.cn (C. Hu).

luminescence features, which correlate with the overlying soil and vegetation changes (McGarry and Baker, 2000; Fairchild and Baker, 2012). Both fluorescence analysis and TOC monitoring of drip water reveal that fulvic acids are the predominant humic substances in drip water and speleothems (van Beynen et al., 2002; Smailer and White, 2013).

The δ^{13} C values of organic matter ($\delta^{13}C_{org}$) in speleothems have considerable importance in paleoenvironmental research (Blyth et al., 2013a.2013b), as they have the potential to directly record how the overlying soil ecosystem responses to paleoclimate changes. Studies using $\delta^{13}C_{org}$ in karst systems are quite limited. A few studies have discussed $\delta^{13}C_{org}$ in cave sediments (Turney et al., 2001; Panno et al., 2004; Polk et al., 2007, 2013). Polk et al. (2013) used carbon isotopes from fulvic acids (FAs) in cave sediments to develop a record of vegetation over the last 3000 yr. Recently Blyth et al. (2013a) developed a novel liquid chromatography-isotope ratio mass spectrometry to determine the δ^{13} C values of non-purgeable organic carbon (NPOC) in speleothems. Immediately, Blyth et al. (2013b) reported a time series of the δ^{13} C of NPOC and long chain *n*-alkanes from a 2000-year old Scotland stalagmite, and demonstrated a negative correlation between NPOC and carbonate isotope values. They hypothesized that this was due to preferential use of ¹²C by microbes during periods of increased activity, resulting in enrichment of the residual OM, and depletion of the CO₂. The above studies highlight that the δ^{13} C of the organic matter pool in cave systems has the potential to track the paleoenvironmental changes.



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^{*} Corresponding author.

In this study, we isolated acid-soluble organic matter (ASOM) from an absolutely dated stalagmite (HS-4) from Heshang Cave and measured the carbon isotope composition using a relatively small amount of rock sample (ca. 2 g). We conducted infrared spectroscopy (IR) analysis to provide evidence that the extracted ASOM mainly originates from the overlying soil organic matter. Then we discuss the plausible processes affecting the ASOM carbon isotope ($\delta^{13}C_{ASOM}$) variations in HS-4 stalagmite over the last 9 ky.

2. Materials and methods

2.1. Locality and sampling

Heshang Cave (30°27′N, 110°25′E, 294 m altitude) is located on the south bank of the Qingjiang River, a large tributary of the Yangtze River in central China (Fig. 1). The geographic and climatic settings of this cave have been shown in detail in Hu et al. (2008a). In general, this region is affected mostly by the East Asian monsoon, with high temperatures and high precipitation in summer months.

The samples used in this study were collected from the HS-4 stalagmite (Hu et al., 2008b). This stalagmite (2.54 m in length) was still actively growing when it was removed. The chronology of HS-4 was established independently by U–Th dating and layer counting (Hu et al., 2008b). In order to avoid organic contamination, all stalagmite samples were selected from the fresh internal parts after discarding the surface layer (Hu et al., 2008b). In total, using a dental-drill, 170 samples were collected from HS-4, all of 1–2 cm thickness and parallel to the growth layers. HS-4 stalagmite has an approximately constant growth rate, averaging 0.27 mm per year (Hu et al., 2008b). Thus the temporal resolution of the samples for ASOM analysis is about 40– 80 years. At one location (173 cm from the top), four samples were collected within a single growth layer.

Surface samples were collected from the overlying soil (0–2 cm; 30°2.334'N, 110°25.148'E, 961 m altitude) above Heshang Cave in July and November 2012. A subsoil sample was collected from the soil pro-file (97–103 cm depth below surface; November 2012). In addition,

two drip water samples (ca. 4 L in a brown bottle) were collected at the former growth site of HS-4 stalagmite in July and November 2012. In addition, a limestone sample was collected from the bedrock of Heshang Cave.

