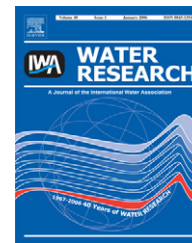


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# Functional variability of dissolved organic matter from the surface water of a productive lake

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## ABSTRACT

Functional variability of dissolved organic matter (DOM) from the surface water of Esthwaite Water (N. England) was investigated using a series of 12 standardised assays, which provide quantitative information on light absorption, fluorescence, photochemical fading, pH buffering, copper binding, benzo(a)pyrene binding, hydrophilicity, and adsorption to alumina. Ten lakewater samples were collected at different times of year during 2003–2005, and DOM concentrates obtained by low-temperature rotary evaporation. Suwannee River Fulvic Acid was used as a quality control standard. For nine of the assays, variability among DOM samples was significantly ( $p < 0.01$ ) greater than could be explained by analytical error. Seasonal trends observed for six of the assays could be explained by a simple mixing model in which the two end-members were DOM from the catchment (allochthonous) and DOM produced within the lake (autochthonous). The fraction of autochthonous DOM predicted by the model is significantly correlated ( $p < 0.01$ ) with chlorophyll concentration, consistent with production from phytoplankton. Autochthonous DOM is less light-absorbing, less fluorescent, more hydrophilic, and possesses fewer proton-dissociating groups, than allochthonous material.

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## 1. Introduction

Dissolved organic matter (DOM) in natural waters participates in many important ecological and geochemical reactions (Perdue and Gjessing, 1990; Kullberg et al., 1993; Hessen and Tranvik, 1998). For example, it controls the transport and fate of heavy metals, aluminium, radionuclides, and organic pollutants, initiates photoreactions, participates in particle surface and colloid chemistry, and affects ionic balance, including pH. Quantitative descriptions of these functional properties are needed for ecology, geochemistry, and to understand and predict the toxicity and fate of pollutants. The need for such descriptions is given extra impetus by the apparent sensitivity of DOM to environmental change, as

shown by long-term increases (Hongve et al., 2004; Evans et al., 2005) or decreases (Schindler et al., 1996) in DOM concentration, and changes in DOM quality (Curtis, 1998; Donahue et al., 1998), attributed to climatic warming and/or declining acid deposition.

Knowledge about the functional properties of DOM has been obtained largely from laboratory experiments with isolated fractions, especially humic and fulvic acids, from different natural environments, and obtained by different methods. Inevitably the data obtained are not systematic, which makes it difficult to apply the available knowledge to field situations. Given that freshwater DOM molecular structure, composition, and size are considered to vary considerably, depending upon (i) source material (Malcolm,

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1990; Curtis, 1998), (ii) differential retention during passage through soils (Kaiser et al., 2002), and (iii) modification in the freshwater system, notably by photolysis (Waiser and Robarts, 2000), it seems inevitable that functional properties will vary as well. However, at present we cannot readily relate DOM function to structure.

To address the issue of functional variability in DOM directly, Thacker et al. (2005) developed standardised assays, that can be applied to DOM isolates in order to quantify variability in the functional properties of DOM. The 11 assays, together with one additional assay, are summarised in Table 1. In each case, solutions of isolated DOM are prepared under standardised conditions, and a functional property is measured. A key feature of the approach is the use of a quality control standard (Suwannee River Fulvic Acid, SRFA) which is analysed alongside each DOM sample. The assays of optical absorbance (1, 2, and 12) characterise the effect of DOC on light penetration of surface waters, while determinations of photodecomposition (assay 4) and fluorescence (assay 3) are relevant to photochemical activity. Assays 5, 6, and 7 quantify interactions of DOM with other solutes, and are relevant to natural water chemistry and the transport and bioavailability of essential and potentially toxic metals and hydrophobic organic contaminants. The hydrophilicity assays (8 and 9) are relevant to aggregation, and sorption processes involving cells and mineral surfaces, while the adsorption assays (10 and 11) deal directly with mineral adsorption.

In lakes, two sources of DOM can broadly be distinguished. Allochthonous DOM (DOM<sub>ALL</sub>) originates from the catchment, mainly through the decay of terrestrial plant material and subsequent leaching of partial decomposition products. Autochthonous DOM (DOM<sub>AUT</sub>) is produced within the lake itself. Thomas (1997) identified three main sources of DOM<sub>AUT</sub>: (i) sloppy feeding or excretion by living organisms (bacteria, phytoplankton, invertebrates, and fish); (ii) bacterial degradation of dead particulate organic matter (in epilimnion,

hypolimnion, and sediment); and (iii) abiotic polymerisation and degradation. Macrophytes may also contribute. "Autochthonous-like" DOM may be produced from DOM<sub>ALL</sub>, due to in-lake chemical alterations, for example, acidification (Donahue et al., 1998) and photobleaching (Waiser and Robarts, 2000). Typically, DOM<sub>AUT</sub> absorbs less UV light, is poorer in aromatic residues, and is more enriched in nitrogen than DOM<sub>ALL</sub> (Tipping et al., 1988; Curtis and Adams, 1995; Curtis, 1998). There are also differences in fluorescence properties, for example, Donahue et al. (1998) reported that, with excitation at 370 nm, the peak emission of DOM<sub>ALL</sub> was at 462 nm, whereas that of DOM<sub>AUT</sub> was at 443 nm. The relative contributions of DOM<sub>ALL</sub> and DOM<sub>AUT</sub> in a lake depend upon hydrological factors and the biological and physico-chemical characteristics of the water body and its surrounding catchment (Thomas, 1997).