2.2. ASOM extraction

The stalagmite samples were air-dried and crushed to pass an 80mesh sieve. The powdered stalagmite samples (ca. 2 g) were dissolved in 6 M HCl and kept overnight. Then the acid solution (pH < 2) was centrifuged at a rate of 8000 r/s to separate the supernatant and insoluble matter. The supernatant was freeze-dried and is referred as ASOM in this study. After acid dissolution, the insoluble fraction was near absent. Thus we only focused on the acid soluble fraction in this study. The soil samples were air-dried at room temperature and then were treated by the same procedure as the stalagmites. Water samples (ca. 0.5 L) were evaporated to dryness at 60 °C and were subsequently treated in the same manner as the soil and stalagmite samples.

Three stalagmite samples (33.0 cm, 118.4 cm, 198.4 cm to the top) were selected for analysis of ASOM concentration using TOC/TC analyzer (Analytik Jena AG Multi N C 3100). The reproducibility was less than 1.5%.

2.3. Carbon isotope composition analysis

The carbon isotopic compositions of the ASOM were determined via continuous flow on a Thermo Flash Elemental Analyzer (EA) coupled with a Thermo MAT-253 isotope ratio mass spectrometer (IRMS) at Nanjing Center, Geological Survey Bureau of China (NJ) and State Key Laboratory of Geological Processes and Mineral Resources, China University of Geosciences at Wuhan (WH). The ASOM was oxidized to CO_2 at 1000 °C with chromium oxide. The carbon isotope composition in the resultant CO_2 was measured by a MAT-253 mass spectrometer. The $\delta^{13}C_{ASOM}$ values were normalized to the VPDB (Vienna Pee Dee Belemnite) standard using a charcoal reference material (GBW04407, with a $\delta^{13}C$ value of -22.43%). The long-term reproducibility of the



Fig. 1. Locality of Heshang Cave in China.

internal standards was better than 0.2‰. It is cautioned that the precipitated acid salts can to some extent affect the EA performance.

2.4. Spectroscopic analyses

2.4.1. IR analyses

IR analyses on selected soil, drip water, bedrock and stalagmite samples were performed on a Thermo Fisher Nicolet 6700 Fourier transform infrared spectrometer (FT-IR) at the School of Material and Chemistry, China University of Geosciences, Wuhan. The freeze-dried ASOM was crushed in a mortar and mixed with potassium bromide (KBr, FT-IR grade) at a ratio of 1:200 and pressed into pellets. These pellets were investigated in transmission mode in the mid-infrared range (4000–400 cm⁻¹) with 32 scans per spectrum and a resolution of 4 cm⁻¹. The spectra were evaluated after height normalization and automatic baseline correction by using the evaluation software provided with the FT-IR spectrometer.

2.4.2. Synchronous fluorescence analysis

Recently, we reported an improved synchronous fluorimetric method for the determination of dissolved organic matter in cave drip water (Li et al., in press). Briefly, the stalagmite samples were dissolved in dilute HCl. Then an aliquot (ca. 10 mL) was transferred to a 25 mL test tube and ascorbic acid was added. Fluorescence analysis was conducted with a PerkinElmer LS-55 Luminescence Spectrophotometer using a 150 W Xe lamp. The synchronous fluorescence intensity (SFI) was measured using a 1 cm quartz cell with wavelength scanning from 200 nm to 500 nm with $\Delta\lambda$ at 20 nm.