Thacker et al. (2005) observed significant differences between functional properties of DOM from a eutrophic lake (Esthwaite Water, EW) and those of DOM from three stream waters, one of which was an inflow to EW. Differences between the two EW samples were attributed to seasonal differences in the content of DOM<sub>AUT</sub> (see also Tipping et al., 1988). In the present work, we investigated the functional properties of DOM in the surface water of EW in more detail, and attempted to explain seasonal variability with a two end-member (DOM<sub>ALL</sub> and DOM<sub>AUT</sub>) mixing model. We applied the 12 assays of Table 1 to a series of samples representative of the mixed surface water of the lake, and collected at different times of year.

## 2. Methods

Heaney et al. (1986) provide a comprehensive description of the physics, chemistry, and biology of EW (54°21'N, 2°59'W). The catchment of the lake has an area of 17.1 km<sup>2</sup> and

**Table 1 – Number and name of each assay, the nature of the assay result, and the abbreviated designation**

Assay no.	Assay	Assay result	Abbreviation
1	Optical absorbance 280 nm	Extinction coefficient <sup>a</sup> at 280 nm (l gC <sup>-1</sup> cm <sup>-1</sup> )	E <sub>280</sub>
2	Optical absorbance 340 nm	Extinction coefficient <sup>a</sup> at 340 nm (l gC <sup>-1</sup> cm <sup>-1</sup> )	E <sub>340</sub>
3	Fluorescence (325/450)	Peak intensity with excitation at 325 nm and emission at 450 nm, per mg DOC l <sup>-1</sup>	F <sub>DOC/325/450</sub>
4	Photochemical fading	% loss in DOM absorbance at 340 nm	A <sub>340</sub> loss%
5	Buffering capacity	Acid groups titrated between pH 4 and 8 (meq/g C)	Ac <sub>4-8</sub>
6	Copper binding	Conditional stability constant (l gC <sup>-1</sup> )	log K <sub>c</sub>
7	Benzo(a)pyrene binding	Partition coefficient (cm <sup>3</sup> g C <sup>-1</sup> )	log K <sub>p</sub>
8	Hydrophilicity (DOC)	% of DOC not adsorbed by DAX-8 resin at pH 2	Hyphil <sub>DOC</sub> %
9	Hydrophilicity (absorbance)	% of DOM absorbance (340 nm) not adsorbed by DAX-8 resin at pH 2	Hyphil <sub>A340</sub> %
10	Alumina adsorption (DOC)	% of DOC adsorbed at pH 4	Ads <sub>DOC</sub> %
11	Alumina adsorption (absorbance)	% of DOM absorbance (340 nm) adsorbed at pH 4	Ads <sub>A340</sub> %
12 <sup>b</sup>	Optical absorbance 254 nm	Extinction coefficient <sup>a</sup> at 254 nm (l gC <sup>-1</sup> cm <sup>-1</sup> )	E <sub>254</sub>

<sup>a</sup> Extinction coefficient: ratio of optical absorbance per cm to DOC concentration in g l<sup>-1</sup>.

<sup>b</sup> Assay 12 is a new assay, in addition to the 11 assays described in Thacker et al. (2005).

receives 1800 mm of rainfall per year on average, of which 60% falls in winter (October–March). The annual mean temperature is ca. 10 °C, with monthly averages that range from ca. 5 °C in January to 15 °C in July. The lake is rarely covered with ice. The lake has a surface area of 1.00 km<sup>2</sup>, mean and maximum depths of 6.4 and 15.5 m, respectively, and a mean residence time of 13 weeks. EW stratifies thermally in summer, and then has an anoxic hypolimnion. There is an annual plankton cycle, estimated by the concentration of the photosynthesis pigment chlorophyll *a*, denoted as [Chl *a*]. During the period of study, phytoplankton was dominated by diatoms (*Asterionella formosa*) in spring, and by blue-green algae such as *Aphanizomenon* sp. and *Woronichinia* sp. in late summer (M. DeVille, pers. comm.). Typical Chl *a* levels range from approximately 1 µg l<sup>-1</sup> in winter to 60 µg l<sup>-1</sup> in late summer. Relevant chemical characteristics of the samples taken in the present work are given in Table 2. These data are representative of the lake at all times, except during short periods in summer when high algal productivity causes higher pH (Maberly, 1996).

Samples (50 l) were collected by wading into the small stream that is the lake outflow. The stream water is representative of either the whole mixed lake (winter) or the epilimnion of the stratified lake (summer). A polyethylene beaker and funnel were used to transfer water to thoroughly rinsed 10-l polyethylene bottles. Collection took approximately 10 min, and was performed between 9.00 and 12.00 h. Samples were returned to the laboratory within 1 h, and stored cold and dark during processing.

The method used to isolate the DOM is described in detail by Thacker et al. (2005) and involved concentrating the filtered (GF/F Millipore, nominal pore size 0.7 µm) sample to approximately 500 cm<sup>3</sup>, using a high-capacity, low-pressure, low-temperature (20 °C), rotary evaporator (Buchi Rotavapor R-220). The sample was then passed through a column of Amberlite IR-120 (in the sodium form) to exchange major cations, and filtered through Whatman GF/F and Millipore 0.22 µm filters. In two cases (EW4 and EW10), a second isolation was carried out, in which the final volume was 1000 cm<sup>3</sup> instead of 500 cm<sup>3</sup>.

The raw water samples and concentrates were analysed within 1 week for pH (Radiometer GK2401C combination glass electrode), DOC (TOC-VC/PN/CPN analyzer, Shimadzu, Kyoto, Japan), absorbance at 340 nm (Hitachi U-2000 Spectrophotometer), and conductivity (Jenway 4510 meter). Stored samples were analysed later for major cations (ICP-OES, Perkin Elmer Optima 4300DV). Raw water samples were also analysed for alkalinity (Gran titration), major anions (Dionex DX100) and Chl *a* by extraction with boiling methanol (Talling, 1974).

The 11 standardised assays, previously tested and described in detail by Thacker et al. (2005), together with one additional optical absorbance assay (Table 1), were applied to the concentrates. For each assay, the DOM was present at a fixed concentration (10–100 mg DOC l<sup>-1</sup> depending upon the measurement), in a solution of defined chemical composition, so that differences in the measured quantity reflected differences in the DOM, and not, for example, in the composition of the raw water sample. A quality control standard, reference SRFA purchased from the International

Table 2 – Chemical compositions of raw samples from Esthwaite water

Sample code	Sampling date	pH	Cond <sup>a</sup> (µS cm <sup>-1</sup> )	DOC (mg l <sup>-1</sup> )	Alk <sup>a</sup> (mg l <sup>-1</sup> )	Na (mg l <sup>-1</sup> )	Mg (mg l <sup>-1</sup> )	Ca (mg l <sup>-1</sup> )	K (mg l <sup>-1</sup> )	E <sub>340</sub> (l g C <sup>-1</sup> cm <sup>-1</sup> )	Chl <i>a</i> (µg l <sup>-1</sup> )
EW1	09/10/03	7.38	119	3.9	31.2	7.21	1.5	12.1	nd <sup>a</sup>	7.4	14.0
EW2	27/07/04	7.87	106	3.7	24.3	6.68	1.4	11.2	0.86	5.1	52.9
EW3	17/01/05	7.50	104	2.9	20.6	6.8	1.3	9.32	1.03	9.9	2.8
EW4	21/02/05	7.64	116	2.8	22.0	7.28	1.4	10.9	1.06	8.7	1.8
EW5	20/04/05	7.63	127	2.6	24.0	7.12	1.4	10.7	0.96	12.5	9.6
EW6	18/05/05	8.00	120	3.3	27.5	7.12	1.5	11.7	1.02	10.3	23.7
EW7	16/06/05	7.59	113	3.4	26.3	6.94	1.5	11.6	0.94	7.6	8.1
EW8	20/07/05	7.87	120	3.4	24.9	7.25	1.5	11.3	0.93	8.2	15.4
EW9	23/08/05	7.82	117	3.6	27.0	7.26	1.5	11.6	0.90	5.6	26.0
EW10	13/09/05	7.84	128	2.8	26.3	7.12	1.5	11.6	0.91	8.8	21.8

<sup>a</sup> Cond = conductivity; Alk = alkalinity; nd = not determined.

Humic Substances Society, was analysed simultaneously with the samples to characterise assay reproducibility.

The extra assay of optical absorbance (at 254 nm) was added to increase the comparability of our results with other published data (e.g. Chin et al., 1994). However, the same numbering system has been maintained for the assays as in the previous work, with the optical absorbance assay at 254 nm numbered as assay 12 (Table 1).

Two modifications were made to the assays described in Thacker et al. (2005). First, an extra quality control standard was formulated for the hydrophilic assay. This was done because the SRFA quality standard is isolated on the basis of its hydrophobic character, i.e., by adsorption onto DAX-8 resin in acid solution, and therefore has a low content of hydrophilic material. To obtain similar results for both standard and samples, to aid statistical analysis, a new quality control standard was prepared by mixing 15 mg DOC l<sup>-1</sup> of SRFA with 5 mg DOC l<sup>-1</sup> of sodium acetate, to provide a hydrophilic component. Second, the assay output for the buffer capacity assay was altered to the number of acid groups titrated between pH 4 and 8, due to the possibility of silicate interference. In Thacker et al. (2005), the number of acid groups was titrated between pH 4 and 9. The results in Thacker et al. (2005) were reanalysed and it was found that variability among the DOM samples is still significantly ( $p < 0.01$ ) greater than can be explained by analytical error, i.e., there is no change in the overall conclusion from the previous work.

It was also found by Thacker et al. (2005) that benzo(a)pyrene binding results for the DOM samples did not vary significantly. To check if this phenomenon could be an artefact of the method, additional measurements were made on a commercially available humic acid (Aldrich Chemical Company), which has a greater affinity for hydrophobic xenobiotics than does natural DOM (Kukkonen, 1991). Aldrich humic acid gave a log  $K_p$  for benzo(a)pyrene binding of 5.11, 0.57 log units higher than the SRFA quality control and 0.49 log units higher than the DOM samples, proving that the lack of variation shown by natural water samples was not an artefact of the method.

### 3. Results

#### 3.1. Esthwaite water

Raw water samples collected from EW during 2003, 2004, and 2005, all have similar chemistries (Table 2). From fortnightly monitoring, Tipping et al. (1988) reported [DOC] in EW to remain relatively constant throughout winter (November–March) with an average of 2.0 mg l<sup>-1</sup> while during summer (May–September) it was higher, with an average of 3.7 mg l<sup>-1</sup>, and the more limited number of observations of the present work are consistent with this pattern. The increase in [DOC] during summer was attributed to within-lake production of DOC as a result of plankton growth and excretion and/or decomposition.

Phytoplankton biomass ( $\mu\text{g Chl } a \text{ l}^{-1}$ ) in EW is highly variable seasonally. Determinations of Chl *a* were made fortnightly during 2003, 2004, and 2005 (M. DeVille, pers. comm.) and the

data show spring and summer maxima. Values of [Chl *a*] determined on samples collected for DOM assays are also shown in Table 2.