3. Results

3.1. ASOM spectroscopic results

The ASOM extracts from the overlying soil, drip water and stalagmite show similar IR spectra, exhibiting a strong and wide absorption band near 3400 cm^{-1} and a secondary band at 1630 cm^{-1} (Fig. 2). The former corresponds with the stretching vibrational absorption of phenolic O– H bond, whereas the latter relates with the stacked absorption peaks of C=C and C=O groups and associated hydrogen bonds (Kim et al., 1990). These spectra are quite typical of those reported for fulvic-acids (Stevenson and Goh, 1971; Senesi et al., 1989; Kim et al., 1990). Comparing the bedrock sample to the soil, water and stalagmite samples, the bedrock shows a quite different IR spectrum, with strong absorption bands near 3667, 3069, 2258 and 1677, and 1601 cm⁻¹ (Fig. 2), which is similar to the spectra of matured kerogen (Lis et al., 2005). During the synchronous fluorescence analysis, the wavelength of maximum fluorescence intensity of the HS-4 stalagmite samples varies between 391 nm and 385 nm and displays a general decreasing trend with time (Fig. 3). The interval between 1.6 and 0 ka has relatively low wavelength of maximum fluorescence.

3.2. Carbon isotopic composition of ASOM

Three samples have been analyzed to determine the concentration of ASOM. These three samples show low but constant ASOM content (0.12–0.14 mg/g calcite). The ASOM concentration is similar with the TOC concentration in Blyth et al. (2013a; 0.11 mg/g calcite), but lower than the TOC concentration in BW-1 stalagmite from Beijing, China (Duan et al., 2014; 0.6–2.5 mg/g calcite).

The isotopic compositions of our samples were analyzed on two different EA-IRMS systems. In order to check for any inter-laboratory differences, three samples (HS4-13, HS4-56 and HS4-133) were selected to be analyzed at both laboratories. These samples exhibit quite similar $\delta^{13}C_{ASOM}$ values, with deviation less than 0.1‰. The reproducibility of our ASOM extraction procedure was evaluated by triplicate treatment of selected samples (HS4-56 and HS4-132) using the same procedure. These results show relatively small derivations (0.1‰ and 0.4‰, respectively). Thus we assume the analytical error for $\delta^{13}C_{ASOM}$ in this study is between 0.2 and 0.4‰, which is relatively higher than the long-term repeatability of $\delta^{13}C_{carb}$ (0.1‰; Hu et al., 2008b).

Four sub-samples (HS4-C1–HS4-C4) were collected across the single growth layer (173 cm from the top) for $\delta^{13}C_{carb}$ and $\delta^{13}C_{ASOM}$



Fig. 2. FT-IR spectra of ASOM isolated from the overlying soil, bedrock, drip water and stalagmite samples.



Fig. 3. Comparison of $\delta^{13}C_{ASOM}$, and the synchronous fluorescence wavelength in HS-4 stalagmite and paleotemperature reconstructed in the Dajiuhu peatland (Huang et al., 2013) over the last 9 ky. The dashed lines above and below the $\delta^{13}C_{ASOM}$ curve refer to the analytical error of 0.4%.

analyses. The $\delta^{13}C_{carb}$ values progressively become larger towards the edge, with a variation of ca. 2‰ (Hu et al., 2008b; Fig. 4). In contrast, $\delta^{13}C_{ASOM}$ shows quite small variation (<0.3‰) in the single growth layer (Fig. 4).

The $\delta^{13}C_{ASOM}$ values of HS-4 range from -25.8 to -22.0%, averaging -24.2% (Fig. 3). The $\delta^{13}C_{ASOM}$ values are relatively more negative during the period from 9 to 4 ky BP (averaging -24.6%) and less negative from 4 to 0 ky BP (averaging -23.8%). In contrast, the $\delta^{13}C_{carb}$



Fig. 4. (Left) The sampling sites along the growing band at 173 cm of HS-4. (Right) Variations of $\delta^{13}C_{ASOM}$ and $\delta^{13}C_{carb}$ values along the individual growing band. The average analytical error is about 0.2–0.4‰ for $\delta^{13}C_{ASOM}$, while the long-term repeatability for $\delta^{13}C_{carb}$ is 0.1‰.

record shows relatively higher values during 9–8 ky BP and then varies around an average value of -11% (Hu et al., 2008b), with a quite different trend than that of $\delta^{13}C_{ASOM}$.