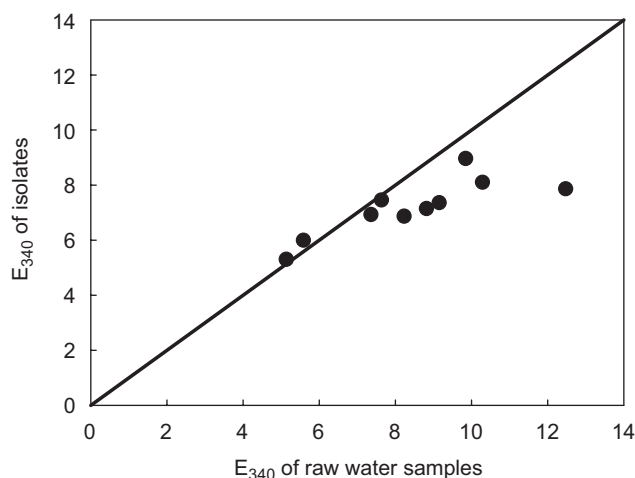
#### 3.2. Isolation and concentration of DOM

The isolation method gave an average DOC yield of 77% (ranging from 70% to 89%). Thacker et al. (2005) concluded that the low recovery is caused by precipitation of calcium carbonate forming during the last stages of concentration and removing some DOM by adsorption or co-precipitation. A strong correlation ( $r = -0.92$ ) was found between  $E_{340}$  values of raw water samples and percentage recovery. Furthermore, samples with the highest raw water  $E_{340}$  values underwent appreciable decreases in  $E_{340}$  on concentration (Fig. 1). These results show that DOM lost during the isolation method is from the most strongly light-absorbing fraction. Therefore, the magnitude of the loss of DOM depends on (i) the proportion of the strongly light-absorbing fraction in the raw water sample, comprising the larger molecules with a higher aromatic and hydrophobic character, and (ii) sufficiently high concentrations of Ca<sup>2+</sup> and CO<sub>3</sub><sup>2-</sup> for precipitation to occur during the concentration process.

To investigate the effect of DOM losses on measured functional properties, in two cases (EW4 and EW10), a second sample was processed, concentrated to 1000 cm<sup>3</sup> instead of the usual 500 cm<sup>3</sup>. By reducing the concentration factor, improved yields were obtained, from 72% to 87% for EW4 and from 78% to 84% for EW10. The less-concentrated samples are referred to as EW4A and EW10A. Assay results for the four concentrates are shown in Table 3.

#### 3.3. Variability in DOM functional properties

Fig. 2 shows that for most of the assays good reproducibility was obtained for the quality control standard, SRFA, with relative standard deviations (RSD) of less than 5%.



**Fig. 1 – Extinction coefficients at 340 nm of raw water samples and their concentrates, following isolation. The line represents a 1:1 relationship.**