The two surface soil samples (0–2 cm) show quite similar $\delta^{13}C_{ASOM}$ results (soil-s: -23.7% in July vs. soil-w: -24.2% in November). The $\delta^{13}C_{ASOM}$ values of the drip waters are -24.8% in July (water-s) vs. -25.3% in November (water-w) 2012, and are similar to the topmost sample of HS-4 stalagmite (-25.4%; ~ 12 yr BP).

4. Discussion

4.1. Origin of ASOM in HS-4 stalagmite

The similar FT-IR spectra of the overlying soil, drip waters and stalagmites clearly provide evidence that the ASOM preserved in HS-4 mainly originates from the overlying soil, with a quite limited contribution from cave bedrock. In addition, the FT-IR spectra of the overlying soils are comparable with those of FAs, suggesting that FAs are the dominant contributor of ASOM. Organic acids, mainly FAs and humic acids (HAs), make up about 60–70% of the total organic carbon in soils (Senesi, 1993; McGarry and Baker, 2000). FAs are soluble in water at any pH, whereas HAs are insoluble in water with pH < 2 (Senesi, 1993). In fact, many studies demonstrate that fulvic-like matter is the dominant natural fluorescent substances in cave drip waters (Baker and Genty, 1999) and speleothems (e.g. Baker et al., 1998; McGarry and Baker, 2000; Blyth et al., 2008). However, we could not exclude the contribution of non-humic and fulvic-like substances to ASOM in this study, and we hence refer to it as ASOM rather than FA.

4.2. Climatic implications of stalagmite $\delta^{13}C_{ASOM}$

Because the $\delta^{13}C_{ASOM}$ values in HS-4 stalagmite are independent of fractionation occurring during degassing and calcite deposition (Fig. 4), the majority of the $\delta^{13}C_{ASOM}$ fluctuations must originate from the overlying vegetation and soil ecosystem. Fluorescence studies of speleothem organic acids also support that soil cover, vegetation and associated climate changes exert more important influence on this stalagmite organic matter proxy than internal cave processes (McGarry and Baker, 2000).

4.2.1. Influence from vegetation

Heshang Cave (294 m above the sea level) forms in Cambrian dolomite and the overlying hillsides have an altitude up to 900 m (Hu et al., 2008a). According to the vegetation in the nearby Houhe National Nature Reserve (29°59′-30°10′N, 110°22′-110°52′E) in the Wufeng Tujiazu Minority Autonomous County of Hubei Province, the region between 400 and 1150 m is dominated by evergreen broadleaf (Li et al., 2005). In the study region, C₃ plants dominate the vegetation, with a minor contribution from C₄ plants (Cui et al., 2008). For C₃ plants, environmental factors, especially temperature and humidity, can mediate the carbon isotope fractionation during photosynthesis (Farquhar et al., 1989; Ménot and Burns, 2001). Vascular plants can adjust their leaf stomata to adapt to humidity changes. An increase of humidity is usually accompanied by an increase of stomatal conductance and $\delta^{13}C_{\text{leaf}}$ (Kohn, 2010). The influence of temperature on C₃ plant $\delta^{13}C$ is relatively complex. Temperature can directly control enzymatic activity in vegetation, affect metabolic capability and finally result in change of δ^{13} C. In addition, temperature can affect other environmental factors and then influence the composition of carbon isotopes in vegetation.

4.2.2. Influence from soil processes

Other than the influence from vegetation changes, microbial activity is another important factor to manifest $\delta^{13}C_{ASOM}$ signals preserved in speleothems.