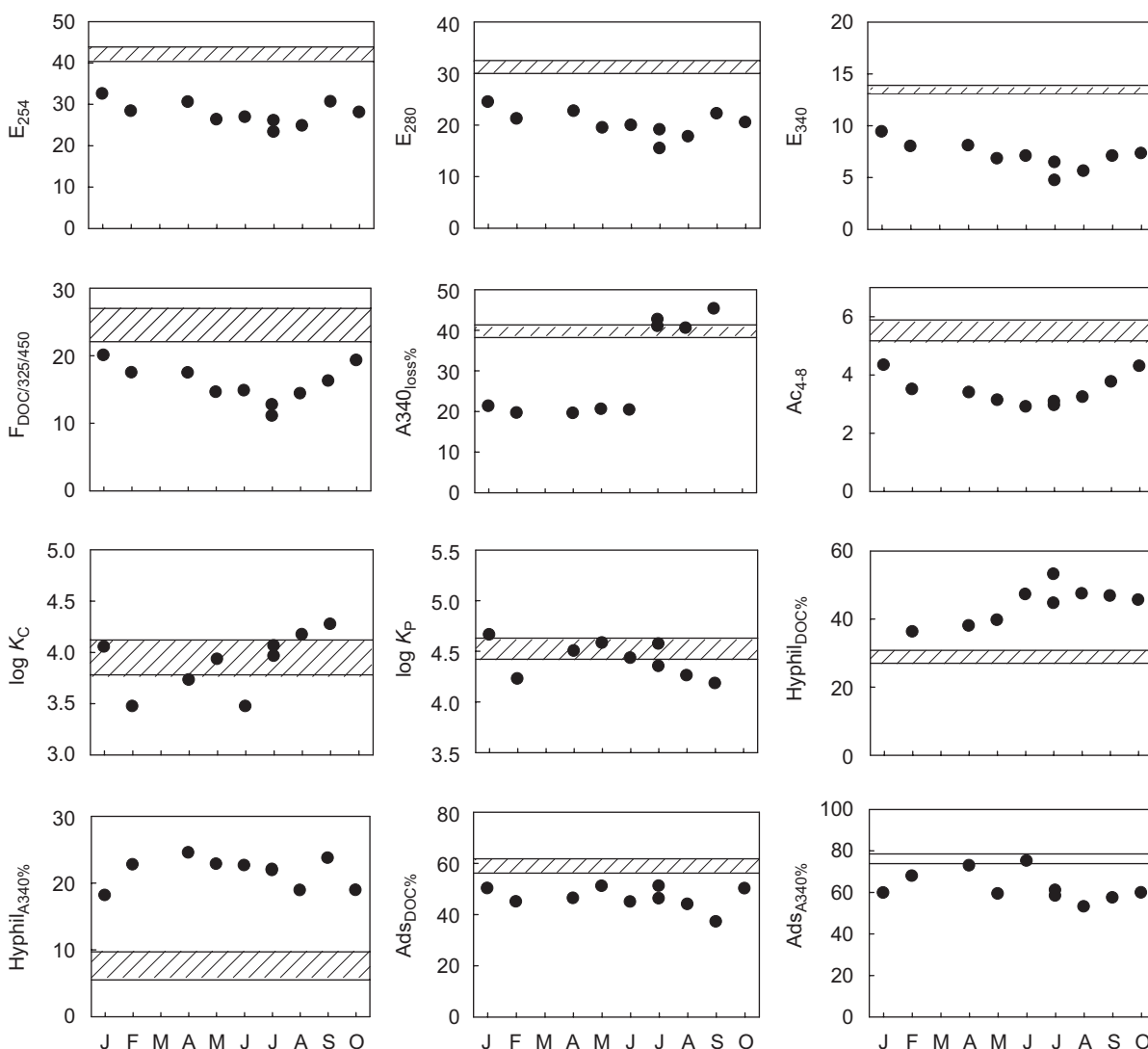


**Table 3 – Assay results for DOM samples concentrated to different extents, and therefore giving different recoveries**

	EW4	EW4A	EW10	EW10A
Recovery %	72	87	78	84
$E_{254}$	28.3	33.2	30.6	28.7
$E_{280}$	21.2	25.1	22.2	20.8
$E_{340}$	8.0	10.0	7.1	6.6
$\log K_C$	3.50	3.08	4.27	4.14
$Ac_{4-8}$	5.00	5.45	5.55	5.31
$F_{DOC/325/450}$	18.9	17.3	18.2	17.2
$Hyphil_{DOC}\%$	37.9	37.0	45.5	45.2
$Hyphil_{A340}\%$	22.7	18.3	23.7	22.9
$A_{340\text{ loss}}\%$	19.6	22.1	45.2	49.9
$Ads_{DOC}\%$	44.9	48.7	37.1	38.6
$Ads_{A340}\%$	72.7	75.5	59.7	61.6
$\log K_p$	4.22	4.62	4.18	4.22

The fluorescence assay gave an RSD of 6.5%, while an RSD of 14.8% was obtained for the assay of hydrophilicity monitored by optical absorption. Results from the quality control standard were used to apply the one-tailed F-test (Snedecor and Cochran, 1967), to assess variability in functional properties of the DOM samples (Thacker et al., 2005). For nine assays, variation among EW DOM samples was significantly greater ( $p < 0.01$ ) than can be explained by analytical error, i.e., by comparison with results for the SRFA standard, but no statistically significant variations were found for the assays of benzo(a)pyrene binding, copper binding, and hydrophilicity monitored by optical absorption.

Several functional properties (all three extinction coefficients, fluorescence, buffer capacity, and hydrophilicity monitored by DOC) show systematic seasonal variations, with a maximum or minimum during the summer months. We therefore attempted analysis of the results with a two-member mixing model, hypothesising that seasonal



**Fig. 2 – Assay results for DOM samples from Esthwaite Water (symbols) and for the quality control standard (shaded areas). Units for the y-axes are given in Table 2.**

variability can be accounted for in terms of mixtures of  $DOM_{AUT}$  and  $DOM_{ALL}$ , the functional properties of  $DOM_{AUT}$  and  $DOM_{ALL}$  being assumed constant. Therefore, a given functional property,  $F$ , of DOM in EW will depend on the proportions of  $DOM_{AUT}$  and  $DOM_{ALL}$ , and can be expressed as

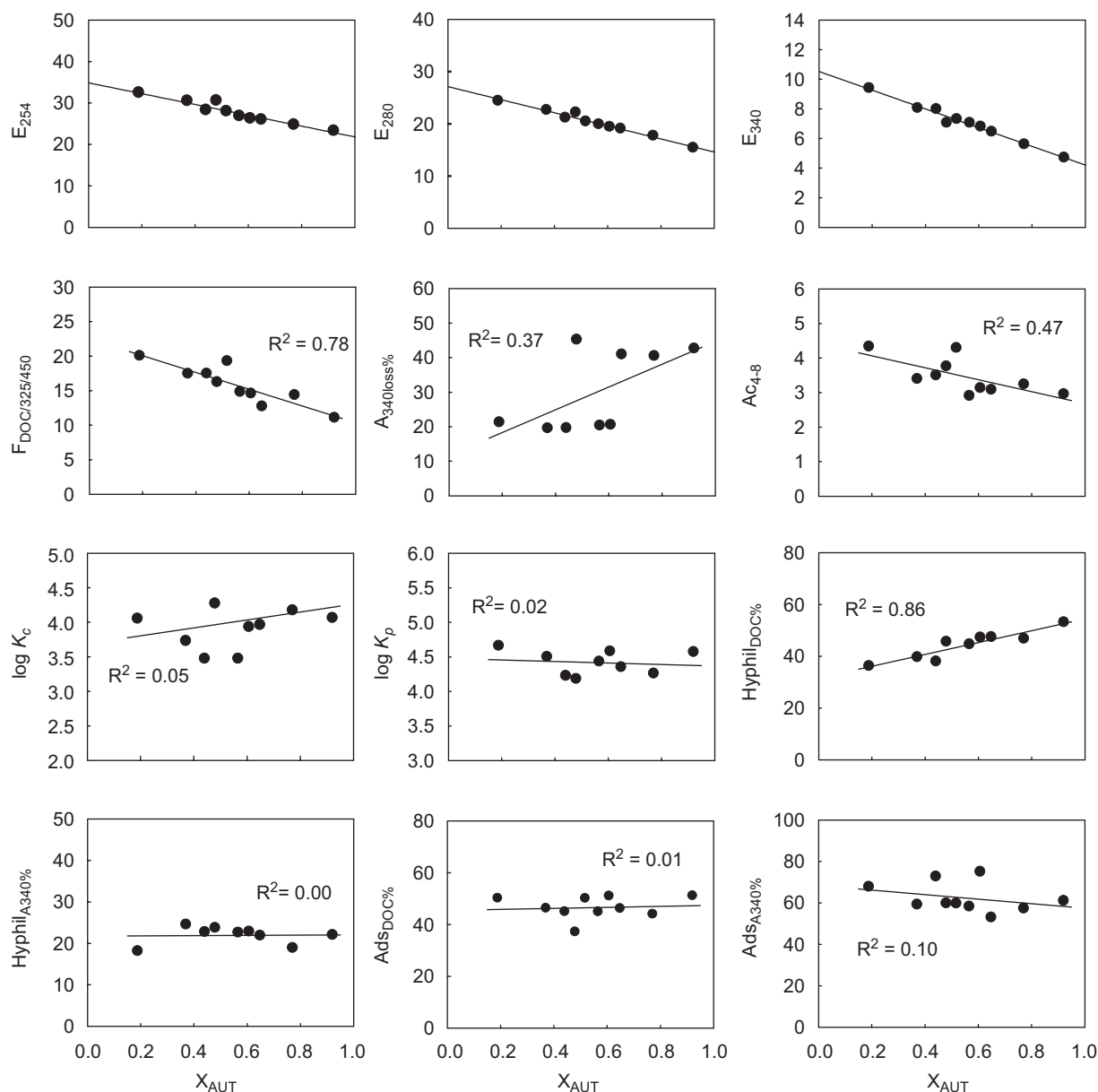
$$F = F_{AUT}X_{AUT} + F_{ALL}X_{ALL}, \quad (1)$$