Dissolved organic matter (DOM) is the main form in which organic carbon is transported downward into the subsoil layers and finally into the groundwater (Batiot et al., 2003; Ban et al., 2008). Soil DOM is generally divided into two groups, the hydrophobic (HPO) and hydrophilic (HPI) fractions (e.g. Aiken and Leenheer, 1993; Nakanishi et al., 2012). The HPO fraction consists mainly of aromatic lignin-derived compounds and has relatively longer fluorescence wavelength than the HPI fraction (Baker et al., 1998; McGarry and Baker, 2000). These hydrophobic materials are relatively stable against biodegradation (Kalbitz et al., 2003). The HPI fraction, in contrast, contains less aromatic structures but more carbohydrates, amino acids and hydroxyl groups. This fraction mainly derives from recent litter and photosynthates (Guggenberger et al., 1994). The HPI fraction is more labile relative to the HPO fraction (Qualls and Haines, 1992; Jandl and Sollins, 1997; Kalbitz et al., 2003). Corresponding with their consistent difference, the carbon isotopic signatures of HPI are commonly heavier than that of HPO (Kaiser et al., 2001, 2004; Kalbitz and Kaiser, 2008; Nakanishi et al., 2012).

If microbial activity can change the HPI/HPO ratio of soil DOM, this activity would finally mediate the carbon isotopic composition of ASOM preserved in stalagmites. Several factors can control microbial degradation on DOM. The first possible factor is the exposure time to microbial degradation. Under conditions with decreased effective precipitation, the residence time of organic matter in the soil and ground water will be extended, increasing the possibility of it being degraded before being preserved in stalagmites. If the exposure time in soil is the dominant process to control the signature of $\delta^{13}C_{ASOM}$, we would expect lighter values of $\delta^{13}C_{ASOM}$ under drier conditions. In such conditions, due to the prolonged residence time, the HPI/HPO ratio will be smaller and the $\delta^{13}C_{ASOM}$ values from 9 to 4 ky BP may indicate that it was relatively dry in the study region. In contrast, the less negative $\delta^{13}C_{ASOM}$ values from 4 to 0 ky BP suggest that it was relatively wet.

Temperature is another important factor to control microbial activity and associated degradation. The rates of microbial activity are normally higher in warmer conditions (Fang and Moncrieff, 2001; Dong et al., 2010). The study region is characterized by warm and humid climate. Consequently, under warmer humid conditions, microbes will preferentially degrade the HPI fraction, resulting in a relatively low HPI/HPO ratio and more negative $\delta^{13}\text{C}_{\text{ASOM}}$. The above deduction is supported by correlation analysis between the HS-4 $\delta^{13}\text{C}_{\text{ASOM}}$ record and the paleo-temperature record derived from the branched fatty alcohol ratio BNA₁₅ recorded in the nearby Dajiuhu peatland (Huang et al., 2013) (Fig. 3). Interestingly, if we take consideration of the dating error and attribute horizons from the two sequences with age difference less than 50 yr as synchronous age, these two sequences correlate negatively (r = -0.48, n = 57, p < 0.001; Fig. 5), with more negative $\delta^{13}C_{ASOM}$ values appearing in warmer conditions. There are also some inconsistencies between the $\delta^{13}C_{ASOM}$ and the BNA₁₅ record. For example, the BNA₁₅ shows a general decreasing trend from 3 ka onwards, while the increasing trend of the $\delta^{13}C_{ASOM}$ is punctuated by an interval with lighter values at ca. 2 ka (Fig. 3).

Caves are often thought of as severely resource-limited settings due to the absence of light. However, recent research highlights that caves also contain high microbial abundance (e.g. Northup and Lavoie, 2001; Novakova, 2009; Cuezva et al., 2012). Thus it is reasonable to deduce that DOM will experience further processing within the cave, which could further decrease the HPI/HPO ratio and $\delta^{13}C_{ASOM}$ values.