where  $F_{AUT}$  and  $F_{ALL}$  are values of the functional properties of the autochthonous and allochthonous end-members, respectively, and  $X_{AUT}$  and  $X_{ALL}$  are the fractions of those end-members. Since the sum of  $X_{AUT}$  and  $X_{ALL}$  must be unity, Eq. (1) can be written as

$$F = F_{AUT}X_{AUT} + F_{ALL}(1 - X_{AUT}). \quad (2)$$

Since there are 12 assays, each applied to 10 samples, there are 120 versions of Eq. (2). Therefore, the total number of parameters to be found is 34, comprising 12 values each of  $F_{AUT}$  and  $F_{ALL}$ , and 10 values of  $X_{AUT}$ . Rather than using the entire data set to extract parameter values, we initially confined the analysis to results for  $E_{254}$ ,  $E_{280}$ , and  $E_{340}$ . Extinction coefficients were chosen firstly because additivity would clearly be expected on mixing the two end-members, and secondly because the measurements are highly precise (quality control RSD <0.5%). The “Solver” facility of Microsoft “Excel” was used to find parameters by least-squares minimisation of the sum of squared residuals between observed and predicted functional assay results.

The mixing model worked well, explaining 99.7% of the variance in the extinction coefficients. Moreover, the derived



**Fig. 3** – Plots of functional assay results against  $X_{AUT}$ , the fraction of autochthonous DOM, derived from the mixing model. Units for the y-axes are given in Table 2. The extinction coefficients at 254, 280, and 340 nm (top three panels) were used to fit the model and derive  $X_{AUT}$ . The remaining panels show regressions of assay results against  $X_{AUT}$ . If  $R^2 > 0.40$ , then  $p < 0.05$ ; if  $R^2 > 0.59$ , then  $p < 0.01$ .

values of  $X_{AUT}$ , from 0.17 to 0.88, indicate that the sampling programme produced an adequate range of mixtures of  $DOM_{ALL}$  and  $DOM_{AUT}$ . The top three panels of Fig. 3 show observed values of  $E_{254}$ ,  $E_{280}$ , and  $E_{340}$  plotted against derived values of  $X_{AUT}$ . The other panels of Fig. 3 show results for the remaining assays plotted against  $X_{AUT}$ , together with the results of regression analysis. In three cases,  $F_{DOC/325/450}$ ,  $Hyphil_{DOC}\%$ , and  $Ac_{4-8}$ , the functional property shows a significant ( $p < 0.01$ ) dependence on  $X_{AUT}$ . Table 4 shows  $F$  values for each assay, for the two end-members.

## 4. Discussion

### 4.1. Isolation of DOM

The method to obtain DOM samples for the assay work is a compromise between full isolation, with removal of all

solutes except DOM, and a mild method that produces a high yield (Thacker et al., 2005). However, the final concentrates obtained from the EW samples with higher  $E_{340}$  values were depleted in the highly coloured aromatic fraction of DOM (Section 3.1, Fig. 1). Because  $DOM_{ALL}$  has higher aromaticity, hydrophobic character, and UV absorbance than  $DOM_{AUT}$  (see Table 3), isolation losses may have selectively affected the  $DOM_{ALL}$  end-member in the final concentrate. The results in Table 4 for samples EW4 (lower yield) and EW4A (higher yield) confirm this to some extent, in that EW4A gave somewhat higher values of  $E_{254}$ ,  $E_{280}$ ,  $E_{340}$ ,  $Ac_{4-8}$ ,  $Ads_{DOC}\%$ , and  $\log K_p$ , and lower values of  $F_{DOC/325/450}$ ,  $Hyphil_{DOC}\%$ , and  $Hyphil_{A340}\%$ . However, the differences are small, and they are not reproduced by samples EW10 and EW10A. Therefore, isolation losses of DOM do not seem to have had a major selective effect on functional properties.

### 4.2. Variability in DOM functional properties

The successful application of the mixing model (Fig. 3, Table 4) permits the distinction of three categories of DOM functional property (Table 5). Category A comprises functional properties that vary significantly both among DOM samples and also with  $X_{AUT}$ . For the six functional properties in this category, some (in five cases, most) of the observed variability can be attributed to variations in  $X_{AUT}$ , and co-variations in  $X_{ALL}$ . As the fraction of  $DOM_{AUT}$  in EW increases, the DOM becomes less light-absorbing and less fluorescent. These results are consistent with the findings of Donahue et al. (1998) and Waiser and Robarts (2004). In addition, the present data show that  $DOM_{AUT}$  is more hydrophilic, and possesses fewer acid-dissociating groups than  $DOM_{ALL}$ . Five of the six functional properties in this category were also found to vary among the samples studied in previous work (Thacker et al., 2005); the  $E_{254}$  was not measured previously.

Category B comprises three functional properties that vary significantly among DOM samples, but do not vary with  $X_{AUT}$ . Two of the three,  $Ads_{DOC}\%$  and  $Ads_{340nm}\%$ , also varied

**Table 4 – Functional properties of  $DOM_{ALL}$  and  $DOM_{AUT}$  derived from the mixing model, mean assay results for two DOM samples from Esthwaite Hall Beck (Thacker et al. (2005)), and SRFA, and extinction coefficients for DOM from Lake Fryxell (Weishaar et al., 2003; Chin et al., 1994)**

	ALL	EHB	AUT	L. Fryxell	SRFA
$E_{254}$	34.8	36.9	21.8	18.0	42.4
$E_{280}$	27.1	28.3	14.6	12.5	31.5
$E_{340}$	10.5	12.4	4.2		13.5
$\log K_C$	3.67	4.30	4.02		3.98
$Ac_{4-8}$	4.22	5.31	2.71		5.42
$F_{DOC/325/450}$	21.7	18.7	9.8		15.8
$Hyphil_{DOC}\%$	32.9	32.8	54.3		12.6
$Hyphil_{A340}\%$	21.0	19.1	22.0		8.1
$A_{340}$ loss %	14.9	31.1	34.1		39.6
$Ads_{DOC}\%$	41.8	59.8	48.7		59.1
$Ads_{A340}\%$	65.4	77.4	57.0		75.8
$\log K_p$	4.42	4.50	4.39		4.51

**Table 5 – Significance of variability in functional properties**

Category	Assay	EW	$X_{AUT}$	SW
A	Optical absorbance 280 nm	S	S	S
	Optical absorbance 340 nm	S	S	S
	Fluorescence (325/450)	S	S	S
	Buffering capacity	S	S	S
	Hydrophilicity (DOC)	S	S	S
	Optical absorbance 254 nm	S	S	Not used
B	Photochemical fading	S	NS	NS
	Alumina adsorption (DOC)	S	NS	S
	Alumina adsorption (absorbance)	S	NS	S
C	Copper binding	NS	NS	NS
	Benzo(a)pyrene binding	NS	NS	NS
	Hydrophilicity (absorbance)	NS	NS	S

The columns headed SW and EW refer to comparisons of assay results with the quality control standard for eight surface waters (SW; Thacker et al., 2005) and EW (present work). The  $X_{AUT}$  column refers to variations of assay results with  $X_{AUT}$  values derived from the mixing model (cf. Fig. 4). S, NS = significant or not significant at the 1% level.

amongst the samples studied by Thacker et al. (2005). The consistent variability of these two related properties is evidently due to factors other than those that control variability within category A. The photochemical fading results for EW differ from those of the other assays, by displaying a step change between June and July, thereby giving rise to a bimodal pattern when plotted against  $X_{AUT}$ , and significant variability. We have no explanation for this phenomenon at present. In the work of Thacker et al. (2005), significant variability in  $A_{340}$  loss% was not found.

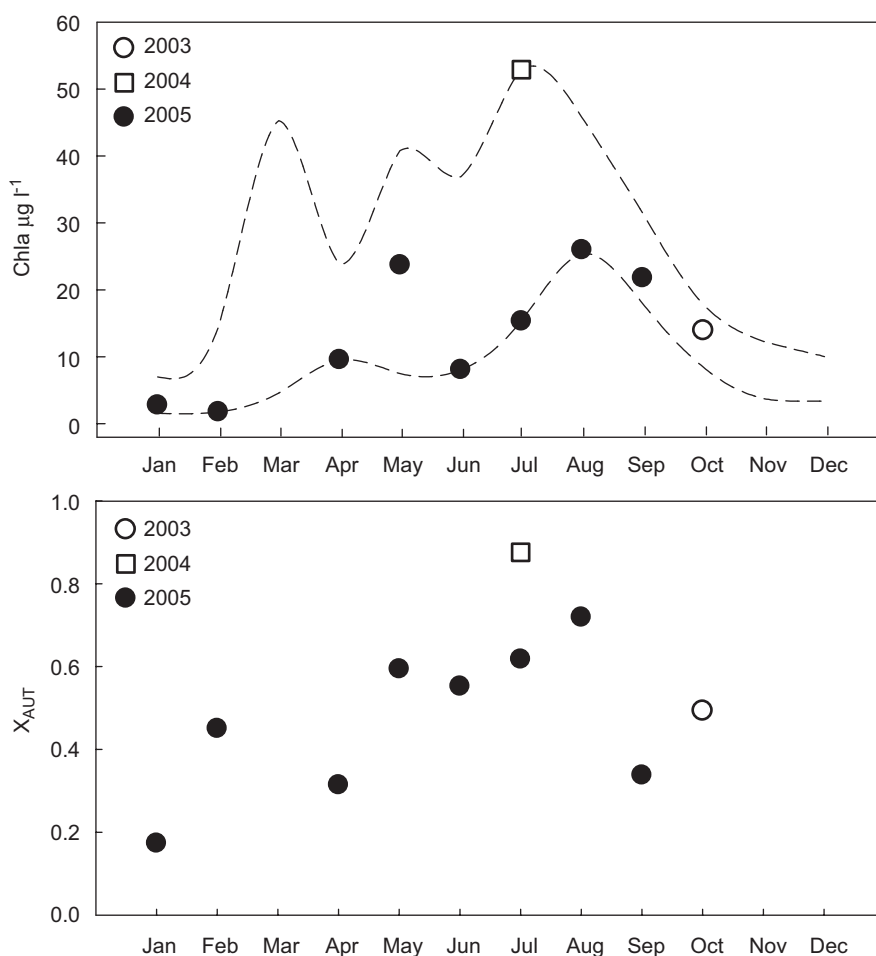
Category C comprises three functional properties that do not vary significantly among the DOM samples, neither do they vary with  $X_{AUT}$ . Thacker et al. (2005) also found that neither copper nor benzo(a)pyrene binding varied amongst surface water samples, but they did find significant variability in hydrophilicity as measured by optical absorbance.