Other than microbial degradation, other soil processes can also affect soil DOM and finally exert controls on $\delta^{13}C_{ASOM}$. In soil horizons, soil organic matter and clay form complexes. Because HPO fractions have relatively high affinity onto the clay surfaces (Kalbitz et al., 2003; Nakanishi et al., 2012), such a clay-organic matter interaction can alter the HPI/HPO ratio and the associated $\delta^{13}C_{ASOM}$ values of the DOM in soils. A recent study conducted by Grybos et al. (2009) highlights that large quantities of DOM are dissolved under elevated pH conditions. The acid-sensitivity of soil DOM has also been proposed as the main cause of widespread increases in concentrations of DOM in the surface



Fig. 5. Correlation analysis (a) between $\delta^{13}C_{ASOM}$ and $\delta^{13}C_{carb}$ (cited from Hu et al., 2008b); (b) $\delta^{13}C_{ASOM}$ and the synchronous fluorescence wavelength; (c) $\delta^{13}C_{ASOM}$ and BNA₁₅ ratio (cited from Huang et al., 2013). Because of the age difference, both $\delta^{13}C_{ASOM}$ and BNA₁₅ records have been interpolated at 100-yr interval.

waters across North America and Northern and Central Europe (Monteith et al., 2007). In this case, the pH-derived dissolution process has the potential to mediate the $\delta^{13}C_{ASOM}$ values in the overlying soils and underlying karst system.

In the monsoonal region of China, soil pH normally correlates with aridity, with relatively high pH values during dry conditions (Huang et al., 2008a; Yang et al., 2012). Soil profiles developed in karst regions normally contain high contents of cations, especially Ca^{2+} and Mg^{2+} , which can buffer the fluctuations of rainfall-derived pH shifts. However, in quite wet intervals or during heavy rain periods, the influence of rainfall can exceed the buffer capacity of soil cations and thus cause soil pH to decrease. Thus we still need to be cautious that, in relatively wet climates, pH in the soil profile overlying Heshang Cave will decrease and a greater proportion of HPO fraction will be absorbed onto clay surfaces. This process finally results in a relatively high HPI/HPO ratio and less negative $\delta^{13}C_{ASOM}$ values in speleothem deposits.

In the above discussion, we use the HPI–HPO model to interpret how microbial activity affects the signature of $\delta^{13}C_{ASOM}$. Further research is necessary to support this model. Interestingly, because of the known relationship between dissolved organic matter fluorescence wavelength and hydrophobicity, we would expect to see a correlation between fluorescence wavelength and $\delta^{13}C_{ASOM}$ if the HPI/HPO ratio is the dominant control on $\delta^{13}C_{ASOM}$. However, for the HS-4 stalagmite, $\delta^{13}C_{ASOM}$ shows

a weak correlation with the synchronous fluorescence wavelength (r = -0.30, p < 0.01; Fig. 5). Such a relatively weak correlation suggests that factors other than the HPI/HPO ratio are also controlling stalagmite $\delta^{13}C_{ASOM}$ in our stalagmite. It is therefore likely that microbial degradation of organic matter is just one competing process controlling the carbon isotope composition of the ASOM, along with the changes in vegetation structure and physiology, and physical factors affecting soil organic matter stability and transport.

5. Conclusions

This study presented first 9000-yr time series of EA-IRMS analysis of acid-solution organic matter isolated from the HS-4 stalagmite collected from Heshang Cave, central China. Spectroscopic analyses support that the ASOM mainly originates from the overlying soil. The $\delta^{13}C_{ASOM}$ varies between -25.8% and -22.0%, with an average of -24.2%. Comparisons between $\delta^{13}C_{ASOM}$ and $\delta^{13}C_{carb}$ in a single growth layer and the time series clearly show that $\delta^{13}C_{ASOM}$ is not affected by kinetic fractionation during stalagmite formation. We hypothesize that the $\delta^{13}C_{ASOM}$ record in HS-4 is controlled by both temperature and effective precipitation. The influence of temperature is mainly on the microbial degradation of soil organic matter, possibly with additional influences on the vegetation physiology. The availability of water can affect the length of time organic matter is exposed to microbial processing in soils. Under drier conditions, the exposure time of soil organic matter will be prolonged and lead to a higher degree of degradation, resulting in relatively low HPI/HPO ratios and more negative $\delta^{13}C_{ASOM}$ values. Further evidence, such as from modern monitoring programs, would help test our hypothesis.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at http://dx. doi.org/10.1016/j.chemgeo.2014.08.029.

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