#### 4.3. Sources of lakewater DOM

The mixing model permits estimation of the functional properties of the two postulated DOM end-members in EW, even though neither can be isolated and characterised in a “pure” state. Table 4 compares the derived properties of  $DOM_{ALL}$  with those determined by Thacker et al. (2005) for

DOM samples from Esthwaite Hall Beck, a stream flowing into EW. The results are very similar for five of the six functional assays,  $E_{254}$ ,  $E_{280}$ ,  $E_{340}$ ,  $F_{DOC/325/450}$ , and  $Hyphil_{DOC}$ . Agreement is less good for  $Ac_{4-8}$  but the result for  $DOM_{ALL}$  is much closer to the value for Esthwaite Hall Beck than is the value for  $DOM_{AUT}$ . Therefore, it can be concluded that  $DOM_{ALL}$  has functional properties consistent with those of DOM entering the lake from its catchment, which is a basic assumption of the mixing model.

A number of studies (Søndergaard et al., 2000; Jørgensen, 1986; Norrman et al., 1995) have implicated phytoplankton in the release of  $DOM_{AUT}$ . We therefore regressed  $X_{AUT}$  against [Chl *a*], as a measure of phytoplankton biomass, and found a significant relationship ( $R^2 = 0.71$ ,  $p < 0.01$ ). Fig. 4 illustrates how the values of  $X_{AUT}$  follow the seasonal pattern of [Chl *a*] in EW. In winter,  $X_{AUT}$  tends to be low, whereas it is high in summer. The sample collected in July 2004 during the period of highest algal biomass, corresponds to the highest value of  $X_{AUT}$  (0.88) predicted by the model. The idea that phytoplankton are the main source of  $DOM_{AUT}$  is supported by the results in Table 4 which show that values of  $E_{254}$  and  $E_{280}$  derived for  $DOM_{AUT}$  are similar to those reported for DOM from Lake Fryxell (Chin et al., 1994; Weishaar et al., 2003). Lake Fryxell is a permanently ice-covered lake in Antarctica,



**Fig. 4 – Seasonal variations in chlorophyll *a* and  $X_{AUT}$ . In the upper panel, dashed lines show the range of [Chl *a*] for 2003–2005, and points are values determined on samples taken for DOM isolation.**



in which DOM is derived mainly from benthic and planktonic microbial populations, with essentially no input of organic material from its surrounding watershed (Aiken et al., 1996).

Another possible source of DOM<sub>AUT</sub> is the in situ degradation and transformation of DOM<sub>ALL</sub> by photolysis and bacterial assimilation. Curtis and Schindler (1997) reported significant losses of both DOC and colour in Canadian lakes, with half-times of 166 and 122 d respectively; during this processing, the characteristics of the DOC would probably move towards those of DOM<sub>ALL</sub>. The average residence time of water in EW is 90 days (Heaney et al., 1986), and values for the summer months tend to be longer. Therefore, degradation of DOM<sub>ALL</sub> might well occur and contribute to DOM<sub>AUT</sub>. However, the fact that concentrations of DOC increase during the summer (see Section 3.1) strongly suggests an internal source, and so conversion of DOM<sub>ALL</sub> cannot be considered the major source of DOM<sub>AUT</sub>.

#### 4.4. Implications of the results

This study and the previous work by Thacker et al. (2005) demonstrate statistically significant variability in a number of functional properties of DOM from surface freshwaters. The results should contribute generally to the understanding of the sources and impacts of DOM in freshwaters, and more specifically to the quantitative description of freshwater systems, through predictive modelling, for example, in estimating the chemical speciation of metals (Tipping, 2002), and their toxicity (Di Toro et al., 2001). The extensive data from laboratory experiments with isolated natural organic matter (mostly fulvic and humic acids) constitute a valuable resource for modelling, but average DOM properties from such studies may not be sufficient. Although it appears from Table 5 that results for SRFA would be satisfactory to predict the interactions of EW DOM with copper and benzo(a)pyrene and its adsorption to mineral surfaces, they would overestimate the absorption of light, especially in surface waters dominated by DOM<sub>AUT</sub>, and also buffering capacity, fluorescence, and hydrophobicity (see also Section 2). Thus, in principle, more precise predictions would result if DOM variability, between and within waters, were taken into account. However, ecosystem modelling inevitably involves approximation, either because of lack of input data, or incomplete process characterisation, and uncertainty arising from variability in DOM properties may be overshadowed by greater uncertainties in other factors. To understand more fully the implications of the variability demonstrated by our results, they need to be incorporated into different ecosystem models, and sensitivity analyses conducted.

## 5. Conclusions

1. The isolation method gave yields of 70–89%, with an average of 77%. The final concentrate had less absorbance per g of DOC than the raw water sample, due to preferential loss of highly coloured material during isolation.
2. For nine of the 12 assays, variability among DOM samples is significantly ( $p < 0.01$ ) greater than can be explained by

analytical error, i.e., by comparison with results from the SRFA quality control standard. The three exceptions are copper binding, benzo(a)pyrene binding, and hydrophilicity monitored by optical absorbance.

3. Six of the 12 functional properties of DOM in EW could be modelled in terms of mixtures of DOM from the catchment (allochthonous) and DOM produced within the lake (autochthonous).
4. Of the two DOM types, autochthonous DOM is less light-absorbing, less fluorescent, more hydrophilic, and possesses fewer proton-dissociating groups.
5. The derived properties of allochthonous DOM are similar to those of DOM in catchment stream water. Autochthonous DOM is mainly derived from phytoplankton.

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